

Structure–affinity relationships of berbines or 5,6,13,13a-tetrahydro-8H-dibenzo[a,g]quinolizines at α -adrenoceptors

G Memetzidis¹, JF Stambach^{1*}, L Jung¹, C Schott², C Heitz², JC Stoclet²

¹Laboratoire de Chimie Thérapeutique, Faculté de Pharmacie, 74, route du Rhin, Illkirch;

²Laboratoire de Pharmacodynamie, Faculté de Pharmacie, 74, route du Rhin, 67401 Illkirch Cedex, France

(Received 28 May 1990; accepted 28 January 1991)

Summary — The synthesis of some derivatives of tetrahydro-8H-dibenzo[a,g]quinolizines or berbines is described. A pharmacological study was carried out at α_1 and α_2 -adrenoceptors using radioligand binding techniques. This study has shown that the aromatic ring A is responsible for the α_2 -affinity of berbines. Furthermore, the aromatic ring D is important for α_1 -affinity. However, in this case, it seems that the planarity of the molecule is a very important structural parameter for affinity. The role of the nitrogen atom is also discussed. A conformational analysis of the partial saturated berbines was established by a ^{13}C NMR study.

Résumé — Relations structure–affinité vis-à-vis des récepteurs α -adrénergiques des berbines ou 5,6,13,13a-tétrahydro-8H-dibenzo[a,g]quinolizines. Ce travail décrit la synthèse de dérivés de la tétrahydro-8H-dibenzo[a,g]quinolizine ou berbine dans le but d'établir des relations structure–activité vis-à-vis des récepteurs α_1 et α_2 -adrénergiques. Une étude pharmacologique a été effectuée à l'aide de la méthode des liaisons spécifiques. Ces études ont montré que le noyau aromatique A est responsable de l'affinité de ces composés pour les récepteurs α_2 -adrénergiques et que le noyau aromatique D est impliqué dans l'affinité pour les récepteurs α_1 -adrénergiques. Le rôle de l'atome d'azote est également discuté. De plus, une étude conformationnelle par RMN du ^{13}C de ces dérivés a été réalisée.

berbines / berbinanes / α_1/α_2 -adrenoceptor selectivity / α -antagonists / structure–affinity relationships

Introduction

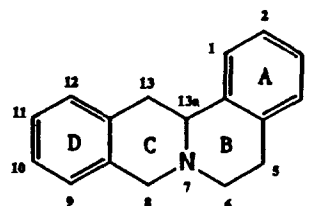
Dibenzo[a,g]quinolizines or berbines are a group of well-known agents acting on α -adrenoceptors [1]. Their 'adrenolytic' properties in anesthetized dogs were first described by Hamet [2] and their antihypertensive effects in man were reported soon after by Facquet *et al* [3].

Previous studies in our laboratory have shown that the affinities of the (+) and (–) enantiomers for α_1 and α_2 binding sites were different and were also differently modified by substituents added to the berbine nuclei, leading to α_1 and α_2 selective compounds [4, 5].

Nevertheless, the rigid tetracyclic structure of berbines provides a basic nitrogen atom, common to all adrenergic ligands, and simultaneously enforces a favorable topographical relationship among this basic nitrogen and the 2 aromatic moieties.

Berbine

Previous studies concerning structure–affinity relationships of α -adrenergic ligands have noted the importance of these structural features [6–9]. In a further extension of our studies in this area, our aim was to gain a better understanding of the structure–affinity relationships of berbines. On this basis, we attempted to acquire some insight into the relative contribution made by the 2 aromatic rings, as well as the nitrogen atom of these molecules, to α_1 and α_2 -adrenoceptor binding.



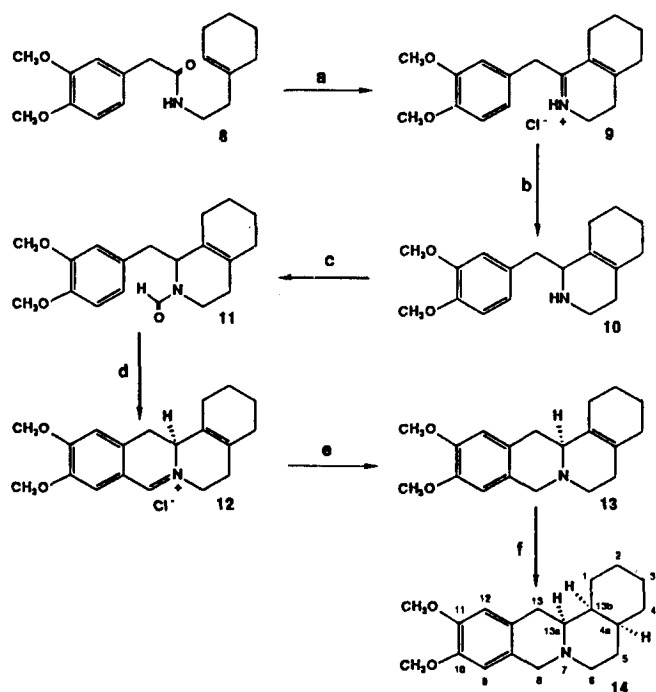
*Correspondence and reprints

Chemistry

The derivatives of berbine **1–6** (table II) substituted in the aromatic rings, were obtained according to our previous synthetic work [10, 11]. Conformational analysis of these berbines by ^1H NMR [12, 13] and IR spectroscopy [14], showed that all these compounds possess a *trans* configuration at the B/C junction.

The synthesis of derivatives of berbine **13**, **14**, saturated in ring A is reported here for the first time; we called them 'berbinanes' (scheme 1). They were prepared according to the classical synthetic strategy [15, 16] using amide **8** as starting material.

The catalytic hydrogenation over Raney Ni (at 85°C , 80 bar) of compounds **13** afforded the decahydro-8*H*-dibenzo[*a,g*]quinolizine **14** as a mixture of *cis/trans* isomers at the A/B junction. The major stereoisomer which was isolated after successive recrystallizations possesses the *cis* configuration at the A/B junction as indicated by the ^{13}C NMR spectra (table I). In fact, C-1 C-3, C-5 and C-13 carbons show an upfield shielding due to γ -steric effects [17]. Furthermore, both **13** and **14** have a *trans* configuration at the B/C junction according to the ^{13}C NMR spectra (table I; see in particular the chemical shifts of C-6, C-8, C-13 and C-13a) [18, 19].



Scheme 1. Reagents: a = $\text{PCl}_5/\text{CHCl}_3$; b = $\text{NaBH}_4\text{CN}/\text{CH}_3\text{OH}$; c = $\text{HCO}_2\text{H}/180^\circ\text{C}$; d = $\text{PCl}_5/\text{CHCl}_3$; e = $\text{NaBH}_4/\text{CH}_3\text{OH}$; f = $\text{H}_2/\text{Ni Raney}$.

Table I. ^{13}C NMR chemical shifts of berbine compounds (δ ppm).

CARBON	COMPOUND					
	1	13	14	20	24 cis	24 trans
C - 1	125.4	25.9	21.2	108.3		
C - 2	126.0	22.6	25.0	147.2		
C - 3	126.0	22.9	20.9	14.2		
C - 4	128.8	30.0	31.7	111.2		
C - 4a	134.5	127.5	36.6	127.0		
C - 5	29.4	30.2	26.8	29.4	23.5	23.8
C - 6	51.2	50.9	57.5	51.2	51.8	60.6
C - 8	58.6	58.1	58.9	59.2	63.4	65.1
C - 8a	134.4	126.7	126.3	126.6		
C - 9	125.8	108.9	108.7	27.3		
C - 10	126.1	147.1	147.4	22.7		
C - 11	126.1	147.2	147.0	22.0		
C - 12	128.7	111.4	110.7	29.0		
C - 12a	134.4	126.5	125.7	126.4		
C - 13	36.6	33.5	32.1	38.2	34.5	29.5
C - 13a	59.8	61.3	61.9	59.5	65.5	66.3
C - 13b	137.8	128.4	40.3	130.2		
CH_3O		55.9(X2)	55.9(X2)	56.0(X2)		
CH_3N					49.8	39.2

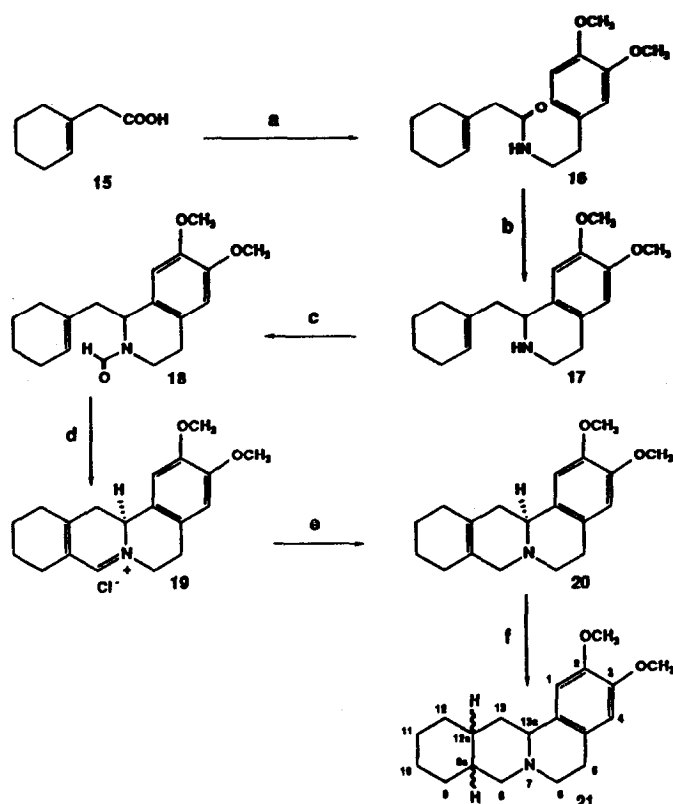
The derivatives of berbine saturated in ring D are known in the literature as 'berbanes' and different methods have been reported for their synthesis [20–22]. In this study the method employed is described in scheme 2. The starting compound **15** was prepared according to the literature [23–25].

The compound **16** was submitted to a Bischler–Napieralski reaction in the presence of phosphorus pentoxide in toluene, rather than phosphorus pentachloride in chloroform, because in the latter case hydrogen chloride is added to the ethylenic double bond. Compound **20** has a *trans* configuration at the B/C junction (see IR, NMR data), while **21** is a mixture of 2 isomers (junction C/D) which could not be separated by using different methods.

The complete reduction of the 2 aromatic rings of **1** was achieved by catalytic hydrogenation over Raney Ni (at 170°C , 150 bar) to afford the perhydroberbine **22**. Compounds **23**, **24**, **25** in which the nitrogen is present in a charged ammonium form, were prepared according to standard literature procedures. However, **23** and **25** are planar structures, while in the case of **24** both *cis* and *trans* isomers were isolated separately (table I) [26].

Biological assays and discussion

Relative affinities of the berbines for central α -adrenoceptors were determined by measuring radioligand displacement from membrane binding sites of rat cerebral cortex.



Scheme 2. Reagents: a = 3,4-dimethoxyphenethylamine/180–200°C; b = P_2O_5 /toluene then $Zn/AcOH$; c = HCO_2H /180°C; d = $PCl_5/CHCl_3$; e = $NaBH_4/CH_3OH$; f = H_2/Ni Raney.

Displacement of [3H]-yohimbine served as a measure of interaction with α_2 -adrenoceptors while [3H]-prazosin displacement was used as an assay for α_1 -adrenoceptor affinity. The results are shown in table II.

Initial inspection of this data indicates that the introduction of large substituents, like carbamate (7), into aromatic ring D had no significant influence on α_1 and α_2 affinity, while introducing the same substituent into ring A (4) produced a 15-fold decrease for both α_1 and α_2 affinity compared to those of compound 3.

Total saturation of the ring A (14) produced a 16-fold decrease for both α_1 and α_2 -affinity compared to those of compound 3. These results indicate that the aromatic moiety A of berbines can be recognized by both α_1 and α_2 -adrenoceptor sites.

However, the introduction of chloro (6) substituents into aromatic ring A reduced affinity for α_2 -sites some 50-fold, compared to the parent compound 1. By

contrast, affinity for α_1 -sites remains unchanged. Partial saturation of this ring A of compound 3 gave 13, in which the α_1 -adrenoceptors affinity was preserved, while α_2 -affinity was 8-fold reduced. Furthermore, the partial saturation of the ring D of 2 gave compound 20, which showed no change in potency for α_1 and α_2 -sites, while its total saturation (21) increased α_1/α_2 -adrenoceptor selectivity by decreasing α_1 -affinity only.

Thus, the major finding of this study is that the aromatic ring A of berbines is essential for the interaction with α_2 -sites, while the ring D could be replaced by a cyclohexane one without causing a significant effect on α_2 -affinity. Otherwise it is known that berbanes, analogues of 21, show a high degree of affinity and selectivity for α_2 -adrenoceptors [27, 28]. A berbane derivative, CH-38083 (7,8-methylenedioxy-14-hydroxyaloberbane) in which the A ring possesses a methylenedioxy group attached to the C_2 - C_3 position, has been shown to be a highly selective antagonist of α_2 -adrenoceptors (table II). This high degree of selectivity of berbanes for α_2 -adrenoceptors is apparently due to the combination of a decrease in α_1 -affinity and an increase in α_2 -affinity when compared to the berbines described in this study.

However, 14, 21, as well as CH-38083, do not have planar structures and therefore show an important decrease in α_1 -affinity, while 13 and 20 which have planar structures preserve α_1 -affinity. According to these results a planar structure is essential for the α_1 -affinity of berbines and berbanes independently of the aromatic ring (A or D) [29, 30].

The permanently charged quaternary analogues 24 and 25 are inactive, suggesting that the presence of a fourth substituent on the nitrogen atom does not permit the approach of the α_1 and α_2 receptor sites because of marked steric constraints. By contrast, 23, which has a planar structure and a positive nitrogen atom preserved some α -adrenergic affinity. For this compound, charge delocalization is extensive, which means that the formal positive centers would be of limited importance for direct receptor interaction, especially in the case of α_1 -adrenoceptors.

In conclusion, the results of this study show that α_2 -activity of the dibenzo[*a,g*]quinolizines depends only on the aromatic ring A. The lack of the aromatic ring D increases the selectivity for the α_2 -sites because this ring is implicated in the α_1 -sites. For their α_1 -activity the planarity of structure is a very important factor. The requirement of at least one aromatic ring for α -adrenergic affinity is indicated by the inactivity of the perhydroberbine 22. The results also indicate that the nitrogen atom is an essential structural feature of berbines and its change can profoundly affect the α -adrenergic affinity, presumably by steric and electronic influences.

Table II. Structure of berbine compounds and their *in vitro* binding affinities to α_1 and α_2 -adrenoceptors.

N°	COMPOUND					K _i nM ^a		SELECTIVITY K _i α_1 / K _i α_2
		R ₁	R ₂	R ₃	R ₄	[³ H] PRAZOSIN (α_1)	[³ H] YOHIMBINE (α_2)	
1		H	H	H	H	880 ± 150	110 ± 14	8.0
2		CH ₃ O	CH ₃ O	H	H	720 ± 130	615 ± 120	1.2
3		H	H	CH ₃ O	CH ₃ O	230 ± 36	180 ± 30	1.3
4		H	NHCO ₂ C ₂ H ₅	CH ₃ O	CH ₃ O	3000 ± 200	3650 ± 1050	0.8
5		H	H	NH ₂ O	CH ₃ O	260 ± 10	260 ± 20	1.6
6		Cl	Cl	CH ₃ O	CH ₃ O	390 ± 60	5100 ± 1700	0.1
7		H	H	NHCO ₂ C ₂ H ₅	CH ₃ O	330 ± 30	630 ± 50	0.5
13						490 ± 40	1400 ± 300	0.35
14						3200 ± 670	3100 ± 410	1.0
20						920 ± 110	630 ± 90	1.5
21						5900 ± 600	420 ± 300	14.0
22						> 10000	> 10000	-
23						2200 ± 380	1400 ± 300	1.6
24					Cis	> 10000	7600 ± 800	-
					Trans	> 10000	7230 ± 900	-
25						> 10000	> 10000	-
	CH - 38083 (réf. 27) 					2600 ^b	1.9 ^b	1368

^aValues are the mean ± SEM of at least 3 separate experiments performed in triplicate.^bThe K_d values of [³H]prazosin and [³H]yohimbine were 0.14 ± 0.02 mmol/l and 10.2 ± 1.6 mmol/l respectively.

Experimental protocols

Chemistry

Melting points were determined on a Koffler hot-stage apparatus and are uncorrected. Spectroscopic data for all compounds were recorded on Beckmann 4230 (IR) and Bruker AC 200 (NMR) instruments. All the ^{13}C NMR spectra were obtained in CDCl_3 and chemical shifts were measured with respect to internal TMS: δ (TMS) = δ (CHCl_3) + 77.2 ppm. Analyses indicated by elemental symbols were within $\pm 0.4\%$ of the theoretical values and were performed by the Central Service of Microanalysis in Vernaion and the Centre Rech Macrom in Strasbourg. All TLC were performed on Merck silica gel F-254 plates ($\text{CHCl}_3/\text{MeOH}$: 17/3).

11-Ethoxycarbamido-10-methoxy-5,6,13,13a-tetrahydro-8H-dibenzo[a,g]quinolizine 7

The amine **5** (500 mg, 1.78 mmol) was dissolved in THF (100 ml) containing 0.2 ml of pyridine (anhydrous). Ethyl chloroformate (0.16 ml) was added slowly and the mixture was stirred at room temperature for 1 h. The solvent was removed under vacuum and the residue was triturated in water, filtered and recrystallized from ethanol. The white crystals (452 mg, 72%) melted at 136–138°C. IR (CHCl_3): 3430 (NH carbamate), 2810–2760 (Bohlmann bands), 1730 cm^{-1} (C=O). ^1H NMR (CDCl_3) δ : 1.32 (t, 3H, CH_2CH_3), 3.35 (dd, 1H, $\text{H}_{13\text{eq}}$), 3.68 (m, 2H, $\text{H}_{8\text{ax}}$ + $\text{H}_{13\text{a}}$), 3.84 (s, 3H, OCH_3), 3.97 (d, 1H, $\text{H}_{8\text{eq}}$), 4.22 (q, 2H, CH_2CH_3), 6.56 (s, 1H, H_9), 7.90 (s, 1H, H_{12}). Anal $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_3$ (C, H, N).

N-[2-(1-Cyclohexenyl)ethyl]-3,4-dimethoxyphenylacetamide 8

A mixture of 3,4-dimethoxyphenylacetic acid (20 g, 102 mmol) and 2-(1-cyclohexenyl)ethylamine (12.82 g, 102 mmol) was heated at 180–200°C (Wood alloy) for 3 h. The solution was cooled and the crude product was crystallized from isopropanol to afford 26.3 g (85%) of white crystals, mp = 104–106°C. IR (CHCl_3): 3410 (NH), 1650 (C=O), 1600 cm^{-1} (C=C). ^1H NMR (CDCl_3) δ : 1.52 (m, 4H, 2CH_2 in β of C=C), 1.80 (m, 4H, 2CH_2 in α of C=C), 1.98 (t, 2H, $\text{CH}_2\text{CH}_2\text{N}$), 3.25 (m, 2H, $\text{CH}_2\text{CH}_2\text{N}$), 3.50 (s, 2H, COCH_3), 3.83 and 3.87 (2s, 6H, 2OCH_3), 5.23 (m, 1H, CH=C), 5.32 (m, 1H, NH).

1-(3,4-Dimethoxybenzyl)-3,4,5,6,7,8-hexahydroisoquinoline HCl 9

The amide **8** (25 g, 82.5 mmol) was dissolved in CHCl_3 (125 ml) and PCl_5 (30 g) was cautiously added. After stirring for 6 h at room temperature, petroleum ether 40–60°C (120 ml) was added and the precipitate was filtered. Recrystallization from $\text{EtOH}/\text{Et}_2\text{O}$ gave 19.1 g (72%) of pale yellow needles, mp = 90–92°C (hygroscopic). IR (CHCl_3): 2700–2300 (N^+H , broad), 1665 ($\text{C}=\text{N}^+$), 1600 cm^{-1} (C=C). ^1H NMR (CDCl_3) δ : 1.60 (m, 4H, $2\text{H}_6+2\text{H}_7$), 2.25 (m, 6H, $2\text{H}_4+2\text{H}_5+2\text{H}_8$), 3.45 (m, 2H, CH_2N), 3.80 and 3.90 (2s, 6H, 2OCH_3), 4.13 (s, 2H, $=\text{CH}_2$), 6.77 (s, 2H, H_2+H_3), 7.00 (s, 1H, H_6), 14.57 (m, 1H, $+\text{NH}$).

1-(3,4-Dimethoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline 10

The above hydrochloride (10 g, 31.1 mmol) was suspended in a mixture of MeOH (50 ml) and THF (200 ml). Sodium cyanoborohydride (3.4 g) was added in several portions and stirring continued for 45 min at room temperature. The solvent was evaporated and the residue was made basic with 3% aqueous NaOH solution. Extraction with Et_2O , washing with water,

drying (MgSO_4) and removal of the solvent gave a liquid which was vacuum distilled (93–95°C/4 mm Hg) to afford 6.7 g (75%) of a colorless oil. IR (CHCl_3): 3320 (NH, weak), 1600 cm^{-1} (C=C). ^1H NMR (CDCl_3) δ : 1.65 (m, 5H, $2\text{H}_6+2\text{H}_7+\text{NH}$), 1.93 (m, 6H, $2\text{H}_4+2\text{H}_5+2\text{H}_8$), 2.82 (m, 5H, $\text{NCHCH}_2+\text{CH}_2\text{N}+\text{CHN}$), 3.90 (s, 6H, 2OCH_3).

1-(3,4-Dimethoxybenzyl)-2-formyl-1,2,3,4,5,6,7,8-octahydroisoquinoline 11

The amine **10** (10 g, 34.8 mmol) was dissolved in a solution of anhydrous formic acid (20 ml). The mixture was heated slowly to 100–110°C (the excess of the acid distilled) and heating continued to 180–190°C for 2–3 h. After cooling the product was crystallized, filtered and recrystallized from diisopropyl-ether/light petroleum ether (1:1) to give 7.68 g (70%) of white prisms, mp = 90–92°C. IR (CHCl_3): 1660 cm^{-1} (HC=O). ^1H NMR (CDCl_3) δ : 3.88 (s, 6H, 2OCH_3), 4.33 and 4.62 (2m, 1H, CHN), 7.45 and 7.93 (2s, 1H, HC=O).

10,11-Dimethoxy-1,2,3,4,5,6,13,13a-octahydrodibenzo[a,g]quinolinium chloride 12

Compound **11** (10 g, 31.7 mmol) was dissolved in CHCl_3 (80 ml) and PCl_5 (11 g) was cautiously added. After stirring for 1 h at room temperature, Et_2O (160 ml) was added and the precipitate was filtered. Recrystallization from water gave 7.43 g (70%) of yellow needles, mp = 170–173°C. IR (CHCl_3): 1635 cm^{-1} (HC=N $^+$). ^1H NMR (CDCl_3) δ : 3.99 (s, 6H, 2OCH_3), 5.02 (m, 1H, $\text{H}_{13\text{a}}$), 6.81 (s, 1H, H_{12}), 7.84 (s, 1H, H_9), 10.50 (s, 1H, H_8).

10,11-Dimethoxy-1,2,3,4,5,6,13,13a-octahydro-8H-dibenzo[a,g]quinolizine 13

5 g (15 mmol) of the quinolinium salt **12** was dissolved in MeOH (200 ml) and NaBH_4 (2.5 g) was added in several portions. After stirring for 1 h at room temperature the solvent was removed and the solution acidified with acetic acid. After basification with diluted NH_4OH solution the product was extracted with Et_2O , washed with water, dried (Na_2SO_4) and evaporated. Recrystallization from cyclohexane gave 3.25 g (73%) of a white powder, mp = 117–119°C. IR (CHCl_3): 2810–2750 cm^{-1} (Bohlmann bands). ^1H NMR (CDCl_3) δ : 2.88 (m, 3H, $\text{H}_{13\text{ax}}+\text{H}_{13\text{eq}}+\text{H}_{13\text{a}}$), 3.52 (d, $J = 14$ Hz, 1H, $\text{H}_{8\text{ax}}$), 3.83 (d, $J = 14$ Hz, 1H, $\text{H}_{8\text{eq}}$), 3.84 (s, 6H, 2OCH_3), 6.53 (s, 1H, H_{12}), 6.59 (s, 1H, H_9). Anal $\text{C}_{19}\text{H}_{25}\text{NO}_2$ (C, H, N).

10,11-Dimethoxy-1,2,3,4,4a,5,6,13,13a,13b-decahydro-8H-dibenzo[a,g]quinolizine 14

Base **13** (2 g, 6.68 mmol) was dissolved in MeOH (100 ml) and hydrogenated over Raney Ni (0.3 g) at 85°C/80 bar for 1.5 h in a stainless steel bomb. Then the reaction mixture was filtered, the solvent evaporated and the residue dissolved in Et_2O . The organic phase was washed with 3% aqueous NaOH solution, then with water, dried (Na_2SO_4) and evaporated. The product was recrystallized in a mixture of $\text{EtOH}/\text{H}_2\text{O}$ (3:1) to give only the major stereoisomer as a white powder, 1.2 g (60%), mp = 92–94°C. IR (CHCl_3): 2810–2750 cm^{-1} (Bohlmann bands). ^1H NMR (CDCl_3) δ : 3.20 (d, $J = 15$ Hz, 1H, $\text{H}_{8\text{ax}}$), 3.83 (s, 6H, 2OCH_3), 3.84 (d, $J = 15$ Hz, 1H, $\text{H}_{8\text{eq}}$), 6.50 (s, 1H, H_{12}), 6.56 (s, 1H, H_9). Anal $\text{C}_{19}\text{H}_{27}\text{NO}_2$ (C, H, N).

N-(3,4-Dimethoxyphenethyl)-2-(1-cyclohexenyl)acetamide 16

This compound was prepared from the acid **15** [23–25] (20 g, 0.14 mol) and 3,4-dimethoxy- β -phenethylamine (25.57 g, 0.14 mol) as described for **8**, to give white needles in 90% yield (Et_2O), mp = 89–91°C. IR (CHCl_3): 3410 (NH), 1650

(C=O), 1600 cm⁻¹ (C=C). ¹H NMR (CDCl₃) δ: 1.53 (m, 4H, 2CH₂ in β of C=C), 1.90 (m, 4H, 2CH₂ in α of C=C), 2.75 (t, 2H, CH₂CH₂N), 2.82 (s, 2H, COCH₃), 3.50 (m, 2H, CH₂CH₂N), 3.88 (s, 6H, 2OCH₃), 5.57 (m, 1H, CH=C), 5.85 (m, 1H, NH).

1-(1-Cyclohexenyl)methyl-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline 17

The above amide **16** (10 g, 33 mmol), 20 g of P₂O₅ and dry toluene (50 ml) were refluxed for 1 h. The solvent was decanted and ice-cold water (100 ml) was cautiously added. The aqueous layer was washed twice with CHCl₃ and conc HCl (100 ml) was added as well as zinc powder (10 g). The whole mixture was refluxed for 2 h and the excess Zn was filtered. After cooling diluted NH₄OH solution was added slowly and the product was extracted with CHCl₃. The organic layer was separated, washed with water, dried (MgSO₄) and evaporated to give 6.6 g (70%) of a reddish oil, which was used with no further purification. IR (CHCl₃): 3320 (NH, weak), 1600 cm⁻¹ (C=C). ¹H NMR (CDCl₃) δ: 2.42 (m, 2H, CH₂CHN), 2.78 (m, 2H, CH₂CH₂N), 3.05 (m, 2H, CH₂N), 3.90 (s, 6H, 2OCH₃), 4.00 (dd, 1H, CHN), 5.61 (s, 1H, CH=C), 6.65 (m, 2H, H₈+H₅).

1-(1-Cyclohexenyl)methyl-2-formyl-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline 18

Using the same procedure as described for the conversion of **10** and **11**, 10 g (34.8 mmol) of **17** gave 8.34 g (76%) of white crystals (EtOH), mp = 149–151°C. IR (CHCl₃): 1660 (HC=O), 1600 cm⁻¹ (C=C). ¹H NMR (CDCl₃) δ: 3.84 (s, 6H, 2OCH₃), 4.49 and 5.42 (2m, 1H, CHN), 5.42 (m, 1H, CH=C), 6.57 (s, 2H, arom), 8.06 and 8.13 (2s, 1H, HC=O).

2,3-Dimethoxy-5,6,9,10,11,12,13,13a-octahydrodibenzo[a,g]-quinolizinium chloride 19

Using the same procedure as described for the conversion of **11** to **12**, 10 g (31.75 mmol) of **18** (except stirring was continued for 3 h), gave 7.62 g (72%) of a yellow powder (EtOH/Et₂O), mp = 206–210°C. IR (CHCl₃): 1650 (HC=N⁺), 1600 cm⁻¹ (C=C, weak). ¹H NMR (CDCl₃) δ: 3.86 (s, 6H, 2OCH₃), 5.22 (m, 1H, H_{13a}), 6.66 (s, 1H, H₄), 6.67 (s, 1H, H₁), 9.25 (s, 1H, H₈).

2,3-Dimethoxy-5,6,9,10,11,12,13,13a-octahydro-8H-dibenzo[a,g]-quinolizine 20

Using the same procedure as described for the conversion of **12** to **13**, 5 g (15 mmol) of **19** gave 3.36 g (75%) of a white powder (*n*-hexane), mp = 89–91°C. IR (CHCl₃): 2810–2750 cm⁻¹ (Bohlmann bands). ¹H NMR (CDCl₃) δ: 3.43 (m, 2H, H_{13a}+H_{8eq}), 3.87 (s, 6H, 2OCH₃), 6.63 (s, 1H, H₄), 6.72 (s, 1H, H₁). Anal C₁₉H₂₅NO₂ (C, H, N).

2,3-Dimethoxy-5,6,8a,9,10,11,12,12a,13,13a-decahydro-8H-dibenzo[a,g]-quinolizine 21

The title compound was prepared from **20** (2 g, 6.7 mmol), as described for **14** (reaction time 5 h), to give 1.2 g (60%) of a white powder (*n*-hexane), mp = 121–128°C [lit [22] 133°C (*cis*), 127°C (*trans*)]. IR (CHCl₃): 2810–2750 cm⁻¹ (Bohlmann bands). ¹H NMR (CDCl₃) δ: 3.82 (2s, 6H, 2OCH₃), 6.50 (s, 1H, H₄), 6.62 (s, 1H, H₁). Anal C₁₉H₂₇NO₂ (C, H, N).

Perhydrodibenzo[a,g]-quinolizine 22

Compound **1** (8 g, 34 mmol) was dissolved in MeOH (150 ml) and hydrogenated over Raney Ni (1.5 g) at 170°C/150 bar for 5 h in a stainless steel bomb. After filtration, the solvent was evaporated and the residue dissolved in ether. The ethereal

phase was washed with 3% aqueous NaOH solution, then with water, dried (MgSO₄) and acidified with gaseous HCl to give white crystals of the hydrochloride. Recrystallization from EtOH/Et₂O (1:10) gave 6.3 g (75%), mp = 198–205°C. ¹H NMR (CDCl₃) (base) δ: 1.48 (m, 24H, CH₂ cycles), 2.50 (m, 5H, CHN+2CH₂N). Anal C₁₇H₂₉N, HCl, H₂O (C, H, N).

5,6-Dihydrodibenzo[a,g]-quinolizinium chloride 23

1-Benzyl-2-formyl-1,2,3,4-tetrahydroisoquinoline* (1 g, 3.96 mmol) was dissolved in CHCl₃ (30 ml) and PCl₅ (1.5 g) was added. The mixture was refluxed overnight, cooled and THF (30 ml) was added. After filtration the solid was recrystallized from EtOH to give 0.557 g (52%) of a white powder, mp = 226–228°C (dec). ¹H NMR (DMSO-*d*₆) δ: 3.38 (m, 2H, 2H₅), 4.96 (m, 2H, 2H₆), 7.57 (m, 3H, H₁+H₂+H₄), 8.03 (t, 1H, H₃), 8.34 (m, 4H, H₈+H₁₀+H₁₁+H₁₂), 9.22 (s, 1H, H₁₃), 10.20 (s, 1H, H₈). Anal C₁₇H₁₄NCl, 2.5 H₂O (C, H, N).

7-Methyl-5,6,13,13a-tetrahydro-8H-dibenzo[a,g]-quinolizinium iodide 24

This compound was prepared from **1** (1.2 g, 5.1 mmol) as described in the literature [31] to afford the *trans* isomer in colourless prisms (H₂O), mp = 229–231°C and the *cis* isomer in white crystals (EtOH/Et₂O) mp = 212–214°C. Anal C₁₈H₂₀NI (C, H, N).

5,6,13,13a-Tetrahydro-8H-dibenzo[a,g]-quinolizine N-oxide 25

Compound **1** (1 g, 4.25 mmol) was dissolved in MeOH (30 ml) and 3 ml of H₂O₂ (30%) was added. Stirring was continued at room temperature for 5 h. Then 100 mg of (5%) Pt/C was added, stirred for a few min and filtered. After evaporation of the solvent the residue was crystallized from MeOH to give 0.85 g (80%) of a white needles, mp = 165–167°C. ¹H NMR (CDCl₃) δ: 2.84 (dd, 1H, H_{13eq}), 3.75 (m, 5H, 2H₅+2H₆+H_{13ax}), 4.55 (d, *J* = 15 Hz, 1H, H_{8ax}), 4.67 (dd, 1H, H_{13a}), 4.73 (d, *J* = 15 Hz, 1H, H_{8eq}), 7.20 (m, 8H, arom). Anal C₁₇H₁₇NO, 2H₂O (C, H, N).

Pharmacology

Radioligand binding studies

Rat brain membranes were prepared as described by Green-grass and Bremner [32] with some modifications (Descombes and Stoclet) [33].

Binding studies were performed with the α₁-selective radioligand [³H]prazosin ([³H]PRA specific activity 644 GBq · mmol⁻¹) and the α₂-selective radioligand [³H]-yohimbine ([³H]YOH specific activity 3126 GBq · mmol⁻¹) supplied by New England Nuclear. The saturation curves were obtained by incubating various concentrations of radioligands (0.05–5 nM [³H]PRA; 0.05–18 nM [³H]YOH with aliquot fractions of membrane suspension (0.5 mg protein). Competition studies were performed with 0.2 nM [³H]PRA or 2 nM [³H]YOH. Non-specific binding was measured in presence of 10 μM phentolamine. All assays were performed in triplicate. In competition studies IC₅₀ (concentration displacing 50% of maximal specific binding) and the slope factor of the displacement curves were calculated using a computerised iterative non-linear curve fitting program (McPherson *et al* [34]). IC₅₀ values were converted to K_i values using the Cheng-Prussoff relation [35].

*Prepared from 1-benzyl-1,2,3,4-tetrahydroisoquinoline as described for compound **11**.

References

- 1 Sasagawa S, Kanetani K, Kiyofuji S (1969) *Chem Pharm Bull* 17, 1–4
- 2 Hamet R (1952) *Bull Acad Méd* 136, 408–410
- 3 Facquet J, Lisle M, Combaz M (1954) *Gaz Méd Fr* 61, 1615–1616
- 4 Schott C, Heitz C, Stoclet JC, Stambach JF, Jung L (1983) *Arch Mal Cœur* (spec issue) 19–22
- 5 Schott C, Tetsi L, Heitz C, Stambach JF, Jung L, Stoclet JC (1988) *Arzneim Forsch (Drug Res)* 38, 1567–1571
- 6 Easson LH, Stedman E (1933) *Biochem J* 27, 1257–1266
- 7 Belleau B (1967) *Ann NY Acad Sci* 139, 580–605
- 8 Kier LB (1969) *J Pharm Pharmacol* 21, 93–96
- 9 Pullman B, Coubeils JL, Courriere P, Gervois JP (1972) *J Med Chem* 15, 17–22
- 10 Stambach JF, Jung L (1985) *Tetrahedron* 41, 169–172
- 11 Memetidis G, Stambach JF, Jung L (1990) *Heterocycles* 31, 341–351
- 12 Tourwe D, Van Binst G, Kametani T (1977) *Org Magn Reson* 9, 341–346
- 13 Yu CK, Maclean BD, Rodrigo RGA, Manske RHF (1970) *Can J Chem* 48, 3673–3678
- 14 Bohlmann F (1958) *Chem Ber* 91, 2157–2167
- 15 Dyke SF (1978) In: *Rodd's Chemistry of Carbon Compounds* (Coffey S, ed), Elsevier, Amsterdam, vol IV H, 110 p
- 16 Pandey GD, Tiwari KP (1980) *Heterocycles* 14, 59–82
- 17 Grant DM, Cheney BV (1967) *J Am Chem Soc* 89, 5315–5318
- 18 Kametani T, Fukumoto K, Ihara M, Ujiie A, Koizumi H (1975) *J Org Chem* 40, 3280–3283
- 19 Van Binst G, Tourwe D (1973) *Heterocycles* 1, 257–265
- 20 Jirkovsky I, Protiva M (1963) *Collect Czech Chem Commun* 28, 2577–2581
- 21 Belleau B, Puranen J (1965) *Can J Chem* 43, 2551–2558
- 22 Szabo L, Honty K, Toke L, Szantay C (1972) *Chem Ber* 105, 3231–3243
- 23 Wallach O (1909) *Liebigs Ann Chem* 365, 255–277
- 24 Wallach O (1906) *Liebigs Ann Chem* 347, 316–386
- 25 Beesley RM, Ingold CK, Thorpe JF (1915) *J Chem Soc* 107, 1080–1160
- 26 Yoshikawa K, Morishima I (1975) *Chem Lett* 961
- 27 Vizi ES, Harsing LG, Gaal J, Kapocsi J, Bernath S, Somogyi GT (1986) *J Pharm Exp Ther* 238, 701–706
- 28 Vizi ES, Toth I, Somogyi GT, Szabo L, Harsing LG, Szantay C (1987) *J Med Chem* 30, 1355–1359
- 29 Ferry N, Goodhardt M, Hanoune J, Sevenet T (1983) *Br J Pharmacol* 78, 359–364
- 30 Campbell SF, Davey MJ, Hardstone JD, Lewis BN, Palmer J (1987) *J Med Chem* 30, 49–57
- 31 Chakravarti SN, Haworth RD, Perkin WH (1927) *J Chem Soc* 2275–2281
- 32 Greengrass P, Bremmer R (1979) *Eur J Pharmacol* 55, 323–326
- 33 Descombes JJ, Stoclet JC (1985) *Naunyn-Schmiedeberg's Arch Pharmacol* 329, 282–288
- 34 McPherson GA, Beart PM (1983) *Eur J Pharmacol* 91, 363–369
- 35 Cheng YC, Prussow WH (1973) *Biochem Pharmacol* 22, 3099–3108