

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 16 (2006) 5700-5703

Inhibition of P-glycoprotein-mediated multidrug efflux by aminomethylene and ketomethylene analogs of reversins

Ali Koubeissi,^a Imad Raad,^a Laurent Ettouati,^{a,*} David Guilet,^a Charles Dumontet^b and Joelle Paris^{a,*}

^aUniversité Claude Bernard Lyon 1, Institut des Sciences Pharmaceutiques et Biologiques, EA 3741 Écosystèmes et Molécules bioactives, 69373 Lyon cedex 08, France ^bUniversité Claude Bernard Lyon 1, Faculté de Médecine, Laboratoire de Cytologie analytique, INSERM U590, 69373 Lyon cedex 08, France

> Received 28 June 2006; revised 17 July 2006; accepted 17 July 2006 Available online 6 September 2006

Abstract—Several aminomethylene analogs and a ketomethylene analog of reversins were synthesized in order to evaluate their ability to inhibit P-glycoprotein-mediated drug efflux in K562/R7 human leukemic cells overexpressing P-glycoprotein. These analogs retained good activity compared to cyclosporin A and the original reversins. © 2006 Elsevier Ltd. All rights reserved.

Multidrug resistance (MDR) to anticancer agents remains a major cause of treatment failure in cancer chemotherapy. MDR describes the cross-resistance of tumor cell lines to several structurally unrelated chemotherapeutic agents after exposure to a single cytotoxic drug. This phenomenon is often associated with overexpression of several proteins.¹ Among them P-glycoprotein (Pgp) is the most important one that belongs to the ABC superfamily of transporters which acts as a drug efflux pump.^{2,3}

Numerous molecules have shown some activity on Pgp.⁴ Among them short linear hydrophobic peptides were described as chemosensitizers.^{5a} Seprõdi et al. showed that small hydrophobic peptide derivatives modulate Pgp-ATPase activity and inhibited the drug extrusion function of Pgp.^{5b,c} These compounds are a family of di- and tripeptide derivatives sharing some common physico-chemical and structural features. Some of them are dimerized aminoacids from diacid derivatives. The enhanced affinity to Pgp of these chemosensitizers finally coined reversins is ascribed to the hydrophobic nature of the side chains protected with bulky aromatic or alkyl groups.⁶ Among them reversin 121 **1a** showed the highest affinity and specificity for Pgp.

At $1-2 \mu M$ this reversin was more effective than cyclosporin A for blocking colchicine transport in isolated membranes and reconstituted systems. In order to improve proteolytic stability and bioavailability of reversins, we planned to synthesize aminomethylene and



Scheme 1. Reagents and condition: (a) NaBH₃CN, MeOH, AcOH, rt, 1 h, 52% (n = 1) and 57% (n = 2).

Keywords: Multidrug resistance; P-glycoprotein; Reversin; Aminomethylene; Ketomethylene; Pseudopeptide; Chemosensitizer.

^{*} Corresponding authors. Tel.: +33 4 78 77 70 82; fax: +33 4 78 77 75 49; e-mail addresses: laurent.ettouati@univ-lyon1.fr; joelle.paris@ univ-lyon1.fr

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2006.07.059

ketomethylene analogs of typical reversin representatives. We chose reversin 121 **1a** as our working model for dipeptide-type reversins and reversin 213 **2a** as a dimerized aminoacid containing succinyl unit. The reduced analogs **3a–b** were synthesized using the classical reductive amination strategy⁷ starting from chiral poolderived aminoaldehydes according to Scheme 1.⁸



As far as reduced analogs of reversin 213 2a were concerned we started from 4-pentenoic acid which was activated as succinimidyl ester and then coupled to Lglutamic acid dibenzyl ester tosylate salt to obtain the amide 4 (Scheme 2). The corresponding aldehyde 6 was obtained by a two-step reaction with osmium tetroxide/4-methylmorpholine N-oxide⁹ followed by oxidative cleavage of the diol 5 with sodium periodate. It is noteworthy that aldehyde 6 exists in equilibrium with the corresponding hemiaminals 6' as previously described in similar reactions.^{10a,b} Structure of the diastereoisomeric hemiaminals 6' was assessed by ESIMS and HSOC and HMBC experiments.^{10c} The last step consisted in obtaining the reduced analogs 8a-c of reversin 213 2a. However in the standard reductive amination conditions, the reaction between diprotected L-glutamic acid 7b and aldehyde 6 did not afford the corresponding secondary amine but instead the pyrrolidinone 9 in 38% yield (Fig. 1). This result is explained by nucleophilic attack of the generated secondary amine on the adjacent benzyl ester side chain as observed for similar derivatives.¹¹ So as to avoid cyclization, several protected derivatives of L-glutamic acid such as the 2,4-dimethyl-3-pentyl $(Dmp)^{12a}$ and cyclohexyl $(cHex)^{12b}$ esters 7c and 7d^{12c} known to prevent aspartimide formation were used to obtain, respectively, analogs 8b and 8c. Moreover by using an excess of aldehyde 6 a double addition product 10 was obtained in 47% yield in the case of reductive amination of L-glu-



Figure 1. Structures of by-products 9 and 10 obtained respectively in the reductive amination of 7b and 7c with aldehyde 6.

tamic acid derivative **7c** (Fig. 1).¹³ Undescribed derivatives **1b** and **1c** along with compound **2b** were also synthesized by standard procedures for sake of comparison.

We thought it would be worthwhile to test the introduction of a spacer between the Asp and Lys residue of reversin 121 1. Thus, we chose to synthesize the protected Asp- ψ (CO-CH₂)Gly-Lys ketomethylene analog 16 as outlined in Scheme 3. The synthesis started from L-homoserine which was protected on the side chain as a tert-butyl dimethyl silyl ether followed by protection of the free α -amine as *tert*-butyloxycarbonvl derivative and finally transformed in Weinreb amide 11 in 40% overall yield over three steps. The alkene 12 was then obtained from Weinreb amide 11 by alkylation with butenyl magnesium bromide as described¹⁴ in 55% yield. Transformation of the protected hydroxyl of 12 in benzyl ester 13 was carried out by oxidation of the deprotected hydroxyl with PDC and then esterification of the cesium salt in 53% overall yield as described.^{15a,15b} Oxidation of alkene 13 with Ru-Cl₃:xH₂O/NaIO₄ afforded free acid 14 in 96% yield. The ketomethylene analog 16 was finally obtained by coupling the succinimidyl derivative 15 and diprotected L-lysine hydrochloride in 94% yield.

The efficiency of reversin analogs to inhibit Pgp-mediated daunorubicin efflux was investigated by monitoring the intracellular accumulation of this drug in K562/R7 human leukemic cells overexpressing Pgp in the presence of daunorubicin.^{16a} Cyclosporin A was used as a positive control (Table 1). These analogs showed strong inhibitory activity comparable to that of reversin 121 **1a** except in the case of the tertiary amine **10** which caused an important decrease in activity. It was noteworthy that the replacement of the amide bond with



Scheme 2. Reagents and conditions: (a) *N*-hydroxysuccinimide, DMAP, DCC, THF, 0 °C, 10 min then rt, 48 h, 94%; (b) DIEA, DMF, rt, 24 h, quant yield; (c) OsO₄ 2.5% in *t*-BuOH, NMO, THF, H₂O, rt, 24 h, 86%; (d) NaIO₄, THF, H₂O, 3 h, quant yield; (e) NaBH₃CN, MeOH, AcOH, rt, 1–2 h.



Scheme 3. Reagents and conditions: (a) TBDMSCl, DBU, CH₃CN, 0 °C then rt, 24 h, 82%; (b) (Boc)₂O, Et₃N, acetone, H₂O, rt, 24 h, 80%; (c) *N*, *O*-dimethylhydroxylamine hydrochloride, TBTU, HOBt, DIEA, CH₂Cl₂, rt, 12 h, 61%; (d) butenyl magnesium bromide, THF, -78 °C then 3.5 h, rt, 30 min, 55%; (e) TBAF, THF, rt, 1 h, 87%; (f) PDC, DMF, rt, 3 h, 71%; (g) i—Cs₂CO₃, MeOH, H₂O then reduced pressure; ii—benzyl bromide, DMF, rt, 4.5 h, 86%; (h) RuCl₃·xH₂O NaIO₄, H₂O, CH₃CN, rt 1 h, 96%; (i) *N*-hydroxysuccinimide, DMAP, DCC, THF, 0 °C, 10 min then rt, 48 h, 49%; (j) DIEA, DMF, rt, 24 h, 94%.

Table 1. Mean inhibitory activity (%) of reversins and synthesized analogs on human leukemic cells $K562/R7^{16b}$

Compound ^a	Mean inhibiting activity $\%$
1a Reversin 121	72.73 (±5.27)
1b	52.70 (±5.27)
1c	85.43 (±5.69)
2a Reversin 213	93.74 (±2.17)
2b	94.45 (±1.36)
3a	74.14 (±4.06)
3b	79.08 (±5.13)
8a	82.22 (±4.57)
8b	54.71 (±3.63)
8c	98.33 (±1.35)
10	3.35 (±3.44)
16	86.62 (±4.86)

^a Compounds were tested at a 10 µM concentration.

 b Cyclosporin A was used as positive control (mean inhibitory activity of 100%) at a final concentration of 2 $\mu M.$ Standard deviation is given in parentheses.

an amino methylene isostere did not impair the capacity to inhibit Pgp-mediated drug efflux (1a vs 3a and 2b vs 8c). Analogs with cyclohexyl ester presented an increased inhibitory activity when compared to 2,4-dimethyl-3-pentyl ester analogs (1b vs 1c and 8b vs 8c). Moreover, exchanging benzyl ester by cyclohexyl ester was tolerated as shown in Table 1 by comparing activity for compounds 2a and 2b. Finally, the inhibitory activity of ketomethylene analog 16 was conserved indicating that inserting a succinyl moiety into reversin 121 1a was not detrimental to the Pgp-mediated drug efflux inhibitory activity.

We have designed aminomethylene and ketomethylene analogs with inhibitory activity of Pgp-mediated drug efflux comparable to that of reversins 121 **1a** and 213 **2a**. We envisage now the synthesis of ketomethylene analogs of reversin 121 and modified side chain analogs to confirm and refine our first results.

References and notes

- Gottesman, M. M.; Fojo, T.; Bates, S. E. Nat. Rev. Cancer 2002, 2, 48.
- 2. Gottesman, M. M.; Pastan, I.; Ambudkar, S. V. Curr. Opin. Genet. Dev. 1996, 6, 610.
- Szakács, G.; Paterson, J. K.; Ludwig, J. A.; Booth-Genthe, C.; Gottesman, M. M. Nat. Rev. Drug Disc. 2006, 5, 219.
- Teodori, E.; Dei, S.; Scapecchi, S.; Gualtieri, F. *Il Farmaco* 2002, 57, 385.
- (a) Sharom, F. J.; DiDiodato, G.; Yu, X.; Ashbourne, K. J. D. J. Biol. Chem. 1995, 270, 10334; (b) Seprödi, J.; Mezõ, I.; Vadász, Zs.; Szabó, K.; Sarkadi, B.; Teplán, I. In Peptides 1996; Proceedings of the 24th European Symposium, 1996, Edimburgh; Ramage, R., Epton, R., Eds; Mayflower Scientific Ltd: Kingswinford, 1998; pp 801–802.; (c) Vadász, Zs.; Szabó, K.; Sarkadi, B.; Teplán, I.; Mák, M.; Miklós, I.; Györffy, E.; Seprödi, J. In Peptides 1998; Proceedings of the 25th European Symposium, 1998, Budapest; Bajusz, S., Hudecz, F., Eds; Akademiai Kiado Ed: Budapest, 1999; pp 640–641.
- Sharom, F. J.; Yu, X.; Lu, P.; Liu, R.; Chu, J. W. K.; Szabó, K.; Müller, M.; Hose, C. D.; Monks, A.; Váradi, A. E.; Seprödi, J.; Sarkadi, B. *Biochem. Pharmacol.* 1999, 58, 571.
- Martinez, J.; Bali, J.-P.; Rodriguez, M.; Castro, B.; Magous, R.; Laur, J.; Lignon, M.-F. J. Med. Chem. 1985, 28, 1874.
- 8. The aminoaldehydes were synthesized by reduction of aminoacids-derived mixed anhydrides with $NaBH_4$ and then oxidized by Swern oxidation.
- 9. VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 17, 1973.
- (a) Baldwin, J. E.; Hulme, C.; Edwards, A. J.; Schofield, C. J.; Parkes, K. E. B. *Tetrahedron Lett.* **1993**, *34*, 1665;
 (b) Delcros, J.-G.; Tomasi, S.; Carrington, S.; Martin, B.; Renault, J.; Blagbrough, I. S.; Uriac, P. J. Med. Chem. **2002**, *45*, 5098; (c) Spectral data of diastereoisomeric hemiaminals **6**': IR (film, *v*, cm⁻¹): 3412 (OH and NH), 3066 (arom CH), 2954 (al CH), 1738 (C=O ester), 1674 (CO amide). ESIMS (negative mode, NaCl): 856,9 (2M+Cl⁻), 446,0 (M+Cl⁻). ¹H NMR (300 MHz, CDCl₃, *δ* ppm): 7.36 (20H, m, 4-Phe), 5.31 (2H, m, 2 CHOH), 5.18

(4H, m, 2 CH₂O Bn), 5.12 (4H, m, 2 CH₂O Bn), 4.77 (1H, dd, J = 4.5 and 9.8 Hz, CH α), 4.64 (1H, t, J = 7.1 Hz, CH α), 4.32 (1H, d, J = 4.5 Hz, exch OH), 3.41 (1H, d, J = 7.8 Hz, exch OH), 1.8–2.8 (16H, m, 2 –CH₂CH₂– and 2 –CH₂CH₂– Glu). ¹³C NMR (75 MHz, CDCl₃, δ ppm): 176.3 (C=O), 175.8 (C=O), 173.7 (C=O), 173.1 (C=O), 172.8 (C=O), 171.0 (C=O), 136.0 (Cq Phe), 135.9 (Cq Phe), 135.5 (Cq Phe), 135.2 (Cq Phe), 129.1 (CH–Phe), 129.07 (CH–Phe), 129.02 (CH–Phe), 128.9 (CH–Phe), 128.8 (CH–Phe), 128.7 (CH–Phe), 128.6 (2 CH–Phe), 84.4 (CHOH), 82 (CHOH), 68.3 (CH₂O), 67.9 (CH₂O), 67.1 (CH₂O), 67.0 (CH₂O), 54.7 (CH α), 54.3 (CH α), 31.3 (CH₂), 30.0 (CH₂), 29.5 (CH₂), 29.3 (CH₂), 29.0 (CH₂), 28.8 (CH₂), 25.7 (CH₂), 24.6 (CH₂).

- Gareau, Y.; Zamboni, R.; Wong, A. W. J. Org. Chem 1993, 58, 1582.
- 12. (a) Karlström, A. H.; Undén, A. E. *Tetrahedron Lett.*1995, 36, 3909; (b) Tam, J. P.; Wong, T.-W.; Riemen, M. W.; Tjoeng, F.-S.; Merrifield, R. B. *Tetrahedron Lett.*1979, 42, 4033; (c) H-Glu(ODmp)-OBn 7c was synthesized from commercial N^αBoc-Glu-OBn by side-chain esterification with DCC/DMAP and 2,4-dimethyl-3-pentanol in dichloromethane in 70% yield and then Boc-deprotected with conc H₂SO₄ in AcOEt in 95% yield. H-Glu(O*C*Hex)-OBn 7d was obtained from commercial N^αBoc-Glu(O-cHex)-OH by esterification of the α-carboxylic acid with

benzyl bromide and cesium carbonate in MeOH/ H_2O in 70% yield and then Boc deprotected in the same conditions as previously in 96% yield.

- 13. See Salvi, J. P.; Walchshofer, N.; Paris, J. *Tetrahedron Lett.* **1994**, *35*, 1181, for similar formation of double condensation product in reductive amination reactions.
- Kaiser, M.; Siciliano, C.; Assfalg-Machleidt, I.; Groll, M.; Milbradt, A. G.; Moroder, L. Org. Lett. 2003, 5, 3435.
- 15. (a) Våbenø, J.; Nielsen, U.; Ingebrigtsen, T.; Lejon, T.; Steffansen, B.; Luthman, K. J. Med. Chem. 2004, 47, 4755;
 (b) As racemization of a similar benzylated ester was noted in the cited reference 15a the enantiomeric purity of 13 was checked and found to be enantiomerically pure by C18 RP-HPLC analysis after deprotection (TFA/CH₂Cl₂) and derivatization with GITC.
- 16. (a) Comte, G.; Daskiewicz, J.-P.; Bayet, C.; Conseil, G.; Viornery-Vanier, A.; Dumontet, C.; Di Pietro, A.; Barron, D. J. Med. Chem. 2001, 44, 763; (b) One million K562/R7 human leukemic cells expressing high levels of P-glycoprotein were incubated for 1 h at 37 °C in 1 mL of RPMI 1640 medium containing a final concentration of 10 μM daunorubicin, in the presence or absence of inhibitor. The cells were then washed twice with ice-cold phosphatebuffered saline (PBSt) and kept on ice until analysis by flow cytometry on a FACS-II. Assays were performed in duplicate, with at least three separate experiments.