Tetrazolyl Peptides and the Tetrazole Analogue of 3-Aminonocardicinic Acid

D.W. Anderson, Malcolm M. Campbell, and Mohammed Malik

School of Chemistry, University of Bath, Claverton Down, Bath BA2 7AY, England.

<u>Abstract</u> The synthesis of the tetrazole analogue of the β -lactam, 3-aminonocardicinic acid, is described. Analogues of seryl p-hydroxy-phenylglycine were also prepared.

Replacement of the carboxyl group in biologically active compounds by the tetrazole group is undertaken because of the close similarity in pKa values, and the steric and electronic similarities.¹ We therefore report the diastereoselective synthesis of the tetrazole (2), designed to mimic 3-aminonocardicinic acid (1), the nucleus of the nocardicin family of antibiotics.² Tetrazole analogues (3) have been reported,³ but are more remote from the natural skeleton. A complementary study addresses the synthesis of the phosphonate analogues.⁴



The N-Boc derivative of D-p-benzyloxyphenylglycine (4) was converted to tetrazole (8) by the sequence shown. Amide (5) was prepared (91%) from acid (4) (EtOCOCl,NH₃ gas), and dehydrated (68%) by POCl₃, giving nitrile (6). Formation of the tetrazole (86%) required NaN₃/NH₄Cl. Formic acid removed the Boc group, giving the formate salt which was transposed into the amine hydrochloride salt of (8) (75%). Tetrazole protection was necessary for the projected synthesis, but isomers were invariably produced, as in benzylation (PhCH₂Br/Et₃N, 80%) of (7) which gave the isomers (10) and (11) which were readily separable. Either (12) or (13) could be hydrogenolysed to the totally deprotected 'D'-aminotetrazole (9) in 60% yield.

Prior to addressing the 3-amino azetidinone target (2), a model cyclisation study was undertaken. β -Bromopropionic acid was coupled with (13) (DCC, 81%) to give (14). Phase transfer cyclisation gave (15) (KOH, nBu₄NBr, CH₂Cl₂, 81%). Hydrogenolysis gave the deprotected β -lactam (16) (H₂, Pd-C, 92%). Note that in other model studies involving nitriles rather than tetrazoles, complications emerged. For example, (17) gave low yields (14% and 41%) of β -lactam (18) and γ -lactam (19) when treated with NaH/DMF/CH₂Cl₂.



DDC coupling procedures gave the dipeptide analogues (20), (21) and (22) which were deprotected, e.g. by catalytic hydrogenolysis of (21) giving the free peptide surrogates (23) and (24), in good yield.

Because of complications encountered in a complementary programme involving cyclisations of aminophosphonate structures closely similar to (20), N-phthaloyl dipeptide (22) was cyclised by the Mitsunobu procedure⁵, (DEAD, Ph₃P, 67%) to give the single diastereoisomer (25). Careful hydrazinolysis (Na₂S 9H₂O, then MeNHNH₂, 55%)⁶ gave free amine (26) which was completely deprotected (H₂/Pd-C,90%) to give (2). By an identical sequence, the benzyltetrazolyl isomer (11) was also progressed to the target β -lactam (2).



14, R¹=R²=PhCH₂

15, $R^1 = R^2 = PhCH_2$ 16, $R^1 = R^2 = H$







20, X=Cl, $R^{1}=Z$, $R^{2}=H$, $R^{3}=R^{4}=PhCH_{2}$ 21, X=OH, $R^{1}=Z$, $R^{2}=H$, $R^{3}=R^{4}=PhCH_{2}$ 22, $R^{1}R^{2}=phthalimide$, X=OH 23, X=Cl, $R^{1}-R^{4}=H$ 24, X=OH, $R^{1}-R^{4}=H$



25, R¹=R²=phthalimide, R³=R⁴=PhCH₂ 26, R¹=R²=H, R³=R⁴=PhCH₂ 2, R¹-R⁴=H

A direct route to the chiral tetrazole analogues of 3-aminocardicinic acids has therefore been established, paralleling our synthesis⁴ of the 3-aminophosphonocardicinic acid. In the key cyclisation steps, different tactics were necessary in each case, emphasising the problems inherent in this approach, and extending other related studies⁷ in the area.

Acknowledgement

We thank Dr. O. Howarth, University of Warwick, and Dr. D.W. Brown, University of Bath, for interpretation of highfield N.M.R. spectra.

New compounds gave satisfactory elemental analyses and/or high resolution mass measurement on

homogeneous samples, and polar compounds gave correct FAB -ve ion M-1 ions.

References

- 1. C.W. Thornber, *Chem. Soc. Revs.*, 1979<u>8</u>, 563. For tetrazole analogues of penams and cephems see: USP, 3957811 (1976) and USP 3966719 (1976).
- M. Hashimoto, T. Komori and T. Kanuja, J. Amer. Chem. Soc., 1976, 98, 3023; H. Aoki, H. Lakai, M. Koshaka, T. Konomi, J. Hosoda, T. Kobochi, E. Isuchi and H. Imanaka, J. Antibiot., 1976, 29, 492, 890; J. Hosoda, T. Konomi, N. Tani, H. Aoki and H. Imanaka, Agric. Biol. Chem., 1977, 41, 2013.
- 3. F.R. Atherton and R.W. Lambert, Tetrahedron, 1983, 39, 2599
- 4. D.W. Anderson, M.M. Campbell, M. Malik, M. Prashad and R.H. Wightman, accompanying paper.
- 5. O. Mitsunobu, M. Wada and J. Sano, J. Amer. Chem. Soc., 1972, <u>94</u>, 679.
- 6. S. Kukolja and S.R. Lambert, J. Amer. Chem. Soc., 1975, <u>97</u>, 5582.
- 7. Phase transfer cyclisations: T. Okawara, Y. Nosuchi, T. Matsuda and F. Furukawa, Chem. Lett., 1981, 185; S. Fletcher and J. Kay, J.C.S. Chem. Commun., 1978, 903; T. Yamazaki, Chem. Pharm. Bull., 1981, 29, 1063; M. Ihara, K. Fukumoto and T. Kametani, Heterocycles, 1982, 19, 1435; C. Cimarusti et al., Tetrahedron, 1983, 39, 2577; DEAD/PPh₃ O. Mitsunobu, M. Wada and T. Sano, J. Amer. Chem. Soc., 1972, 94, 679; M. Wada and O. Mitsunobu, Tetrahedron Letters, 1972, 1279; J.F. Kerwin, P.G. Mattingly, M.J. Miller and M.A. Morrison, J. Amer. Chem. Soc., 1980, 102, 7026, 7027; M.J. Miller and P.G. Mattingly, Tetrahedron, 1983, 39, 2563; M.J. Miller, A. Biswas and M.A. Kiock, ibid., 2571; M.J. Miller and S.R. Woolfe, Tetrahedron Letters, 1984, 3293; A.K. Bose, M.S. Maukes, D.P. Sahn and V.R. Hegde, Can. J. Chem., 1984, 62, 2498; M.J. Miller and M. Jung, Tetrahedron Letters, 1985, 977. Sodium hydride: J.E. Baldwin and M.A. Christie, J. Amer. Chem. Soc., 1978, 100, 4597; G.A. Koppel, L. McShane, F. Jose and R.V.G. Cooper, J. Amer. Chem. Soc., 1978, 100, 3933; H.H. Wasserman, D.J. Hlasta, A.W. Tremper and J.S. Wu, Tetrahedron Letters, 1979, 549; B. Resul, B. Ringdal and R. Dahlbohm, Eu. J. Med. Chem-Chim. Ther., 1981, 16, 379.

(Received in UK 8 February 1990)