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Synthesis and DNA damaging ability of enediyne model compounds possessing photo-triggering devices

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Abstract—Enediyne model compounds possessing photo-triggering devices were developed. These enediynes afforded biradicals by UV irradiation and showed DNA cleaving activity. The DNA damage was confirmed to be mainly caused by the biradical, not singlet oxygen.

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For many decades, DNA damage induced by photoirradiation has attracted considerable attention and several types of DNA photocleavers have been exploited. Since the photochemical processes of these agents are generally compatible with other biological processes, they have significant potential as biological probes of DNA, and drugs for photochemotherapy in which ultraviolet, visible, and infrared lights are used together with administered photoactive agents.¹ Now with the recent rapid progress of laser and fiber technology it is becoming possible to introduce an intense beam of these radiations into the depths of the human body and the importance of photochemotherapy has been correspondingly increasing.² Since Shiraki and Sugiura reported the photocleavage of DNA by Dynemicin A in 1990, this new class of photocleaver has attracted a great deal of attention from researchers, and several enedivne model compounds bearing photo-triggering devices have been developed.³ In the course of our studies for developing enediyne model compounds generating bioactive carbon biradicals through a characteristic triggering action,⁴ we have also investigated the design and synthesis of enediynes activated photochemically. Here, we report the synthesis and DNA cleaving abilities of the enediyne model compounds 1, 2a, and 2b that we designed, activated by photo-irradiation and, by comparison with reference compounds 3 and 4, we also confirm that the DNA damage was mainly caused by the biradical. The molecular design of targeting arenediynes

is depicted in Figure 1. Photochemical removal of the protecting group on the cyanohydrin moiety initiates the reaction cascade including regeneration of the carbonyl group and isomerization to an arenyne–allene, which cycloaromatizes to generate the α ,3-didehydrotoluene biradical.⁵

Initially, the syntheses of the alkynes 9a and 9b were undertaken as shown in Scheme 1. Hydroxy ester 5⁶ was reacted with 2-nitrobenzyl chloromethyl ether⁷ giving acetal 6, which was treated with aq NH₃ in EtOH affording amide and the following dehydration reaction using Burgess reagent⁸ gave nitrile **9a**. Carbonate **9b** was also synthesized from ester 5. THP protection of 5 afforded acetal 7, which was converted to nitrile 8 in two steps. The removal of the THP protection followed by the introduction of the carbonate moiety using *p*-nitrophenyl 2-(2-nitrophenyl)propyl carbonate⁹ afforded **9b**. Diiodobenzene was coupled with methyl propargyl ether in the presence of Pd catalyst affording arenyne 10, and the sequential coupling reaction of 10 with 9a or 9b under the Sonogashira conditions gave enediynes 1 and 2a. Enediyne 2b was also synthesized from 11 in a similar fashion to the synthesis of 2a.

First, we examined the cycloaromatization reactions of enediynes **1**, **2a**, and **2b** in the presence of 1.1 equiv of Et₃N and excess amounts (50 equiv) of 1,4-cyclohexadiene under UV irradiation ($\lambda > 365$ nm) in degassed MeOH. The results are summarized in Table 1. While carbonate **2a** and **2b** were photolyzed completely within 30 min, a considerably longer reaction time (0.5 h vs 17 h) was needed for the deprotection of acetal **1**.^{9b} In

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Figure 1.



Scheme 1. Reagents and conditions: (a) 2-Nitrobenzyl chloromethyl ether, NaH/THF, rt. (b) aq NH₃/EtOH, rt. (c) $Et_3N^+SO_2N^-CO_2Me/THF$, rt. (d) DHP, *p*-TsOH/C₆H₆, rt. (e) Amberlyst 15/MeOH, rt. (f) *p*-Nitrophenyl 2-(2-nitrophenyl)propyl carbonate, Py/CH₂Cl₂, rt. (g) Methyl propargyl ether, Pd(PPh₃)₄, CuI, Et₃N/toulene, rt. (h) 2-Propyn-1-ol, Pd(PPh₃)₄, CuI, Et₃N/toulene, rt. (i) TBSCl, imidazole/CH₂Cl₂, rt. (j) **9a** (or **9b**), Pd(PPh₃)₄, CuI, Et₃N/toulene, rt. (k) aq HF/MeCN, 0 °C.

both enediynes 1 and 2a, the reactions proceeded, giving the products 13a, 14, and 15a accompanied by considerable amounts of unknown polar materials (entries 1 and 2). Adduct 13a was obtained as an isomeric mixture and then was converted to 16 by catalytic hydrogenation to confirm the structure. In the cycloaromatization reactions of 2b, 13b was obtained along with the trace amounts of 15b (entry 3). 13b and 15b were converted to 17 and 15a, respectively, to confirm the structure. To exclude any possibility that the cascade reaction was initiated by the base promoted deprotection of 2, we prepared reference enediyne 3, bearing a photostable protecting group. When cycloaromatization of the reference enediyne 3 was carried out under the same

Table 1. The cycloaromatization reactions of enediynes 1 and 2



Enediynes, Et₃N (1.1 equiv), and 1,4-cyclohexadiene (50 equiv) were dissolved in MeOH and photo-irradiated using a MUV-202U (MORITEX, 2000 mW cm⁻², $\lambda > 365$ nm) for the indicated times.

^a Compound 13a was obtained as an isomeric mixture and then was converted to 16 by catalytic hydrogenation to confirm the structure.

^bCompound 13b was converted to 17 in two steps and the structure was confirmed.

 $^{\rm c}$ The structure of 15b was confirmed by converting to 15a using $\rm Me_2SO_4$ and aq NaOH.

conditions, enediyne **3** was recovered almost completely (entry 4). These results indicate that the cascade reaction was initiated photochemically to form the α ,3-didehy-drotoluene biradical. In addition, the enediyne skeleton itself might undergo a photo-Bergman cyclization reaction,^{3d,g} however, in our enediyne systems, no product arising from Bergman cycloaromatization was obtained.

A plausible mechanism of the formation of **13–15** is depicted in Figure 2. Photo-irradiation initiates the reaction cascade, consisting of the deprotection, isomerization, and cycloaromatization processes, giving the biradical. The resulting biradical abstracts hydrogen from 1,4-CHD, and finally should afford **13**. The biradical also reacts with adventitious oxygen to give a peroxide, which would decompose to aldehyde **14**.¹⁰ In addition, the intermediary arenyne–allene degrades with the conjugate addition of solvent MeOH affording **15**. Next, we carried out DNA cleaving assays of synthetic enediynes. Enediynes 2a and 2b were incubated with Φ X174R-1 plasmid DNA in a phosphate buffer solution (pH 8.0, containing 20% of acetonitrile) and the mixture was irradiated ($\lambda > 365 \text{ nm}$). The resultant DNA fragments were separated by electrophoresis on agarose gel and visualized by ethidium bromide staining: the results are summarized in Table 2. Prolonged irradiation time (30 min vs 60 min) and elevated temperature (25 °C vs 37 °C) were effective in improving the DNA damaging abilities of 2b (entries 4–6), however, in the case of 2a, these modifications were not so effective (entries 1-3). Although 2a and 2b showed a similar degree of facility in the deprotection step (Table 1, entries 2 and 3) and disappeared almost completely after the irradiation, 2a showed only low activity compared with 2b. This difference would be attributed to the low solubility of 2a in the buffer solution. Indeed, enediyne 2b could dissolve



Entry	Enediyne	Time (min)	Temperature (°C)	% Cleavage ^{b,c}
1	2a ^a	30	25	11 ± 3
2	2a ^a	60	25	12 ± 4
3	2a ^a	30	37	15 ± 4
4	2b	30	25	27 ± 4
5	2b	60	25	40 ± 3
6	2b	30	37	47 ± 7
7	3	30	25	<5
8	4	30	25	11 ± 2

Table 2. DNA cleaving abilities of synthetic enediynes 1-3 and the reference 4

 Φ X174R-1 DNA (0.46 µg, WAKO Pure Chemical Industries, Ltd) in 20 µL phosphate buffer solution (pH 8.0, containing 20% acetonitrile) with drugs (500 µM) was incubated and photo-irradiated using a VL-30L (VILBER LOURMAT, 1820 µW cm⁻², λ > 365 nm) for indicated times at 25 or 37 °C. Immediately, 15 µL samples were loaded into 1% agarose gel. The running buffer was 20 mM TAE (pH 7.8). Electrophoresis was at 50 V for 8 h. After electrophoresis, gel was stained for 1 h in ethidium bromide (1 µg/mL) and de-stained for 5 min in water. Relative amounts of DNA in Form I, Form II, and Form III were determined by densitometry.

^a Enediyne 2a was suspended in phosphate buffer solution.

^b Values presented are mean value \pm SD of three runs. A control reaction mixture without the addition of any drug was incubated and photoirradiated. The mean value of three runs was used as the background to be subtracted from the obtained values.

^cNo isolable product could be detected on TLC from the reaction mixtures.

into a buffer solution up to a concentration of $500 \,\mu M$, while 2a did not dissolve completely even below 100 μ M, becoming a suspension. The observed DNA cleavage could be caused by the biradical through the phototriggered cycloaromatization reaction, however, the possibility of cleavage being due to the singlet oxygen that was yielded by the photosensitization of a nitrophenyl moiety could not be excluded. To clarify this point, we performed a DNA cleaving assay of the reference compounds 4 under the same conditions. Although the reference carbonate 4 damaged DNA to some extent, the degree of cleavage was around 10% (entry 8) and therefore the potent DNA cleavage exhibited by enediyne 2b should be mainly attributed to the oxidative damage caused by the photoreleased biradicals.¹¹

In conclusion, we developed enediyne model compounds bearing photo-triggering devices and showed that these synthetic enediynes exhibited potent DNA damaging activity under UV irradiation conditions. We also clarified that the biradical formation played the important role in the cleavage of DNA in our systems.¹² Further improvement of the model compounds is now underway.

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- 11. Mechanistically, the σ -radical part of the biradical instead of the resonance-stabilized π -radical part would be responsible for this DNA scission, although the alkylation pathway cannot be ruled out.
- 12. Since the DNA damage caused by **2a** was of the similar degree to the case of **4**, we cannot attribute the DNA damage to only the biradical.