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Chiral zinc(II) and copper(II)-catalyzed asymmetric ring-opening reactions of *meso*-epoxides with aniline and indole derivatives

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ABSTRACT

The ring-opening reactions of *meso*-epoxides with aniline and indole derivatives proceeded smoothly in water in the presence of Zn(II) and Cu(II) surfactant-type catalysts to afford the corresponding products in moderate to high yields with good to excellent enantioselectivities. Opposite enantiomers were obtained by using Sc(III) and Zn(II) or Cu(II) with the same chiral ligand. Crystal structures of these catalysts may explain the reversal of the enantioselectivity. Some reactions were also tested in dichloromethane (DCM), and it was revealed that the reactions proceeded faster in water than in DCM. Finally, several non-linear effect experiments suggested unique structure of these chiral catalysts.

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1. Introduction

Most organic reactions have been carried out in organic solvents because most organic materials are not soluble in water and most reactive species are not stable in the presence of water. This is also the case for chiral catalysts, which are often unstable in the presence of water and thus are difficult to conduct catalytic asymmetric reactions in the presence of water. On the other hand, from an environmentally point of view, recent demand is development of benign chemical processes.¹ In this context, while several types of reactions in aqueous media have been reported,² organic solvents are required as co-solvents to dissolve organic materials in many cases. To overcome this problem, we developed Lewis acid-surfactant-combined catalysts (LASCs) to conduct organic reactions in water as the sole solvent. Hydrophobic substrates are taken into micellous emulsions created by LASCs and the hydrophobic substrates, where reactants, substrates, and the catalyst are concentrated, and the desired reactions proceed very efficiently.³ Very recently, we have also shown that even hydrophilic substrates can be used in this system.^{3e}

Catalytic asymmetric ring-opening reactions of *meso*-epoxides provide a useful method for the synthesis of chiral 1,2-bifunctional alcohols. Based on this method, a number of protocols have been developed for the synthesis of enantiomerically enriched 1,2-azido alcohols,⁴ 1,2-amino alcohols,⁵ 1,2-diol derivatives,⁶ 1,2-cyano alcohols,⁷ 1,2-mercaptoalcohols,⁸ 1,2-halohydrines,⁹ and 1,2-seleno

alcohols.¹⁰ Our group has mainly contributed to this field by exploiting efficient asymmetric reactions in water, and reported the first examples of chiral scandium and bismuth-catalyzed ringopening reactions of *meso*-epoxides with aniline derivatives in water. We have also developed the first catalytic asymmetric ringopening reactions of *meso*-epoxides with indole derivatives in water. In the course of our investigations to develop more efficient catalysts in water, we have discovered unique catalysis of zinc(II) and copper(II). We describe zinc(II) and copper(II) catalyzed asymmetric ring-opening reactions of *meso*-epoxides with aniline and indole derivatives in water.

2. Results and discussion

At the outset, we carried out the ring-opening reaction of *meso*epoxide **1** with aniline in the presence of various metal surfactantcombined catalysts in water (Table 1). Sc(III), Zn(II), and Cu(II) catalysts gave the desired product **4a** in good to excellent yields with good to excellent enantioselectivities (entries 1, 7, 8). It is noted that opposite enantiomers were obtained by using Sc(III) and Zn(II) or Cu(II) with the same chiral ligand; while Sc(III) gave one enantiomer of the product, Zn(II) or Cu(II) gave the opposite enantiomer of the product. On the other hand, the reactions also proceeded in dichloromethane (DCM), and the Sc(III) and Zn(II) catalysts gave the desired product in 85% and 60% yields with 93% and 90% ees, respectively, although Cu(II) catalyst was not so effective.

We then carefully examined the reaction of *cis*-stilbene oxide (1) with aniline in the presence of 5 mol% of the Zn(II) or Cu(II) catalysts and 6 mol% of chiral bipyridine **3** in water and DCM at room





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Ring-opening reaction of meso-epoxide 1 with a	niline

		M(OTf) _n (DCM)	
		M[O ₃ S(CH ₂) ₁₀ CH ₃] _n (H ₂ O)	
		(10 mol%)	
Ph		(<i>S</i> , <i>S</i>)- 3 (12 mol%)	Ph、_OH
$\sum 0$	+ PhNH ₂		× Ť
Ph	E.	H ₂ O or DCM, 0.1 M, rt, 22 h	Ph ^{_/} "NHPh
1	2a (1.5 equi	iv)	4a

Entry	М	n	In H ₂ O		In DCM		
			Yield ^a (%)	ee ^b (%)	Yield ^a (%) 1	ee ^b (%)	
1	Sc	3	87	95 (1 <i>S</i> ,2 <i>S</i>)	85	93 (1 <i>S</i> ,2 <i>S</i>)	
2	Y	3	9	62 (1S,2S)	_	_	
3	Yb	3	27	82 (1 <i>S</i> ,2 <i>S</i>)	_	_	
4	In	3	6	32 (1 <i>S</i> ,2 <i>S</i>)	_	_	
5	Mn	2	6	81 (1 <i>S</i> ,2 <i>S</i>)	_	_	
6	Ni	2	Trace	_	_	_	
7	Cu	2	82	80 (1R,2R)	18	80 (1R,2R)	
8	Zn	2	97	92 (1R,2R)	60	90 (1R,2R)	
9	Ag	1	Trace	_	_	—	

^a Isolated yield.

^b ee was determined by chiral HPLC analysis.

temperature. It is noted that $Zn[O_3S(CH_2)_{10}CH_3]_2$ or $Cu[O_3S(CH_2)_{10}CH_3]_2$ and **3** in water gave higher yields than $Zn(OTf)_2$ or $Cu(OTf)_2$ and **3** in DCM. In addition, $Zn[O_3S(CH_2)_{10}CH_3]_2$ or $Cu[O_3S(CH_2)_{10}CH_3]_2$ and **3** in water showed higher reactivity than $Zn(OTf)_2$ or $Cu(OTf)_2$ and **3** in DCM (Fig. 1).

After optimization of the reaction conditions, we surveyed substrate scope of several *meso*-epoxides and aniline derivatives using Zn(II) or Cu(II) as a catalyst (Table 2). While stilbene oxide derivatives showed good selectivities with various aniline derivatives (entries 1–8, 13–20), alkyl epoxide derivatives gave only moderate selectivities (entries 9–12, 21). It is interesting to find that Cu(OTf)₂ had almost no activity in DCM, but that Zn(OTf)₂ catalyzed the *meso*-epoxides ring-opening reactions in DCM (entries 22–29). The reactions proceeded slower in DCM than in water, while the desired products were obtained with good selectivities.

Next, we carried out the ring-opening reactions of *meso*-epoxides with indole derivatives using Sc(III), Zn(II), and Cu(II) catalysts in water and DCM (Table 3). Nitrogen-containing heterocycles and their derivatives are well-known structures in natural products and medicines.^{11,12} The synthesis of chiral *N*-heteroaromatic derivatives in optically active form is especially important in this area. Gratifyingly, Sc(III) and Cu(II) gave the product in good yields with excellent enantioselectivities in water. In this reaction, enantiofacial selectivities between Sc(III) and Cu(II) were reversed, and the reaction proceeded sluggishly in the presence of Zn(II). On the other hand, when the reaction was carried out in DCM, only Cu(OTf)₂ gave the desired product in moderate vield with high enantioselectivity. We then examined several reaction conditions with the Cu(II) catalyst in water (Table 4). As for the equivalents of indole, 1.2 equiv gave the best yield and selectivity (entry 2). The concentration was found to give almost no influence on yield and selectivity (entries 2, 4-6). The yield dropped when the catalyst loading decreased (entries 2, 7-9). When 2 mol% of the catalyst was used, the desired product was obtained in 73% yield with 90% ee.

The reaction of *cis*-stilbene oxide (1) with indole (**5a**) also proceeded in the presence of 5 mol % of Cu(OTf)₂ catalysts and 6 mol % of chiral bipyridine **3** in DCM. While the reaction rate in DCM was almost the same as that in water (Fig. 2), the enantioselectivity was higher in water than in DCM.

Several examples of *meso*-epoxide ring-opening reactions with indole derivatives are shown in Table 5. In cases of stilbene oxide (**1**) with substituted indoles, the reactions proceeded smoothly in most cases to afford the corresponding indole adducts in moderate to good yields with excellent enantioselectivities (entries 3–6). As for *meso*-epoxides, aromatic ones reacted with indole (**5a**) to afford the desired compounds in moderate yields with good selectivities (entries 7–9). However, alkyl epoxide derivatives showed poor reactivity (entries 10, 11). On the other hand, when the reactions were conducted in DCM, the rates were slower than those in water, and the reaction mixtures were messy. In some cases, only a trace amount of the desired product was obtained (entries 15, 17, and 18).

To obtain information on the chiral Cu complex, X-ray crystal structural analysis was performed. Single crystals suitable for X-ray analysis were obtained from a (S,S)-**3**–CuBr₂ complex (Fig. 3). The complex adopts square pyramidal structure in which two pyridines and one hydroxyl group of **3** coordinated to copper in a tridentate manner. Formation of this structure may be a key for obtaining high enantioselectivity. In addition, difference of the structure between chiral scandium^{2c} and copper may cause the difference of absolute configurations of the products.



Figure 1. Plot of yield versus time for the ring-opening reactions of *cis*-stilbene oxide (1) with aniline (2a) in the presence of Zn(II) catalyst (1–a) or Cu(II) catalyst (1–b) and 3 in water (pink) and DCM (blue).

Table 2

Ring-opening reaction of meso-epoxides with aniline derivatives

R		M(O ₃ SC ₁₂ H ₂₅) ₂ (5 mol%) (<i>S</i> , <i>S</i>)- 3 (6 mol%)	Ph_OH
R	+ NHRR'	H ₂ O or DCM, conc., rt, 22 h	Ph ''NRR'
1	2 (1.5 equ	iv)	4

Entry	R	NHRR′	Metal	Solvent	Conc (M)	Yield (%) ^a		ee (%) ^b
1	Ph	Aniline	Zn	H ₂ O	0.1	4a	97	92
2	Ph	2-Trifluoromethylaniline	Zn	H ₂ O	1.0	4b	100	92
3	Ph	1-Naphthylamine	Zn	H ₂ O	1.0	4c	55	95
4	Ph	N-Methylaniline	Zn	H ₂ O	1.0	4d	56	95
5	Ph	2-Methoxyaniline	Zn	H ₂ O	1.0	4e	43	91
6	3-MePh	Aniline	Zn	H ₂ O	1.0	4f	95	93
7	4-MePh	Aniline	Zn	H ₂ O	1.0	4g	82	93
8	4-BrPh	Aniline	Zn	H ₂ O	1.0	4h	44	90
9	Me	Aniline	Zn	H ₂ O	1.0	4i	80	54
10	Me	2-Naphthylamine	Zn	H ₂ O	1.0	4j	100	67
11 ^c	-(CH ₂) ₄ -	Aniline	Zn	H ₂ O	0.5	4k	94	66
12 ^c	-(CH ₂) ₄ -	2-Naphthylamine	Zn	H ₂ O	0.5	41	77	79
13	Ph	Aniline	Cu	H_2O	1.0	4a	82	80
14	Ph	2-Trifluoromethylaniline	Cu	H_2O	1.0	4b	100	78
15	Ph	1-Naphthylamine	Cu	H ₂ O	1.0	4c	56	75
16	Ph	N-Methylaniline	Cu	H ₂ O	1.0	4d	88	91
17	Ph	2-Methoxyaniline	Cu	H ₂ O	1.0	4e	44	70
18	4-MePh	N-Methylaniline	Cu	H ₂ O	1.0	4m	78	90
19	4-BrPh	N-Methylaniline	Cu	H ₂ O	1.0	4n	72	92
20	2-Naphthyl	N-Methylaniline	Cu	H_2O	1.0	40	81	90
21	-(CH ₂) ₄ -	2-Naphthylamine	Cu	H ₂ O	0.5	41	84	44
22	Ph	Aniline	Zn	DCM	0.1	4a	60	90
23	Ph	2-Trifluoromethylaniline	Zn	DCM	0.1	4b	92	90
24	Ph	1-Naphthylamine	Zn	DCM	0.1	4c	69	87
25	Ph	N-Methylaniline	Zn	DCM	0.1	4d	63	85
26	Ph	2-Methoxyaniline	Zn	DCM	0.1	4e	10	69
27	4-BrPh	Aniline	Zn	DCM	0.1	4h	20	86
28	-(CH ₂) ₄ -	Aniline	Zn	DCM	0.1	4k	77	74
29	Me	Aniline	Zn	DCM	0.1	4i	74	70

^a Isolated yield.

^b ee was determined by chiral HPLC analysis.

^c Reaction was conducted at 10 °C.

We also conducted the reaction with Zn(II) and chiral bipyridine derivatives **7** and **8** (Table 6). Judging from these results, both hydroxyl groups are necessary for reactivity and selectivity. One hydroxyl group may be needed to coordinate to zinc and the other needed to coordinate to a substrate.

Finally, we examined non-linear effect (NLE) of these catalysts (Figs. 4 and 5). In Cu-catalyzed reactions, NLE was not observed both in water and in DCM. On the other hand, in Sc-catalyzed reactions, while NLE was not observed in water, positive NLE was appeared in DCM. These results suggest that monomeric structures

Table 3

Sc(III), Zn(II), and Cu(II)-catalyzed ring opening of meso-epoxide ${\bf 1}$ with indole ${\bf 5a}$



Entry	Metal	п	In H ₂ O		In DCM	
			Yield ^a (%)	ee ^b (%)	Yield ^a (%)	ee (%) ^b
1	Sc	3	58	92 (1R,2R)	Trace ^c	_
2	Zn	2	8	80 (1 <i>S</i> ,2 <i>S</i>)	Trace ^c	_
3	Cu	2	80	96 (1 <i>S</i> ,2 <i>S</i>)	60	86 (1 <i>S</i> ,2 <i>S</i>)

^a Isolated yield.

^b ee was determined by chiral HPLC analysis.

^c By-product was generated.

of Cu catalysts both in water and DCM and Sc catalysts in water are predominant. Interestingly, when Sc–(S,S)-**3** and Sc–(R,R)-**3** complexes were prepared independently and combined appropriately to make catalysts with low ees in DCM, NLE was not observed (Fig. 6). It was assumed from these experiments that the Sc(III)-**3** complex would not dissociate after formation of the complex in DCM. On the other hand, formation of dimeric or oligomeric structure would be suppressed in water.

Table 4

Effect of concentrations and catalyst loadings



Entry	Indole (equiv)	Cone (M)	x (mol %)	Yield ^a (%)	ee ^b (%)
1	1.1	1.0	10	80	88
2	1.2	1.0	10	80	96
3	2.0	1.0	10	80	88
4	1.2	0.50	10	80	93
5	1.2	0.17	10	79	93
6	1.2	0.10	10	75	93
7	1.2	1.0	5	71	96
8	1.2	1.0	2	54 (73) ^c	92 (90)
9	1.2	1.0	1	48	86

^a Isolated yield.

^b ee was determined by chiral HPLC analysis.

^c Reaction time was 43 h.



Figure 2. Plot of yield versus time for the ring-opening reactions of *cis*-stilbene oxide (1) with indole (**5a**) in the presence of Cu(II) catalyst and **3** in water (pink) and DCM (blue).

Table 5

Ring opening of meso-epoxides with indole derivatives

3. Conclusion

We have developed the ring-opening reaction of *meso*-epoxide with aniline and indole derivatives using surfactant-type chiral zinc and copper catalysts, and the desired ring-opening products were obtained in moderate to high yields with good to excellent enantioselectivities. Opposite enantiomers were obtained by using Sc(III) and Zn(II) or Cu(II) with the same chiral ligand; while Sc(III) gave one enantiomer of the product, Zn(II) or Cu(II) gave the opposite enantiomer of the product. Crystal structures of Sc(III) and Cu(II) catalysts may explain the reversal of the selectivity. Some reactions were tested not only in water but also in dichloromethane (DCM), and it was revealed that the reactions proceeded faster in water than in DCM. Catalytic activity of Zn(II) and Cu(II) as well as Sc(III) is remarkable for the reactions of *meso*-epoxides with anilines and indoles. Finally, several non-linear effect experiments suggested unique structure of these chiral catalysts.



Entry	R	R ¹	R ²	Solvent	Concn (M)	Yield ^a (%)		ee ^b (%)
1	Ph	Н	Н	H ₂ O	1.0	6a	80	96 (1 <i>S</i> ,2 <i>S</i>)
2 ^c	Ph	Н	Н	H ₂ O	1.0	6a	71	96 (1 <i>S</i> ,2 <i>S</i>)
3	Ph	Н	OMe	H ₂ O	1.0	6b	45	92 (1 <i>S</i> ,2 <i>S</i>)
4	Ph	Н	Me	H ₂ O	1.0	6c	81	92 (1 <i>S</i> ,2 <i>S</i>)
5	Ph	Н	Br	H ₂ O	1.0	6d	58	90 (1 <i>S</i> ,2 <i>S</i>)
6	Ph	Me	Н	H ₂ O	0.1	6e	61	95 (1 <i>S</i> ,2 <i>S</i>)
7	2-Naphthyl	Н	Н	H ₂ O	0.1	6f	43	87
8	4-MePh	Н	Н	H ₂ O	0.1	6g	43	84 (1 <i>S</i> ,2 <i>S</i>)
9	4-Br-Ph	Н	Н	H ₂ O	0.5	6h	53	92 (1 <i>S</i> ,2 <i>S</i>)
10	Me	Н	Н	H ₂ O	0.1	6i	Trace	_
11	-(CH ₂) ₄ -	Н	Н	H ₂ O	0.1	6j	Trace	_
12	Ph	Н	Н	DCM	0.5	6a	60	92 (1 <i>S</i> ,2 <i>S</i>)
13	Ph	Н	OMe	DCM	0.5	6b	49	90 (1 <i>S</i> ,2 <i>S</i>)
14	Ph	Н	Me	DCM	0.5	6c	46	47 (1S,2S)
15	Ph	Н	Br	DCM	0.5	6d	Trace	_
16	2-Naphthyl	Н	Н	DCM	0.5	6f	33	70
17	4-MePh	Н	Н	DCM	0.5	6g	Trace	_
18	-(CH ₂) ₄ -	Н	Н	DCM	0.5	6j	Trace	—

^a Isolated yield.

^b ee was determined by chiral HPLC analysis.

^c The reaction was conducted in the presence of 2 mol % of the catalyst.



Figure 3. X-ray structure of $[CuBr_2 \cdot 3]$ (left) and $[ScBr_2 \cdot H_2O \cdot 3]^+$ moiety in the X-ray structure of $[ScBr_2 \cdot H_2O \cdot 3] \cdot Br \cdot H_2O \cdot 2^{-c}$ Hydrogen atoms are omitted for clarity.

Table 6

Ring-opening reactions of meso-epoxide (1) with Zn(II) and chiral bipyridine derivatives



^a Isolated yield.

^b ee was determined by chiral HPLC analysis.



Figure 4. Non-linear effect experiments of ring-opening reaction of *cis*-stilbene (1) with aniline using Cu(II) catalyst in water (light blue) and in DCM (red).



Figure 5. Non-linear effect experiments of ring-opening reaction of *cis*-stilbene with aniline using Sc(III) catalyst in water (light blue) and in DCM (red).



Figure 6. Non-linear effect experiments using pre-mixed catalysts in DCM.

4. Experimental section

4.1. General

IR spectra were measured with JASCO FT/IR-610 spectrometers. Data are represented as frequency of absorption (cm⁻¹). ¹H and ¹³C NMR spectra were recorded on a JEOL JNM-LA 400, LA-500, or LA-600 spectrometer in CDCl₃ unless otherwise noted. Tetramethylsilane (TMS) or chloroform served as internal standard (δ =0 or 7.26) for ¹H NMR and CDCl₃ as internal standard (δ =77.0) for ¹³C NMR. Mass spectra were recorded on Brucker Daltonics BioTOF II spectrometer (ESI). Optical rotations were measured on a JASCO P1010 polarimeter using a 2 mL cell with 1 dm path length. Analytical high performance liquid chromatography (HPLC) was performed on a Shimadzu VP-series instrument equipped with a variant wavelength detector (D lump; 190–600 nm) using Daicel columns (0.46 cm ID×25 cm) as chiral stationary phase. Preparative thin-layer chromatography (TLC) was carried out using Wakogel B-5F. Column chromatography was carried out using Kanto silica gel 60 N (spherical, neutral, 40–100 mesh).

4.2. Materials

Chiral bipyridine ligand **3**, **7**, and **8** were prepared according to the reported procedure.¹³ Aromatic epoxides were prepared by *m*CPBA oxidation of the corresponding *cis*-alkenes according to the reported procedures.^{14,15} $M[O_3S(CH_2)_{10}CH_3]_n$ were prepared by reported procedures using MCl_n and $CH_3(CH_2)_{10}SO_3Na$ in water.^{3b} All other compounds were commercially available and used as received except aniline and substituted anilines, which were distilled from calcium hydride before use.

4.2.1. Characterization of surfactant-type catalysts. Elementary analysis of $M[O_3S(CH_2)_{10}CH_3]_n$:

Sc[O₃S(CH₂)₁₀CH₃]₃·3H₂O (white powder): Calcd C: 51.04, H: 9.64; found C: 51.31, H: 9.40.

Zn[O₃S(CH₂)₁₀CH₃]₂·5H₂O (white powder): calcd C: 44.06, H: 9.24; found C: 44.61, H: 8.97.

Cu[O₃S(CH₂)₁₀CH₃]₂·2H₂O (pale blue powder): calcd C: 48.17, H: 9.10; found C: 47.91, H: 8.93.

4.3. General procedure for *meso*-epoxide-opening reaction with anilines

A mixture of $Zn[O_3S(CH_2)_{10}CH_3]_2$ or $Cu[O_3S(CH_2)_{10}CH_3]_2$ and bipyridine ligand **3** in water was stirred for 1 h at room temperature. *meso*-Epoxide and aniline were then added to the mixture. The mixture was further stirred for 22–48 h at room temperature. The resulting mixture was quenched with satd NaHCO₃ aq and brine. The aqueous layer was extracted with dichloromethane (three times), and the combined organic layers were washed with brine, and dried over Na₂SO₄. After filtration, the solvent was removed under reduced pressure. The residue was purified by preparative TLC (elution *n*-hexane/ethyl acetate=3/1) to give the corresponding amino alcohols.

4.3.1. Physical data of amino alcohols.

4.3.1.1. (1R,2R)-1,2-Diphenyl-2-(phenylamino)-ethanol (4a). ¹H NMR δ 2.75 (br s, 1H), 4.50 (d, J=5.5 Hz, 1H), 4.55 (br s, 1H), 4.85 (d, J=5.5 Hz, 1H), 6.56-6.58 (m, 1H), 6.57-6.58 (m, 1H), 6.83-6.84 (m, 1H), 7.06-7.08 (m, 2H), 7.18-7.28 (m, 10H); ¹³C NMR δ 64.4, 77.9, 110.6, 114.1, 116.6, 118.1, 126.5, 127.2, 127.7, 128.1, 128.4, 128.7, 129.5, 129.7, 139.7, 140.5, 147.5; IR (KBr): 3406, 3065, 3031, 1616, 1513, 1492, 1342, 1164, 1121, 1068, 698 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₀H₂₀NO [M+H]⁺ 290.1545, found 290.1517; [α]_D²⁶ +46.4 (c 1.00, CH₂Cl₂, 92% ee (1R,2R)); lit.^{5d} [α]_D²⁴ -45.2 (c 0.52, CH₂Cl₂, 92% ee (1S,2S)); HPLC (Daicel Chiralpak AD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=25.5 min (minor, (1S,2S)); *t*_R=29.8 min (major, (1R,2R))).

4.3.1.2. 1,2-Diphenyl-2-(3-trifluorophenyl)-ethanol (**4b**). ¹H NMR δ 2.75 (br s, 1H), 4.53 (d, *J*=6.2 Hz, 1H), 4.54 (br s, 1H), 4.88 (d,

J=6.2 Hz, 1H), 6.52–6.54 (m, 2H), 6.53–6.65 (m, 1H), 7.03–7.06 (m, 2H), 7.20–7.27 (m, 9H); ¹³C NMR δ 64.8, 78.0, 114.2, 118.0, 126.6, 127.3, 127.5, 127.9, 128.2, 128.6, 129.0, 140.2, 140.6, 147.2; IR (KBr): 3407, 3058, 3028, 1502, 1453, 1317, 1262, 1155, 1051 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₁H₁₉F₃NO [M+H]⁺ 358.1418, found 358.1457; $[\alpha]_D^{26}$ +42.1 (*c* 0.99, CH₂Cl₂, 92% ee); HPLC (Daicel Chiralpak AD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=15.9 min (minor); *t*_R=18.5 min (major)).

4.3.1.3. (1R,2R)-2-(Naphthalen-1-ylamino)-1,2-diphenylethanol (**4c**). ¹H NMR δ 2.54 (br s, 1H), 4.68 (d, *J*=9.6 Hz, 1H), 4.99(d, *J*=9.6 Hz, 1H), 5.51 (br s, 1H), 6.27-6.28 (m, 1H), 7.06-7.41 (m, 14H), 7.72-7.77 (m, 1H), 7.96-7.97 (m, 1H); ¹³C NMR δ 64.5, 78.2, 106.7, 117.7, 120.0, 124.0, 124.8, 125.6, 126.3, 126.5, 127.2, 127.5, 128.0, 128.3, 128.6, 128.7, 134.2, 139.9, 140.7, 142.1; IR (KBr): 3406, 3059, 3030, 1580, 1523, 1476, 1453, 1407, 1342, 1280, 1032, 768, 700 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₄H₂₂NO [M+H]⁺ 340.1701, found 340.1680; [α]_D²⁶ +126.6 (*c* 1.07, CH₂Cl₂, 95% ee (1*R*,2*R*)); lit.^{5d} [α]_D²² -144.9 (*c* 0.39, CH₂Cl₂, 91% ee (1*S*,2*S*)); HPLC (Daicel Chiralpak OD, eluent *n*-hexane/*i*-PrOH=4/1, flow rate 1.0 mL/min (major, (1*R*,2*R*)), *t*_R=11.4 min; *t*_R=21.0 min (minor, (1*S*,2*S*))).

4.3.1.4. (1*R*,2*R*)-2-(*N*-Methyl-*N*-phenylamino)-1,2-diphenylethanol (**4d**). ¹H NMR δ 2.70 (s, 3H), 3.95 (br s, 1H), 4.86 (d, *J*=9.7 Hz, 1H), 5.27(d, *J*=9.7 Hz, 1H), 6.88–6.90 (m, 1H), 6.90–7.00 (m, 4H), 7.11–7.27 (m, 8H), 7.36–7.38 (m, 2H); ¹³C NMR δ 32.8, 71.6, 73.6, 117.7, 120.3, 127.6, 127.7, 127.8, 128.0, 128.2, 128.8, 129.1, 134.8, 140.7, 151.4; IR (KBr): 3423, 3060, 3030, 1597, 1561, 1451, 1320, 1188, 1079, 1055, 1030, 935, 754, 698 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₁H₂₂NO [M+H]⁺ 304.1701, found 304.1739; [α]₂²⁶ –171.6 (*c* 0.99, CH₂Cl₂, 95% ee (1*R*,2*R*)); lit.^{5d} [α]₂²² +171.7 (*c* 0.53, CH₂Cl₂, 96% ee, (1*S*,2*S*)); HPLC (Daicel Chiralpak ASH, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 0.8 mL/min, *t*_R=15.2 min (major, (1*R*,2*R*)); *t*_R=21.0 min (minor, (1*S*,2*S*))).

4.3.1.5. (1R,2R)-2-(2-Methoxyphenylamino)-1,2-diphenylethanol (**4e**). ¹H NMR δ 2.66 (br s, 1H), 3.85 (s, 3H), 4.50 (d, J=6.2 Hz, 1H), 4.86 (d, J=6.2 Hz, 1H), 5.12 (br s, 1H), 6.37–6.39 (m, 1H), 6.58–6.74 (m, 3H), 7.14–7.26 (m, 10H); ¹³C NMR δ 55.6, 65.0, 78.3, 109.7, 111.8, 117.2, 121.1, 126.7, 127.3, 127.8, 128.0, 128.4, 137.1, 140.7, 147.4; IR (KBr): 3403, 3061, 3027, 1601, 1509, 1454, 1428, 1342, 1224, 1125, 1048, 1026, 768, 738, 700 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₁H₂₂NO₂ [M+H]⁺ 320.1650, found 320.1665; $[\alpha]_D^{26}$ +51.9 (c 1.08, CH₂Cl₂, 95% ee (1R,2R)); lit.^{5d} $[\alpha]_D^{22}$ -48.0 (c 0.54, CH₂Cl₂, 93% ee, (1S,2S)); HPLC (Daicel Chiralpak ASH, eluent n-hexane/i-PrOH=19/1, flow rate 0.8 mL/min, t_R =20.4 min (minor, (1S,2S)); t_R =24.9 min (major, (1*R*,2*R*))).

4.3.1.6. (1R,2R)-2-(Phenylamino)-1,2-di(m-tolyl)-ethanol (**4f**). ¹H NMR δ 2.28 (s, 3H), 2.31 (s, 3H), 2.44 (br s, 1H), 4.48 (d, J=5.5 Hz, 1H), 4.55 (br s, 1H), 4.85 (d, J=5.5 Hz, 1H), 6.50–6.52 (m, 2H), 6.60–6.64 (m, 1H), 7.02–7.17 (m, 10H); ¹³C NMR δ 21.4, 64.6, 77.9, 114.1, 117.8, 123.5, 124.3, 127.9, 128.1, 128.3, 128.4, 128.6, 129.0, 137.9, 138.2, 140.3, 140.6, 147.3; IR (KBr): 3406, 3049, 3021, 1602, 1503, 1430, 1317, 1267, 1155, 1048, 789, 749, 706 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₂H₂₄NO [M+H]⁺ 318.1858, found 318.1827; $[\alpha]_{D}^{27}$ +41.3 (c 0.05, CH₂Cl₂, 95% ee (1R,2R)); Iit.¹⁷ $[\alpha]_{D}^{23}$ +50.2 (c 0.89, CH₂Cl₂, 95% ee (1R,2R)); HPLC (Daicel Chiralpak OD, eluent *n*-hexane/*i*-PrOH=9/1, flow rate 0.8 mL/min, t_{R} =12.1 min (major, (1R,2R)); t_{R} =15.9 min (minor, (1S,2S))).

4.3.1.7. (1*R*,2*R*)-2-(Phenylamino)-1,2-di(p-tolyl)-ethanol (**4g**). ¹H NMR δ 2.28 (s, 3H), 2.31 (s, 3H), 2.44 (br s, 1H), 4.48 (d, *J*=5.5 Hz, 1H), 4.55 (br s, 1H), 4.81 (d, *J*=5.5 Hz, 1H), 6.49–6.51 (m, 2H), 6.60–6.64 (m, 1H), 7.01–7.22 (m, 10H); ¹³C NMR δ 21.1, 64.3, 77.8, 114.1, 117.8, 126.5, 127.2, 128.9, 129.0, 129.2, 137.0, 137.3, 137.4, 137.7, 147.7; IR (KBr): 3398, 3020, 2920, 1602, 1502, 1318, 1048, 819, 749, 691 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₂H₂₄NO [M+H]⁺ 318.1858,

found 318.1827; $[\alpha]_D^{26}$ +74.4 (*c* 1.01, CH₂Cl₂, 95% ee (1*R*,2*R*)); lit.^{5d} $[\alpha]_D^{24}$ -47.3 (*c* 0.54, CH₂Cl₂, 90% ee, (1*S*,2*S*)); HPLC (Daicel Chiralpak ADH, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, t_R =25.2 min (minor, (1*S*,2*S*)); t_R =30.6 min (major, (1*R*,2*R*))).

4.3.1.8. (1R,2R)-2-(*Phenylamino*)-1,2-*di*(*p*-bromophenyl)ethanol (**4h**). ¹H NMR δ 2.58 (br s, 1H), 4.40 (d, *J*=6.2 Hz, 1H), 4.52 (br s, 1H), 4.73 (d, *J*=6.2 Hz, 1H), 6.48–6.49 (m, 2H), 6.66–6.67 (m, 1H), 7.02–7.07 (m, 6H), 7.34–7.40 (m, 4H); ¹³C NMR δ 64.3, 77.3, 114.2, 118.4, 121.5, 122.0, 128.3, 129.0, 129.1, 131.4, 131.7, 138.9, 139.3, 146.7; IR (KBr): 3398, 3049, 2363, 2344, 1601, 1501, 1486, 1430, 1403, 1315, 1071, 1008, 829, 751, 691 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₀H₁₈Br₂NO [M+H]⁺ 447.9735, found 447.9763; [α]₂²⁶ +72.0 (*c* 1.01, CH₂Cl₂, 90% ee); HPLC (Daicel Chiralpak AD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=32.3 min (minor, (15,2S)); *t*_R=41.5 min (major, (1*R*,2*R*))).

4.3.1.9. (1*R*,2*R*)-3-(*Phenylamino*)-2-*butanol* (**4i**). ¹H NMR δ 1.14 (d, *J*=6.9 Hz, 3H), 1.25 (d, *J*=6.9 Hz, 3H), 2.62 (br s, 1H), 3.29–3.34 (m, 1H), 3.61–3.65 (m, 1H), 6.66–6.74 (m, 3H), 7.15–7.23 (m, 2H); ¹³C NMR δ 17.3, 19.5, 56.0, 71.3, 114.3, 118.2, 129.3, 147.8; IR (KBr): 3398, 2973, 2929, 1602, 1503, 1318, 1256, 750, 693 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₁₀H₁₆NO [M+H]⁺ 166.1232, found 166.1259; [α]_D²⁵ –50.1 (*c* 0.99, CH₂Cl₂, 90% ee (1*R*,2*R*)); lit.¹⁶ [α]_D²⁵ –38.3 (*c* 0.95, benzene, >99.9% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak OD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=26.0 min (major (1*R*,2*R*)); *t*_R=29.9 min (minor (1*S*,2*S*))).

4.3.1.10. 3-(1-Naphthylamino-1-yl)-2-butanol (**4***j*). ¹H NMR δ 1.21 (d, J=6.4 Hz, 3H), 1.28 (d, J=6.4 Hz, 3H), 2.60 (br s, 1H), 3.49–3.56 (m, 1H), 3.75–3.82 (m, 1H), 4.24 (br s, 1H), 6.68 (d, J=7.2 Hz, 1H), 7.20–7.44 (m, 4H), 7.75–7.82 (m, 2H); ¹³C NMR δ 17.0, 19.8, 55.3, 71.4, 105.9, 117.9, 119.9, 124.0, 124.8, 125.7, 126.5, 128.7, 134.4, 142.6; IR (KBr): 3424, 2970, 2927, 1580, 1526, 1408, 1121, 1011, 769 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₁₄H₁₈NO [M+H]⁺ 216.1388, found 216.1400; [α] $^{26}_{16}$ –64.6 (*c* 1.00, CH₂Cl₂, 67% ee); HPLC (Daicel Chiralpak AD, eluent *n*-hexane/*i*-PrOH=30/1, flow rate 1.0 mL/min, *t*_R=15.0 min (minor); *t*_R=17.0 min (major)).

4.3.1.11. (1R,2R)-2-Phenylamino-1-cyclohexanol (**4k**). ¹H NMR δ 1.10–1.08 (m, 1H), 1.27–1.43 (m, 3H), 1.40–1.43 (m, 2H), 1.70–1.78 (m, 2H), 3.00 (br s, 1H), 3.11–3.16 (m, 1H), 3.33–3.37 (m, 1H), 6.70–6.76 (m, 2H), 7.16–7.25 (m, 2H); ¹³C NMR δ 24.2, 25.0, 31.5, 33.1, 60.2, 74.5, 114.4, 118.4, 129.3, 147.7; IR (KBr): 3394, 2924, 2858, 1599, 1513, 1497, 1449, 1321, 1052, 864, 741, 689 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₁₂H₁₈NO [M+H]⁺ 192.1388, found 192.1377; [α]_D²⁶ –44.7 (*c* 0.99, CH₂Cl₂, 90% ee (1*R*,2*R*)); lit.¹⁷ [α]_D²⁵ –37.7 (*c* 0.54, CH₂Cl₂, 54% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak OD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=18.9 min (minor, (1*S*,2*S*)); *t*_R=22.3 min (major, (1*R*,2*R*))).

4.3.1.12. 2-(1-Naphthylamino-1-yl)-1-cyclohexanol (**4I**). ¹H NMR δ 1.05–1.10 (m, 1H), 1.29–1.44 (m, 3H), 1.69–1.78 (m, 2H), 2.11–2.23 (m, 2H), 2.85 (br s, 1H), 3.29–3.36 (m, 1H), 3.46–3.52 (m, 1H), 6.76 (d, *J*=7.2 Hz, 1H), 7.29–7.44 (m, 4H), 7.72–7.82 (m, 1H); ¹³C NMR δ 24.2, 25.0, 31.5, 33.1, 60.2, 74.5, 114.4, 118.4, 129.3, 147.7; IR (KBr): 3515, 3399, 3049, 2922, 2855, 1576, 1537, 1489, 1446, 1411, 1282, 1064, 766 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₁₆H₂₀NO [M+H]⁺ 242.1545, found 242.1552; $[\alpha]_D^{26}$ –63.0 (*c* 0.99, CH₂Cl₂, 90% ee); HPLC (Daicel Chiralpak OD, eluent *n*-hexane/*i*-PrOH=19/1, flow rate 1.0 mL/min, *t*_R=18.9 min (minor); *t*_R=22.3 min (major)).

4.3.1.13. 2-(*N*-Methyl-*N*-phenylamino)-1,2-di(p-tolyl)-ethanol (**4m**). ¹H NMR δ 2.20 (s, 3H), 2.24 (s, 3H), 2.64 (s, 3H), 3.97 (br s, 1H), 4.85 (d, *J*=10.3 Hz, 1H), 5.23 (d, *J*=10.3 Hz, 1H), 6.85–6.93 (m, 5H), 7.00–7.04 (m, 4H), 7.21–7.29 (m, 4H); ¹³C NMR δ 21.0, 32.4, 71.1,

73.3, 111.7, 120.2, 127.5, 128.6, 128.9, 129.1, 131.6, 137.2, 137.3, 137.6, 151.5; IR (KBr): 3422, 3023, 2918, 2882, 1597, 1501, 1451, 1379, 1318, 1180, 1032, 816, 751, 694 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₃H₂₆NO [M+H]⁺ 332.2014, found 332.1999; $[\alpha]_D^{26} - 134.3$ (*c* 1.00, CH₂Cl₂, 90% ee); HPLC (Daicel Chiralpak ASH, eluent *n*-hexane/*i*-PrOH=40/1, flow rate 1.0 mL/min, t_R =16.1 min (major); t_R =22.6 min (minor)).

4.3.1.14. (1R,2R)-2-(N-Methyl-N-phenylamino)-1,2-di(4-bromophenyl)ethanol (**4n**). ¹H NMR δ 2.67 (s, 3H), 3.88 (br s, 1H), 4.67 (d, J=9.6 Hz, 1H), 5.17 (d, J=9.6 Hz, 1H), 6.78–6.81 (m, 2H), 6.93–6.97 (m, 3H), 7.21–7.36 (m, 8H); ¹³C NMR δ 32.8, 70.8, 73.7, 118.2, 121.1, 121.8, 122.0, 129.2, 130.3, 130.9, 131.3, 131.4, 132.4, 133.0, 139.4, 150.8; IR (KBr): 3425, 2883, 1699, 1594, 1490, 1380, 1311, 1185, 1069, 1009, 820, 753, 694 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₁H₂₀Br₂NO [M+H]⁺ 461.9891, found 461.9894; [α]_D²⁷ –61.5 (*c* 1.02, CH₂Cl₂, 90% ee (1R,2R)); HPLC (Daicel Chiralpak ASH, eluent *n*-hexane/*i*-PrOH=40/1, flow rate 1.0 mL/min, *t*_R=26.8 min (major, (1*R*,2*R*)); *t*_R=31.3 min (minor, (15,2S))).

4.3.1.15. 2-(*N*-Methyl-*N*-phenylamino)-1,2-di(2-naphthyl-1-yl)ethanol (**4o**). ¹H NMR δ 2.76 (s, 3H), 3.88 (br s, 1H), 5.16 (d, *J*=9.6 Hz, 1H), 5.59 (d, *J*=9.6 Hz, 1H), 6.92–6.94 (m, 1H), 7.07–7.09 (m, 2H), 7.11–7.18 (m, 4H), 7.28–7.68 (m, 8H), 7.88–7.97 (m, 4H); ¹³C NMR δ 32.9, 71.6, 73.7, 117.8, 120.5, 122.7, 125.1, 125.8, 127.5, 127.8, 127.9, 128.0, 129.2, 133.1, 134.5, 138.1, 151.3; IR (KBr): 3422, 3054, 1690, 1596, 1501, 1347, 1269, 1168, 1119, 859, 820, 749 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₉H₂₆NO [M+H]⁺ 404.2014, found 404.2005; $[\alpha]_D^{26}$ –60.2 (*c* 0.99, CH₂Cl₂, 90% ee); HPLC (Daicel Chiralpak ASH, eluent *n*-hexane/*i*-PrOH=40/1, flow rate 1.0 mL/min, *t*_R=44.4 min (major); *t*_R=50.1 min (minor)).

4.4. General procedure for *meso*-epoxide ring-opening reaction with indole

A mixture of $Cu[O_3S(CH_2)_{10}CH_3]_2$ and bipyridine ligand **3** in water was stirred for 1 h at room temperature. *meso*-epoxide and indole were then added to the mixture. The mixture was further stirred for 22–48 h at room temperature. The resulting mixture was quenched with satd NaHCO₃ aq and brine. The aqueous layer was extracted with dichloromethane (three times), and the combined organic layers were washed with brine, and dried over Na₂SO₄. After filtration, the solvent was removed under reduced pressure. The residue was purified by preparative TLC (elution *n*-hexane/ ethyl acetate=3/2) to give the product.

4.4.1. Physical data of the amino indole adducts.

4.4.1.1. (15,2S)-2-(1H-Indole-3-yl)-1,2-diphenylethanol (**6a**). ¹H NMR δ 2.56 (br s, 1H), 4.55 (d, *J*=8.2 Hz, 1H), 5.28 (d, *J*=8.2 Hz, 1H), 6.99–7.20 (m, 14H), 7.42–7.43 (m, 1H), 8.09 (br s, 1H); ¹³C NMR δ 52.0, 77.6, 111.1, 115.2, 119.3, 119.6, 122.2, 122.4, 126.3, 126.8, 127.3, 127.5, 127.9, 128.0, 128.6, 136.3, 141.7, 142.4; IR (KBr): 3348, 3026, 2873, 1490, 1457, 1340, 1224, 1042, 744, 698 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₂H₂₀NO [M+H]⁺ 314.1545, found 314.1578; [α]_D²⁷ +61.3 (*c* 1.08, CHCl₃, 96% ee (15,2S)); lit.^{8d} [α]_D²⁴ –63.4 (*c* 1.08, CHCl₃, 93% ee, (1*R*,2*R*)); HPLC (Daicel Chiralpak ODH, eluent *n*-hexane/*i*-PrOH=4/1, flow rate 0.8 mL/min, *t*_R=34.6 min (minor, (1*R*,2*R*)); *t*_R=48.3 min (major, (15,2S))).

4.4.1.2. (1S,2S)-2-(5-Methoxy-1H-indole-3-yl)-1,2-diphenylethanol (**6b**). ¹H NMR δ 2.04 (br s, 1H), 3.72 (s, 3H), 4.53 (d, J=7.6 Hz, 1H), 5.33 (d, J=7.6 Hz, 1H), 6.77-6.82 (m, 2H), 7.08-7.30 (m, 12H), 8.02 (br s, 1H); ¹³C NMR δ 52.0, 55.8, 77.3, 101.4, 111.7, 112.6, 115.0, 123.4, 126.3, 126.7, 127.3, 127.9, 128.0, 128.1, 128.6, 131.5, 141.9, 142.6, 154.1; IR (KBr): 3518, 3369, 3314, 3025, 1623, 1583, 1485, 1453, 1437, 1213, 1166, 1024, 926, 823, 808, 754, 727, 695 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₃H₂₂NO₂ [M+H]⁺ 344.1651, found 344.1679; $[\alpha]_D^{28}$ +14.0 (*c* 0.96, CHCl₃, 92% ee (1*S*,2*S*)); lit.^{8d} [α]_D²⁴ –15.0 (*c* 0.96, CHCl₃, 92% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ADH, eluent *n*-hexane/*i*-PrOH=7/3, flow rate 1.0 mL/min, *t*_R=21.8 min (major, (1*S*,2*S*)); *t*_R=28.5 min (minor, (1*R*,2*R*))).

4.4.1.3. (1*S*,2*S*)-2-(5-*Methyl*-1*H*-*indole*-3-*yl*)-1,2-*diphenylethanol* (**6**c). ¹H NMR δ 2.34 (s, 3H), 2.49 (br s, 1H), 4.56 (d, *J*=8.3 Hz, 1H), 5.31(d, *J*=9.6 Hz, 1H), 6.97–6.99 (m, 1H), 7.06–7.30 (m, 13H), 8.03 (br s, 1H); ¹³C NMR δ 21.5, 52.1, 77.7, 110.7, 113.5, 114.8, 119.0, 122.6, 124.0, 126.3, 126.8, 127.3, 127.9, 128.1, 128.6, 128.9, 134.7, 141.8, 142.4; IR (KBr): 3490, 3285, 3025, 1490, 1454, 1225, 1111, 1073, 1024, 797, 761, 729, 697, 647 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₃H₂₂NO [M+H]⁺ 328.1701, found 328.1715; [α]₃³⁰ +33.7 (*c* 0.99, CHCl₃, 92% ee (15,25)); lit.^{8d} [α]₆²⁴ –137.6 (*c* 0.97, CHCl₃, 85% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ODH, eluent *n*-hexane/*i*-PrOH=7/3, flow rate 0.7 mL/min, t_R =29.6 min (major, (1*S*,2*S*)); t_R =36.9 min (minor, (1*R*,2*R*))).

4.4.1.4. (1S,2S)-2-(5-Bromo-1H-indole-3-yl)-1,2-diphenylethanol (**6d**). ¹H NMR δ 2.03 (br s, 1H), 4.51 (d, *J*=7.6 Hz, 1H), 5.27 (d, *J*=7.6 Hz, 1H), 7.06–7.34 (m, 13H), 7.47–7.48 (m, 1H), 8.18 (br s, 1H); ¹³C NMR δ 51.6, 77.6, 79.0, 112.5, 112.9, 114.9, 121.9, 123.8, 125.1, 126.5, 126.6, 126.9, 127.4, 127.9, 128.0, 128.1, 128.2, 128.5, 129.4, 134.8, 141.4, 142.3; IR (KBr): 3396, 3347, 3243, 1603, 1491, 1455, 1332, 1048, 1034, 887, 798, 751, 741, 699 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₂H₁₉BrNO [M+H]⁺ 392.0630, found 392.0646; [α]₀³⁰ +15.4 (*c* 1.00, CHCl₃, 95% ee (1*S*,2*S*)); lit.^{8d} [α]₀²² –13.0 (*c* 0.43, CHCl₃, 90% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ODH, eluent *n*-hexane/ *i*-PrOH=7/3, flow rate 1.0 mL/min, *t*_R=12.8 min (major, (1*S*,2*S*)); *t*_R=22.7 min (minor, (1*R*,2*R*))).

4.4.1.5. (1S,2S)-2-(2-Methyl-1H-indole-3-yl)-1,2-diphenylethanol (**6e**). ¹H NMR δ 2.03 (s, 3H), 2.54 (br s, 1H), 4.10 (d, *J*=7.2 Hz, 1H), 5.69 (d, *J*=7.2 Hz, 1H), 7.03–7.21 (m, 13H), 7.66 (d, *J*=7.2 Hz, 1H), 7.93 (br s, 1H); ¹³C NMR δ ; IR (KBr): 3400, 3058, 3027, 1617, 1491, 1458, 1387, 1303, 1246, 1185, 1079, 1051, 909, 740, 699 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₃H₂₂NO [M+H]⁺ 328.1701, found 328.1687; $[\alpha]_{10}^{30}$ +89.8 (*c* 1.09, CHCl₃, 95% ee (1*S*,2*S*)); lit.^{8d} [α]_{12}^{22} –153.5 (*c* 0.68, CHCl₃, 98% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ADH, eluent *n*-hexane/*i*-PrOH=9/1, flow rate 1.0 mL/min, *t*_R=43.8 min (major, (1*S*,2*S*)); *t*_R=46.8 min (minor, (1*R*,2*R*))).

4.4.1.6. 2-(1*H*-Indole-3-yl)-1,2-di(naphthalene-2-yl)ethanol (**6f**). ¹H NMR δ 2.61 (br s, 1H), 4.91 (d, J=7.6 Hz, 1H), 5.64 (d, J=7.6 Hz, 1H), 6.96–6.99 (m, 1H), 7.12–7.715 (m, 1H), 7.31–7.46 (m, 9H), 7.59–7.74 (m, 8H), 8.12 (br s, 1H); ¹³C NMR δ 51.8, 77.5, 111.1, 115.1, 119.7, 122.4, 122.8, 124.8, 125.4, 125.6, 125.7, 127.1, 127.5, 127.6, 127.8, 128.0, 132.2, 132.9, 133.1, 133.4, 136.3, 139.3, 139.9; IR (KBr): 3423, 3335, 3051, 2879, 1599, 1508, 1457, 1420, 1342, 1271, 1122, 1100, 1034, 908, 815, 745 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₃₀H₂₄NO [M+H]⁺ 414.1858, found 414.1858; [α]²⁹₂+74.1 (*c* 0.96, CHCl₃, 78% ee); HPLC (Daicel Chiralpak ADH, eluent *n*-hexane/*i*-PrOH=4/1, flow rate 1.0 mL/min, *t*_R=49.6 min (major); *t*_R=66.1 min (minor)).

4.4.1.7. (1*S*,2*S*)-2-(1*H*-Indole-3-*y*)-1,2-*d*i-*p*-tolylethanol (**6g**). ¹H NMR δ 2.21 (s, 3H), 2.26 (br s, 1H), 2.27 (s, 3H), 4.55 (d, *J*=7.6 Hz, 1H), 5.28 (d, *J*=7.6 Hz, 1H), 6.94 (d, *J*=8.3 Hz, 2H), 7.00–7.04 (m, 5H), 7.11–7.17 (m, 3H), 7.22–7.24 (m, 2H), 7.45 (d, *J*=6.9 Hz, 1H), 8.10 (br s, 1H); ¹³C NMR δ 20.9, 21.1, 51.3, 60.4, 77.5, 111.0, 115.6, 119.4, 119.5, 122.2, 122.5, 126.7, 126.8, 127.0, 127.6, 128.5, 128.6, 128.8, 129.0, 135.6, 136.3, 136.8, 138.9, 139.5; IR (KBr): 3412, 3020, 2918, 1511, 1489, 1419, 1339, 1189, 1036, 813, 742, 576 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₄H₂₄NO [M+H]⁺ 342.1858, found 342.1822; [*α*]^{*b*}/_{*D*} +43.8 (*c* 0.98, CHCl₃, 70% ee (1*S*,2*S*)); lit.^{8d} [*α*]²⁴/_{*D*} –54.5 (*c* 0.81, CHCl₃, 90% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ODH, eluent *n*-hexane/ *i*-PrOH=4/1, flow rate 1.0 mL/min, *t*_R=23.4 min (major, (1*S*,2*S*)); *t*_R=27.9 min (minor, (1*R*,2*R*))). 4.4.1.8. (15,25)-1,2-Bis(4-bromophenyl)-2-(1H-indole-3-yl)ethanol (**6h**). ¹H NMR δ 2.60 (br s, 1H), 4.42 (d, *J*=8.3 Hz, 1H), 5.17 (d, *J*=8.3 Hz, 1H), 6.93 (d, *J*=8.9 Hz, 2H), 7.01–7.04 (m, 3H), 7.15–7.38 (m, 8H), 8.18 (br s, 1H); ¹³C NMR δ 51.4, 76.8, 111.3, 114.1, 119.1, 119.8, 120.3, 121.3, 122.3, 122.5, 127.2, 128.4, 128.6, 130.3, 131.1, 131.3, 136.3, 140.3, 141.0; IR (KBr): 3411, 3055, 2892, 1702, 1591, 1486, 1456, 1405, 1339, 1100, 1071, 1040, 1009, 907, 833, 817, 781, 743 cm⁻¹; HRMS (ESI, Pos.): Calcd for C₂₂H₁₈Br₂NO [M+H]⁺ 471.9735, found 471.9711; [α]₂^{D9} +78.7 (*c* 1.02, CHCl₃, 90% ee (15,25)); lit.^{8d} [α]₂^{D8} –28.9 (*c* 0.90, CHCl₃, 92% ee (1*R*,2*R*)); HPLC (Daicel Chiralpak ADH, eluent *n*-hexane/*i*-PrOH=7/3, flow rate 1.0 mL/min, *t*_R=14.9 min (major, (15,25)); *t*_R=18.7 min (minor, (1*R*,2*R*))).

4.5. Crystallization of [3 CuBr₂] complexes

Crystallization of $[3 \cdot \text{CuBr}_2]$ complex was carried out as follows: To a solution of CuBr₂ (9.6 mg, 0.043 mmol) in DME (1.5 mL) was added **3** (14.1 mg, 0.043 mmol) at room temperature. The solution was stirred at 25 °C. Water (0.07 mL) was then added to the mixture, which was cooled to room temperature. After 3 days, crystals suitable for X-ray analysis were formed.

Crystallographic data for the structure reported in this paper have been deposited with the Cambridge Crystallographic Data Center as supplementary publication number CCDC-698334. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 J 1EZ, UK (fax: +44 1223 336 033; e-mail: deposit@ccdc.cam.ac.uk).

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