Diastereoselectivity in the synthesis of D-glycero-D-aldoheptoses by 2-trimethylsilylthiazole homologation from hexodialdo-1,5-pyranose derivatives

NAVEEN K. KHARE,¹ RAMESH K. SOOD,² AND GERALD O. ASPINALL³

Department of Chemistry, York University, North York, Toronto, ON M3J 1P3, Canada

Received May 20, 1993

This paper is dedicated to Professors David B. MacLean and Ian D. Spenser

NAVEEN K. KHARE, RAMESH K. SOOD, and GERALD O. ASPINALL. Can. J. Chem. 72, 237 (1994).

An exploration of the synthesis of D-glycero-D-altro-heptose, a constitutent of O antigen chains in lipopolysaccharides from *Campylobacter jejuni* serotypes O:23 and O:36 led to a study of the 2-trimethylsilylthiazole homologation procedure for heptose synthesis. In contrast to the diastereoselective formation of a 1,2:3,4-di-O-isopropylidene-D-glycero- α -D-galacto-heptopyranose derivative from 1,2:3,4-di-O-isopropylidene- α -D-galacto-hexodialdo-1,5-pyranose, methyl 2,3,4-tri-O-benzyl-D-hexodialdo-1,5-pyranosides with the gluco and manno configurations showed no preference for the formation of compounds with the D-glycero configuration. Attempts to achieve high diastereoselectivity in the conversion of L-glycero into the D-glycero isomers by oxidation at C-6 followed by reduction with L-selectride were unsuccessful with the thiazole adducts, but the desired products were formed in similar reactions of methyl 2,3,4-tri-O-benzyl-7-O-tert-butyldimethylsilyl-D-heptopyranosides. The approach to homologation in the altro series was thwarted by epimerization at C-5 in the attempted formation of methyl 2,3,4-tri-O-benzyl- α -D-altro-hexodialdo-1,5-pyranoside. The successful synthesis of methyl D-glycero- α -D-altro-heptopyranoside from methyl α -D-glucopyranoside was achieved by homologation followed by configurational alteration from the D-gluco to the D-altro series.

NAVEEN K. KHARE, RAMESH K. SOOD et GERALD O. ASPINALL. Can. J. Chem. 72, 237 (1994).

Une exploration de la synthèse du D-glycéro-D-altro-heptose, un constituant des chaînes O antigéniques des lipopolysaccharides des sérotypes O:23 et O:36 du Campylobacter jejuni a conduit à étudier la possibilité d'utiliser la procédure d'homologation du 2-triméthylsilylthiazole pour réaliser la synthèse d'heptoses. Par opposition à la formation diastéréosélective d'un dérivé 1,2:3,4-di-O-isopropylidène-D-glycéro- α -D-galacto-heptopyranose à partir 1,2:3,4-di-O-isopropylidène- α -D-galacto-hexodialdo-1,5-pyranose, les 2,3,4-tri-O-benzyl-D-hexodialdo-1,5-pyranosides de méthyles avec des configurations gluco et manno ne présentent aucune préférence pour la formation de composés avec la configuration D-glycéro. Les essais effectués sur des adduits thiazoles dans le but d'obtenir une grande diastéréosélectivité lors de la conversion des isomères L-glycéro en D-glycéro par le biais d'une oxydation en C-6 suivie par une réduction à l'aide de L-sélectride se sont avérés vains; toutefois, les produits désirés se forment lors de réactions semblables avec des 2,3,4-tri-O-benzyl-7-O-tert-butyldiméthylsilyl-D-heptopyranosides de méthyle. L'approche à l'homologation dans la série altro ne peut être réalisée à cause d'une épimérisation en C-5 lorsqu'on essaie de former le 2,3,4-tri-O-benzyl- α -D-altro-hexodialdo-1,5-pyranoside de méthyle. On a réalisé la synthèse du D-glycéro- α -D-altro-heptopyranoside de méthyle à partir de l' α -D-glucopyranoside de méthyle en procédant à une homologation suivie d'une modification de configuration de la série D-gluco à D-altro.

[Traduit par la rédaction]

Introduction

L-glycero-D-manno-Heptose is a commonly encountered component of the core region of lipopolysaccharides (LPS) on Gram-negative bacteria (1). In view of the immunological importance of this region of LPS structure considerable attention has been directed to efficient methods for the preparation of derivatives of the sugar for incorporation into synthetic oligosaccharides (2-6). Methods of choice have been those involving a one-carbon extension from benzyl (1) or methyl (2) 2,3,4-tri-O-benzyl-α-D-manno-hexodialdo-1,5-pyranoside which that involving reaction with (phenyldimethylsilyl)magnesium chloride (4) proceeds with high diastereoselectivity to give derivatives of L-glycero-D-manno-heptose from which glycosides of the parent sugar may be readily obtained. In a recent study of the O antigen chains of LPSs of Campylobacter jejuni serotypes O:23 and O:36 unusual heptose sugars were found as four variants with the same configurationally unusual altro ring configuration (7). LPS preparations all contained trisaccharide,

 $-3-\beta$ -D-GlcNAc- $(1\rightarrow 3)-\alpha$ -D-Gal- $(1\rightarrow 2)-\alpha$ -Hep 1-, repeating units with the same linkages and anomeric configurations, but with the heptose residue present in varying proportions as 6-deoxy-D-altro-heptose (8) and 6-deoxy-3-O-methyl-D-altroheptose (unpublished synthesis with G.V. Reddy), for which enantiomeric configurations have been confirmed, and D-glycero-D-altro-heptose and 3-O-methyl-D-glycero-D-altro-heptose or their respective enantiomers. In view of the close structural, and presumably also biosynthetic, relationship between the four variants it was highly probable that all had the α -D-altro ring configuration. It was still necessary to confirm the absolute configurations of the two latter sugars through the synthesis of defined reference compounds before making preparative quantities for oligosaccharide assembly. The only reported C-6 homologation of a D-hexodialdo-1,5-pyranose derivative proceeding with high diastereoselectivity for the formation of the *D*-glycero product is that reported by Dondoni et al. (5, 6) using 2-(trimethylsilyl)thiazole (TMSthiazole) as a one-carbon synthon in reaction with 1,2:3,4-di-O-isopropylidene- α -D-galacto-hexodialdo-1,5-pyranose (3). This procedure therefore seemed worthy of exploration for the synthesis of D-glycero-D-altro-heptose derivatives. Two alternative overall strategies seemed possible starting from D-glucopyranose derivatives: (A) 2,3-inversion to gain entry into the Daltropyranose series, followed by C-6 homologation; and (B)

¹On leave of absence from the Department of Chemistry, Lucknow University, Lucknow, India, 1991–1992.

²Present address: Univax Biologicals Inc., 12280 Wilkins Ave., Rockville, MD 20852, U.S.A.

³Author to whom correspondence may be addressed.

CAN. J. CHEM. VOL. 72, 1994

Parent sugar	Relative retention times ^a of heptitol acetates with respect to		
	Glucitol hexaacetate	Galactitol hexaacetate	
L-glycero-D-manno-Heptose	1.90	2.08	
D-glycero-D-manno-Heptose	1.65	1.80	
L-glycero-D-gluco-Heptose	2.07	2.26	
D-glycero-D-gluco-Heptose	1.75	1.90	
L-glycero-D-galacto-Heptose	2.06	2.28	
D-glycero-D-galacto-Heptose	1.90	2.08	
D-glycero-D-altro-Heptose	1.47	1.61	
3-O-Methyl-D-glycero-D-altro-heptose	1.52	1.66	

TIDICI	Cas abromata anombia	omolucio	of homeital	
IABLE I.	Gas chromatographic	analysis	or nephtor	acetates

^aGas chromatography using program A.

C-6 homologation in the D-glucopyranose series, followed by 2,3-inversion of the D-glycero-D-gluco-heptopyranose product. In the event unexpected problems were encountered in both proposed routes. Route A was rejected at an early stage when it was observed that oxidation of methyl 2,3,4-tri-O-benzyl-a-D-altropyranoside to give the dialdohexo-1,5-pyranoside was accompanied by substantial epimerization at C-5 with formation of the corresponding β -L-galacto derivative. Route B ultimately proved successful for the preparation of the desired heptoses but no diastereoselectivity was observed in the C-6 homologation reaction. We report herein syntheses of these heptoses in confirmation of their absolute configurations, but discuss first diastereoselectivity in the TMSthiazole homologation together with experiments related to the C-5 epimerization accompanying oxidation at C-6 of both altrose and galactose derivatives.

Results and discussion

Configurational assignments for heptose derivatives epimeric at C-6

All homologations were performed and the thiazole adducts were converted into heptose derivatives as described by Dondoni et al. (5, 6) and O-debenzylated. Since all 10 diastereomeric heptitol heptaacetates are separable by gas-liquid chromatography (glc) (9) (see Table 1 for examples now reported), formation of these derivatives by hydrolysis of glycosides, reduction, and acetylation was the simplest method for distinguishing the pairs of D-glycero/L-glycero epimers that may result from C-6 homologations. Other, sometimes quantitatively dominant, compounds are produced in varying amounts due to accompanying 1,6- and (or) 1,7-anhydroheptose formation under hydrolysis conditions, as in the case of D-glycero-Daltro-heptose (7). In the example reported by Dondoni et al. (5, 6), the absolute configuration at C-6 of the thiazole adduct, (6S)-1,2:3,4-di-O-isopropylidene-6-(2-thiazolyl)-α-D-galactohexo-1,5-pyranose (4), formed as the sole product from 1,2:3,4di-O-isopropylidene-D-galacto-hexodialdo-1,5-pyranose (3) was established by conversion into a previously characterized D-glycero- α -D-galacto-heptose derivative (10). However, derivatives of defined configuration from rarely encountered heptoses are unlikely to be available for comparison. Furthermore for the compounds of present interest, we have found that the differences in nmr parameters for C-6 and H-6 in epimeric heptose derivatives are insufficiently consistent to provide a basis for configurational assignments.

Diastereoselectivity in C-6 homologation of mannose and glucose derivatives

With access to heptitol heptaacetates derived from L-glyceroand D-glycero-D-manno-heptoses for comparison, the TMSthiazole homologation reaction was examined as an alternative to the above-mentioned homologation of 1 with (phenyldimethyl)silylmethylmagnesium chloride, which had been used in the preparation of derivatives of L-glycero-D-manno-heptose with high diastereoselectivity (3, 4). Diastereoselectivity in homologation was first examined for methyl 2,3,4-tri-O-benzyl- α -Dmanno-hexodialdo-1,5-pyranoside (2), prepared as required on Swern oxidation (11) of methyl 2,3,4-tri-O-benzyl-a-D-mannopyranoside with oxalyl chloride and methyl sulfoxide. Reaction of 2 with TMSthiazole gave the chromatographically separable 6-(2-thiazolyl)-derivatives (5 and 6) in respective yields of 55 and 37%. Conversion of 5 and 6 into the respective methyl 2,3,4-tri-O-benzylheptopyranosides (7 and 8), in four steps with overall yields of 60-70%, by successive N-methylation, reduction with NaBH₄, hydrolysis in the presence of $HgCl_2$, and reduction again with NaBH₄. Since this research was completed Dondoni et al. (12) have reported an improved procedure for the thiazole to formyl deblocking protocol. Configurational assignments for 7 and 8 were made by conversion into L-glycero- and D-glycero-D-manno-heptitol heptaacetates.

TMSthiazole homologation of the hexodialdo-1,5-pyranoside (9) formed on oxidation of methyl 2,3,4-tri-O-benzyl- α -Dglucopyranoside was next explored with very similar results to those in the mannose series. Reaction of 9 with TMSthiazole furnished a mixture of 6-(2-thiazolyl) derivatives from which 10 and 11 were obtained in chromatographically pure form in yields of 54 and 18%. These compounds were converted into the corresponding methyl 2,3,4-tri-O-benzylheptopyranosides 12 and 13, as described above, and thence after O-debenzylations into the respective methyl glycosides 14 and 15. Compounds 14 and 15 (and similarly others elsewhere) were converted into their acetylated derivatives 16 and 17 to provide satisfactory elemental analyses and to give well-dispersed one-dimensional ¹H nmr spectra for structural assignments. Relative configurations were assigned after conversion of 14 and 15 into heptitol heptaacetates that were indistinguishable on glc from those derived respectively from L-glycero- and D-glycero-D-gluco-heptitols.

Configurational inversions at C-6 of heptose derivatives

In the light of the preceding results attention was turned to the



Can. J. Chem. Downloaded from www.nrcresearchpress.com by 120.117.138.77 on 11/09/14 For personal use only.

possibility of achieving a high-yield conversion of L-glycero into the epimeric D-glycero compounds. Dondoni et al. (13) have described inversion of configuration at C-6 of the (2-thiazolyl) derivative 4 by oxidation to the 6-ulose followed by stereoselective reduction with L- or K-selectride to give the (6R) epimer (18).

With formation of the L-glycero compound 7 as the major product of TMSthiazole homologation in the gluco series the possibility of achieving improved overall yields of D-glycero-D-gluco-heptose derivatives by configurational inversions at C-6 was examined for three derivatives. The 6-(2-thiazolyl) derivative 10 was oxidized with periodinane (14) and the resulting 6-ulose, without purification, was treated with L-selectride to regenerate a mixture of thiazolyl derivatives 10 and 11, now in the proportions of 2:3. Alternative methods for the configurational inversion were explored for 7-tert-butyldiphenylsilyl (TBDPS) and 7-tert-butyldimethylsilyl (TBDMS) derivatives of 12 and 13. In the case of a preliminary experiment performed on TBDPS derivatives from an unresolved mixture of 12 and 13, Swern oxidation (11) to give the 6-ulose followed by reduction with L-selectride gave back derivatives of 12 and 13 in the same proportions. Samples of the respective TBDMS derivatives (19 and 20) of 12 and 13 were prepared so that the reduction of the 6-ulose could be monitored, and complete conversion of 19 to 20 was achieved when 19 was submitted to the oxidation-reduction sequence. A single product was detected by tlc as reduction of the 6-ulose with L-selectride proceeded, but when the reaction mixture was quenched with alkaline hydrogen peroxide, a mixture of products was formed. The chromatographic mobilities of these substances indicated retention of a silvl substituent. Since treatment of these substances with tetrabutylammonium fluoride gave 13 as the sole product, it is probable that intramolecular silyl migration, from O-7 to O-6, had taken place.

In the manno series, preparation of the TBDMS derivative 21 from 7 was followed by Swern oxidation (11) to the 6-ulose, reduction with L-selectride, and O-desilylation to give the D-glycero epimer 8 as the only detectable product and in 80% isolated yield.

Diastereoselectivity in C-6 homologation of altrose and galactose derivatives

Treatment of methyl 2,3,4-tri-O-benzyl- α -D-altropyranoside with oxalyl chloride and methyl sulfoxide gave a mixture of hexodialdo-1,5-pyranosides, as judged by the appearance of two aldehydic protons in the ¹H nmr spectrum and, most prominently in the ¹³C nmr spectrum, of two sets of resonances assignable to ring carbons of the expected methyl 2,3,4tri-O-benzyl-a-D-altro-hexodialdo-1,5-pyranoside (22) and the C-5 epimer, methyl 2,3,4-tri-O-benzyl-B-L-galacto-hexodialdo-1,5-pyranoside (23) in the approximate ratio of 1:4. The correctness of this conclusion was confirmed by reduction of the mixture of hexodialdo-1,5-pyranosides with NaBH₄ followed by acetylation to give a mixture of methyl 6-O-acetyl-2,3,4-tri-O-benzyl-hexopyranosides in the somewhat altered ratio of 2:3 with glc retention times coincident with those of standard α -D-altrose and β -D-galactose derivatives. Confirmation of the L-galacto configuration for the main component was obtained by O-debenzylation of the mixture of tri-O-benzyl glycosides followed by hydrolysis and conversion of the resulting reducing sugars into acetates of chiral 2-butyl glycosides (15). These results confirmed that epimerization at C-5 had occurred, leading to the formation of the β -L-galacto isomer as the major component of the mixture of hexodialdo-1,5-pyranosides. Mixtures of 22 and 23 in the same proportions were obtained on oxidation of methyl 2,3,4-tri-Obenzyl- α -D-altropyranoside with periodinane (14), indicating that the epimerization was not just a base-catalyzed reaction promoted by the use of triethylamine in the Swern oxidation.

A rapid interconversion of 22 and 23, with the composition of isomers on further reaction dependent on the relative rates of formation of different products, was indicated in the abovementioned reduction to give stereoisomeric products in changed proportions. In an exploration of the stereochemical consequences of the C-6 homologation with TMSthiazole, reaction of the mixture of 22 and 23 gave only one detectable product, which was assigned the structure methyl (6R)-2,3,4-tri-O- benzyl-6-(2-thiazolyl)- β -L-galactopyranoside (24) on the basis of the following evidence. The ¹³C nmr spectra of 24 and the derived methyl 2,3,4-tri-O-benzyl-L-glycero-



 β -L-galacto-heptopyranoside (25) showed resonances for carbon atoms of the β -galacto ring configuration and none for those of the α -altro configuration. O-Debenzylation of 25 followed by hydrolysis, reduction, and acetylation gave a single heptitol heptaacetate coincident on glc with that formed from the enantiomeric L-glycero-D-manno-heptose (= D-glycero-D-galacto-heptose) and therefore confirmed the relative configuration of the chiral centres.

The reversibility of the *altro-galacto* interconversion was also examined in the formation of hexodialdo-1,5-pyranose derivatives in the more accessible D-galactose series. In a reexamination of the oxidation of 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose as reported by Dondoni et al. (5, 6), the ¹H and ¹³C nmr spectra of the corresponding hexodialdo-1,5-pyranose (3) showed only the galacto isomer with no trace of the altro derivative. TMSthiazole homologation likewise gave only the reported (6S)-1,2:3,4-di-O-isopropylidene-6-(2-thiazolyl)- α -D-galactopyranose (4). In confirmation of this observation, independent evidence for the configurational assignments at C-6 in 4 and in its C-6 epimer (18) was obtained by their conversions into 1,2:3,4-di-O-isopropylidene-D-glycero-D- galactoheptopyranose (26) and the corresponding L-glycero-D-galacto compound (27), and thence into heptitol heptaacetates that were identical (gc) to samples prepared from the corresponding D-glycero-D-galacto-heptose and L-glycero-D-gluco-heptose (= L-glycero-D-galacto-heptose).

As expected, a parallel series of oxidation and TMSthiazole homologation reactions starting from methyl 2,3,4-tri-Obenzyl- β -D-galactopyranoside afforded compounds enantiomeric with those described above that were derived from methyl 2,3,4-tri-O-benzyl- α -D-altropyranoside. Comment here is restricted to a few salient points and details are given later. Oxidation of the D-galactopyranoside gave the corresponding mixture of *galacto* and *altro* dialdohexopyranosides in the ratio of 3:1 and, as expected, only a single product (**28**) with the D-glycero- α -D-galacto configuration was isolated from the ensuing TMSthiazole homologation. Although there were small differences in the magnitudes of the optical rotations of **28** and the corresponding methyl 2,3,4-tri-O-benzylheptopyranoside **29**, compared with those of the L-galacto compounds **24** and **25**, the enantiomeric pairs were chromatographically indistinguishable and ¹H and ¹³C nmr spectra showed no significant differences.

The stereoselectivity of configurational inversion at C-6 of 28 by oxidation and subsequent reduction with L-selectride was also examined. Two fractions were separated by preparative tlc and the structures of the products, which were obtained in insufficient quantity and purity for elemental analysis, were established from nmr spectra data and conversions to the corresponding heptitol heptaacetates. The nmr spectra of the derivative from the first fraction showed the presence of a single component with the galacto configuration, and further processing furnished L-glycero-D-galacto-heptitol heptaacetate derived from the product of C-6 inversion. The ¹³C nmr spectrum of the second fraction showed the presence of compounds with α -altro and β -galacto configurations, and further processing of the fraction afforded a mixture of two heptitol heptaacetates. Comparison of their glc retention times with those of reference compounds with the D-glycero-D-galacto and the D-glycero-D-altro configurations showed that the second fraction of the oxidation-reduction sequence consisted of the product from unchanged starting compound 28 and only one of two possible products arising from epimerization at C-5 of the 6-ulose, that with the L-glycero-L-altro configuration.

Synthesis of derivatives of D-glycero-D-altro-heptose

The preceding studies of C-6 homologations of D-hexodialdo-1,5-pyranose derivatives provided abundant evidence that route B is the only practical strategy for the synthesis of D-glycero-D-altro-heptose derivatives and ruled out any reac-



tions involving carbonyl groups at C-6 of compounds having the altro ring configuration. Methyl 7-tert-butyldiphenylsilyl-D-glycero-a-D-gluco-heptopyranoside (31) was best prepared by O-silvlation of methyl glycoside 15, since silvlation of 13 to give the TBDPS derivative (30) followed by hydrogenolysis was accompanied by $\sim 25\%$ O-desilylation. Reaction of 31 with benzaldehyde dimethylacetal in the presence of catalytic amounts of p-toluenesulfonic acid gave the 4,6-O-benzylidene derivative 32. Several methods for the preparation of methyl 2,3-anhydro-4,6-O-benzylidene-D-glycero-a-D-manno-heptopyranoside (35) were explored. In the course of these experiments samples of the 2-tolylsulfonyl ester 33 and the O-silyl epoxide 36 were obtained in adequate purity for structural characterization by nmr but not for elemental analysis. These samples were of value in monitoring the procedure of Hicks and Fraser-Reid (16) in which treatment of 32 with sodium hydride and tosyl imidazole led to the formation in a single operation of a derivative with the desired 2,3-anhydro-*manno* configuration but this reaction was accompanied by O-desilvlation to give the epoxide 35. The initially formed tosyl ester 33 was rapidly converted into the silvlated epoxide 34, but by the time all starting material (32) had reacted, much 34 had undergone O-desilylation. Prolongation of the reaction led to complete conversion into the epoxide 35. This further reaction turned out to be advantageous since the 7-O-silyl expoxide (34) proved to be quite unreactive under normal conditions of epoxide opening. The O-desilylated epoxide 35 underwent normal diaxial opening on reaction with aqueous sodium hydroxide to yield methyl 4,6-O-benzylidene-D-glycero-α-D-altro-heptopyranoside (36), and O-debenzylidenation gave methyl D-glycero- α -D-altro-heptopyranoside (38). The nmr data for the respective acetylated derivatives (37 and 39) of these two compounds supported the gluco to altro configurational change. Confirmation of the relative configuration was obtained by hydrolysis of the glycoside 38, followed by reduction to give the heptitol and a mixture of 1,6- and 1,7-anhydroheptoses, whose acetylated derivatives were coincident on glc with those derived from the C. jejuni O:36 LPS (7). A sample of the heptose was converted into the (R)- and (S)-2-butyl glycosides whose acetylated derivatives were identical (glc) with those similarly derived from the polysaccharide, thus confirming the absolute configuration of the naturally occurring sugar as D-glycero-D-altro-heptose. In like manner, methoxide opening of epoxide **35** afforded methyl 4,6-O-benzylidene-3-O-methyl-D-glycero-D-altro-heptopyrano side (**40**) and the derived compounds **41–43** were characterized in the same way as for compounds **36–38**. Conversion of a sample of methyl glycoside **41** into (R)- and (S)-2-butyl glycosides followed by acetylation for glc comparison with those derived from the *C. jejuni* O:23 LPS (7) confirmed that the naturally occurring heptose was 3-O-methyl-D-glycero-D-altro-heptose.

ÓMe

ÓМе

In summary, these studies have shown that the high diastereoselectivity observed by Dondoni et al. (5, 6) in the C-6 homologation of a hexodialdo-1,5-pyranose derivative using 2-trimethylsilylthiazole as a one-carbon synthon is restricted to the galactose series. The unidirectional attack at the si face of the 6-aldehyde in this series is presumably governed by the steric effect of the axial oxygen substituent at C-4. No such stereoselectivity was found with glucose and mannose derivatives. In the galactose series a further observation was made that might have escaped attention if experiments had not also been performed in the altrose series. Oxidation of methyl 2,3,4-tri-O-benzyl- α -D-altropyranoside was accompanied by epimerization at C-5 with the formation of an equilibrium mixture of 5-epimeric hexodialdo-1,5-pyranosides favouring the β -L-galacto isomer 23 over the α -D-altro isomer 22 in the ratio of 3:1. In the opposite direction, oxidation of the enantiomeric β-D-galactoside gave the corresponding mixture of products with the α -L-altropyranoside as the minor component. For the monocyclic methyl 2,3,4-tri-O-benzyl hexopyranosides, rapid equilibration may be presumed with product composition on further reaction being kinetically determined. Thus rapid and irreversible reduction with sodium borohydride gave back a mixture of hexopyranosides in approximately the same proportions from glc analysis as indicated by nmr for the hexodialdo-1,5-pyranosides. In contrast the TMSthiazole addition, of presumably reversible character, gave a single detectable product with the β -galacto ring configuration. In the D-series high diastereoselectivity, as reported by Dondoni et al. (5, 6), led to the formation of the 6-(S) thiazolyl derivative 28 and thence of the *D*-glycero-*D*-galacto-heptopyranoside 29. In reexamining the preparation of 1,2;3,4-di-O-isopropylidene- α -D-galacto-hexodialdo-1,5-pyranose (3), nmr showed no evidence for epimerization at C-5 in this conformationally more rigid fused ring derivative. The enolization mechanism for epimerization would require the adoption of a trigonal carbon at C-5 with energetically unfavourable conformational changes in the fused ring system. It is noteworthy that no evidence was obtained for C-5 epimerization in the preparation of hexodialdo-1,5-pyranosides in the mannose and glucose series.

The conversion of (6S)- (4) into the epimeric (6R)-1,2;3,4di-O-isopropylidene-6-(2-thiazolyl)- α -D-galactopyranose (18) by oxidation to the thiazolyl heptose-6-ulose followed by stereoselective reduction with L-selectride (5, 6) appeared to establish in principle a route for conversion of D-glycero- to the 6-epimeric L-glycero-D-heptopyranose derivatives. In our hands this stereoselectivity is not general and seems to be restricted to the conformationally rigid fused ring system in 18 where reduction of the thiazolyl heptose-6-ulose would require unidirectional attack on the re face at C-6 analogous to that on the si face of the hexodialdo-1,5-pyranose 3 in the formation of thiazolyl derivative 4. Reaction starting from the (6S)-(2-thiazolyl)- β -D-galactopyranoside 28 gave a mixture of products that could be converted into derivatives of D-glycero- and L-glycero- β -D-galactopyranosides, and L-glycero- α -L-altropyranosides. The analogous reaction sequence with the corresponding thiazolyl derivatives in the glucose and mannose series showed no stereoselectivity. Stereoselective reductions were achieved here, but in the opposite direction when the heptoside-6-uloses from the severely hindered 7-TBDMS L-glycero-a-D-manno-(19) and α -D-gluco- (20) heptopyranosides were treated with L-selectride.

In reaching our initial objective, that of developing a synthesis of D-glycero-D-altro-heptose to confirm the absolute configuration of the O antigen constituent in the LPS of C. jejuni serotype O:36, the C-6 homologation was achieved first with the formation of derivatives of D-glycero-D-gluco-heptose. The well-known series of transformations from D-glucopyranosides to D-altropyranosides with inversions at C-2 and C-3 presented unexpected difficulties in the formation, and subsequent ring opening, of the intermediate 2,3-anhydro-manno-heptose derivative bearing a 7-O-TBDMS substituent until that protecting group could be removed after its purpose had been served.

Experimental

General methods

Can. J. Chem. Downloaded from www.nrcresearchpress.com by 120.117.138.77 on 11/09/14 For personal use only.

The following solvents were dried and purified by distillation over the reagents specified in parentheses: dichloromethane (phosphorus pentoxide), oxolane (lithium aluminum hydride), methanol (magnesium-iodine), toluene (sodium). Light petroleum (bp $30-60^{\circ}$ C) and ethyl acetate used as chromatographic solvents were freshly distilled immediately before use. Acetonitrile, *N*,*N*-dimethylformamide, and methyl sulfoxide were purchased already dried and glass-distilled. Evaporations were conducted under diminished pressure at <40°C. Thin-layer chromatography (tlc) was conducted on plates coated with Silica Gel $60F_{254}$ (Merck). Optical rotations were measured with a Perkin–Elmer 141 polarimeter, for solutions in CHCl₃ at ~20°C unless otherwise stated.

¹H, ¹³C (often in the JMODXH mode), and 2D ¹H nmr spectra were recorded with a Bruker AM 300 spectrometer. Solutions in chloroform-*d* were referenced to internal tetramethylsilane as internal reference. Solutions in D₂O were referenced to internal acetone ($\delta_{\rm H}$ 2.17 ppm and $\delta_{\rm C}$ 30.5 ppm). Chemical shifts and coupling constants are first-order values and assignments in ¹H nmr spectra on the basis of double resonance or COSY experiments are specified in parentheses. Spectra are reported only in sufficient detail to substantiate chemical changes effected and for configurational assignments. The following characteristic resonances were observed consistently and are not cited individually other than when the groups were first incorporated: for thiazolyl derivatives at $\delta_{\rm H} \sim 7.7$ (H-4') and 7.3 (H-5'), and $\delta_{\rm C} \sim 171$ (C-2'), 142 (C-4') and 119 (C-5'), for benzyl ethers at $\delta_{\rm H} \sim 7.4-7.2$ (Ph-H) and 5.0–4.8 (CH₂Ph), and $\delta_{\rm C} \sim 137$ and 127 (aromatic C) and 75–72 (Ph-CH₂); and for *O*-isopropylidene derivatives at $\delta_{\rm H} 1.5-1.3$ ((CH₃)₂C), and $\delta_{\rm C} \sim 108$ ((CH₃)₂C) and 26–24 ((CH₃)₂C). The glc analyses were carried out on a Hewlett–Packard model 5890 A chromatograph. Separations were performed using a DB-23 (30 m × 0.25 mm) column with the following programs: (A) for alditol acetates, isothermally at 220°C unless specified at 200°C; (B) for 2-butyl glycoside acetates, 200°C (10 min), 200° \rightarrow 240°C at 2°/min. Microanalyses were by Canadian Microanalytical Service, Ltd., Delta, B.C.

Preparations of hexodialdo-1,5-pyranose derivatives

Typical oxidations were performed by the addition of freshly distilled methyl sulfoxide (0.23 mL, 3 mmol) in dichloromethane (4 mL) to oxalyl chloride (1.2 mL, 2.4 mmol) in dichloromethane (10 mL) at -60° C. After 5 min substrate (~0.5 g, 1 mmol) in dichloromethane was added dropwise. After 30 min the reaction was quenched by the addition of triethylamine (2 mL) in dichloromethane (5 mL), and the solution was allowed to rise to room temperature, washed twice with water, dried, and concentrated. Completeness of reaction was checked by tlc for disappearance of initial substrate and, if necessary, by nmr (appearance of an aldehydic proton singlet at $\delta \sim 9.7$ in the ¹H spectrum, appearance of c-6 resonance of substrate at $\delta \sim 62$ in the ¹³C spectrum). The hexodialdo-1,5-pyranose derivative was used directly without further purification in the TMSthiazole homologation reaction.

Addition of 2-(trimethylsilyl)thiazole to methyl 2,3,4-tri-O-benzyl- α -pmanno-hexodialdo-1,5-pyranoside 2

2-(Trimethylsilyl)thiazole (0.4 mL) was added to 2 (from methyl 2,3,4-tri-O-benzyl- α -D-mannopyranoside (17) (0.5 g)) in dichloromethane (10 mL) at 0°C under nitrogen. The mixture was kept at 0°C overnight and then at room temperature for 3 h. Solvents were evaporated, and the residue in oxolane (10 mL) was treated with 1 M tetrabutylammonium fluoride (2.8 mL) for 30 min. Solvent was partly evaporated and the residue was treated with aqueous sodium hydrogen carbonate and extracted with ethyl acetate. The extract was dried, concentrated, and chromatographed on silica gel (dichloromethane containing 3% of acetone) to give, in order of elution, 5 (320 mg, 55%) and **6** (216 mg, 37%). Methyl (6*R*)-2,3,4-tri-O-benzyl-6-(2-thiazolyl)- α -D-mannopyranoside (5) had $[\alpha]_D$ +29 (c 1.0); ¹H nmr, δ: 5.33 (s, 1H, H-6), 4.64 (s, 1H, H-1), 3.04 (s, 3H, OCH₃); ¹³C nmr, δ_C: 99.3 (C-1), 80.1, 74.7, 74.2, 73.6, 70.3 (C-2 to C-6), 54.7 (OCH₃). Anal. calcd. for C₃₁H₃₃NO₆S: C 67.98, H 6.07, N 2.55, S 5.85; found: C 67.96, H 5.98, N 2.55, S 5.85. Methyl (6S)-2,3,4-tri-O-benzyl-6-(2-thiazolyl)-α-Dmannopyranoside (6) had $[\alpha]_D$ +31 (c 1.0); ¹H nmr, δ : 5.39 (d, 1H, $J_{5,6}$ 3.4 Hz, H-6), 4.75 (d, 1H, J_{1,2} 1.7 Hz, H-1), 3.28 (s, 3H, OCH₃); ¹³C nmr, δ_{C} : 98.9 (C-1), 80.1, 75.1, 74.6, 73.8, 72.3 (C-2 to C-6), 54.9 (OCH₃). Anal. calcd. for C₁₃H₃₃NO₆S: C 67.98, H 6.07, N 2.55, S 5.85; found: C 67.75, H 6.00, N 2.55, S 5.85.

Methyl 2,3,4-tri-O-benzyl-L-glycero- (7) and methyl 2,3,4-tri-O-

benzyl-D-glycero- (8) α-D-manno-heptopyranoside

Formyl deblocking from the thiazole ring and subsequent reduction were carried in four successive steps. (1) Compounds 5 and 6 (150 mg, 0.27 mmol each) were separately treated with methyl iodide (0.17 mL, 2.7 mmol) in boiling acetonitrile (7 mL) for 24 h. (2) After removal of solvent, the residues in methanol (5 mL) were each treated with NaBH₄ (21 mg, 2 equiv.) at -10° C for 30 min, after which acetone (2 mL) was added to destroy excess of hydride. Solvent was evaporated, the residue were treated with saturated aqueous NaCl and extracted into ethyl acetate, and the extracts were concentrated. (3) The separate residues in acetonitrile (0.4 mL) were added to solutions of $HgCl_2$ (90 mg, 1.2 equiv.) in acetonitrile-water (4:1, 5 mL), and the solutions were stirred at room temperature for 15 min. Reaction mixtures were filtered, and the filtrates were concentrated. The residues were treated with saturated aqueous NaCl, extracted with chloroform, and the extracts were concentrated. (4) NaBH₄ (21 mg each, 2 equiv.) was added portionwise during 30 min to the crude aldehydic products in methanol (10 mL) at room temperature. Acetic acid – water (1:1) was added dropwise to each solution to destroy excess of hydride and bring the solutions to pH 7, the respective solutions were concentrated, and the residues were chromatographed on silica gel.

Methyl 2,3,4-tri-*O*-benzyl-L-*glycero*-α-D-*manno*-heptopyranoside (7) (82 mg, 61%), formed from 5, was eluted with dichloromethane containing 6% of acetone and had $[\alpha]_D + 25$ (*c* 1.0) (lit. $[\alpha]_D + 30$ (18), $[\alpha]_D + 23$ (19)); ¹H nmr, δ: 4.65 (s, 1H, H-1), 4.11 (t, 1H, $J_{3,4} \sim J_{4,5} \sim$ 9.5 Hz, H-4), 3.94 (m, 1H, H-6), 3.84 (dd, 1H, $J_{2,3}$ 3.0 Hz, $J_{3,4}$ 9.4 Hz, H-3), 3.80–3.60 (m, 3H, H-2, H-7), 3.55 (dd, 1H, $J_{5,6}$ 1.4 Hz, $J_{4,5}$ 9.7 Hz, H-5); ¹³C nmr, δ_C : 99.5 (C-1), 80.0, 74.5, 74.3, 72.5, 69.4 (C-2 to C-6), 65.0 (C-7) (lit. (17, 18) for similar nmr data).

Methyl 2,3,4-tri-*O*-benzyl-*D*-glycero-α-*D*-manno-heptopyranoside (8) (76 mg, 57%), formed from 6, was eluted with dichloromethane containing 4% of acetone and had $[\alpha]_D$ +35 (*c* 1.0); ¹H nmr, δ: 4.68 (d, 1H, $J_{1,2}$ 2.1 Hz, H-1), 4.02 (t, 1H, $J_{3,4} ~ J_{4,5} ~ 9.3$ Hz, H-4), 3.93 (m, 2H, H-3, H-6), 3.80–3.63 (m, 4H, H-2, H-5, H-7); ¹³C nmr, δ_C : 99.0 (C-1), 80.5, 76.9, 74.4, 72.5, 71.1 (C-2 to C-6), 62.9 (C-7) (lit. (20) for similar data).

Samples of 7 and 8 were each hydrogenolyzed in ethanol over palladium 5% on charcoal, the resulting methyl glycosides were hydrolyzed, and the heptoses were reduced and acetylated. The glc analysis showed the formation, respectively, of heptaacetates of L-glycero-Dmanno-heptitol and D-glycero-D-manno-heptitol.

Addition of 2-(trimethylsilyl)thiazole to methyl 2,3,4-tri-O-benzyl-a-Dgluco-hexodialdo-1,5-pyranoside 9

Treatment of **9** (from methyl 2,3,4-tri-*O*-benzyl- α -D-glucopyranoside (21) (18 g, 38.8 mmol)) in dichloromethane (125 mL) with TMSthiazole (6.5 mL, 39.2 mmol) with subsequent processing, as described for **2**, afforded a mixture of **10** and **11** that was chromatographed on silica gel (light petroleum – ethyl acetate, 2:1). Methyl (6*R*)-2,3,4-tri-*O*-benzyl-6-(2-thiazolyl)- α -D-glucopyranoside (**10**) (11.57 g, 54.3%) had mp 110–112°C, and [α]_D +35 (*c* 1.0); ¹H nmr, δ : 5.24 (bs, 1H, H-6), 4.48 (d, 1H, $J_{1,2}$ 3.4 Hz, H-1), 4.16 (m, 1H, H-5), 4.06 (t, 1H, *J* 9.3 Hz, H-3 or H-4), 3.77 (t, 1H, *J* 9.6 Hz, H-4 or H-3), 3.51 (dd, 1H, $J_{1,2}$ 3.5 Hz, $J_{2,3}$ 9.5 Hz, H-2); ¹³C nmr, δ_{C} : 98.1 (C-1), 81.9, 79.6, 76.9, 72.4, 69.8 (C-2 to C-6). Anal. calcd. for C₃₁H₃₃NO₆S: C 67.98, H 6.07, N 2.55, S 5.85; found: C 67.77, H 5.89, N 2.53, S 6.05.

Methyl (6*S*)-2,3,4-tri-*O*-benzyl-6-(2-thiazolyl)- α -D-glucopyranoside (**11**) (3.84 g, 18%) had $[\alpha]_D$ +33 (*c* 1.0); ¹H nmr, δ : 5.30 (d, 1H, $J_{5,6}$ 3.5 Hz, H-6), 4.60 (d, 1H, $J_{1,2}$ 3.8 Hz, H-1), 4.11 (dd, 1H, $J_{5,6}$ 3.5 Hz, $J_{4,5}$ 10 Hz, H-5), 4.04 (t, 1H, J 9.2 Hz, H-3 or H-4), 3.67 (t, 1H, J 9.4 Hz, H-4 or H-3), 3.43 (dd, 1H, $J_{1,2}$ 3.4 Hz, $J_{2,3}$ 9.5 Hz, H-2); ¹³C nmr, δ_C : 97.8 (C-1), 82.0, 79.9, 78.6, 72.4, 71.9 (C-2 to C-6). Anal. calcd. for C₃₁H₃₃NO₆S: C 67.98, H 6.07, N 2.55, S 5.85; found: C 67.89, H 5.93, N 2.50; S 6.20.

Methyl 2,3,4-tri-*O*-benzyl-L-*glycero*-α-D-*gluco*-heptopyranoside (**12**) (4.5 g, 71%), formed from **10** (7 g), as described for **7** and **8**, was eluted with dichloromethane containing 8% of acetone and had $[\alpha]_D$ +53 (*c* 1.0); ¹H nmr, δ: 4.54 (d, 1H, $J_{1,2}$ 3.5 Hz, H-1), 3.97–3.50 (m, 7H, H-2 to H-7'); ¹³C nmr, δ_C : 97.4 (C-1), 81.1, 79.0, 76.5, 69.9, 68.3 (C-2 to C-6), 64.0 (C-7). Anal. calcd. for C₂₉H₃₄O₇: C 70.42, H 6.92; found: C 70.49, H 7.06.

Methyl 2,3,4-tri-*O*-benzyl-D-*glycero*-α-D-*gluco*-heptopyranoside (13) (1.1 g, 71%), formed from 11 (2 g), as described for 7 and 8, was eluted with dichloromethane containing 8% of acetone and had $[\alpha]_D$ +55 (*c* 1.0); ¹H nmr, δ: 4.53 (d, 1H, $J_{1,2}$ 3.4 Hz, H-1), 4.05–3.50 (m, 7H, H-2 to H-7'); ¹³C nmr, δ_C : 97.7 (C-1), 82.1, 80.2, 79.7, 72.6, 69.8 (C-2 to C-6), 62.8 (C-7). Anal. calcd. for C₂₉H₃₄O₇: C 70.42, H 6.92; found: C 70.41, H 6.82.

Methyl L-glycero- α -D-gluco-heptopyranoside (14) (40 mg), from hydrogenolysis of 12 (100 mg) in ethanol (50 mL) over palladium 5%

on charcoal, had mp 153–155°C, $[\alpha]_D$ +157 (*c* 1.0 MeOH); ¹H nmr (D₂O), δ : 4.73 (d, 1H, $J_{1,2}$ 3.66 Hz), 3.32 (s, 3H, OCH₃). The derived pentaacetate (**16**) had $[\alpha]_D$ +86 (*c* 1.0); ¹H nmr, δ : 5.46 (t, 1H, $J_{2,3} \sim J_{3,4} \sim 9.8$ Hz, H-3 or H-4), 5.27 (m, 1H, H-6), 5.04 (t, 1H, $J_{3,4} \sim J_{4,5} \sim 9.8$ Hz, H-4 or H-3), 4.99 (d, 1H, $J_{1,2}$ 3.7 Hz, H-1), 4.90 (dd, 1H, $J_{1,2}$ 3.7 Hz, $J_{2,3}$ 10 Hz, H-2), 4.29 (dd, 1H, $J_{6,7}$ 5.7 Hz, $J_{7,7}$ 11.2 Hz, H-7), 4.21 (dd, 1H, $J_{6,7}$ 7.5 Hz, $J_{7,7'}$ 11.2 Hz, H-7), 4.08 (dd, 1H, $J_{5,6}$ 2.0 Hz, $J_{4,5}$ 10.2 Hz, H-5), 3.40 (s, 3H, OCH₃), 2.12, 2.08, 2.05, 2.01, 2.00 (5 s, 15H, OCOCH₃). Anal. calcd. for C₁₈H₂₆O₁₂: C 49.76, H 6.03; found: C 49.56, H 5.86.

Methyl D-glycero- α -D-gluco-heptopyranoside (15) (20 mg), from hydrogenolysis of 13 (60 mg) in ethanol (25 mL) over palladium 5% on charcoal, had [α]_D +107 (*c* 1.0, MeOH); ¹H nmr (D₂O), δ : 4.72 (d, 1H, $J_{1,2}$ 3.4 Hz), 3.35 (s, 3H, OCH₃). The derived pentaacetate (17) had [α]_D +127 (*c* 1.0); ¹H nmr, δ : 5.44 (t, 1H, J 10.1 Hz, H-3 or H-4), 5.22–5.17 (overlapping ddd, 1H, $J_{5,6}$ 2.8 Hz, $J_{6,7}$ 4.1 Hz, $J_{6,7}$, 7.2 Hz, H-6), 5.08 (dd, 1H, J 9.2 and 10.2 Hz, H-4 or H-3), 4.94 (d, 1H, $J_{1,2}$ 3.6 Hz, H-1), 4.84 (dd, 1H, $J_{1,2}$ 3.7 Hz, $J_{2,3}$ 10.3 Hz, H-2), 4.36 (dd, 1H, $J_{7,7'}$ 11.8 Hz and $J_{6,7}$ 4.0 Hz, H-7a), 4.19 (dd, 1H, $J_{7,7'}$ 11.8 and $J_{6,7'}$ 7.23 Hz, H-7'), 4.01 (dd, 1H, $J_{5,6}$ 2.8 Hz, $J_{4,5}$ 10.1 Hz, H-5), 3.40 (s, 3H, OCH₃), 2.12, 2.08, 2.05, 2.01, 2.00 (5 s, 15H, OCOCH₃). Anal. calcd. for C₁₈H₂₆O₁₂: C 49.76, H 6.03; found: C 49.57, H 5.92.

Methyl glycosides 14 and 15 were hydrolyzed, and the heptoses were reduced and acetylated. The glc analysis showed the formation, respectively, of heptaacetates of L-glycero-D-gluco-heptitol and D-glycero-D-gluco-heptitol.

Configurational inversions at C-6 of derivatives of methyl 2,3,4-tri-Obenzyl-L-glycero-α-D-gluco-heptopyranoside by an oxidationreduction sequence

(A) Thiazolyl derivative 10 (3.7 g, 6 mmol) in dichloromethane (10 mL) was added to periodinane (3.4 g, 8 mmol) at 0°C and the solution was kept for 30 min at 0°C and then at room temperature for 3.5 h. The solution was diluted with ether (100 mL) and poured with stirring into saturated aqueous NaHCO₃ (75 mL) containing Na₂S₂O₃•5H₂O, and shaken for 5 min. The organic layer was separated, washed successively with saturated aqueous NaHCO3, water, and aqueous sodium chloride, dried, and concentrated. L-Selectride (1 M in oxolane, 14 mL) was added to the resulting syrup (3.7 g) in oxolane at -78° C and stirred for 1 h. The reaction was quenched with a solution of aqueous 10% NaOH (20 mL) and 30% hydrogen peroxide (10 mL), and the mixture was stirred at room temperature for 2 h. Solvents were evaporated, the mixture was extracted with ethyl acetate, and the extract was washed with saturated aqueous NaCl, dried, and concentrated. Column chromatography, as above, furnished 10 (1.6 g) and 11 (1.4 g), characterized by tlc and their ¹H and ¹³C nmr spectra, and incompletely separated mixture (0.5 g).

(B) Methyl glycoside (12) (4 g, 8 mmol) in *N*,*N*-dimethylformamide (30 mL) containing imidazole (1.2 g, 17.8 mmol) was treated with *tert*-butyldimethylsilyl chloride (1.34 g, 8.8 mmol) for 1 h at room temperature. The solution was diluted with ether, washed with water, dried, and concentrated. The product was chromatographed on silica gel (light petroleum – ethyl acetate, 2:1) to yield methyl 2,3,4-tri-*O*-benzyl-7-*O*-*tert*-butyldimethylsilyl-L-*glycero*- α -D-*gluco*-heptopyra noside (19) (4.9 g, 99%), [α]_D +7.4; ¹H nmr, δ : 4.52 (d, 1H, $J_{1,2}$ 3.47 Hz, H-1), 3.26 (s, 3H, OCH₃), 0.82 (s, 9H, (CH₃)₃Si), -0.012, -0.014 (2 s, 6H, CH₃Si). Anal. calcd. for C₃₅H₄₈O₇Si: C 69.04, H 7.94; found: C 69.00, H 7.85.

Methyl 2,3,4-tri-*O*-benzyl-7-*O*-tert-butyldimethylsilyl-D-glycero- α -D-gluco-heptopyranoside (**20**), similarly prepared as a reference sample, had [α]_D +11 (c 1.0); ¹H nmr, δ : 4.56 (d, 1H, $J_{1,2}$ 3.4 Hz, H-1), 3.37 (s, 3H, OCH₃), 0.85 (s, 9H, (CH₃)₃Si), 0.00 (2 s, 6H, CH₃Si). Anal. calcd. for C₃₅H₄₈O₇Si: C 69.04, H 7.94; found: C 68.94, H 7.74.

Oxidation of 19 (4.9 g, 7.9 mmol) with oxalyl chloride (9 mL, 18 mmol) and methyl sulfoxide (1.63 mL, 24.5 mmol) as described above and monitored by tlc afforded the 6-ulose, which was treated directly in oxolane (30 mL) with L-selectride (1 M in oxolane, 16 mL). After 30 min tlc showed the formation of an altered product with the mobility of 20 and only trace amounts of 19. When the reaction was

quenched with alkaline hydrogen peroxide, tlc showed the formation of modified products without loss of silyl substituent. *O*-Desilylation (22) on treatment of the mixture in oxolane (30 mL) with tetrabutylammonium fluoride (1 M in oxolane, 15 mL) gave **13** (3.6 g, 90%), $[\alpha]_D$ +55 (*c* 1.0), identical (tlc, ¹H and ¹³C nmr, and diastereomeric purity by conversion to the heptitol heptaacetate for glc analysis) to the previously prepared sample.

Configurational inversion at C-6 of methyl 2,3,4-tri-O-benzyl-L-

glycero- α -*p*-manno-*heptopyranoside by oxidation–reduction* Methyl heptoside 7 (47 mg, 0.09 mmol) in *N*,*N*-dimethylformamide (3 mL) containing imidazole (14 mg, 0.2 mmol) was treated with *tert*-butyldimethylsilyl chloride (16 mg, 0.1 mmol) for 4 h at room temperature. Thin-layer chromatography showed that reaction was incomplete, and further TBDMS (16 mg) and imidazole (14 mg) were added and reaction was continued for another 2 h. The solution was diluted with ether, washed with water, dried, concentrated, and chromatographed on silica gel (light petroleum – ethyl acetate, 3:1) to give methyl 2,3,4-tri-*O*-benzyl-7-*O*-*tert*-butyldimethylsilyl-L-*glycero*- α -*Dmanno*-heptopyranoside (**21**) (53 mg, 90%), [α]_D +24 (*c* 1.0); ¹H nmr, δ : 4.56 (d, 1H, $J_{1,2}$ 2.78 Hz, H-1), 3.22 (s, 3H, OMe), 0.83 (s, 9H, (CH₃)₃Si), 0.00, -0.01 (2 s, 6H, CH₃Si). Anal. calcd. for C₃₅H₄₈O₇Si: C 69.04, H 7.94; found: C 68.92, H 7.97.

Oxidation of **21** (40 mg, 0.06 mmol) with oxalyl chloride (184 μ L) and methyl sulfoxide (33 μ L) in dichloromethane at -78°C, as described above and monitored by tlc, afforded the 6-ulose, which was treated directly in oxolane (5 mL) with L-selectride (1 M in oxolane, 136 μ L) for 1 h. The reaction was quenched by the addition of alkaline hydrogen peroxide (10% aqueous NaOH and 30% H₂O₂, 2:1) to give a mixture of compounds that was treated with tetrabutylammonium fluoride (1 M in oxolane, 133 μ L). The residue was chromatographed on silica gel to give **8** (26 mg, 80%), [α]_D +33 (*c* 1.0), as the sole detectable product, which was indistinguishable (tlc, ¹H and ¹³C nmr) from the previous sample, and was further characterized by conversion into D-glycero-D-manno-heptitol heptaacetate (glc analysis). Treatment of the above-mentioned 6-ulose with sodium borohydride in methanol gave configurationally altered silyl ether (DDManHep) and **21** (LDMan-Hep) in the approximate ratio of 9:1.

Oxidation of methyl 2,3,4-tri-O-benzyl-α-p-altropyranoside and addition of 2-(trimethylsilyl)thiazole to the resulting hexodialdo-1,5-pyranoside mixture

Methyl 2,3,4-tri-O-benzyl- α -D-altropyranoside (23) (250 mg, 0.53 mmol) was oxidized with oxalyl chloride (0.6 mL, 1.2 mmol) and methyl sulfoxide (0.11 mL, 1.43 mmol), as described previously, and gave a mixture of products (22 and 23) as indicated by the appearance of two aldehydic protons in the ¹H nmr spectrum at δ 9.76 and 9.59 in the approximate ratio of 1:4. The ¹³C nmr spectrum showed inter alia two sets of resonances in the same relative proportions corresponding to those of ring carbon atoms of methyl glycosides having the β -galactopyranose (δ_C 104.6 (C-1), 80.8, 78.8, 78.5, and 74.7 (C-2 to C-5)) and α -altropyranose (δ_C 101.1 (C-1), 75.8, 74.6, 74.0, and 71.5 (C-2 to C-5)) configurations. Part of the mixture was reduced with $NaBH_4$ in methanol. One portion of the reduction product was acetylated and gc analysis on column B showed the presence of two components with the retention times of 6-O-acetyl-2,3,4-tri-O-benzyl derivatives of methyl β -D-galactopyranoside and methyl α -Daltropyranoside in the approximate ratio of 3:2. Another portion of the reduction product was O-debenzylated for hydrolysis, followed by (a) reduction and acetylation to give the corresponding mixtures of galactitol and altritol hexaacetates, and (b) glycosidation with (R)- and (S)-2-butanol and acetylation to give products with the retention times (glc on a DB-5 (15 m \times 0.25 mm) column, 230°C (5 min), 230° \rightarrow 260°C, 2°/min, 260°C) of derivatives of L-galactose and D-altrose.

2-TMSthiazole (0.12 mL was added to the remaining mixture of hexodialdo-1,5-pyranosides **22** and **23** (150 mg) as described previously, and chromatography on silica gel (light petroleum – ethyl acetate, 2:1) gave methyl (6*R*)-2,3,4-tri-*O*-benzyl-6-(2-thiazolyl)- β -L-galactopyranoside (**24**) (103 mg, 59%), [α]_D -4 (*c* 1.0); ¹H nmr, δ :

4.26 (d, 1H, $J_{1,2}$ 7.7 Hz, H-1), 3.87 (dd, 1H, $J_{1,2}$ 7.7 Hz, $J_{2,3}$ 9.7 Hz, H-2), 3.53 (s, 3H, OCH₃); ¹³C nmr, δ_C : 105.2 (C-1), 81.8, 79.3, 75.9, 73.8, 70.1 (C-2 to C-6), 57.4 (OCH₃). Anal. calcd. for C₃₁H₃₃NO₆S: C 67.98, H 6.07, N 2.55; found: C 67.82, H 6.15, N 2.55.

Methyl 2,3,4-tri-*O*-benzyl-L-*glycero*-α-L-*galacto*-heptopyranoside (**25**) (50 mg, 74%), formed from **24** (75 mg), as described for **7** and **8**, was chromatographed on silica gel with dichloromethane containing 8% of acetone and had $[\alpha]_D$ +13 (*c* 1.0); ¹H nmr, δ: 4.24 (d, 1H, $J_{1,2}$ 7.6 Hz, H-1), 3.51 (s, 3H, OCH₃); ¹³C nmr, δ_C : 105.0 (C-1), 82.1, 79.5, 74.0, 71.9, 69.1 (C-2 to C-6), 63.8 (C-7). Anal. calcd. for C₂₉H₃₄O₇: C 70.42, H 6.92; found: C 70.28, H 6.94.

A sample of 25 was hydrogenolyzed in ethanol over palladium 5% on charcoal, the resulting methyl glycoside was hydrolyzed, and the heptose was reduced and acetylated. The glc analysis showed the formation, respectively, of a heptitol heptaacetate with the retention time of the corresponding derivative of L-glycero-D-manno-heptitol (= enantiomer of L-glycero-L-galacto-heptitol).

2-(Trimethylsilyl)thiazole homologation of 1,2:3,4-di-O-isopro-

pylidene- α -p-galacto-hexadialdo-1,5-pyranose 3 and configurational inversion of product 4 at C-6

A solution in dichloromethane (10 mL) of compound **3** (1 g, 3.84 mmol), from oxidation of 1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose with oxalyl chloride and methyl sulfoxide showed ¹³C nmr, $\delta_{\rm C}$: 199.0 (CHO), 108.3 (C-1), 95.5, 72.5, 70.9, 69.7 (C-2 to C-5), and was reacted with TMS thiazole (0.72 mL, 4.37 mmol), as described previously, and silica gel chromatography (light petroleum – ethyl acetate, 2:1) furnished (6*S*)-1,2:3,4-di-*O*-isopropylidene-6-(2-thiazolyl)- α -D-galactopyranose (4) (780 mg, 60%), mp 178–180°C (lit. (6) mp 170–172°C), $[\alpha]_{\rm D}$ –67 (*c* 1.0); ¹H nmr, δ : 5.57 (d, 1H, $J_{1,2}$ 4.9 Hz, H-1), 5.14 (d, 1H, $J_{5,6}$ 7.9 Hz, H-6), 4.65 (dd, 1H, $J_{2,3}$ 2.3 Hz, H-2), 4.54 (dd, 1H, $J_{4,5}$ 1.3 Hz, H-4), 4.33 (dd, 1H, H-2), 4.10 (dd, 1H, H-5); ¹³C nmr, $\delta_{\rm C}$: 96.2 (C-1), 70.7, 70.4, 70.3, 69.4, 68.0 (C-2 to C-6).

1,2:3,4-Di-*O*-isopropylidene-D-glycero- α -D-galacto-heptopyranose (**26**) (182 mg, 72%), formed from **4** (300 mg), as described for **7** and **8**, was chromatographed on silica gel (dichloromethane containing 10% of acetone) and had $[\alpha]_D = 57 (c \ 1.0)$; ¹H nmr: $\delta : 5.50 (d, 1H, J_{1,2} 4.9 Hz, H-1)$, 4.63 (dd, 1H, $J_{2,3} 2.3 Hz, J_{3,4} 7.9 Hz, H-3)$, 4.46 (dd, 1H, $J_{4,5} 1.6 Hz, H-4$), 4.30 (dd, 1H, H-2), 3.88–3.68 (m, 4H, H-5, 6, 7, 7'); ¹³C nmr, δ_C : 96.2 (C-1), 70.7, 70.6, 70.5, 69.9, 67.3 (C-2 to C-6), 63.8 (C-7). Anal. calcd. for C₁₃H₂₂O₇: C 53.78, H 7.64; found: C 53.46, H 7.74. Compound **26** was hydrolyzed, and the resulting heptose was reduced and acetylated. The glc analysis showed the formation, respectively, of the heptaacetate of D-glycero-D-galacto-heptitol (= L-glycero-D-manno-heptitol).

Thiazole adduct **4** (49 mg) was oxidized with oxalyl chloride and methyl sulfoxide and the product, without further purification, was reduced with L-selectride, as reported by Dondoni et al. (13), and the isolated (6*R*)-1,2:3,4-di-*O*-isopropylidene-6-(2-thiazolyl)- α -D-galactopyranose (**18**) (30 mg, 60%) had mp 102–104°C (lit. (13) mp 98–100°C); ¹H nmr, δ : 5.57 (d, 1H, $J_{1,2}$ 4.8 Hz, H-1), 5.34 (d, 1H, $J_{5,6}$ 4.8 Hz, H-6), 4.63 (dd, 1H, $J_{2,3}$ 2.4 Hz, $J_{3,4}$ 8.0 Hz, H-3), 4.46 (dd, 1H, $J_{4,5}$ 1.6 Hz, H-4), 4.32 (dd, 1H, H-2), 4.15 (dd, 1H, H-5); ¹³C nmr, δ _C: 96.4 (C-1), 72.4, 71.9, 70.9, 70.8, 69.8 (C-2 to C-6).

1,2:3,4-Di-*O*-isopropylidene-L-*glycero*-α-D-*galacto*-heptopyranose (27) (13 mg, 63%), formed from 18 (25 mg), as described for 7 and 8, was chromatographed on silica gel (dichloromethane containing 10% of acetone) and had mp 98–100°C, $[\alpha]_D - 41$ (*c* 1.0); ¹H nmr, δ: 5.59 (d, 1H, $J_{1,2}$ 5.0 Hz, H-1), 4.62 (dd, 1H, $J_{2,3}$ 2.3 Hz, $J_{3,4}$ 7.9 Hz, H-3), 4.36–4.33 (m, 2H), 3.79–3.77 (m, 4H); ¹³C nmr, δ_C : 96.4 (C-1), 71.6, 71.3, 70.8, 70.5, 67.5 (C-2 to C-6), 62.4 (C-7). Anal. calcd. for C₁₃H₂₂O₇: C 53.78, H 7.64; found: C 53.93, H 7.63. Compound 27 was hydrolyzed, and the resulting heptose was reduced and acetylated. The glc analysis showed the formation of the heptaacetate of L-*glycero*-D-*galacto*-heptitol (= L-*glycero*-D-*gluco*-heptitol).

Oxidation of methyl 2,3,4-tri-O-benzyl-β-D-galactopyranoside and addition of 2-(trimethylsilyl)thiazole to the resulting hexodialdo-1,5-pyranoside mixture

Methyl 2,3,4-tri-O-benzyl-β-D-galactopyranoside (24) (1.09 g,

KHARE ET AL.

2.3 mmol) was oxidized with oxalyl chloride (2.4 mL, 4.8 mmol) and methyl sulfoxide (0.47 mL, 6.1 mmol), as described previously, and gave a mixture of products corresponding to enantiomers 22 and 23, as indicated by the appearance of two aldehydic protons in the ¹H nmr spectrum at δ 9.68 and 9.53 in the approximate ratio of 1:6. The ¹³C nmr spectrum showed inter alia two sets of resonances in the same relative proportions corresponding to those of ring carbon atoms of methyl glycosides having the β -galactopyranose (δ_C 104.6 (C-1), 80.8, 78.8, 78.5, and 74.7 (C-2 to C-5)) and α -altropyranose ($\delta_{\rm C}$ 101.1 (C-1), 75.8, 74.6, 74.0, and 71.5 (C-2 to C-5)) configurations. A portion of the mixture was reduced with NaBH4 in methanol, the reduction product was acetylated, and glc analysis on column B showed the presence of components with the retention times of 6-O-acetyl-2,3,4-tri-O-benzyl derivatives of methyl β -D-galactopyranoside and methyl α -D-altropyranoside in the approximate ratio of 3:1. 2-TMSthiazole (0.45 mL) was added to the remaining mixture of hexodialdo-1,5-pyranosides (950 mg) was described previously, and chromatography on silica gel (light petroleum - ethyl acetate, 2:1) gave methyl (6S)-2,3,4-tri-O-benzyl-6-(2-thiazolyl)-β-D-galactopyranoside (28) (853 mg, 76%), $[\alpha]_{\rm D}$ +2.5 (c 1.0); ¹H nmr, δ : 4.20 (d, 1H, $J_{1,2}$ 7.6 Hz, H-1), 3.46 (s, 3H, OCH₃); ¹³C nmr, δ_C: 104.7 (C-1), 81.4, 78.9, 75.8, 73.5, 69.3 (C-2 to C-6), 56.9 (OCH₃). Within experimental limits these spectra were identical to those of the enantiomer 24. Anal. calcd. for C₃₁H₃₃NO₆S: C 67.98, H 6.07, N 2.55; found: C 67.92, H 6.14, N 2.53

Methyl 2,3,4-tri-*O*-benzyl-D-*glycero*- α -D-*galacto*-heptopyranoside (**29**) (220 mg, 60%), formed from **28** (407 mg), as described for **7** and **8**, was eluted from silica gel with dichloromethane containing 5% of acetone and had mp 101–103°C and $[\alpha]_D = 18 (c \ 1.0)$; ¹H nmr, δ : 4.21 (d, 1H, $J_{1,2}$ 7.6 Hz, H-1), 3.47 (s, 3H, OCH₃); ¹³C nmr, δ_C : 104.8 (C-1), 81.9, 79.3, 73.8, 72.4, 69.0 (C-2 to C-6), 63.6 (C-7). Anal. calcd. for C₂₉H₃₄O₇: C 70.42, H 6.92; found: C 70.41, H 6.88. A sample of **29** was hydrogenolyzed in ethanol over palladium 5% on charcoal, the resulting methyl glycoside was hydrolyzed, and the heptose was reduced and acetylated. The glc analysis showed the formation of a heptitol heptaacetate with the retention time of the corresponding derivative of L-*glycero*-D-*manno*-heptitol (= enantiomer of L-*glycero*-L-*galacto*-heptitol).

Configurational inversions at C-6 and C-5 of methyl (6S)-2,3,4-tri-Obenzyl-6-(2-thiazolyl)-β-D-galactopyranoside (28)

Thiazolyl derivative 28 (85 mg, 0.15 mmol) was oxidized as described previously with oxalyl chloride (0.175 mL, 0.35 mmol) and methyl sulfoxide (0.03 mL, 0.45 mmol). The product (84 mg), without purification, was reduced with L-selectride (0.3 mL, 0.33 mmol) in oxolane. The resulting syrup was separated by preparative layer chromatography (multiple development in light petroleum – ether-methanol, 14:6:1) to give two main fractions.

Fraction 1 (15 mg) had $[\alpha]_D$ +5 (c 1.0); ¹H nmr, δ : 5.25 (d, 1H, $J_{5,6}$ 3.58 Hz, H-6), 4.22 (d, 1H, J_{1,2} 7.7 Hz, H-1), 3.84 (d, 1H, J_{2,3} 9.7 Hz and $J_{1,2}$ 7.7 Hz, H-2), 3.52 (dd, 1H, $J_{2,3}$ 9.7 Hz and $J_{3,4}$ 2.8 Hz, H-3), 3.41 (s, 3H, OCH₃); ¹³C nmr, δ_{C} : 105.1 (C-1), 81.9, 79.1, 75.9, 75.6, 72.4 (C-2 to C-5), 57.0 (OCH₃). The sequence of operations, formyl deblocking, O-debenzylation, hydrolysis, reduction, and acetylation, gave a single heptitol heptaacetate coincident on glc analysis (column A) with the corresponding derivative from L-glycero-D-galacto-heptose. The ¹³C nmr spectrum of fraction 2 (39 mg) showed the presence of signals assignable to anomeric C-1 resonances of compounds with β -galacto (δ_C 104.3) and α -altro (δ_C 99.6) ring configurations and, without individual specifications, to two sets of carbon atoms 2 to 6. Formyl deblocking furnished a mixture of methyl tri-O-benzyl heptopyranosides whose ¹³C nmr spectrum likewise showed anomeric carbon resonances at δ_C 105.1 and 100.5 consistent with the presence of β -galacto and α -altro configurational isomers together with two sets of other carbon resonances. This mixture of compounds was O-debenzylated by formic acid with 5% palladium on charcoal as catalyst, hydrolyzed, reduced, and acetylated. The glc analysis (column A) showed the presence of two heptitol heptaacetates coincident with those from D-glycero-D-galacto-heptose and D-glycero-D-altroheptose.

Conversion of methyl D-glycero-D-gluco-heptopyranoside into methyl D-glycero-D-altro-heptopyranoside derivatives

Methyl 2,3,4-tri-*O*-benzyl-7-*O*-tert-butyldiphenylsilyl-D-glycero- α -D-gluco-heptopyranoside (**30**) was prepared as described for **19** and had [α]_D +9 (c 1.0); ¹H nmr, δ : 7.63–7.10 (m, 25H, Ph-H), 4.61 (d, 1H, $J_{1,2}$ 3.60 Hz, H-1), 3.24 (s, 3H, OCH₃), 1.05 (s, 9H, (CH₃)₃Si) Anal. calcd. for C₄₅H₅₂O₇Si: 73.73, H 7.15; found: C 73.45, H 7.04.

Similarly, silylation of methyl D-glycero- α -D-gluco-heptopyranoside (15) (2 g, 8.9 mmol) followed by chromatography on silica gel (dichloromethane-methanol, 9:1) afforded methyl 7-O-tert-butyldiphenylsilyl-D-glycero- α -D-gluco-heptopyranoside (31) (2.9 g, 71%), [α]_D+59 (c 1.0, methanol); ¹H nmr, δ : 7.68–7.32 (m, 10H, Ph-H), 4.61 (d, 1H, J_{1,2} 3.6 Hz, H-1), 3.24 (s, 3H, OCH₃). Exact Mass calcd. for C₂₄H₃₄O₇Si + H: 463.2135; found: 463.2152; calcd. for C₂₄H₃₄O₇Si + Na: 485.1941; found: 485.1971.

A solution of **31** (1 g, 2.16 mmol), benzaldehyde dimethyl acetal (0.67 mL, 2 equiv., 3.9 mmol), and *p*-toluenesulfonic acid (50 mg) in toluene (50 mL) was kept at 30°C for 1 h under reduced pressure (aspirator), to ensure removal of methanol. Solid NaHCO₃ was added, the solution was concentrated, the residue was partitioned between dichloromethane and water, and the organic layer was dried and concentrated. The residue was chromatographed on silica gel (dichloromethane containing 1% of methanol) to give methyl 4,6-*O*-benzylidene-7-*O*-tert-butyldiphenylsilyl-D-glycero- α -D-gluco-heptopyranoside (**32**) (913 mg, 76%), [α]_D +94 (c 1.0); ¹H nmr, δ : 7.60–7.22 (m, 15H, Ph-H), 5.57 (s, 1H, CHPh), 4.69 (d, 1H, $J_{1,2}$ 3.10 Hz, H-1), 3.34 (s, 3H, OCH₃), 1.03 (s, 9H, (CH₃)₃Si). Exact Mass calcd. for C₃₁H₃₈O₇Si + H: 551.2465; found: 551.2465.

Compound 32 (630 mg, 1.14 mmol) was added to sodium hydride (55 mg, 2 equiv.) in N,N-dimethylformamide (10 mL) and the mixture was stirred for 30 min. N-Tosylimidazole (280 mg, 1.1 equiv.) was added and the mixture was stirred at room temperature for 4 days. Thin-layer chromatography showed the successive formation after 1 h of 2-O-tosyl derivative (33) and O-silyl epoxide (34), and, after 24 h, complete reaction of 32 and conversion of 33 into 34, but now the appearance of O-desilylated epoxide (35). After 84 h conversion of 34 into 35 was complete. The reaction mixture was poured into ice-water, the product was extracted with dichloromethane, and the organic layer was washed with water, concentrated, and chromatographed on silica gel (light petroleum – ethyl acetate, 9:1) to give methyl 2,3-anhydro-4,6-O-benzylidene-D-glycero- α -D-manno-heptopyranoside (35) (236 mg, 70%), $[\alpha]_D$ +20 (c 1.0); ¹H nmr (including ¹H-¹H COSY), δ: 7.52-7.25 (m, 5H, Ph-H), 5.69 (s, 1H, CHPh), 4.92 (d, 1H, H-1), 3.49 (d, 1H, J_{2,3} 3.7 Hz, H-3), 3.34 (s, 3H, OCH₃), 3.18 (d, 1H, $J_{2,3}$ 3.7 Hz, H-2), 1.03 (s, 9H, (CH₃)₃Si). Anal. calcd. for C₁₅H₁₈O₆: C 61.21, H 6.16; found: C 61.16, H 6.18.

The ¹H nmr spectrum of **33** showed inter alia resonances at δ 7.8– 7.2 (m, 14H, Ph-H), 5.57 (s, 1H, acetal CHPh), 4.83 (d, 1H, $J_{1,2}$ 3.7 Hz, H-1), 4.39 (dd, 1H, $J_{1,2}$ 3.8 Hz, $J_{2,3}$ 9.4 Hz, H-2), 4.16 (t, 1H, $J_{2,3} \sim J_{3,4}$ = 9.3 Hz, H-3), 3.31 (s, 3H, OCH₃), 2.43 (s, 3H, CH₃-C₆H₄SO₂), 1.03 (s, 9H, (CH₃)₃C). On acetylation the resonance of H-3 is shifted to δ 5.60. The ¹H nmr spectrum of **34** showed inter alia resonances at δ 7.7–7.2 (m, 10H, Ph-H), 5.66 (s, 1H, acetal CHPh), 4.89 (s, 1H, H-1), 3.51 (d, 1H, $J_{2,3}$ 3.7 Hz, H-3), 3.40 (s, 3H, OCH₃), 3.17 (d, 1H, $J_{2,3}$ 3.7 Hz, H-2), 1.06 (s, 9H, (CH₃)₃C).

Compound **35** (151 mg) was heated at 100°C in 1 M aqueous sodium hydroxide for 12 h. Liquid–liquid extraction of the solution with chloroform afforded methyl 4,6-*O*-benzylidene-D-glycero- α -D-altro-heptopyranoside (**36**) (140 mg, 87%), which crystallized from ether–methanol and had mp 209–211°C, $[\alpha]_D$ +92 (*c* 1.0, methanol); ¹H nmr (D₂O), δ : 7.55–7.42 (m, 5H, Ph-H), 5.85 (s, 1H, Ph-CH), 4.67 (s, 1H, H-1), 3.35 (s, 3H, OMe); ¹³C nmr, δ : 129.4, 128.2, 126.0 (aromatic C), 101.4 (Ph-CH), 101.0 (C-1), 79.0, 74.3, 69.5, 67.5, 58.3 (C-2 to C-6), 60.1 (C-7), 54.7 (OCH₃). Anal. calcd. for C₁₅H₂₂O₇: C 57.68, H 6.45; found: C 57.34, H, 6.33. Acetylation of **36** under standard conditions afforded the tri-*O*-acetyl derivative **37**, which had $[\alpha]_D$ +92 (*c* 1.0); $[\alpha]_D$ +55 (*c* 1.0); ¹H nmr (1 D with decoupling and ¹H–¹H COSY), δ : 7.47–7.26 (m, 5H, Ph-H), 5.23 (1H, br s, H-3), 5.00 (d, 1H, J_{2,3} 2 Hz, H-2), 4.60 (s, 1H, H-1), 4.49 (d, 1H, J_{7,7}' 12 Hz, H-7), 4.27

Can. J. Chem. Downloaded from www.nrcresearchpress.com by 120.117.138.77 on 11/09/14 For personal use only. (dd, 1H, $J_{7,7'}$ 12 Hz, $J_{6,7'}$ 5 Hz, H-7'), 4.06 (m, 3H, H-4, H-5, H-6), 3.36 (s, 3H, OMe), 2.14, 2.12, 2.10 (3 s, 9H, OAc).

O-Benzylidene compound **36** (110 mg) was heated in acetic acid – water (4:1) for 2 h at 80°C and concentration of the solution gave methyl D-*glycero*-α-D-*altro*-heptopyranoside (**38**) (70 mg, 86%), $[\alpha]_D$ +68 (*c* 1.0, methanol); ¹H nmr (D₂O), δ: 4.61 (s, 1H, H-1), 3.24 (s, 3H, OMe); ¹³C nmr, δ_C: 100.4 (C-1), 70.8, 70.0, 69.4, 69.1, 65.0 (C-2 to C-6), 61.3 (C-7), 55.0 (OCH₃). The penta-*O*-acetyl derivative **39** had $[\alpha]_D$ +50 (*c* 1.0); ¹H nmr (1 D with decoupling and ¹H–¹H COSY), δ: 4.94 (dd, 1H, J_{1,2} 1.7 Hz, J_{2,3} 4.1 Hz, H-2), 4.62 (d, 1H, J_{1,2} 1.3 Hz, H-1), 4.43 (dd, 1H, J_{7,7} 12 Hz, J_{6,7} 3.8 Hz, H-7), 4.18 (dd, 1H, J_{7,7} 12 Hz, J_{6,7} 7.2 Hz, H-7'), 3.40 (s, 3H, OMe), 2.13, 2.12, 2.10, 2.07, 2.05 (5 s, 15H, OAc); ¹³C nmr, δ_C: 99.2 (C-2), 69.8, 69.4, 67.3, 66.6, 65.8 (C-2 to C-6), 61.9 (C-7), 55.6 (OCH₃). Anal. calcd. for C₁₈H₂₆O₁₂: C 49.76, H 6.03; found: C 49.83, H 5.95.

Hydrolysis of methyl heptoside (38), followed by treatment of the product with NaBH₄ and acetylation, gave products that were coincident on gc with the heptitol heptaacetate formed from the O:36 glycan with accompanying 1,6- and 1,7-anhydroheptose tetraacetates. Further samples of the hydrolyzate were converted into glycoside acetates on reaction with (R)- and (S)-2-butanol followed by acetylation. Comparison (gc on column A) of these glycoside acetates with those similarly derived from the hydrolysate of the O:36 glycan showed that the natural heptose was D-glycero-D-altro-heptose.

Anhydro glycoside 35 (34 mg) was boiled with methanolic 2 M sodium methoxide (10 mL) for 16 h. Solvent was evaporated and the residue in dichloromethane was washed successively with water, saturated NaCl, and again water, dried, concentrated, and chromatographed on silica gel (light petroleum - ethyl acetate, 4:1, later 2:1) to give methyl 4,6-O-benzylidene-3-O-methyl-D-glycero-a-D-altro-heptopyranoside (40) (28 mg, 74%), $[\alpha]_D$ +81 (c 1.0); ¹H nmr, δ : 7.52-7.26 (m, 5H, Ph-H), 5.67 (s, 1H, Ph-CH), 4.60 (s, 1H, H-1), 3.58, 3.41 (2 s, 6H, OMe). Acetylation of 40 gave the di-O-acetyl derivative 41, $[\alpha]_{D}$ +45 (c 0.5); ¹H nmr (1 D with decoupling and ¹H–¹H COSY), δ : 7.52–7.26 (m, 5H, Ph-H), 5.67 (s, 1H, Ph-C-H), 5.09 (d, 1H, J_{2.3} 2.5 Hz, H-2), 4.58 (s, 1H, H-1), 4.47 (dd, 1H, J_{7,7}, 12 Hz, J_{6,7} 1.7 Hz, H-7), 4.26 (dd, 1H, $J_{7,7'}$ 12 Hz, $J_{6,7}$ 5.9 Hz, H-7'), 4.15 (t, 1H, $J_{4,5} \sim J_{5,6} \sim$ 9.6 Hz, H-5), 4.05–3.99 (m, 1H, H-6), 3.95 (dd, 1H, J_{4.5} 9.5 Hz, J_{3.4} 2.9 Hz, H-4), 3.71 (t, 1H, J 2.4 Hz, H-3), 3.58, 3.39 (2 s, 6H, OMe), 2.14, 2.09 (2 s, 6H, OAc); ${}^{13}C$ nmr, δ_C : 137.2 (quat. aromatic C), 129.2, 128.3, 126.4 (aromatic C), 101.9 (Ph-C-H), 99.4 (C-1), 77.5, 76.3, 75.6, 70.0, 60.1 (C-2 to C-6), 63.2 (C-7), 59.2, 55.8 (2 OCH₃). Anal. calcd. for C₂₀H₂₆O₉: C 58.51, H 6.38; found: C 58.24, H 6.35.

Hydrolysis of **40** (27 mg) in acetic acid – water (4:1) for 1 h at 80°C gave methyl heptoside (**42**) (18 mg), $[\alpha]_D +92$ (*c* 1.0, methanol), which was converted into the acetylated derivative (**43**), $[\alpha]_D +86$ (*c* 1.0); ¹H nmr (1 D with decoupling and ¹H–¹H COSY), δ : 5.33 (ddd (apparent quintet), 1H, $J_{5,6} = J_{6,7'} \sim 3.8$ Hz, H-6), 5.11 (dd, 1H, $J_{3,4}$ 3.5 Hz, $J_{4,5}$ 9.1 Hz, H-4), 5.04 (dd, 1H, $J_{1,2}$ 1.7 Hz, $J_{2,3}$ 4.5 Hz, H-2), 4.60 (s, 1H, H-1), 4.43 (dd, 1H, $J_{6,7}$ 3.6 Hz, $J_{7,7'}$ 12.0 Hz, H-7), 4.28 (dd, 1H, $J_{4,5}$ 9.1 Hz, $J_{5,6}$ 4.0 Hz, H-5), 4.18 (dd, 1H, $J_{6,7'}$ 7.4 Hz, $J_{7,7'}$ 12.0 Hz, H-7'), 3.66 (t, 1H, $J_{2,3} = J_{3,4} \sim 3.9$ Hz, H-3), 3.45, 3.40 (2 s, 6H, OCH₃), 2.14, 2.13, 2.10, 2.05 (4 s, 12H, OAc). Anal. calcd. for C₁₇H₂₆O₁₁: C 50.24, H 6.45; found: C 50.44, C 6.47.

Hydrolysis of methyl heptoside (42), followed by treatment of the product with $NaBH_4$ and acetylation, gave products that were coincident on gc with the *O*-methylheptitol hexaacetate formed from the

O:23 glycan with accompanying 1,6- and 1,7-anhydro-3-O-methylheptose triacetates. Further samples of the hydrolyzate were converted into glycoside acetates on reaction with (R)- and (S)-2-butanol followed by acetylation. Comparison (gc on column A) of these glycoside acetates with those similarly derived from the hydrolysate of the O:23 glycan showed that the natural heptose was 3-O-methyl-D-glycero-D-altro-heptose.

Acknowledgements

We thank the Natural Sciences and Engineering Research Council of Canada for financial support through its strategic and research grants programs. We thank Dr. A.G. McDonald for assistance in glc analyses, and Dr. C.P.J. Glaudemans (National Institutes of Health, Bethesda, Maryland) for reference samples.

- L. Kenne and B. Lindberg. In The polysaccharides. Vol. 2. Edited by G.O. Aspinall. Academic Press, New York. 1983. pp. 287–363.
- 2. K. Dziewiszek and A. Zamojski. Carbohydr. Res. 150, 163 (1986).
- G.J.P.H. Boons, P.A.M. Van der Klein, G.A. Van Der Marel, and J.H. Van Boom. Recl. Trav. Chim. Pays-Bas, 107, 507 (1988).
- G.J.P.H. Boons, G.A. Van Der Marel, and J.H. Van Boom. Tetrahedron Lett. 30, 229 (1989).
- A. Dondoni, G. Fantin, M. Fogagnolo, and A. Medici. Tetrahedron, 43, 3533 (1987).
- A. Dondoni, G. Fantin, M. Fogagnolo, A. Medici, and P. Pedrini, J. Org. Chem. 54, 693 (1989).
- G.O. Aspinall, A.G. McDonald, and H. Pang. Carbohydr. Res. 231, 13 (1992).
- G.O. Aspinall, A.G. McDonald, and R.K. Sood. Can. J. Chem. 72, 247 (1994).
- 9. R. Young and G.A. Adams. Can. J. Chem. 44, 32 (1966).
- S.J. Danishefsky, W.H. Pearson, D.F. Harvey, C.J. Maring, and J.P. Springer. J. Am. Chem. Soc. 107, 1256 (1985).
- 11. K. Omura and D. Swern. Tetrahedron, 34, 1651 (1978).
- A. Dondoni, A. Marra, and D. Perrone. J. Org. Chem. 58, 275 (1993).
- A. Dondoni, G. Fantin, M. Fogagnolo, A. Medici, and P. Pedrini. J. Org. Chem. 54, 702 (1989).
- 14. D.B. Dess and J.C. Martin. J. Org. Chem. 48, 410 (1983).
- G.J. Gerwig, J.P. Kamerling, and J.F.G. Vliegenthart. Carbohydr. Res. 62, 349 (1978).
- 16. D.R. Hicks and B. Fraser-Reid. Synthesis, 203 (1974).
- H.B. Boren, K. Eklind, P.J. Garegg, B. Lindberg, and A. Pillotti. Acta Chem. Scand. 26, 4143 (1972).
- G.J.P.H. Boons, G.A. Van Der Marel, J.T. Poolman, and J.H. Van Boom. Recl. Trav. Chim. Pays-Bas, 108, 339 (1989).
- P.J. Garegg, S. Oscarson, and M. Szönyi. Carbohydr. Res. 205, 125 (1990).
- G.J.P.H. Boons. Ph.D. Thesis, University of Leiden, The Netherlands, 1991.
- 21. A. Lipták, I. Jodal, and P. Nanasi. Carbohydr. Res. 44, 1 (1975).
- 22. S. Hanessian and P. Lavallée. Can. J. Chem. 53, 2975 (1975).
- 23. R. Julina and A. Vasella. Helv. Chim. Acta, 68, 819 (1985).
- 24. P.A.J. Gorin. Carbohydr. Res. 101, 13 (1982).