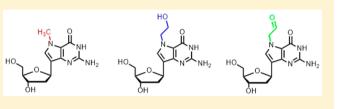
Synthesis of N^7 -Alkyl-9-deaza-2'-deoxyguanosines Containing Polar N⁷ Chains. Examples of Chemically Stable Analogues of N^7 -Hydroxyethyl and N^7 -Oxoethyl Adducts of 2'-Deoxyguanosine

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Supporting Information

ABSTRACT: Development of chemically stable analogues of unstable DNA lesions enables accurate study of polymerase bypass. We report the design and synthesis of N^7 -hydroxyethyl-9-deaza-2'-deoxyguanosine and N^7 -oxoethyl-9-deaza-2'-deoxyguanosine as the analogues of N^7 -hydroxyethyl-2'-deoxyguanosine and N^7 -oxoethyl-2'-deoxyguanosine, respectively. We also developed the synthesis of these two nucleosides whose



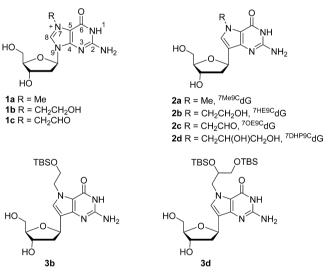
N⁷ side chains are protected by TBS for the convenience of conversion to phosphoramidites.

INTRODUCTION

DNA alkylation is one of the most common forms of DNA damage.¹ Many alkylating agents that are found in consumer products or released from industrial plants may present a threat to public health. At the molecular level, these agents or their metabolites are known to react with DNA nucleobases to form alkyl adducts.²⁻⁷ The biological effects of DNA alkyl adducts, however, are difficult to evaluate at the cellular level due to the uncontrolled distribution of the adducts within the whole genome and the involvement of multiple DNA polymerases and repair enzymes.^{6,8,9} Furthermore, N^7 -alkylguanines, the major reaction products produced by most alkylating agents, are converted to highly mutagenic abasic sites and weakly mutagenic N^7 -alkyl-FAPY-guanines through spontaneous or enzymatic hydrolysis.¹⁰⁻¹⁵ The conversion rate is dependent on the conditions of the study and the repair activities of the cell line. Therefore, comprehensive understanding of the biological picture of N^7 -alkylguanines at the molecular level, especially how fast and accurately polymerases bypass the lesions, is essential for connecting the chemistry of DNA damage to its biological consequences.

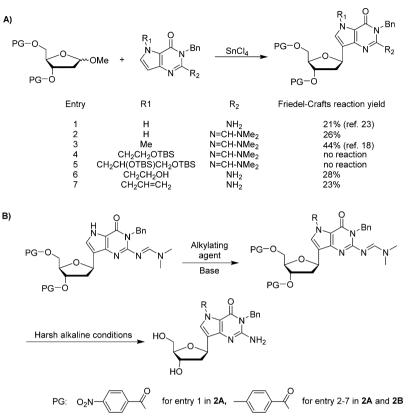
Synthesis of oligonucleotides that contain a single N^7 alkylguanine has become a vital tool to achieve the above goal. However, the instability of N^7 -alkyguanines is not compatible with the phosphoramidite chemistry used by solid-phase oligonucleotide synthesis. Two studies have been reported in which a short oligonucleotide containing a single guanine reacted with alkylating agents to form the desired N^7 -adducts.^{16,17} However, depurination may occur if the N^7 -alkylguaninecontaining oligonucleotide needs to be ligated to a long template and annealed to a primer for a standard polymerase kinetic assay where the incorporation accuracy and rate can be determined. Furthermore, the requirement for a single G site limited the possibility of investigating different local sequences. In the past,

Scheme 1. Compounds of Interest



we have used N^7 -methyl-9-deazaguanine (2a) as the model of N^7 -methylguanine (1a) to study its polymerase replication (Scheme 1).¹⁸ We now would like to expand the DNA lesions of interest to the adducts formed between guanine and epoxides, particularly those generated in vivo from various olefins. Specifically we are interested in N^7 -hydroxyethyl-2'-deoxyguanine (1b) and N^7 -oxoethyl-2'-deoxyguanosine (1c) (Scheme 1). The former is formed via a reaction with ethylene oxide.¹⁹ The latter can be generated from multiple sources in the chemical industry, among which the most widely used are vinyl chloride, vinyl bromide, urethane, and acrylonitrile.^{20–22} Herein we report

Received: August 26, 2016 Published: November 2, 2016 Scheme 2. (A) Formation of 9-Deazaguanine Nucleosides through Friedel–Crafts Reactions.^{*a*}(B) Alternative Method To Introduce N^7 -Alkyl Groups after C-Glycosidic Bond Formation



^aPG in entry 1 is a *p*-nitrobenzoyl group. PG in entries 2–7 is a *p*-toluoyl group. Reaction details are described in Results and Discussion.

the synthesis of N^7 -hydroxyethyl-9-deaza-2'-deoxyguanosine (^{7HE9C}dG, **2b**) and N^7 -oxoethyl-9-deaza-2'-deoxyguanosine (^{7OE9C}dG, **2c**) as two N^7 -alkyl-dG analogues that are resistant to glycosidic bond cleavage (Scheme 1). We also report the side-chain-protected nucleosides (**3b** and **3d**) that can be readily converted to phosphoramidites for oligonucleotide synthesis (Scheme 1). Notably, ^{7OE9C}dG contains an aldehyde group and is not compatible with solid-phase oligonucleotide synthesis. Therefore, its vicinal dihydroxyl precursor (**3d**) was the focus of the synthesis.

TBS is an evident choice of protecting group for the side chains of **3b** and **3d**, as it is commonly used to protect the 2'-OH of ribonucleosides in RNA synthesis. However, our preliminary studies showed that TBS is not compatible with the Friedel–Crafts reaction that is responsible for the formation of C-glycosidic bonds of 9-deaza-dG (Scheme 2A, entries 4 and 5).^{18,23} We examined several possible solutions to this problem. The immediate alternative solution was to directly introduce alkyl group to the N⁷ position of fully protected 9-deaza-dG (Scheme 2B). Using this strategy, **2b**, **2d**, and **2c** were prepared successfully.

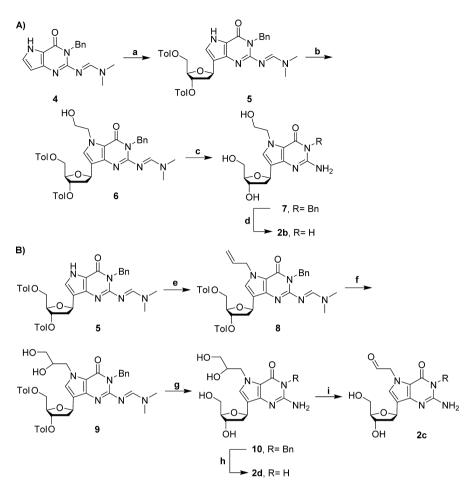
Unfortunately, we experienced difficulties when preparing **3b** and **3d**. It is well documented that the *N*,*N*-dimethylformamidine (dmf) group is a mild protecting group for the exocyclic amine of nucleobases.^{24,25} However, during the course of our study of 7Me9C dG (**2a**), we found that when N¹ of 9-deazaguanine was protected with a benzyl (Bn) group, strong basic conditions and high temperature were required for complete removal of N^2 -dmf.¹⁸ Under the same conditions, TBS was also cleaved from the side chains of **3b** and **3d**. Therefore, we attempted alkylation using dmf-free 9-deaza-dG.^{23,26} The reaction conditions used

to introduce the 7-hydroxyethyl group to 9-deaza-dG caused serious epimerization, while in the other case, the 7-allyl group was added to 9-deaza-dG nonselectively at the N² and N⁷ positions at the same time. As the last resort, we revisited the possibility of using alkylated 9-deazaguanines that do not contain TBS or dmf groups for C-glycosidic bond formation. Here, we are pleased to report the successful Friedel-Crafts reactions of N^7 -hydroxyethyl-9-deazaguanine and N^7 -allyl-9-deazaguanine (Scheme 2A, entries 6 and 7). To our surprise, the unprotected hydroxyl group of N^7 -hydroxyethyl-9-deazaguanine did not significantly affect the yield of glycosylation. The free hydroxyl group was then protected as a TBS ether. Compound 3b was obtained by selectively removing the O3' and O5' toluoyl and the N^1 -benzyl groups. The product in entry 7 contained an allyl group, which was later dihydroxylated to generate a vicinal diol (Scheme 2A).²⁷⁻²⁹ The diol was protected by a pair of TBS groups, yielding compound 3d after removal of the toluoyl and benzyl groups.

RESULTS AND DISCUSSION

The N^2 -dmf-protected 9-deazaguanine 4, previously obtained from 2-amino-4-hydroxy-6-methylpyrimidine in five steps,²³ underwent Lewis-acid-mediated Friedel–Crafts reaction with a 1-*O*-methyl-3,5-O-ditoluated deoxyribose derivative to yield C-nucleoside 5 (Scheme 3A). This coupling mainly afforded the β -anomer in 26% yield. This yield is comparable to the reaction of N²- and N⁷-naked 9-deazaguanine (Scheme 2A, entry 1)²³ but significantly lower than what we observed for N²-dmf-N⁷-methyl-9-deazaguanine (Scheme 2A, entry 3).¹⁸ The hydroxylethyl group was introduced to the deprotonated N⁷

Scheme 3^{*a*}



^{*a*}(a) 1-(*α*,*β*)-*O*-methyl-3,5-di-(*O*-*p*-toluoyl)-2-deoxy-D-ribose, SnCl₄, 1:1 (v/v) dry CH₃CN-CH₂Cl₂, 65 °C, 16 h, 26%; (b) ethylene carbonate, DBU, dry DMF, 90 °C, 4 h, 49%; (c) 1 M NaOH, 2:1 (v/v) CH₃OH-H₂O, 70 °C, 16 h, 72%; (d) ammonium formate, Pd/C, CH₃OH, 75 °C, 16 h, 48%; (e) allyl bromide, NaH, dry THF, rt, 16 h, 48%; (f) osmium tetraoxide, TBHP, TBAF, 4:1 (v/v) acetone-H₂O, rt, 76%; (g) 1 M NaOH, 6:1 (v/v) CH₃OH-H₂O, 70 °C, 16 h, 54%; (h) ammonium formate, Pd/C, CH₃OH, 75 °C, 16 h, 74%; (i) KIO₄, 1:1 (v/v) CH₃OH-H₂O, rt, 30 min, 85%.

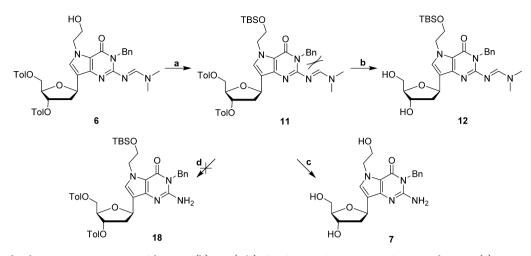
position using ethylene carbonate as the alkylating agent, and then the toluoyl and dmf groups were removed via strong alkaline hydrolysis to afford 7. Pd/C-catalyzed hydrogenolysis by ammonium formate was employed to deprotect the N^1 -Bn group under mild heating, which was previously demonstrated by us to be an efficient way to remove the N^1 -Bn group.¹⁸ This reaction lead to the formation of ^{7HE9C}dG (**2b**). The synthesis of ^{7OE9C}dG followed the same strategy. Allylation was performed on **5** to generate N^7 -allyl nucleoside **8**. Then the olefin underwent dihydroxylation to form a vicinal diol (**9**). After deprotection of dmf, toluoyl, and benzyl groups successively, ^{7DHP9C}dG (**2d**) was obtained. The vicinal diol was cleaved by potassium periodate to generate ^{7OE9C}dG (**2c**) (Scheme 3B). Nucleoside ^{7OE9C}dG was not very stable at room temperature. It was generated and purified immediately before characterization.

The stabilities of the glycosidic bonds of ^{7HE9C}dG and ^{7DHP9C}dG were examined after incubation under the following three conditions for 8 h: (i) HCl (pH 2.5), rt, (ii) NaOH (pH 11.7), rt, and (iii) phosphate buffer (pH 7.2), 70 °C. The NMR results confirmed that the glycosidic bond was stable under these conditions, and no sign of epimerization was detected. This result demonstrated the feasibility of using these structures as stable analogues of N^7 -alkyl-dG. ^{7OE9C}dG was excluded from the study due to the chemical instability of the aldehyde group.

Having completed the synthesis of ^{7HE9C}dG and ^{7OE9C}dG, we sought to add TBS groups to the 7-alkyl chains of 6 and 9 to generate side-chain-protected nucleosides. However, efforts to hydrolyze the toluoyl and dmf groups simultaneously while keeping the OTBS ether intact were futile (Scheme 4). Different bases (concentrated ammonia, diluted or 1 M KOH/NaOH), temperatures (0-75 °C), and solvent ratios (water-ethanol-THF mixture) were tested. We found that in general, mild conditions only enabled the removal of the toluoyl groups, while harsher conditions caused deprotection of toluoyl, dmf, and TBS at the same time. This was consistent with our past observation that hydrolysis of N^2 -dmf in the presence of N^1 -Bn was difficult.¹⁸ Anticipating that removal of the N^1 -Bn group first should facilitate the removal of the dmf group, we treated 11 with Pd/C-ammonium formate in refluxing methanol overnight. Unfortunately, the same reaction that worked efficiently in the syntheses of 2b and 2d did not show any effect on 11. Change of the conditions to refluxing DMF resulted in partial removal of the N^2 -dmf group (48 h, less than 10% conversion). The N^1 -Bn group, however, remained intact. Therefore, it appears that N^1 -Bn and N^2 -dmf of 11 have remarkable steric effect on each other, which blocks facile deprotection of either group.

To circumvent this problem, we decided to remove the dmf group prior to alkylation, on condition that the unprotected N^2 -amino group would not compete with the N⁷ alkylation under

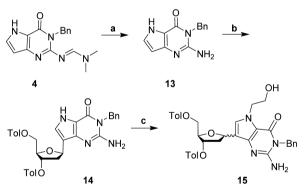
Scheme 4^a



^{*a*}(a) TBSCl, imidazole, 4-DMAP, DMF, rt, 16 h, 75%; (b) 5:1 (v/v) CH₃OH–H₂O, 0.05 M NaOH, rt, 1 h, 75%; (c) 1 M NaOH, 2:1 (v/v) CH₃OH-H₂O, 70 °C, 16 h, 72%; (d) different bases (ammonia, KOH, or NaOH), temperatures (0–75 °C), reaction times (1–48 h), and solvent ratios (water–ethanol–THF mixture).

basic conditions. The dmf-free 9-deaza-dG (14) was treated with DBU, followed by the addition of ethylene carbonate. Surprisingly, the reaction did not proceed under the previously adopted hydroxyethylation temperature (90 °C). This indicated that removal of N^2 -dmf group rendered the N^7 -H more difficult to dissociate. At an elevated temperature (110 °C), the desired hydroxyethylated product was obtained (Scheme 5), however, as

Scheme 5^{*a*}



^{*a*}(a) 1 M NaOH, 1:4 (v/v) H₂O–CH₃OH, reflux, 20 h, 75%; (b) 1-($\alpha_{\beta}\beta$)-O-methyl-3,5-di-(*O*-*p*-toluoyl)-2-deoxy-D-ribose, SnCl₄, 1:1 (v/v) dry CH₃CN–CH₂Cl₂, 65 °C, 16 h, 26%; (c) ethylene carbonate, DBU, dry DMF, 110 °C, 4 h, 46%, $\alpha:\beta = 1:3$.

a mixture of inseparable α/β isomers (1:3, ¹H NMR integration). Therefore, an alternative solution to the synthesis of **3b** was then developed.

It is interesting to compare the epimerization of 14 with other C-nucleosides reported by us and others.^{18,23,26,31–33} Hamm et al. first reported epimerization of unalkylated 9-deaza-dG resulting from treatment in concentrated ammonia at 55 °C for 15 h. The same observation was not found at room temperature.²⁶ In contrast, we previously observed that N^7 -methyl-9-deaza-dG (2a) was not prone to epimerization under strong alkaline conditions at high temperature.¹⁸ Therefore, we concluded that deprotonating N^7 -H through either general base catalysis or specific base catalysis is required for the epimerization of 9-deazaguanine nucleosides. Here we further provide evidence to show that 9-deaza-dG epimerizes at 110 °C in the presence of only 1 mol equiv of DBU, while the same treatment at 90 °C did not result in detectable epimerization. Furthermore, the less electron-donating effect exerted by N^2 -dmf can attenuate epimerization. This electronic effect is consistent with the literature record of the *C*-glycoside of 6-amino-pyridone.³⁴ In short, the availability of N^7 -H, alkaline pH, high temperature, and electron-donating groups on the nucleobase have in combination created a favorable condition for the epimerization of 9-deaza-dG.

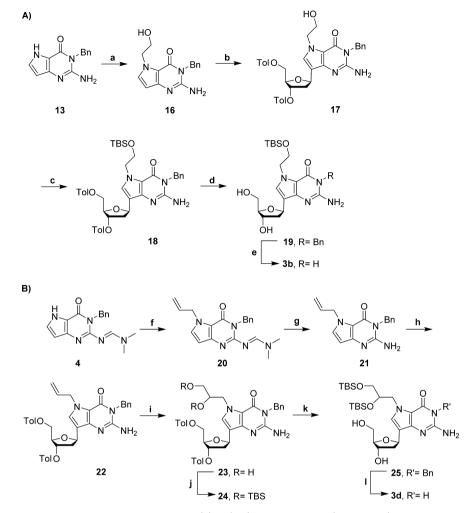
Nonreacting hydroxyl groups are often protected in a Lewis acid-promoted glycosylation reaction. However, when the glycosyl donor is an *O*-glycoside, attack of the alkoxyl group to the oxocarbenium intermediate is reversible. As such, formation of a more stable C-glycosidic bond will be favored over an O-glycosidic bond. On the basis of this analysis, we treated the unprotected N^7 -hydroxyethyl-9-deazaguanine (16) with the sugar donor successfully with no significantly lower yield (Scheme 6A). The hydroxyl group was then protected as a TBS ether, followed by removal of the two toluoyl groups at room temperature (19). The N^1 -Bn group of 19 was deprotected smoothly using Pd/C-catalyzed hydrogenolysis to afford 3b.

The synthesis of 3d was carried out using a similar strategy (Scheme 6B). As we observed that allylation of 13 produced a mixture of N^2 - and N^7 -allyl products, the N^2 -dmf-protected 9-deazaguaine (4) was used as the starting material instead. After the allyl group was specifically introduced to N^7 , the N^2 -dmf group was removed under strong alkaline conditions (21). The N^7 -allyl group survived the next Friedel–Crafts reaction. Then the olefin was dihydroxylated to form a vicinal diol 23, followed by TBS protection of both hydroxyl groups (24). The two toluoyl groups and the N^1 -Bn were successively removed under the same conditions as described for the synthesis of 3b.

CONCLUSION

In summary, we have successfully synthesized two polar-chaincontaining N^7 -alkyl-9-deaza-dGs and expanded the series of chemically stable analogues of N^7 -alkyl-dGs. The C-glycosidic bonds of the two new compounds were proved to be stable under strong acidic and basic conditions and at high temperatures. In addition, we overcame the difficulties in stereoselectivity and regioselectivity when the exocyclic amine of 9-deazaguanine was

Scheme 6^{*a*}



^{*a*}(a) Ethylene carbonate, DBU, dry DMF, 90 °C, 4 h, 47%; (b) 1-(*α*,*β*)-*O*-methyl-3,5-di-(*O*-*p*-toluoyl)-2-deoxy-D-ribose, SnCl₄, 1:1 (v/v) dry CH₃CN-CH₂Cl₂ reflux, 16 h, 28%; (c) TBSCl, imidazole, 4-DMAP, DMF, rt, 16 h, 93%; (d) 0.05 M NaOH, 10:1 (v:v) CH₃OH-H₂O, rt, 1 h, 74%; (e) ammonium formate, Pd/C, CH₃OH, 75 °C, 16 h, 43%; (f) allyl bromide, NaH, dry THF, rt, 4 h, 64%; (g) 1 M NaOH, 1:4 (v/v) H₂O-CH₃OH, 70 °C, 16 h, 70%; (h) 1-(*α*,*β*)-*O*-methyl-3,5-di-(*O*-*p*-toluoyl)-2-deoxy-D-ribose, SnCl₄, 1:1 (v/v) dry CH₃CN-CH₂Cl₂, 65 °C, 16 h, 23%; (i) osmium tetraoxide, TBHP, TBAF, 4:1 (v/v) acetone-H₂O, rt, 16 h, 40%; (j) TBSCl, imidazole, 4-DMAP, DMF, rt, 16 h, 76%; (k) 0.1 M NaOH, 10:1 (v:v) CH₃OH-H₂O, rt, 1 h, 68%; (l) ammonium formate, Pd/C, CH₃OH, 75 °C, 16 h, 56%.

not protected and developed an efficient synthesis of the two nucleosides whose N⁷ polar side chains were protected by TBS groups. These side-chain-protected nucleosides can be readily converted to phosphoramidites for solid-phase oligonucleotide synthesis, which will play essential roles in elucidating the polymerase actions on N⁷-hydroxyethyl and N⁷-oxoethyl adducts of 2'-deoxyguanosine.

EXPERIMENTAL SECTION

General Information. ¹H and ¹³C NMR spectra were recorded on a NMR spectrometer operating at 500 or 600 MHz for ¹H and 125 or 150 MHz for ¹³C using the solvent as an internal reference. The coupling constants (*J*) for ¹H NMR are recorded in hertz. High resolution mass spectra (HRMS) of compounds **9**, **11**, **12**, **18**, and **24** were obtained using a MALDI-TOF spectrometer and all others with an ICR (ESI) spectrometer. Melting points were recorded on a microscopic instrument.

THF was distilled freshly from LiAlH₄. Acetonitrile, dichloromethane, and DMF were distilled freshly from CaH₂. 1-Benzyl-9-deazaguanine (4) was prepared following Rana's protocol, and its characterization was described previously by Gibson et al.^{18,23} 1-(α , β)-O-Methyl-3,5-di(O-*p*-toluoyl)-2-deoxy-D-ribose was prepared according to a reported procedure.³⁰

Stability Test. ^{7HE9C}dG and ^{7DHP9C}dG were incubated under the following three conditions for 8 h: (i) HCl (pH 2.5), rt; (ii) NaOH (pH 11.7), rt; (iii) phosphate buffer (pH 7.2), 70 °C. After that, the reaction mixtures were neutralized, concentrated, and passed through a silica gel column using a relatively polar eluent (methanol/ethyl acetate 1:2, compared to 1:5 for **2b** and **2d**) to elute possible cleaved products. The NMR spectra were compared with the spectra of **2b** and **2d** to determine any possible bond cleavage and epimerization.

(2R,3S,5R)-5-(3-Benzyl-2-(((dimethylamino)methylene)amino)-4oxo-4,5-dihydro-3*H*-pyrrolo[3,2-*d*]pyrimidin-7-yl)-2-(((4methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (5). To a suspension of *N'*-(3-benzyl-4-oxo-4,5-dihydro-3*H*-pyrrolo-[3,2-*d*]pyrimidin-2-yl)-*N*,*N*-dimethylformimidamide 4 (3.52 g, 12.0 mmol) and 1-(α , β)-O-methyl-3,5-di(*O*-*p*-toluoyl)-2-deoxy-D-ribose (6.6 g, 17.2 mmol) in a mixture of methylene chloride (20.0 mL) and acetonitrile (20.0 mL) was added a solution of SnCl₄ (23.2 mL, 23.2 mmol, 1 M in CH₂Cl₂). The reaction mixture was heated at 65 °C for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with sat. NaHCO₃ and brine. The organic layer was separated and dried over MgSO₄. The solution was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with hexane/ethyl acetate 2:1 to 1:2) to give compound **5** as a light yellow solid (1.96 g, 26%). mp 92–93 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.36 (s, 3H), 2.42 (s, 3H), 2.49–2.53 (m, 1H), 2.91–2.95 (m, 1H), 3.05 (s, 3H), 3.12 (s, 3H), 4.50 (s, 1H), 4.58 (dd, J = 10.5, 5.0 Hz, 1H), 4.66 (dd, J = 11.0, 4.5 Hz, 1H), 5.49–5.56 (m, 3H), 5.74 (s, 1H), 7.15– 7.36 (m, 10H), 7.90 (d, J = 7.0 Hz, 2H), 8.01 (s, 2H), 8.58 (s, 1H), 10.36 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) 21.6, 21.7, 35.0, 37.9, 40.8, 45.6, 60.4, 64.8, 73.8, 81.8, 114.4, 116.2, 126.8, 127.0, 127.2, 127.23, 128.0, 128.1, 129.1, 129.14, 129.7, 129.8, 138.9, 143.6, 144.0, 153.7, 156.2, 156.8, 166.2, 166.5. ESI-MS (M + H)⁺ for C₃₇H₃₇N₅O₆: expected 648.2822, found 648.2852.

(2R,3S,5R)-5-(3-Benzyl-2-(((dimethylamino)methylene)amino)-5-(2-hydroxyethyl)-4-oxo-4,5-dihydro-3H-pyrrolo[3,2-d]pyrimidin-7yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (6). To a solution of 5 (0.52 g, 0.8 mmol) in dry DMF (5.0 mL) were added DBU (280.0 µL, 1.8 mmol) and ethylene carbonate (0.21 g, 2.4 mmol). The reaction was heated under 90 °C for 4 h. After removal of DMF, the reaction mixture was diluted with ethyl acetate and washed successively with water and brine. The solution was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 1:4 to1:8) to give compound 6 as a white solid (0.27 g, 49%). mp 75-76 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.40 (s, 3H), 2.44 (s, 3H), 2.52-2.57 (m, 1H), 2.85-2.89 (m, 1H), 3.05 (s, 3H), 3.15 (s, 3H), 3.90 (s, 2H), 4.46 (s, 2H), 4.51 (s, 1H), 4.59 (dd, J = 11.0, 4.0 Hz, 1H), 4.70 (dd, J = 11.3, 4.5 Hz, 1H), 5.53 (s, 3H), 5.76 (s, 1H), 7.09 (s, 1H), 7.19-7.21 (m, 3H), 7.26-7.29 (m, 4H), 7.35 (d, J = 5.0 Hz, 2H), 7.94 (d, J = 7.5 Hz, 2H), 8.01 (d, J = 7.5 Hz, 2H), 8.60 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 21.5, 21.6, 34.9, 37.8, 40.7, 45.2, 50.8, 63.0, 64.5, 73.5, 81.7, 113.4, 115.2, 126.6, 127.0, 127.04, 127.6, 128.0, 129.0, 129.4, 129.6, 130.7, 138.6, 143.6, 143.8, 153.7, 156.3, 156.6, 166.0, 166.3. ESI-MS (M + H)⁺ for C₃₉H₄₁N₅O₇: expected 692.3084, found 692.3122.

2-Amino-3-benzyl-7-((2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-5-(2-hydroxyethyl)-3,5-dihydro-4H-pyrrolo[3,2d]pyrimidin-4-one (7). A suspension of 6 (0.15 g, 0.22 mmol) in 1 M sodium hydroxide in a mixture of methanol (2.0 mL) and water (1.0 mL) was heated under 70 $^{\circ}\mathrm{C}$ for 16 h and allowed to cool to room temperature. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ ethyl acetate 1:5 to 1:10) to give 7 as a white solid (0.063 g, 72%). mp 99–100 °C. ¹H NMR (500 MHz, CD₃OD) δ ppm 2.08 (dd, J = 13.0, 5.0 Hz, 1H), 2.46–2.52 (m, 1H), 3.69 (d, J = 12.0 Hz, 1H), 3.80–3.85 (m, 3H), 4.02 (s, 1H), 4.36-4.41 (m, 3H), 4.45 (d, J = 5.0 Hz, 1H), 5.28–5.32 (m, 3H), 7.22 (d, J = 7.5 Hz, 2H). 7.25 (d, J = 7.5 Hz, 2H), 7.31 (t, J = 8.0 Hz, 2H). ¹³C NMR (125 MHz, CD₃OD) δ ppm 42.1, 43.7, 50.5, 61.7, 63.4, 74.4, 74.7, 87.9, 112.4, 112.8, 126.17, 127.2, 128.4, 131.7, 135.7, 142.6, 151.0, 154.7. ESI-MS $(M + H)^+$ for $C_{20}H_{24}N_4O_5$: expected 401.1825, found 401.1839.

2-Amin o-7-((2*R*,4*S*,5*R*)-4-hydr oxy-5-(hydr oxymethyl)tetrahydrofuran-2-yl)-5-(2-hydroxyethyl)-3,5-dihydro-4*H*-pyrrolo[3,2*d*]pyrimidin-4-one (**2b**). To a mixture of 7 (0.063 g, 0.16 mmol) and 10% palladium on carbon (0.03 g) in methanol (2.0 mL) was added ammonium formate (0.10 g, 1.5 mmol) under Ar. The reaction mixture was heated under 75 °C for 16 h and then filtered through Celite. The filtrate was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with methanol/ethyl acetate 1:50 to 1:5) to give **2b** as a white solid (0.024 g, 48%). mp 124–125 °C. ¹H NMR (S00 MHz, CD₃OD) δ 2.04 (dd, *J* = 11.0, 4.5 Hz, 1H), 2.43 (ddd, *J* = 11.0, 9.5, 4.5 Hz, 1H), 3.70 (dd, *J* = 10.0, 2.0 Hz, 1H), 3.79–3.83 (m, 3H), 4.00 (s, 1H) 4.34–4.37(m, 2H), 4.44 (d, *J* = 4.5 Hz, 1H), 5.26 (dd, *J* = 9.5, 4.5 Hz, 1H). 7.20 (s, 1H). ¹³C NMR was not available due to the poor solubility. ESI-MS (M + H)⁺ for C₁₃H₁₈N₄O₅: expected 311.1355, found 311.1377.

(2R,3S,5R)-5-(5-Allyl-3-benzyl-2-(((dimethylamino)methylene)amino)-4-oxo-4,5-dihydro-3*H*-pyrrolo[3,2-*d*]pyrimidin-7-yl)-2-(((4methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (8). To a suspension of 5 (1.61 g, 2.4 mmol) in dry THF (25.0 mL) was added sodium hydride (0.1 g, 2.3 mmol, 60% in mineral oil). The mixture was stirred for 15 min before allyl bromide (150 μ L, 3.5 mmol) was added. The mixture was stirred at room temperature for 16 h. After removal of solvent, the reaction was diluted with methylene chloride and washed successively with water and brine. The organic layer was separated, dried over MgSO₄, concentrated, and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with hexane/ethyl acetate 5:1 to 3:1) to give compound **8** as a white solid (0.82 g, 48%). mp 94–95 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.41 (s, 3H), 2.45 (s, 3H), 2.53–2.57 (m, 1H), 2.89–2.92 (m, 1H), 3.04 (s, 3H), 3.12 (s, 3H), 4.52–4.55 (m, 1H), 4.60 (dd, *J* = 11.5, 4.0 Hz, 1H), 4.71 (dd, *J* = 11.5, 5.0 Hz, 1H), 5.01–5.09 (m, 2H), 5.13–5.23 (m, 2H), 5.55 (s, 3H), 5.77 (t, *J* = 5.0 Hz, 1H), 6.00–6.05 (m, 1H), 7.06 (s, 1H), 7.19–7.23 (m, 3H), 7.26–7.29 (m, 4H), 7.38–7.39 (m, 2H), 7.95 (d, *J* = 7.5 Hz, 2H), 8.01 (d, *J* = 7.5 Hz, 2H), 8.59 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 21.8, 21.83, 35.2, 38.4, 41.0, 45.3, 50.6, 64.8, 68.6, 73.8, 82.1, 114.0, 117.6, 126.8, 127.3, 127.9, 128.3, 129.0, 129.2, 129.23, 129.26, 129.3, 129.85, 129.9, 130.1, 134.5, 143.8, 144.0, 154.2, 166.5. ESI-MS (M + H)⁺ for C₄₀H₄₁N₅O₆: expected 688.3135, found 688.3202.

(2R,3S,5R)-5-(3-Benzyl-5-(2,3-dihydroxypropyl)-2-(((dimethylamino)methylene)amino)-4-oxo-4,5-dihydro-3H-pyrrolo-[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (9). To the suspension of 8 (0.82 g, 1.2 mmol) in a mixture of acetone (4.0 mL) and water (1.0 mL) were added TBHP (220 µL, 1.1-1.3 mmol, 5.0-6.0 M in decane), TBAF (0.028 g, 0.11 mmol), and OsO₄ (trace). The reaction was stirred at room temperature for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with water and brine. The organic layer was separated, dried over MgSO4, concentrated, and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with hexane/ethyl acetate 1:1 to 1:3) to give compound 9 as a light yellow solid (0.66 g, 76%). mp 80-81 °C. ¹H NMR (500 MHz, CDCl₂) δ ppm 2.40 (s, 3H), 2.44 (s, 3H), 2.52-2.60 (m, 1H), 2.84-2.93 (m, 1H), 3.06 (s, 3H), 3.15 (s, 3H), 3.51 (s, 2H), 3.88-3.94 (m, 2H), 4.30-4.34 (m, 1H), 4.51-4.60 (m, 3H), 4.69-4.72 (m, 1H), 5.54 (s, 3H), 7.11 (s, 1H), 7.19–7.35 (m, 9H), 7.93 (d, J = 8.0 Hz, 2H), 8.01 (s, 2H), 8.58 (s, 1H). $^{13}\mathrm{C}$ NMR for both diastereomers (125 MHz, CDCl_3) δ ppm 21.0, 21.6, 35.0, 37.8, 40.8, 45.4, 50.0, 50.1, 60.4, 62.9, 63.0, 64.6, 71.8, 73.5, 73.6, 81.8, 113.8, 126.8, 127.08, 127.1, 127.7, 128.1, 129.0, 129.09, 129.1, 129.2, 129.68, 129.7, 129.72, 131.7, 138.7, 143.7, 143.9, 153.8, 156.8, 166.4. MALDI-MS $(M + H)^+$ for $C_{40}H_{43}N_5O_8$: expected 722.3190, found 722.3193.

2-Amino-3-benzyl-5-(2,3-dihydroxypropyl)-7-((2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-3,5-dihydro-4Hpyrrolo[3,2-d]pyrimidin-4-one (10). A suspension of 9 (0.66 g, 0.91 mmol) in 1 M sodium hydroxide in a mixture of methanol (3 mL) and water (0.5 mL) was heated under 70 °C for 16 h and allowed to cool to room temperature. The reaction mixture was concentrated and purified by column chromatography (SiO $_2$, 0.06–0.20 mm, eluting with methanol/ethyl acetate 1:20 to 1:10) to give 10 as a white solid (0.21 g, 54%). mp 93–94 °C. ¹H NMR (500 MHz, CD₃OD) δ ppm 2.07 (dd, J = 13.0, 5.5 Hz, 1H), 2.51 (ddd, J = 13.0, 11.5, 5.5 Hz, 1H), 3.48 (ddd, J = 11.5, 5.5, 3.0 Hz, 1H), 3.54 (ddd, J = 11.5, 5.0, 2.0 Hz, 1H), 3.69 (dd, J = 9.5, 2.5 Hz, 1H), 3.82 (dd, J = 9.5, 2.5 Hz, 1H), 3.93-3.97 (m, 1H), 4.00–4.02 (m, 1H), 4.26 (dt, J = 14.0, 5.0 Hz, 1H), 4.45–4.46 (d, *J* = 5.0 Hz, 1H), 4.51–4.56 (m, 1H), 5.29–5.35 (m, 3H), 7.23–7.29 (m, 4H), 7.32-7.35 (m, 2H). ¹³C NMR for both diastereomers (125 MHz, CD₃OD) δ ppm 44.18, 44.2, 45.9, 52.6, 62.2, 65.4, 65.5, 73.9, 76.5, 76.8, 90.1, 114.9, 115.1, 128.3, 129.3, 130.5, 134.3, 137.9, 145.3, 153.1, 157.2. ESI-MS $(M + H)^+$ for $C_{21}H_{26}N_4O_6$: expected 431.1931, found 431.1966.

2-Amino-5-(2,3-dihydroxypropyl)-7-((2*R*,4*S*,5*R*)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-3,5-dihydro-4*H*-pyrrolo[3,2-*d*]-pyrimidin-4-one (**2d**). To a mixture of **10** (0.21 g, 0.49 mmol) and 10% palladium on carbon (0.1 g) in methanol (5 mL) was added ammonium formate (0.29 g, 4.5 mmol) under Ar. The reaction mixture was heated under 75 °C for 16 h and then filtered through Celite. The filtrate was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with methanol/ethyl acetate 1:20 to 1:5) to give **2d** as a white solid (0.12 g, 74%). mp 135–136 °C. ¹H NMR (500 MHz, *d*₆-DMSO) δ ppm 1.90 (dd, *J* = 13.8, 4.8 Hz, 1H), 2.11–2.17 (m, 1H), 3.27 (t, *J* = 5.0 Hz, 2H), 3.45 (s, 2H), 3.71 (s, 2H), 4.03–4.09 (m, 1H), 4.18 (s, 1H), 4.32–4.35 (m, 1H), 4.66 (s, 1H), 4.91 (s, 2H), 5.07 (dd, *J* = 11.2, 6.0 Hz, 1H), 5.20 (br, 1 H), 5.72 (s, 2H), 7.15 (s, 1H), 10.53 (s, 1H). ¹³C NMR for both of the diastereomers (150 MHz, *d*₆-DMSO)

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 δ ppm 42.7, 51.6, 59.9, 63.9, 64.4, 72.3, 73.5, 74.0, 88.4, 113.3, 114.4, 131.3, 132.3, 151.6, 156.3. ESI-MS (M + H)^+ for C_{14}H_{20}N_4O_6: expected 341.1461, found 341.1486.

2-(2-Amino-7-((2*R*,4*S*,5*R*)-4-hydroxy-5-(hydroxymethyl)-tetrahydrofuran-2-yl)-4-oxo-3,4-dihydro-5*H*-pyrrolo[3,2-*d*]pyrimidin-5-yl)acetaldehyde (**2c**). To compound **2d** (0.050 g, 0.15 mmol) in a mixture of methanol (1 mL) and water (1 mL) was added potassium periodate (0.035 g, 0.15 mmol). The reaction mixture was stirred under room temperature for 30 min, concentrated, and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with methanol/ethyl acetate 1:20 to 1:5) to give **2c** as a white solid (0.039 g, 85%). mp 117–118 °C. ¹H NMR (600 MHz, *d*₆-DMSO) δ ppm 1.91–1.99 (m, 1H), 2.10–2.13 (m, 1H), 3.44 (s, 2H), 3.72 (s, 1H), 4.20–4.22 (m, 1H), 4.93–4.95 (m, 1H), 5.07 (s, 2H), 5.84–6.01 (m, 2H), 7.16 (s, 1H), 9.60 (s, 1H), 10.66–10.72 (s, 1H). ¹³C NMR was not available due to the poor solubility. ESI-MS (M + H)⁺ for C₁₃H₁₆N₄O₅: expected 309.1199, found 309.1220.

(2R,3S,5R)-5-(3-Benzyl-5-(2-((tert-butyldimethylsilyl)oxy)ethyl)-2-(((dimethylamino)methylene)amino)-4-oxo-4,5-dihydro-3H-pyrrolo-[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (11). To a solution of 6 (0.45 g, 0.65 mmol) in DMF were added imidazole (0.13 g, 1.9 mmol), 4-DMAP (0.002 g, 0.018 mmol), and TBSCl (0.29 g, 1.9 mmol). The reaction was stirred under room temperature for 16 h. After removal of the solvent, the mixture was diluted by methylene chloride and washed with water and brine. The solution was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ ethyl acetate 5:1 to 3:1) to give compound 11 as a light yellow solid (0.43 g, 82%). mp 50–51 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm -0.12 (s, 6H), 0.83 (s, 9H), 2.40 (s, 3H), 2.44 (s, 3H), 2.49-2.52 (m, 1H), 2.87-2.89 (m, 1H), 3.03 (s, 3H), 3.10 (s, 3H), 3.93 (s, 2H), 4.44 (s, 2H), 4.52 (s, 1H), 4.60 (dd, J = 11.3, 4.5 Hz, 1H), 4.66 (dd, J = 11.5, 5.0 Hz, 1H), 5.56 (s, 3H), 5.78 (s, 1H), 7.12 (s, 1H), 7.19-7.21 (m, 3H), 7.25–7.29 (m, 4H), 7.35–7.37 (m, 2H), 7.96 (d, J = 7.5 Hz, 2H), 8.01 (d, J = 6.5 Hz, 2H), 8.59 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ ppm -5.4, 18.4, 21.9, 21.94, 26.1, 31.8, 35.2, 41.0, 45.3, 51.3, 63.5, 65.0, 73.6, 82.1, 113.0, 114.7, 126.8, 127.46, 127.5, 127.9, 128.3, 129.3, 129.4, 129.96, 130.0, 139.3, 143.1, 143.9, 144.1, 154.2, 156.0, 156.8, 166.4, 166.6. MALDI-MS $(M + H)^+$ for $C_{45}H_{55}N_5O_7Si$: expected 806.3943, found 806.3944.

N'-(3-Benzyl-5-(2-((tert-butyldimethylsilyl)oxy)ethyl)-7-((2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-4oxo-4,5-dihydro-3H-pyrrolo[3,2-d]pyrimidin-2-yl)-N,N-dimethylformimidamide (12). To compound 11 (0.43 g, 0.53 mmol) in a mixture of methanol (5 mL) and water (1 mL) was added sodium hydroxide (0.05 M). The reaction mixture was stirred under room temperature for 1 h. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with methanol/ethyl acetate 1:50 to 1:20) to give 12 as a light yellow solid (0.20 g, 72%). mp 65–66 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm –0.11 (s, 3H), -0.09 (s, 3H), 0.83 (s, 9H), 2.04 (dd, J = 11.0, 5.0 Hz, 1H), 2.60-2.66 (m, 1H), 3.03 (s, 3H), 3.14 (s, 3H), 3.72-3.75 (m, 1H), 3.86-3.90 (m, 3H), 4.12 (s, 1H), 4.32–4.35 (m, 1H), 4.56–4.63 (m, 2H), 5.32 (dd, J= 11.0, 5.0 Hz, 1H), 5.49 (s, 2H), 7.06 (s, 1H), 7.19-7.33 (m, 5H), 8.28 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ ppm -5.4, 18.4, 26.1, 35.1, 41.0, 43.8, 45.6, 51.1, 63.5, 64.1, 75.7, 88.5, 113.5, 115.1, 127.0, 127.9, 128.4, 131.4, 138.7, 154.8, 155.8, 157.1. MALDI-MS (M + H)⁺ for C₂₉H₄₃N₅O₅Si: expected 570.3112, found 570.3108.

2-Amino-3-benzyl-3,5-dihydro-4*H*-pyrrolo[3,2-d]pyrimidin-4-one (13). A suspension of **6** (3 g, 10.2 mmol) in 1 M sodium hydroxide in a mixture of methanol (12 mL) and water (6 mL) was heated under 70 °C for 20 h and allowed to cool to room temperature. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with ethyl acetate) to give 13 as a white solid (1.86 g, 75%). The characterization of this compound was previously described by Gibson et al.²³

(2R,3S,5R)-5-(2-Amino-3-benzyl-4-oxo-4,5-dihydro-3*H*-pyrrolo-[3,2-*d*]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (14). To compound 13 (0.31 g, 1.3 mmol) and 1- (α,β) -O-methyl-3,5-di(O-p-toluoyl)-2-deoxy-D-ribose

(0.7 g, 1.9 mmol) in a mixture of methylene chloride (3.0 mL) and acetonitrile (3.0 mL) was added a solution of SnCl₄ (2.5 mL, 2.5 mmol, 1 M in CH₂Cl₂). The reaction mixture was heated at 65 °C for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with sat. NaHCO3 and brine. The organic layer was separated and dried over MgSO4. The solution was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 1:1 to 1:2) to give compound 14 as a light yellow solid (0.20 g, 26%). mp 84–85 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.37 (s, 3H), 2.41 (s, 3H), 2.60–2.63 (m, 2H), 4.53 (s, 1H), 4.64 (dd, J = 12.0, 4.0 Hz, 1H), 4.78 (dd, J = 11.5, 4.5 Hz, 1H), 5.34-5.42 (m, J = 11.5, 4.5 Hz, 1H)2H), 5.47-5.50 (m, 1H), 5.65-5.66 (m, 1H), 7.18-7.33 (m, 10H), 7.91 (d, J = 7.5 Hz, 2H), 7.98 (d, J = 8.0 Hz, 2H), 10.54 (s, 1H).¹³C NMR (125 MHz, CDCl₃) δ ppm 21.9, 22.0, 30.0, 39.3, 45.1, 65.1, 73.6, 82.9, 113.0, 113.7, 127.0, 127.2, 127.3, 128.5, 129.3, 129.39, 129.4, 129.5, 130.2, 134.8, 143.7, 144.1, 144.2, 151.3, 166.5, 166.8. ESI-MS (M + H)⁺ for C₃₄H₃₂N₄O₆: expected 593.2401, found 593.2422.

(2R,3S,5R,5S)-5-(2-Amino-3-benzyl-5-(2-hydroxyethyl)-4-oxo-4,5dihydro-3H-pyrrolo[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (15). To compound 14 (0.100 g, 0.17 mmol) in dry DMF (2 mL) were added DBU (28 μ L, 0.18 mmol) and ethylene carbonate (0.04 g, 0.45 mmol). The reaction was heated under 110 °C for 4 h. After removal of the solvent, the reaction mixture was diluted with ethyl acetate and washed with water and brine. The residue was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ ethyl acetate 1:5 to 1:8) to give compound 15 as a white solid (0.050 g, 46%). The ratio of the epimers ($\alpha:\beta = 1:3$) was determined by ¹H NMR. ¹H NMR for both epimers (600 MHz, CDCl₃) δ ppm 2.38 (s, 4.78H), 2.41 (s, 3H), 2.50 (dd, I = 11.2, 5.6 Hz, 1.35H), 2.62–2.65 (m, 1.38H), 3.35 (s, 0.66H), 3.67 (s, 2H), 3.89 (t, J = 5.6 Hz, 2.88H), 4.42 (t, J = 5.6 Hz, 2H), 4.46–4.47 (m, 1.59H), 4.51–4.53 (m, 0.53H), 4.55 (dd, J = 11.2, 5.6 Hz, 1H), 4.64–4.65 (m, 0.39H), 4.75 (dd, J = 11.2, 5.6 Hz, 1H), 5.11 (s, 0.61H), 5.20–5.29 (m, 3.21H), 5.42 (dd, J = 9.6, 4.8 Hz, 1H), 5.48 (t, J = 6.6 Hz, 0.38H), 5.52–5.58 (m, 0.34H), 5.64 (d, J = 5.4 Hz, 1H), 7.08 (s, 1H), 7.16–7.36 (m, 13.72H), 7.78 (d, J = 7.2 Hz, 0.62H), 7.93 (d, J = 7.8 Hz, 2.47H), 7.97 (d, J = 8.2 Hz, 2H). ¹³C NMR for both epimers (150 MHz, CDCl₃) δ ppm 21.80, 21.82, 42.2, 44.6, 51.3, 62.2, 64.7, 73.2, 83.2, 110.6, 110.9, 112.4, 126.9, 127.1, 127.3, 128.6, 129.2, 129.3, 129.8, 129.9, 130.1, 130.2, 131.2, 133.4, 144.0, 144.1, 144.2, 151.36, 151.40, 152.5, 166.5, 166.6, 166.7.

2-Amino-3-benzyl-5-(2-hydroxyethyl)-3,5-dihydro-4*H*-pyrrolo[3,2*d*]pyrimidin-4-one (**16**). To compound **13** (1.3 g, 4.4 mmol) in dry DMF (5 mL) were added DBU (280 μ L, 1.83 mmol) and ethylene carbonate (1.0 g, 11.7 mmol). The reaction was heated under 90 °C for 4 h. After removal of the solvent, the reaction mixture was diluted with ethyl acetate and washed with water and brine. The residue was chromatographed by ethyl acetate to give compound **16** as a white solid (0.6 g, 47%). mp 168–169 °C. ¹H NMR (500 MHz, *d*₆-DMSO) δ ppm 3.65 (dd, *J* = 9.5, 5.0 Hz, 2H), 4.30 (t, *J* = 4.5 Hz, 2H), 5.23 (s, 2H), 5.90 (s, 1H), 6.25 (s, 1H), 7.21–7.26 (m, 4H), 7.32 (t, *J* = 6.0 Hz, 2H). ¹³C NMR (150 MHz, CD₃OD) δ ppm 45.1, 51.9, 63.2, 100.3, 112.7, 127.5, 128.5, 129.8, 134.6, 137.3, 146.9, 152.8, 156.2. ESI-MS (M + H)⁺ for C₁₅H₁₆N₄O₂: expected 285.1352, found 285.1337.

(2R,3S,5S)-5-(2-Amino-3-benzyl-5-(2-hydroxyethyl)-4-oxo-4,5-dihydro-3*H*-pyrrolo[3,2-*d*]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (17). To compound 16 (0.6 g, 2.2 mmol) and $1-(\alpha,\beta)$ -O-methyl-3,5-di(O-p-toluoyl)-2-deoxy-Dribose (2.4 g, 3.3 mmol) in a mixture of methylene chloride (5.0 mL) and acetonitrile (5.0 mL) was added a solution of SnCl₄ (4.5 mL, 4.5 mmol, 1 M in CH₂Cl₂). The reaction mixture was heated at 65 °C for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with sat. NaHCO3 and brine. The organic layer was separated and dried over MgSO4. The solution was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with hexane/ethyl acetate 1:1 to 1:2) to give compound 17 as a white solid (0.39 g, 28%). mp 89–90 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.40 (s, 3H), 2.43 (s, 3H), 2.50 (dd, J = 13.0, 5.0 Hz, 1H), 2.67 (ddd, J = 10.5, 10.5, 4.5 Hz, 1H), 3.60 (s, 1H), 3.90 (s, 2H), 4.42 (s, 3H), 4.55 (dd, J = 10.5, 4.5 Hz, 1H), 4.73 (dd, J = 10.5, 4.5 Hz, 1H), 5.02 (s, 2H), 5.19–5.26 (m, 2H), 5.45 (dd, *J* = 10.5, 5.0 Hz, 1H), 5.65 (d, *J* = 5.0 Hz, 1H), 7.09 (s, 1H), 7.20–7.32 (m, 9H), 7.95 (d, *J* = 8.0 Hz, 2H), 7.99 (d, *J* = 8.0 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 21.87, 21.9, 38.6, 44.8, 51.3, 63.0, 65.0, 73.5, 82.6, 112.8, 113.1, 126.7, 127.3, 127.4, 128.2, 129.38, 129.4, 129.9, 130.0, 131.0, 135.3, 144.1, 144.3, 150.9, 155.2, 166.4, 166.7. ESI-MS (M + H)⁺ for C₃₆H₃₆N₄O₇: expected 637.2663, found 637.2697.

(2R,3S,5S)-5-(2-Amino-3-benzyl-5-(2-((tert-butyldimethylsilyl)oxy)ethyl)-4-oxo-4,5-dihydro-3H-pyrrolo[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (18). To compound 17 (0.39 g, 0.6 mmol) in DMF were added imidazole (0.13 g, 1.9 mmol), 4-DMAP (0.002 g, 0.018 mmol), and TBSCl (0.3 g, 2.0 mmol). The reaction was stirred under room temperature for 16 h. After removal of the solvent, the mixture was diluted by methylene chloride and washed with water and brine. The solution was concentrated and purified by column (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 5:1 to 3:1) to give compound 18 as a gummy solid (0.42 g, 93%). mp 55–56 °C. 1 H NMR $(500 \text{ MHz}, \text{CDCl}_3) \delta \text{ ppm} - 0.11 (d, J = 5.0 \text{ Hz}, 6\text{H}), 0.82 (s, 9\text{H}), 2.39$ (s, 3H), 2.41 (s, 3H), 2.44 (dd, J = 11.5, 4.5 Hz, 1H), 2.62 (ddd, J = 11.5, 11.5, 4.5 Hz, 1H), 3.90 (t, J = 3.5 Hz, 2H), 4.31–4.37 (m, 2H), 4.44 (dt, *J* = 11.5, 3.5 Hz, 1H), 4.51 (dd, *J* = 9.5, 3.5 Hz, 1H), 4.67 (dd, *J* = 9.5, 4.0 Hz, 1H), 5.14–5.28 (m, 3H), 5.45 (dd, J = 9.5, 4.5 Hz, 1H), 5.65 (d, J = 4.5 Hz, 1H), 7.08 (s, 1H), 7.19–7.30 (m, 9H), 7.94 (d, J = 7.0 Hz, 2H), 7.97 (d, J = 6.5 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ ppm –5.6, 18.1, 21.6, 21.65, 25.8, 37.8, 44.3, 51.1, 53.5, 63.1, 64.7, 72.6, 82.3, 111.7, 112.2, 126.1, 127.1, 127.3, 127.8, 129.0, 129.1, 129.7, 129.74, 130.9, 135.3, 143.7, 144.0, 151.4, 154.7, 166.0, 166.2. MALDI-MS (M + H)+ for C₄₂H₅₀N₄O₇Si: expected 751.3528, found 751. 3505.

2-Amino-3-benzyl-5-(2-((tert-butyldimethylsilyl)oxy)ethyl)-7-((2S,4S,5R)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-3,5-dihydro-4H-pyrrolo[3,2-d]pyrimidin-4-one (19). To compound 18 (0.42 g, 0.56 mmol) in a mixture of methanol (10.0 mL) and water (1.0 mL) was added sodium hydroxide (0.05 M). The reaction mixture was stirred under room temperature for 1 h. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with methanol/ethyl acetate 1:50 to 1:20) to give 19 as a white solid (0.21 g, 74%). mp 57-58 °C. ¹H NMR (500 MHz, CD₃OD) δ ppm -0.09 (d, J = 10.0 Hz, 6H), 0.82 (s, 9H), 2.02 (dd, J = 11.0, 5.0 Hz, 1H), 2.51–2.56 (m, 1H), 3.68 (d, J = 12.0 Hz, 1H), 3.87– 3.90 (m, 2H), 4.08 (s, 1H), 4.31 (dt, J = 14.0, 5.0 Hz, 1H), 4.53-4.58 (m, 2H), 5.13 (d, J = 16.0 Hz, 1H), 5.26 (dd, J = 11.5, 5.5 Hz, 1H), 5.40 (d, J = 16.0 Hz, 1H), 7.06 (s, 1H), 7.22–7.35 (m, 5H). ¹³C NMR (125 MHz, CD₃OD) δ ppm -4.5, 19.7, 27.2, 44.4, 45.7, 52.6, 64.9, 65.5, 76.7, 89.9, 114.3, 114.7, 128.2, 129.2, 130.4, 134.0, 137.7, 144.9, 152.9, 156.6. ESI-MS $(M + H)^+$ for C₂₆H₃₈N₄O₅Si: expected 515.2690, found 515.2706.

2-Amino-5-(2-((tert-butyldimethylsilyl)oxy)ethyl)-7-((2S,4S,5R)-4hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-3,5-dihydro-4Hpyrrolo[3,2-d]pyrimidin-4-one (3b). To compound 19 (0.21 g, 0.41 mmol) and 10% palladium on carbon (0.11 g) in methanol (5.0 mL) was added ammonium formate (0.28 g, 4.5 mmol) under Ar. The reaction mixture was heated under 75 °C for 16 h and then filtered through Celite. The filtrate was concentrate and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with methanol/ethyl acetate 1:20 to 1:5) to give 3b as a white gummy solid (0.075 g, 43%). mp 49–50 °C. ¹H NMR (600 MHz, d_6 -DMSO) δ ppm –0.13 (s, 6H), 0.79 (s, 9H), 1.88 (dd, J = 12.2, 5.4 Hz, 1H), 2.06-2.11 (m, 1H), 3.42 (s, 2H), 3.72 (s, 1H), 3.81 (t, J = 5.4 Hz, 2H), 4.17 (s, 1H), 4.25 (t, J = 4.8 Hz, 2H), 4.90 (s, 1H), 5.07 (dd, J = 12.0, 5.4 Hz, 1H), 5.71 (s, 1H), 7.12 (s, 1H), 10.78 (s, 1H). ¹³C NMR (150 MHz, d_6 -DMSO) δ ppm -4.7, 18.8, 26.7, 42.8, 51.1, 63.9, 73.4, 74.0, 88.4, 112.8, 114.5, 130.9, 145.8, 151.6, 155.6. ESI-MS $(M + H)^+$ for $C_{19}H_{32}N_4O_5Si$: expected 425.2221, found 425.2260.

N'-(5-Allyl-3-benzyl-4-oxo-4,5-dihydro-3*H*-pyrrolo[3,2-*d*]pyrimidin-2-yl)-*N*,*N*-dimethylformimidamide (**20**). To a suspension of compound **6** (2.0 g, 6.8 mmol) in dry THF (25.0 mL) was added sodium hydride (0.28 g, 6.7 mmol, 60% in mineral oil). The mixture was stirred for 15 min before allyl bromide (330.0 μ L, 8.0 mmol) was added. The mixture was stirred at room temperature for 16 h. After removal of solvent, the reaction was diluted with methylene chloride and washed successively with water and brine. The organic layer was separated, dried over MgSO₄, concentrated, and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with hexane/ethyl acetate 5:1 to 3:1) to give compound **20** as a light yellow solid (1.4 g, 64%). mp 93–94 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 3.06 (s, 3H), 3.15 (s, 3H), 5.06–5.09 (m, 3H), 5.19 (d, *J* = 10.5 Hz, 1H), 5.52 (s, 2H), 6.05 (ddt, *J* = 16.5, 10.5, 5.0 Hz, 1H), 6.31 (s, 1H), 7.03 (s, 1H), 7.19–7.21 (m, 1H), 7.27 (t, *J* = 7.5 Hz, 2H), 7.37 (d, *J* = 7.5 Hz, 2H), 8.61 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 35.0, 40.8, 45.1, 50.4, 101.3, 114.3, 116.9, 126.7, 127.9, 128.1, 130.4, 134.8, 139.0, 144.4, 154.3, 155.8, 156.2. ESI-MS (M + H)⁺ for C₁₉H₂₁N₅O: expected 336.1824, found 336.1832.

5-Allyl-2-amino-3-benzyl-3,5-dihydro-4*H*-pyrrolo[3,2-*d*]pyrimidin-4-one (**21**). To compound **20** (1.0 g, 3 mmol) in a mixture of methanol (20.0 mL) and water (5.0 mL) was added sodium hydroxide (1.0 M). The reaction mixture was heated under 70 °C for 16 h. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with ethyl acetate) to give **21** as a white solid (0.58 g, 70%). mp 121–122 °C. ¹H NMR (500 MHz, CD₃OD) δ ppm 4.99–5.05 (m, 3H), 5.16 (dd, *J* = 10.5, 1.5 Hz, 1H), 5.33 (s, 2H), 6.04–6.12 (m, 2H), 7.21–7.30 (m, 4H), 7.35 (t, *J* = 7.5 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 44.5, 50.5, 101.3, 112.3, 117.0, 126.5, 127.9, 129.1, 131.1, 134.6, 135.7, 144.9, 151.1, 154.8. ESI-MS (M + H)⁺ for C₁₆H₁₆N₄O: expected 281.1403, found 281.1387.

(2R,3S,5R)-5-(5-Allyl-2-amino-3-benzyl-4-oxo-4,5-dihydro-3Hpyrrolo[3,2-*d*]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (22). To compound 21 (0.58 g, 2.1 mmol) and 1-(α , β)-O-methyl-3,5-di(O-p-toluoyl)-2-deoxy-D-ribose (2.3 g, 3.1 mmol) in a mixture of methylene chloride (5.0 mL) and acetonitrile (5.0 mL) was added a solution of SnCl₄ (4.2 mL, 4.2 mmol, 1 M in CH₂Cl₂). The reaction mixture was heated at 65 °C for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with sat. NaHCO3 and brine. The organic layer was separated, dried over MgSO₄, concentrated, and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 2:1 to 1:2) to give compound 22 as a light yellow solid (0.30 g, 23%). mp 59–60 °C. ¹H NMR (600 MHz, CDCl₃) δ ppm 2.40 (s, 3H), 2.41 (s, 3H), 2.56–2.59 (m, 2H), 4.47 (dt, J = 4.2, 1.8 Hz, 1H), 4.57 (dd, J = 12.0, 4.2 Hz, 1H), 4.76 (dd, J = 12.0, 4.2 Hz, 1H), 4.94-4.96 (m, 2H), 5.09 (d, J = 10.4 Hz, 1H), 5.17 (d, J = 10.4 Hz, 1H), 5.29 (s, 2H), 5.48 (t, J = 8.4 Hz, 1H), 5.64-5.66 (m, 1H), 5.95-6.02 (m, 1H), 7.04 (s, 1H), 7.23–7.29 (m, 7H), 7.31–7.34 (m, 2H), 7.94 (d, J = 8.4 Hz, 2H), 7.97 (d, J = 8.4 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ ppm 21.75, 21.8, 38.6, 44.5, 50.6, 64.8, 73.2, 77.4, 82.5, 112.8, 113.1, 117.6, 126.4, 127.2, 127.3, 128.0, 129.0, 129.2, 129.23, 129.8, 134.2, 135.4, 143.8, 144.1, 151.3, 154.7, 166.2, 166.4. ESI-MS $(M + H)^+$ for $C_{37}H_{36}N_4O_6$: expected 633.2714, found 633.2745.

(2R,3S,5R)-5-(2-Amino-3-benzyl-5-(2,3-dihydroxypropyl)-4-oxo-4,5-dihydro-3H-pyrrolo[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3-yl 4-Methylbenzoate (23). To the suspension of compound 22 (0.30 g, 0.48 mmol) in a mixture of acetone (4.0 mL) and water (1.0 mL) were added TBHP (100 μ L, 0.5– 0.6 mmol, 5.0-6.0 M in decane), TBAF (0.012 g, 0.048 mmol), and OsO_4 (trace). The reaction was stirred at room temperature for 16 h. The reaction mixture was diluted with methylene chloride and washed successively with water and brine. The organic layer was separated, dried over MgSO₄, and concentrated. The solution was purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 1:1 to 1:3) to give compound 23 as a light yellow solid (0.13 g, 40%). mp 61–62 °C. ¹H NMR (500 MHz, CDCl₃) δ ppm 2.36 (s, 3H), 2.39 (s, 3H), 2.50-2.58 (m, 2H), 3.46 (dd, J = 12.0, 5.5 Hz, 1H), 3.52 (ddd, J = 11.5, 4.5, 1.5 Hz, 1H), 3.88-3.93 (m, 1H), 4.18 (dt, J = 14.0, 7.0 Hz, 1H), 4.40 (dt, J = 5.0, 2.0 Hz, 1H), 4.44 (dd, J = 14.0, 3.5 Hz, 1H), 4.54 (ddd, J = 12.0, 5.0, 2.0 Hz, 1H), 4.67 (dt, J = 12.0, 5.4 Hz, 1H), 5.26 (s, 2H), 5.43 (dd, J = 10.0, 6.0 Hz, 1H), 5.59-5.60 (m, 1H), 7.19–7.28 (m, 10H), 7.89 (d, J = 8.5 Hz, 2H), 7.94 (d, J = 8.0 Hz, 2H). ¹³C NMR for both diastereomers (125 MHz, CDCl₃) δ ppm 22.5, 22.56, 46.0, 52.79, 52.8, 65.40, 65.44, 66.7, 73.8, 73.85, 75.5, 79.5, 84.6, 114.1, 114.3, 128.3, 129.0, 129.1, 129.40, 130.6, 131.1, 131.5, 131.53,

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133.7, 137.7, 146.2, 146.3, 153.4, 156.9, 168.4, 168.6. ESI-MS $(M + H)^+$ for $C_{37}H_{38}N_4O_8$: expected 667.2769, found 667.2786.

(2R,3S,5R)-5-(2-Amino-3-benzyl-5-(2,3-bis((tertbutyldimethylsilyl)oxy)propyl)-4-oxo-4,5-dihydro-3H-pyrrolo[3,2-d]pyrimidin-7-yl)-2-(((4-methylbenzoyl)oxy)methyl)tetrahydrofuran-3yl 4-Methylbenzoate (24). To compound 23 (0.13 g, 0.19 mmol) in DMF were added imidazole (0.08 g, 1.1 mmol), 4-DMAP (0.002 g, 0.018 mmol), and TBSCl (0.17 g, 1.1 mmol). The reaction was stirred under room temperature for 16 h. After removal of the solvent, the mixture was diluted by methylene chloride and washed with water and brine. The solution was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with hexane/ethyl acetate 5:1 to 3:1) to give compound 24 as a white gummy solid (0.13 g, 76%). mp 44-45 °C. ¹H NMR (600 MHz, CDCl₃) δ ppm -0.31 (s, 3H), -0.07 (d, J = 10.2 Hz, 3H), 0.07 (s, 6H), 0.78 (d, J = 2.4 Hz, 9H), 0.91 (s, 9H), 2.39 (s, 3H), 2.40 (s, 4H), 2.42-2.44 (m, 1H), 3.52-3.55 (m, 1H), 3.61 (dd, J = 10.2, 3.6 Hz, 1H), 3.99-4.14 (m, 2H), 4.50-4.52 (m, 1H), 4.60–4.64 (m, 2H), 4.77 (dd, J = 11.4, 4.8 Hz, 1H), 5.39–5.43 (m, 3H), 5.61–5.63 (m, 1H), 7.11 (s, 1H), 7.18–7.20 (m, 2H), 7.24– 7.27 (m, 4H), 7.32–7.35 (m, 5H), 7.91 (d, J = 7.8 Hz, 2H), 8.02 (d, J = 7.8 Hz, 2H). ¹³C NMR for both diastereomers (150 MHz, CDCl₃) δ ppm -5.1, -5.0, -4.5, 18.2, 18.6, 21.9, 22.0, 26.1, 26.2, 29.9, 38.5, 44.6, 52.3, 52.6, 65.2, 65.8, 72.9, 73.0, 73.6, 82.6, 112.2, 112.3, 126.7, 126.8, 127.4, 127.5, 128.3, 129.4, 129.5, 130.0, 130.1, 131.4, 131.6, 135.5, 143.9, 144.2, 150.9, 151.2, 166.4, 166.6. MALDI-MS (M + H)⁺ for C49H66N4O8Si2: expected 895.4498, found 895.4525.

2-Amino-5-(2,3-bis((tert-butyldimethylsilyl)oxy)propyl)-7-((2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)-3,5dihydro-4H-pyrrolo[3,2-d]pyrimidin-4-one (3d). To compound 24 (0.13 g, 0.14 mmol) in a mixture of methanol (5 mL) and water (0.5 mL) was added sodium hydroxide (0.1 M). The reaction mixture was stirred under room temperature for 1 h. The reaction mixture was concentrated and purified by column chromatography (SiO₂, 0.06-0.20 mm, eluting with methanol/ethyl acetate 1:50 to 1:20) to give 25 (0.065 g, 68%). To compound 25 (0.065 g, 0.10 mmol) and 10% palladium on carbon (0.030 g) in methanol (3.0 mL) was added ammonium formate (0.063 g, 1.0 mmol) under Ar. The reaction mixture was heated under 75 °C for 16 h and then filtered through Celite. The filtrate was concentrated and purified by column chromatography (SiO₂, 0.06–0.20 mm, eluting with methanol/ethyl acetate 1:20 to 1:5) to give 3d as a white gummy solid (0.031 g, 56%). mp 53-54 °C. ¹H NMR (600 MHz, d_6 -DMSO) δ ppm -0.28 (d, J = 6.0 Hz, 3H), -0.08 (s, 3H), 0.05 (s, 6H), 0.79 (s, 9H), 0.89 (s, 9H), 1.88-1.91 (m, 1H), 2.02–2.11 (m, 1H), 3.46–3.50 (m, 2H), 3.70 (d, J = 15.6 Hz, 1H), 3.97–4.03 (m, 2H), 4.18 (d, J = 12.0 Hz, 1H), 4.34 (d, J = 10.2 Hz, 1H), 4.90 (s, 1H), 5.06–5.10 (m, 1H), 5.69 (d, J = 20.0 Hz, 1H), 7.02 (s, 1H), 10.50 (d, J = 15.2 Hz, 1H). ¹³C NMR (150 MHz, d_6 -DMSO) δ ppm -4.5, -3.9, 18.6, 19.0, 26.7, 49.6, 52.1, 52.2, 63.9, 64.0, 66.3, 73.6, 73.9, 74.2, 88.3, 113.2, 114.8, 131.2, 145.8, 151.4, 155.3. ESI-MS (M + H)⁺ for C₂₆H₄₈N₄O₆Si₂: expected 569.3191, found 569.3234.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b02110.

¹H and ¹³C NMR spectra (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

- (1) Gates, K. S. Chem. Res. Toxicol. 2009, 22, 1747.
- (2) Hecht, S. S. Chem. Res. Toxicol. 2008, 21, 160.
- (3) Melnick, R. L. Ann. N. Y. Acad. Sci. 2002, 982, 177.
- (4) Peterson, L. A. Chem. Res. Toxicol. 2013, 26, 6.
- (5) Pratt, M. M.; John, K.; MacLean, A. B.; Afework, S.; Phillips, D. H.; Poirier, M. C. Int. J. Environ. Res. Public Health **2011**, *8*, 2675.
- (6) Shrivastav, N.; Li, D.; Essigmann, J. M. Carcinogenesis 2010, 31, 59.
 (7) Smela, M. E.; Currier, S. S.; Bailey, E. A.; Essigmann, J. M. Carcinogenesis 2001, 22, 535.
- (8) Eoff, R. L.; Choi, J.-Y.; Guengerich, F. P. J. Nucleic Acids 2010, 830473.
- (9) Zhang, H.; Eoff, R. L.; Kozekov, I. D.; Rizzo, C. J.; Egli, M.; Guengerich, F. P. J. Biol. Chem. 2009, 284, 3563.
- (10) Tang, M.; Pham, P.; Shen, X.; Taylor, J.-S.; O'Donnell, M.; Woodgate, R.; Goodman, M. F. *Nature* **2000**, *404*, 1014.
- (11) Boudsocq, F.; Iwai, S.; Hanaoka, F.; Woodgate, R. *Nucl. Acids. Res.* **2001**, *29*, 4607.
- (12) Haracska, L.; Washington, M. T.; Prakash, S.; Prakash, L. J. Biol. Chem. 2001, 276, 6861.
- (13) Huang, H.; Greenberg, M. M. J. Am. Chem. Soc. 2008, 130, 6080.
- (14) Asagoshi, K.; Terato, H.; Ohyama, Y.; Ide, H. J. Biol. Chem. 2002, 277, 14589.

(15) Christov, P. P.; Yamanaka, K.; Choi, J.-Y.; Takata, K.-i.; Wood, R. D.; Guengerich, F. P.; Lloyd, R. S.; Rizzo, C. J. *Chem. Res. Toxicol.* **2012**, 25, 1652.

(16) Philippin, G.; Cadet, J.; Gasparutto, D.; Mazon, G.; Fuchs, R. P. DNA Repair **2014**, *22*, 133.

(17) Giri, I.; Johnston, D. S.; Stone, M. P. Biochemistry 2002, 41, 5462.
(18) Rana, J.; Huang, H. Bioorg. Med. Chem. 2014, 22, 2825; Bioorg. Med. Chem. 2014, 22, 2825.

(19) Tompkins, E. M.; McLuckie, K. I.; Jones, D. J.; Farmer, P. B.; Brown, K. Mutat. Res., Genet. Toxicol. Environ. Mutagen. 2009, 678, 129.
(20) Scherer, E.; Der Van Laken, C.; Gwinner, L.; Laib, R.; Emmelot, P. Carcinogenesis 1981, 2, 671.

(21) Scherer, E.; Winterwerp, H.; Emmelot, P. IARC Sci. Publ. 1985, 109.

- (22) Solomon, J. J.; Segal, A. Environ. Health Perspect. 1989, 81, 19.
- (23) Gibson, E. S.; Lesiak, K.; Watanabe, K. A.; Gudas, L. J.; Pankiewicz, K. W. Nucleosides Nucleotides 1999, 18, 363.
- (24) Vu, H.; McCollum, C.; Jacobson, K.; Theisen, P.; Vinayak, R.; Spiess, E.; Andrus, A. *Tetrahedron Lett.* **1990**, *31*, 7269.
- (25) Seela, F.; Shaikh, K. I.; Budow, S.; Jawalekar, A. M. Nucleosides, Nucleotides Nucleic Acids 2005, 24, 851.

(26) Hamm, M. L.; Parker, A. J.; Steele, T. W.; Carman, J. L.; Parish, C. A. J. Org. Chem. **2010**, *75*, 5661.

- (27) Huang, H.; Greenberg, M. M. J. Org. Chem. 2008, 73, 2695.
- (28) Shishkina, I. G.; Johnson, F. Chem. Res. Toxicol. 2000, 13, 907.
- (29) Kim, J.; Weledji, Y. N.; Greenberg, M. M. J. Org. Chem. 2004, 69, 6100.
- (30) Hoffer, M. Chem. Ber. 1960, 93, 2777.
- (31) Pankiewicz, K.; Matsuda, A.; Watanabe, K. A. J. Org. Chem. 1982, 47, 485.
- (32) Voegel, J. J.; Benner, S. A. Helv. Chim. Acta 1996, 79, 1863.
- (33) Jiang, Y. L.; Stivers, J. T. Tetrahedron Lett. 2003, 44, 85.

(34) Wellington, K. W.; Benner, S. A. Nucleosides, Nucleotides Nucleic Acids 2006, 25, 1309.