J.C.S. Снем. Сомм., 1980

## Synthesis of Hepta- and Penta-saccharides, Part of the Complex-type Carbohydrate Portion of Glycoproteins

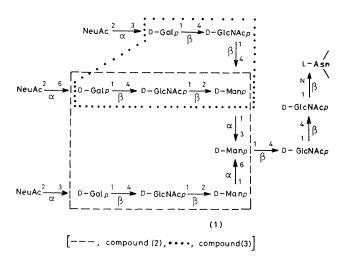
By JAN ARNARP and JORGEN LONNGREN\*

(Department of Organic Chemistry, Arrhenius Laboratory, University of Stockholm, S-106 91 Stockholm, Sweden)

pyranoside gave a protected pentasaccharide and a protected heptasaccharide in 56% and 43% yield, respectively, after deblocking the free penta- and hepta-saccharides were obtained

THE oligosaccharides that are N-glycosidically linked to Lasparagine residues in glycoproteins are of two major types, the 'high-mannose type' which contain D-mannosyl- and N-acetyl-D-glucosaminyl-residues and the 'complex type' which contain D-mannosyl-, D-galactosyl-, N-acetyl-Dglucosaminyl-, and sialyl-residues <sup>1</sup> The 'complex type' has been found with different degrees of branching The Nglycosidically linked carbohydrate portion of fetuin (the predominant glycoprotein of fetal calf serum), which has the structure (1),<sup>2</sup> is a representative example

These oligosaccharides are assumed to be involved in different biological functions<sup>1</sup> and the synthesis of oligosaccharides which form parts of these structures is a matter of some interest. We now report the synthesis of the reduc-

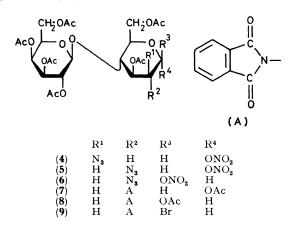


ing heptasaccharide (2) [indicated by the dashed line in structure (1)] and the reducing pentasaccharide (3) [indicated by the dotted line in structure (1)]

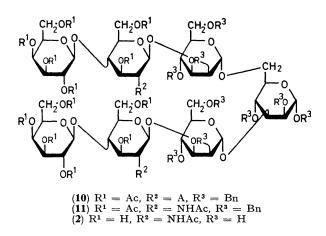
Hexa-O-acetyl-D-lactal<sup>3</sup> (20 g) was subjected to azidonitration<sup>4</sup> by treatment with ceric ammonium nitrate and sodium azide in acetonitrile to give, after work-up, a crystalline mixture (16 5 g)  $\dagger$  According to the <sup>1</sup>H n m r spectrum

<sup>†</sup> Yields are not optimized

and to the sugar analysis<sup>5</sup> of a sample that had been subjected to hydrogenation over palladium-charcoal, this mixture contained mainly compounds (4), (5), and (6) in the approximate proportions 1:4:8. The mixture was treated

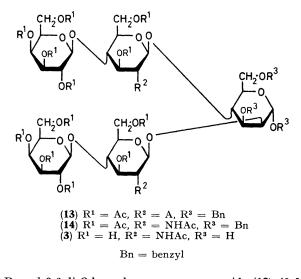


sequentially with hydrogen over palladium-charcoal in ethyl acetate, phthalic anhydride (6 equiv.) in 90% aqueous ethanol adjusted to pH ca. 9 (r.t., 2 h), and acetic anhydridepyridine (r.t., 12 h; 100 °C, 1 h) to give, after silica gel chromatography and crystallization, (7)<sup>‡</sup> (m.p. 229-230 °C) and (8) (m.p. 263-265 °C). In pilot experiments compounds (7) and (8) were separately transformed into the same crystalline bromide (9) (m.p. 109-110 °C) by treatment with hydrogen bromide in methylene chloride (r.t., 4 h). For preparative purposes the crude mixture of (7) and (8) was used to give (9) [34% overall yield from a mixture of (4), (5)and (6)]. Sugar analysis of (9) showed D-galactose and Dglucosamine, but no D-mannosamine. The <sup>1</sup>H n.m.r. spectrum of (9) showed, inter alia, a signal for the anomeric proton of the D-glucosaminyl residue at  $\delta$  6.39 (1 H, d, J 10 Hz) indicating that the  $\beta$ -bromide was obtained. Glycosidation with 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-Dglucopyranosyl bromide<sup>6</sup> in the presence of silver trifluoromethanesulphonate is known<sup>6,7</sup> to give a high yield of  $\beta$ -glucoside and (9) could be expected to behave analogously. For the synthesis of compound (2), benzyl 2,4-di-Obenzyl-3,6-di-O-(3,4,6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl)- $\alpha$ -D-mannopyranoside<sup>8</sup> (0.22 mmol) was condensed with (9)



‡ Satisfactorily n m.r. data were obtained for all compounds.

(0.7 mmol) in methylene chloride, using silver trifluoromethanesulphonate-s-collidine (0.7 mmol) as a promotor<sup>7</sup> (-50-+20 °C for 16 h). Compound (10) { $[\alpha]_{589}$  + 10° (CHCl<sub>3</sub>) }, isolated as a syrup in 43% yield after silica gel chromatography, was treated subsequently with sodium methoxide in methanol (r.t., 16 h), hydrazine hydrate<sup>7</sup> (10 equiv. in boiling ethanol, 16 h), and acetic anhydridepyridine (r.t., 16 h) to give, after silica gel chromatography, compound (11) { $[\alpha]_{589}$  + 11° (CHCl<sub>3</sub>)} in 55% yield. Compound (11) was finally subjected to de-O-acetylation and de-O-benzylation by catalytic hydrogenation (Pd-C catalyst) to give, after gel-filtration (Sephadex G-25), compound (2)  $\{[\alpha]_{589} + 14^{\circ} (H_2O)\}$  in 46% yield. The <sup>1</sup>H n.m.r. spectrum (200 MHz, D<sub>2</sub>O, 85 °C) of (2) showed, inter alia, signals for anomeric protons at  $\delta$  5.14 (1.7 H, J small;  $\alpha$ -D-mannoseresidue and 1,2-linked  $\alpha$ -D-mannosyl-residue), 4.89 (1.3 H, J small;  $\beta$ -D-mannose-residue and 1,2-linked  $\alpha$ -D-mannosylresidue), 4.60 br (2 H, d; N-acetyl-B-D-glucosaminyl-residues), and 4.46 (2 H, d,  $\int 7.5 \text{ Hz}$ ;  $\beta$ -D-galactosyl-residues). Methylation analysis<sup>9</sup> of the alditol of (2) showed 2,3,4,6tetra-O-methyl-D-galactose, 3,4,6-tri-O-methyl-D-mannose, 3,6-di-O-methyl-N-methyl-N-acetyl-D-glucosamine, and 1,2,4,5-tetra-O-methyl-D-mannitol.



Benzyl 3,6-di-O-benzyl- $\alpha$ -D-mannopyranoside (12) {[ $\alpha$ ]<sub>589</sub> +  $42^{\circ}$  (CHCl<sub>3</sub>) was prepared in 40% yield from benzyl  $\alpha$ -D-mannopyranoside by tributylstannylation<sup>10</sup> and benzylation. Compound (12) (0.38 mmol) and (9) (1.1 mmol) were condensed, as described above, to give (13)  $\{[\alpha]_{589} + 12^{\circ}\}$ (CHCl<sub>3</sub>) } in 56% yield after silica gel chromatography. Compound (13) was then transformed into (14)  $\{[\alpha]_{589} - 1^{\circ}\}$ (CHCl<sub>3</sub>) } in 74% yield, as described for the corresponding transformation (10) to (11). Removal of the O-acetyl- and O-benzyl- groups of (14) gave (3)  $\{[\alpha]_{589} - 13^{\circ} (H_2O)\}$  in 61% yield. The <sup>1</sup>H n.m.r. spectrum (200 MHz, D<sub>2</sub>O, 85 °C) of (3) showed, inter alia, signals for anomeric protons at δ 5·16 (0·8 H, 4d, J 1·5 Hz; α-D-mannose-residue), 4·90 (0·2 H, 4d, J < 1 Hz;  $\beta$ -D-mannose-residue), 4.58 br (2 H, d; Nacetyl- $\beta$ -D-glucosaminyl-residues), and 4.47 (2 H, d, J 7.5 Hz;  $\beta$ -D-galactosyl-residues). Methylation analysis<sup>9</sup> of the alditol of (3) showed 2,3,4,6-tetra-O-methyl-D-galactose.

3,6-di-O-methyl-N-methyl-N-acetyl-D-glucosamine, and 1,3,5,6-tetra-O-methyl-D-mannitol

The <sup>1</sup>H n.m r. data given above for (2) and (3) are in good agreement with those reported for related natural oligosaccharides 11

We thank Professors Per J Garegg and Bengt Lindberg for their interest and the Swedish Natural Science Research Council for financial support

(Received, 21st July 1980, Com. 788)

- <sup>1</sup> R G Spiro, Adv Protein Chem, 1973, 27, 349

<sup>1</sup> R G Spiro, Adv Protein Chem, 1973, 27, 349
<sup>2</sup> B Nilsson, N E Nordén, and S Svensson, J Biol Chem, 1979, 254, 4545
<sup>3</sup> W N Haworth, E L Hirst, M M T Plant, and R J W Reinolds, J Chem Soc, 1930, 2647
<sup>4</sup> R U Lemieux and R M Ratcliffe, Can J Chem, 1979, 57, 1244
<sup>5</sup> J S Sawardeker, J H Sloneker, and A R Jeanes, Anal Chem, 1965, 37, 1602
<sup>6</sup> R U Lemieux, T Takeda, and B Y Chung, Am Chem Soc Symposium Series, 1976, 39, 90
<sup>7</sup> D R Bundle and S Josephson, J Chem Soc, Perkin Trans 1, 1979, 2736
<sup>8</sup> J Arnarp and J Lonngren, Acta Chem Scand, Ser B, 1978, 32, 696
<sup>9</sup> P E Jansson, L Kenne, H Liedgren, B Lindberg, and J Lonngren, Chem Commun, Univ Stockholm, 1976, 8
<sup>10</sup> T Ogawa and M Matsui, Carbohydr Res, 1978, 62 Cl
<sup>11</sup> L Dorland, J Haverkamp, J F G Vliegenthart, G Strecker, J -C Michalski, B Fournet, G Spik, and J Montreuil, Eur J Biochem, 1978, 87, 323

1002