SYNTHESES OF BIOLOGICALLY ACTIVE SIALOSYLGLYCEROL DERIVATIVES

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New sialosylglycerol derivatives were synthesized and found to inhibit the phospholipase A_2 and C activities

KEYWORDS sialosylglycerol derivative; phospholipase A, inhibitor; phospholipase C inhibitor; polysaccharide; hexadecanoic acid

A constituent of bacterial cell wall has various biological activities such as immnological response, phage receptor, endotoxin etc.. In 1981, the capsular polysaccharide of Neisseria meningitidis, which plays the main antigen role, was structurally defined (Fig 1). The polysaccharide consists of a polymer of ($\alpha 2 \rightarrow 9$) sialic acid, which has many important functions as constituents of glycoconjugate, and phosphoglycerolipid. Recently we have synthesized a series of biologically active compounds designed on the basis of the chemical structure of bacterial cell wall. Here, we describe the synthesis of the sialosylglycerolipids ($1\alpha_{n-d}$, $1\beta_{n-d}$, $2\alpha_n$, and $2\beta_n$) which imitate the partial structure of the capsular polysaccharide. The synthetic design of these compounds was determined as follows. The absolute configuration of the glycerol C-2 (S) was the same as that of the natural product and the 2-hydroxyl glycerolipid (lyso type) was expected to inhibit phospholipase A, by feedback regulation. Four kinds of fatty acid (a-d) were studied to determine the differences among their biological activities due to the fatty acid type. Similarly, sialosyl-(R)-glycerol derivatives were studied with regard to their palmitoyl type.

HOW OH COOH

HOW OH

ACN
HO WHO WHO

Fig 1

COOH

$$COOH$$
 $COOH$
 OOH
 OOH

Chart 1 shows the synthetic route of the sialosyl-(S)-glycerol derivatives. (S)-1-0-Acetyl-2-0-benzylglycerol (3), 3) the chiral starting material was treated with trityl chloride and pyridine to give the 3-tritylated compound (4; 72.5%, oil, $[a]_D$ +11.3°). Alkaline hydrolysis of the acetate (4) led to the (R)-1-0-trityl-2-0-benzylglycerol derivatives (5; 77.4%, mp 59-62 C°, $[a]_D$ +22.5°). The 3-hydroxyl compound (5) was acylated with acyl chlorides and triethylamine to yield $6_{\bullet-4}$ (6_{\bullet} ; 85.0%, oil,

Chart 1

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2260 Vol. 37, No. 8

 $[a]_D + 12.6^\circ$, 6_b ; 93.7%, oil, $[a]_D + 11.6^\circ$, 6_c ; 86.0%, oil, $[a]_D + 4.7^\circ$, 6_d ; 88.8%, oil, $[a]_D + 9.3^\circ$). The trityl group of 6_{a-d} was removed by hydrolysis with 80% aqueous acetic acid at 80°C to afford (S)-1-0-acyl-2-0-benzylglycerols (S-7_{a-d}, S-7_a; 74.6%, oil, $[a]_D$ -4.8°, S-7_b; 80.3%, oil, $[a]_D$ -5.7°, S-7_b; 80.3%, oil, $[a]_D$ -5.8°, $[a]_D$ 7_c ; 73.9%, oil, $[a]_D$ -6.4°, S-7_a; 85.8%, oil, $[a]_D$ -8.0°). These were used as the glycosyl acceptor. (R)-Glycerol derivative (R- 7_a) was synthesized in the following way. The 1-O-hydroxyl compound (5) was chloroacetylated to give 8 (91.5%, oil, $[a]_D$ +10.7°). Detritylation of the chloroacetyl compound (8) gave the 3-hydroxyl compound (9, 69.0%, oil, $[a]_D$ -3.1°). The compound (10, 70.1%, oil, $[a]_D$ +2.8°) was obtained by treating 9 with palmitoyl chloride and triethylamine. Dechloroacetylation of 10 was achieved with diisopropylethylamine and thiourea to yield the (R)-glycosyl acceptor (R- 7_a , 92.7%, oil, $[a]_D$ +4.7°). S-7_{a-d} was glycosylated with the glycosyl donor (11) in the presence of $Hg(CN)_2$, $HgBr_2$ and molecular sieves 4A to give $12t_{a-d}$ and $12t_{a-d}$, respectively. anomeric mixture was separated by preparative TLC (CHCl₃-MeOH = 20:1) and their structures were confirmed on the basis of the chemical shift of the H-3eq atom of $12a_{a-d}$ and $12\beta_{a-d}$ in the ¹H-NMR spectrum: the H-3eq chemical shift of a glycoside was lower than that of β glycoside (12a, 26.9%, amorphous, $[a]_D$ -8.5°, $12\beta_a$; 32.4%, amorphous, $[a]_D$ -5.5°, $12\alpha_b$; 10.8%, amorphous, $[a]_D$ -3.7°, $12\beta_b$; 11.5%, amorphous, $[a]_D$ -6.6°, $12a_c$; 15.6%, amorphous, $[a]_D$ -18.2°, $12\beta_c$; 27.0%, amorphous, $[a]_D$ - 10.7° , $12\alpha_{d}$; 17.1%, amorphous, $[\alpha]_{D}$ -16.7°, $12\beta_{d}$; 23.5%, amorphous, $[\alpha]_{D}$ -8.2°). $12\alpha_{d-d}$ and $12\beta_{d-d}$ were hydrogenolyzed with Pd(OH)₂/C in methanol to give the sialosyl-(S)-glycerol derivatives (1g_{a-d} and $1\beta_{a-d}$, $1\alpha_a$; 71.2%, amorphous, $[\alpha]_D$ -11.3°, $1\beta_a$; 74.1%, amorphous, $[\alpha]_D$ -12.6°, $1\alpha_b$; 95%, amorphous, $[a]_D$ -8.0°, $1\beta_D$; 87%, amorphous, $[a]_D$ -10.0°, $1\alpha_C$; 95%, amorphous, $[a]_D$ -7.8°, $1\beta_C$; 96%, amorphous, $[a]_D$ -5.1°, $[a]_B$ -6.0°).4-5)

Sialosyl-(R)-glycerol derivatives (2α and 2β) were synthesized in the same manner. The (R)-1-0-hexadecanoyl-2-0-benzylglycerol (R-7_a) was glycosylated with 11 in the presence of Hg(CN)₂, HgBr₂, and molecular sieves 4A to give an anomeric mixture of sialosylglycerol compounds (13α and 13β , 13α ; 24.6%, amorphous, $[\alpha]_D$ -10.8°, 13β ; 20.9%, amorphous, $[\alpha]_D$ -7.0°). 13α and 13β were each hydrogenolyzed with Pd(OH)₂/C to yield the sialosyl-(R)-glycerol derivatives (2α and 2β , 2α ; 86.2%, amorphous, $[\alpha]_D$ -3.6°, 2β ; 84.8%, amorphous, $[\alpha]_D$ -4.8°).

Preliminary examination of the biological activities revealed that the lysosialosylpalmitoylglycerol derivatives ($1a_a$, $1\beta_a$, $2a_a$, and $2\beta_a$) have the most powerful phospholipase A₂ and phospholipase C inhibitory activities among the investigated sialosyl derivatives.⁶⁻⁷⁾

REFERENCES AND NOTES

- 1. E.C. Gotschlich, B.A. Frasser, O. Nashimura, J.B. Robbins, and T.-Y. Lui, J. Biol. Chem. <u>256</u>, 8915 (1981).
- 2. C. Shimizu, K. Ikeda, and K. Achiwa, Chem. Pharm. Bull., 36, 1772 (1988) and the references cited therein.
- 3. Y. Terao, M. Murata, K. Achiwa, T. Nishio, M. Akamatsu, and M. Kamimura, Tetrahedron Lett. $\underline{29}$, 5173 (1988).
- 4. ¹H-NMR of H-3eq (ppm, J=Hz): $12\alpha_a$; 2.59 (1H, dd, J=4.9, 12.9), $12\alpha_b$; 2.61 (1H, dd, J=4.4, 12.4), $12\alpha_c$; 2.61 (1H, dd, J-4.9, 12.4), $12\alpha_d$; 2.61 (1H, dd, J=4.6, 12.4), 13α ; 2.61 (1H, dd, J=4.6, 12.7): Because H-3eq signals of $12\beta_{a-d}$ and 13β were overlapped by the methylene protons of fatty acid, the evidence of β linkage was the fact that H-3eq signals were not found at downfield less than 2.49 ppm. 5. J. P. Kamerling, L. Dorland, H. van Halbeek, J. F. G. Vliegenthart, Carbohydr. Res. 100, 331 (1989).
- 6. T. Miyazawa, S. Toyoshima, T. Osawa, I. Suda, M. Ito, and K. Tomita, Abstracts of Paper 109th Annual Meeting of the Pharmaceutical Society of Japan, V, p. 57 (1989).
- 7. C. Shimizu, K. Achiwa, T. Miyazawa, S. Toyoshima, and T. Osawa, Abstracts of Ith Japanese Carbohydrate Symposium, pp. 77-78 (1989).

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