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Catalytic Detoxification of Organophosphorus Nerve Agents by Butyrylcholinesterase-Polymer-Oxime Bioscavengers

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ABSTRACT

Organophosphorus nerve agents (OPNA), used in chemical warfare, irreversibly inhibit essential cholinesterases (ChEs) in the cholinergic neurotransmission system. Several potent nucleophilic oximes have been approved for the treatment of acute poisoning by OPNAs, but they are rapidly cleared from blood circulation. Butyrylcholinesterase (BChE) stoichiometrically binds nerve agents, but since the molecular weight of a nerve agent is

about 500-fold less than the enzyme, the bioscavenger has had limited utility. We have synthesized BChE-polymer-oxime conjugates using atom transfer radical polymerization (ATRP) and azide-alkyne "click" chemistry. The activity of the BChE-polymer-oxime conjugates was dependent on the degree of oxime loading within the copolymer side chains. The covalent modification of oxime-containing copolymers prolonged the activity of BChE in the presence of the VX- and cyclosarin-fluorogenic analogues EMP-MeCyC and CMP-MeCyC, respectively. After complete inactivation by VX and cyclosarin fluorogenic analogues, the conjugates demonstrated efficient self-reactivation of up to 80% within 3-6 hours. Repeated inhibition and high-level self-reactivation assays revealed that the BChE-polymer-oxime conjugates were excellent reactivators of OPNA-inhibited BChE. Recurring self-reactivation of BChE-polymer-oxime conjugates following repeated BChE inhibition by Flu-OPs opens the door to developing the next generation of nerve agent "catalytic" bioscavengers.

INTRODUCTION

Initially developed in the 1930s, OPNAs have been used as pesticides, tested on criminal subjects, used in terrorist acts and employed as chemical warfare agents.[1] Tabun and sarin, for example, were used in the Iran-Iraq war in the1980s, sarin was used against civilians in Syria and by a terrorist sect in the Tokyo subway, and VX was used to assassinate the North Korean, Kim Jong-nam, in Malaysia (2017).[2-5] Accidental and self-poisoning by organophosphorus pesticides results in around 100,000 deaths and affects more than 1 million people each year worldwide.[6] OPNAs inhibit cholinesterases (ChEs) in the cholinergic neurotransmission system[7] by phosphorylation of a catalytic serine residue in the active site. OPNA toxicity results from the accumulation of neurotransmitters in cholinergic synapses leading to paralysis, hypotension, breathing failure and death.[8-11] Hydrophobic OPNAs cross the blood-brain barrier (BBB) and inactivate acetylcholinesterase (AChE) in the central nervous system (CNS). Accumulated OPNAs, retained in the CNS, can also partition back into blood causing "cholinergic crisis" and irreversible brain damage.[12, 13]

Currently approved treatment of acute OPNA poisoning includes administration of a mixture of a pyridinium aldoxime, an anticonvulsant (benzodiazepine) and a muscarinic acetylcholine receptor antagonist (atropine).[14, 15] Since the seminal work of Wilson and colleagues in the mid-1950s, thousands of aldoximes have been synthesized and screened.[16, 17] Few molecules, for example pralidoxime (2-PAM), asoxime (HI-6), and obidoxime (toxogonin), have been developed as OPNA poisoning therapeutics.[14, 15] Aldoximes, in the charged oximate nucleophilic state, attack the phosphoryl bond of covalently-inhibited ChEs at the active site serine, releasing the OP moiety from the active site and restoring ChE activity.[18-20] Over several decades, numerous imidazolium oximes have been developed as OP-ChE reactivators.[12, 21-24] Some of these derivatives were shown to be efficient reactivators in specific *in vitro* and *in vivo* reactivation assays using VX- and tabun-inhibited ChEs.[23-27] Antidotal therapy by oximes, unfortunately, requires repeated administration because low molecular weight oximes have very short half-lives in circulation.[28, 29] This short half-life also limits the use of oximes as effective long-term prophylactic drugs.

A new concept of acute OPNA poisoning treatment, by the administration of bioscavengers, emerged at the end of the 1980s.[30-35] Bioscavengers are proteins that react with and neutralize OPNAs. Human butyrylcholinesterase (BChE) is effective as a bioscavenger at both pre- and post-OPNA exposure.[36-40] The enzyme is stable in human blood circulation, with a half-life of 12 days, in concentration ranges between 3.5 and 9.3 mg/L.[41-45] BChE is found in plasma, organs, and tissues including liver, skin, striated muscle, smooth muscle, lung, and brain.[46] Administration of BChE has proven to be safe with no toxic, immunogenic or behavioral effects.[47] As a stoichiometric bioscavenger, with a molecular weight of about 85 kDa, 500-600 times higher than that of OPNAs, a relatively high dose of BChE (100-500 mg) is required to effectively protect a 70 kg patient.[48] Unfortunately, this dose costs more than $2,000, and therefore BChE therapy has not been deployed as a prophylactic drug.[30]

Grafting poly(ethylene glycol) (PEG)[49-52] and poly(sialic acid) to the surface of BChE has been used to increase the half-life of the enzyme in the blood.[53-56] Nevertheless, even extended lifetime BChE does not overcome the inherent limitations of a stoichiometric

bioscavenger. We have become interested in whether polymers could serve to increase BChE protein lifetime while also enabling reactivation and protection of the enzyme, thereby defeating the stoichiometric limitations that drive up the necessary dose. Such a self-reactivating BChE protein-polymer conjugate could also be used as a critical component in decontamination of broad areas.

A “grafting from” high yield approach to protein-polymer conjugate synthesis, involving the *in situ* growth of polymers from biomacromolecular initiators, emerged in the early 2000s.[57-59] Developed in the early 1990s, atom transfer radical polymerization (ATRP) has proved to be a controlled and powerful technique to prepare well-defined polymers and protein-polymer conjugates.[51, 60-65] With a long term interest in polymer-based protein engineering, we have generated a large library of protein-polymer conjugates.[66-69]

In the past decade, first generation “pseudo-catalytic" bioscavenger systems, composed of BChE delivered with an efficient oxime reactivator, were developed to reduce the amount of BChE needed for efficient protection.[23, 24, 36, 70] Unfortunately, the fast elimination of oxime reactivators from blood circulation, requiring continuous infusion of

the reactivator to sustain reactivation kinetics, limited the use of pseudo-catalytic bioscavengers *in vivo*.[30] Most recently, we have used ATRP to synthesize BChE-polymer conjugates with multiple pendant azido groups. Azide-alkyne "click" chemistry was then used to covalently couple oximes to the surface of the enzyme.[71] The BChE-polymer-oximes exhibited a slowed rate of inactivation by organophosphates and some evidence of inter- and intramolecular reactivation. Herein, fluorogenic analogues of VX and cyclosarin (EMP-MeCyC and CMP-MeCyC, respectively)[72] were used to study the kinetics of inactivation and reactivation of next-generation BChE-polymer-oxime conjugates. The BChE-polymer-oxime conjugate recovered about 85% activity after complete inactivation induced by fluorogenic analogues of VX in three successive inactivation-reactivation "catalytic" cycles. The conjugates should also be useful in surface decontamination of OPNAs.

EXPERIMENTAL SECTION

Materials

Common reagents and solvents were purchased from Fisher Scientific (Pittsburgh, PA) and used as received unless otherwise specified. Dr. Oksana Lockridge (Eppley Institute, University of Nebraska Medical Center) kindly provided the human BChE. EMP-MeCyC and CMP-MeCyC were prepared we have described previously.[72] MA-PEG_6-N_3 monomer and NHS-Br (ATRP initiator) were prepared in the same way as our previous reports.[71, 73] Copper(II) bromide ($CuBr_2$), copper(II) sulfate ($CuSO_4$), hydroxylamine hydrochloride, triethylamine, sodium ascorbate (NaAsc), 2-imidazolecarboxaldehyde, *S*-butyrylthiocholine iodide (BTC), 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB), *N,N*-dimethylacrylamide (DMAA), 1,1,4,7,10,10-hexamethyltriethylenetetramine (HMTETA), tris[2-(dimethylamino)ethyl] amine (Me_6TREN), isonicotinaldehyde, 1,4-diiodobutane, 6-iodohex-1-yne, and 4-chlorobenzyl bromide were purchased from Sigma-Aldrich. DMAA, Me_6TREN and HMTETA were purified with a basic alumina column before using. 2-(4-

((bis((1-(tert-butyl)-1H-1,2,3-triazol-4-yl)methyl)amino)methyl)-1H-1,2,3-triazol-1-yl) acetic acid (BTTAA) and Azide-PEG-Amine were purchased from Click Chemistry Tools LLC. *N,N*-dimethylprop-2-yn-1-amine was purchased from Fisher Scientific.

Characterization

Nuclear magnetic resonance (NMR) analysis

A 400 MHZ spectrometer (Bruker Avance) in the Center for Molecular Analysis, Carnegie Mellon University, Pittsburgh, PA was used to collect ^{1}H NMR spectra.

Ultraviolet-visible (UV-VIS) and fluorescence spectrophotometry

UV−VIS spectra were analyzed by a UV−VIS spectrophotometer (Lambda 45, PerkinElmer). Fluorescence spectra were analyzed by a plate reader (Bio-Tek Synergy H1).

GPC analysis

The molecular weight (M_n) and the polydispersity ($Đ$) of azide containing copolymers were determined by GPC with 3-columns on a Waters 2695 Series system (Waters Ultrahydrogel Linear, 500 and 250) as our previous report.[71] The BChE, BChE-Br, BChE-PDMAA-N_3 and BChE-PDMAA-OX conjugates were analyzed by the same GPC system with 4-columns.

Protein analysis by the Bicinchoninic acid (BCA) assay

The BChE-PDMAA-OX conjugate (1-2 mg/mL, protein, 20 or10 μL) in BP buffer (50 mM, pH 7.4) was mixed with 200 μL BCA working reagent solution (Reagent A/Reagent B = 50/1). The mixture was incubated at 60°C for 15 min. 562 nm absorbance of the samples was recorded by a plate reader (Bio-Tek Synergy H1). Standard curves of PEG-IO, PEG-PO and native BChE were used to determine the protein concentration of the conjugates (**Figure S1**).

SDS-PAGE analysis

BChE, BChE-Br, or BChE-polymer-oxime conjugate solution (1.0 mg/mL protein, 10 μL) was mixed with SDS-PAGE loading buffer (2X, 10 μL) and heated at 95 °C for 10 min. 10

μL of the sample was loaded onto 4-15% precast gel and run at 200 V for 30 min. The gel was washed with distilled water three times and stained with 50 mL PageBlue staining solution. 1 hour later, the gel was de-stained overnight by distilled water.

Methods

Synthesis of alkyne-imidazolium-oxime

Alkyne-imidazolium-oxime was synthesized as described previously.[71] Briefly, 2-imidazolecarboxaldehyde (12 mmol) and K_2CO_3 (24 mmol) were mixed in 30 mL DMF. 4-Chlorobenzyl bromide (24 mmol) was added and stirred at room temperature overnight. The reaction mixture was filtered, and 250 mL distilled water was added to the filtrate. The resulting solution was extracted with ethyl acetate (200 mL × 3), then the organic phase was dried with $MgSO_4$. After removing the solvent by rotary evaporator, the alkylimidazole-2-carbaldehyde yield was 74%.

Hydroxylamine hydrochloride (13.5 mmol) was dissolved in 30 mL water, and added to Na_2CO_3 (13.5 mmol) and alkylimidazole-2-carbaldehyde (9 mmol). The reaction mixture was stirred for 3.5 h at room temperature, and the precipitate was filtered and washed by

distilled water three times and Et_2O four times. The product was dried over a vacuum, generating an 87% yield.

6-Iodohex-1-yne (1.5 mmol) in 10 mL ACN and 1 mL DMSO were added to the alkylimidazole-2-carbaldehyde oxime (0.4 mmol). The reaction mixture was stirred at 67 ˚C for 5 days. Removed the solvent by rotary evaporator. The crude product was purified by silica gel column (from hexane/acetone = 1/1 to acetone/MeOH = 3/1). White powder, 48% yield. ^{1}H NMR (400 MHz, MeOD-d4) δ 8.52 (s, 1H, CH), 7.82 (d, J = 3, 1H, Ar H), 7.74 (d, J = 3, 1H, Ar H), 7.44 (d, J = 3, 2H, Ar H), 7.35 (d, J = 3, 2H, Ar H), 5.63 (s, 2H; CH_2), 4.43 (t, J = 3, 2H; CH_2), 3.36 (s, 1H; CH), 2.27 (d, J = 3, 2H, CH_2), 2.02-2.00 (m, 2H; CH_2), 1.62-1.59 (m, 2H; CH_2) (**Figure S2**).

Synthesis of PEG-IO

PEG-IO was synthesized as described previously.[71] Briefly, BTTAA (2.4 mg in 120 µL H_2O) and sodium ascorbate (NaAsc, 2 mg in 100 µL H_2O) was added to a $CuSO_4$ solution (100 mM, 60 µL). Alkyne-imidazolium-oxime (22 mg) and NH_2-PEG_6-N_3 (20 mg in 300 µL DMF) were added to the solution. The reaction mixture was incubated at room

temperature for 1 hour. The product was purified by precipitation in ether/acetone = 10/1 four times and dried by a vacuum. Dark oil, 75% yield. ^{1}H NMR (400 MHz, MeOD-d4) δ 8.60 (s, 1H; CH), 7.94 (s, 1H, Ar H), 7.92 (s, 1H, Ar H), 7.81 (d, J = 3, 1H, Ar H), 7.49 (d, J = 3, 2H, Ar H), 7.33 (d, J = 6, 2H, Ar H), 5.59 (s, 2H; CH_2), 4.45 (t, J = 3, 2H; CH_2), 4.35 (t, J = 3, 2H; CH_2), 3.78 (s, 2H; CH_2), 3.50-3.44 (m, 24H; $12CH_2$), 2.62 (d, J = 3, 2H, CH_2), 1.81-1.79 (m, 2H; CH_2), 1.59 (d, J = 3, 2H, CH_2) (**Figure S2**). PEG-IO was used as a standard for the calculation of oxime to BChE ratios of BChE-polymer-oxime conjugates.

Synthesis of alkyne-pyridinium-oxime

Isonicotinaldehyde (50 mmol), hydroxylamine hydrochloride (55 mmol) and K_2CO_3 (7.6 g, 55 mmol) were mixed in 250 mL MeOH at room temperature overnight. After removing the solvent by rotary evaporator, the residue was diluted with water (250 mL) and extracted with ethyl acetate (5 × 150 mL). The combined extract was washed with brine (2 × 150 mL) and dried over $MySO_4$. Remove the solvent by rotary evaporator and got the product isonicotinaldehyde oxime, 84% yield.

Isonicotinaldehyde oxime (10 mmol) and 1,4-diiodobutane (30 mmol) were dissolved in

50 mL ACN and stirred at room temperature for 5 days. The resulting precipitate was collected by filtration, then concentrated and added to 50 mL ethyl acetate. We next collected and combined the precipitate, then purified by silica gel (EA/MeOH/HCOOH = 4/1/0.25) to generate the product 4-((hydroxyimino)methyl)-1-(4-iodobutyl)pyridin-1-ium iodide in 77% yield.

4-((hydroxyimino)methyl)-1-(4-iodobutyl)pyridin-1-ium iodide (0.5 mmol) was dissolved in 5 mL ACN and 400 µL DMSO. *N,N*-dimethylprop-2-yn-1-amine (1 mmol) was added to the ACN/DMSO solution and stirred at room temperature for 2 days. The solvent was removed by rotary evaporation. The product alkyne-pyridinium-oxime was purified by precipitation in ethyl acetate/acetone (2/1) three time and dried by vacuum. Yellow powder, 65% yield. ^{1}H NMR (400 MHz, MeOD-d4) δ 8.82 (d, J = 6, 2H, Ar H), 8.35 (s, 1H, Ar H), 8.19 (d, J = 6, 2H, Ar H), 4.68-4.63 (m, 2H; CH_2), 4.23 (d, J = 3, 2H, CH_2), 3.49-3.46 (m, 2H; CH_2), 3.23 (t, J = 3, 1H; CH), 3.15 (s, 6H; $2CH_3$), 2.18-2.07 (m, 2H; CH_2), 1.91-1.87 (m, 2H; CH_2) (**Figure S3**).

Synthesis of PEG-PO

PEG-PO was synthesized as described previously.[71] Briefly, BTTAA (13 mg in 300 μL H_2O) and sodium ascorbate (NaAsc, 20 mg in 200 μL H_2O) were added to a $CuSO_4$ solution (100 mM, 300 μL). Alkyne-pyridinium-oxime (52 mg) and NH_2-PEG_3-N_3 (55 mg in 600 μL DMF) were then added to the solution and the reaction mixture was incubated at room temperature overnight. The product was purified by precipitation in ether/acetone = 10/1 four times and dried by a vacuum. Dark oil, 53% yield. ^{1}H NMR (400 MHz, MeOD-d4) δ 8.82 (s, 1H; Ar H), 8.80 (s, 1H; Ar H), 8.34 (d, J = 3, 2H, Ar H), 8.17 (d, J = 6, 2H, Ar H), 4.69-4.62 (m, 6H; $3CH_2$), 3.95 (t, J = 3, 2H, CH_2), 3.70 (t, J = 3, 2H, CH_2), 3.65-3.59 (m, 8H; $4CH_2$), 3.35-3.32 (m, 2H; CH_2), 3.16 (t, J = 3, 2H, CH_2), 3.04 (s, 6H; $2CH_3$), 2.08-2.03 (m, 2H; CH_2), 2.01-1.95 (m, 2H; CH_2) (**Figure S3**). PEG-PO was used as a standard for the calculation of oxime to BChE ratios of BChE-polymer-oxime conjugates.

Synthesis of BChE-polymer-oxime conjugates

BChE-PDMAA-N_3 conjugate was synthesized as described previously.[71] Alkyne-imidazolium-oxime or alkyne-pyridinium-oxime was attached to the side chains of BChE-

PDMAA-N_3 conjugate by "click" chemistry.[74] NaAsc (10 mM × 35 μL), $CuSO_4$ (10 mM × 17.6 μL), and BTTAA (10 mM × 35 μL) in distilled water were added into a BChE-PDMAA-N_3 (27 μM BChE, 650 μL) solution. Alkyne-imidazolium-oxime or alkyne-pyridinium-oxime (50 mM) in distilled water was added in different oxime to BChE ratios (40-, 80- or 160-fold). The reaction mixture was incubated at room temperature for 3 h. The BChE-PDMAA-OX conjugates were purified by ultrafiltration (50 kDa cut-off membrane) four times. The number of oximes per BChE monomer of the conjugates was determined by BCA assay and UV absorbance at 280 nm (**Figure S1**). BChE-PDMAA-OX conjugates were characterized by GPC and SDS-PAGE analysis.

Synthesis of PDMAA-IO and PDMAA-PO

To synthesize the azide containing copolymer, PDMAA-N_3, DMAA (1.20 mmol), MA-PEG_6-N_3 (60 μmol) and ATRP initiator (1 μmol) were mixed in 2 mL of PB buffer (50 mM, pH 7.4), sealed, and bubbled with argon for 30 min at room temperature. Deoxygenated Me_6TREN (8 μmol), $CuBr_2$ (3 μmol) and NaAsc (4 μmol) in distilled water (1 mL) were added to the initiator-monomer solution, sealed, and stirred for 2 h at room temperature.

The copolymer PDMAA-N_3 was purified by dialysis (8 kDa cut-off membrane) in distilled water and lyophilization. The molecular weight (M_n) and dispersity ($Đ$) of PDMAA-N_3 were 64.4 kDa and 1.87, respectively (**Figure S4** and **S5**). GPC calibration was based on poly(ethylene glycol) standards.

Alkyne-imidazolium-oxime or alkyne-pyridinium-oxime was attached to the side chains of PDMAA-N_3 conjugate by "click" chemistry to prepare PDMAA-IO or PDMAA-PO.[74] NaAsc (10 mM × 80 μL), $CuSO_4$ (10 mM × 40 μL), and BTTAA (10 mM × 80 μL) in distilled water were added into a PDMAA-N_3 (14 mg in 650 μL H_2O) solution. Alkyne-imidazolium-oxime or alkyne-pyridinium-oxime (2 mg in 80 μL DMSO) was added and incubated at room temperature overnight. PDMAA-IO or PDMAA-PO was purified by ultrafiltration (30 kDa cut-off membrane) four times.

The oxime content of PDMAA-IO and PDMAA-PO, determined from the UV-VIS spectra, were 12.5 wt% and 4.5 wt%, respectively (**Figure S1** and **S4**).

Activity assay of BChE and BChE-PDMAA-OX conjugates

BTC was used as a substrate to determine the activity of BChE and BChE-PDMAA-OX

conjugates at room temperature. 1 mL of BTC (1 mM) and DTNB (Ellman assay reagent, 0.1 mM) in PB buffer (50 mM, pH 7.4) was added to a 1.5 mL cuvette. Native BChE or BChE-PDMAA-OX conjugates (20 nM protein, 100 μL) was then added to the cuvette.[75] The activity of BChE or BChE-PDMAA-OX conjugates was evaluated by monitoring BTC hydrolysis, resulting in an increase in absorbance at 412 nm from TNB (extinction coefficient of 14,000 M^{-1} cm^{-1}), using a Lambda 2 PerkinElmer UV-VIS spectrophotometer.

Inhibition assay of BChE and BChE-polymer-oxime conjugates

BChE or BChE-PDMAA-OX conjugate (20 or 10 nM) was incubated with a 5-fold stoichiometric excess of EMP-MeCyC or CMP-MeCyC at room temperature. An aliquot was diluted 10 or 20 times in PB buffer (50 mM, pH 7.4) at specified time intervals. After addition of BTC (1 mM) and DTNB (0.1 mM), we measured the enzymatic activity of each group. The uninhibited BChE or BChE-PDMAA-OX conjugate was treated as 100% activity at the time zero. Activity evaluation was performed in PB buffer (50 mM, pH 7.4) by the Ellman assay using a Lambda 2 PerkinElmer UV-VIS spectrophotometer.[75]

Flu-OPNAs degradation assay

A mixture of BChE, BChE-PDMAA-IO_{172} conjugate (or BChE-PDMAA-PO_{216} conjugate) and PDMAA-IO (or PDMAA-PO) at various ratios (protein 50 nM; Oxime/BChE = 43 or 54) were incubated with a 5-fold excess of EMP-MeCyC (or CMP-MeCyC) in PB buffer at pH 7.4. The total volume of the mixture was 100 µL. The fluorescence intensity (ex 400 nm; em 446 nm) was measured using a Bio-Tek Synergy H1 plate reader at specified time intervals.

Reactivation assay of BChE and BChE-PDMAA-OX conjugates

A reactivation assay was carried out with BChE or BChE-PDMAA-OX conjugates (1 µM) in 1 mg/mL bovine serum albumin (BSA) solution in PB buffer (50 mM, pH 6.0) that we mixed with a 10-fold molar excess of EMP-MeCyC or CMP-MeCyC in PB buffer (50 mM, pH 6.0) for 10 min. The completely inactive BChE or conjugates were transferred into an ultrafiltration tube (50 kDa cut-off membrane) to remove the excess of EMP-MeCyC or CMP-MeCyC (10,000 × g for 5 min, three times), then diluted 50-fold in PB buffer (50 mM, pH 8.0) at room temperature. At specified time intervals, the activities of BChE or

conjugates were measured by Ellman assay using BTC (1 mM) and DTNB (0. 1 mM). The activity of uninhibited BChE or conjugates were used to determine the 100% activity.

Inactivation and reactivation recycle of BChE-PDMAA-OX conjugates

BChE-PDMAA-OX conjugates 1, 2 or 3 (1 μM) in 1 mg/mL bovine serum albumin (BSA) in PB buffer (50 mM, pH 6.0) solution was mixed with a 10-fold molar excess of EMP-MeCyC in PB buffer (50 mM, pH 6.0) for 10 min. The completely inactive conjugates were purified by ultrafiltration tube (50 kDa cut-off membrane, 10,000 × g for 5 min, three times), then diluted 50-fold in PB buffer (50 mM, pH 8.0) and incubated at room temperature for 200 min. The conjugates were buffer-exchanged with PB buffer (50 mM, pH 6.0) and concentrated to 1 μM, then mixed with a 10-fold molar excess of EMP-MeCyC in PB buffer (50 mM, pH 6.0) and incubated at room temperature for 10 min. The completely inactive conjugates were purified by ultrafiltration tube (50 kDa cut-off membrane, 10,000 × g for 5 min, three times), then diluted 50-fold in PB buffer (50 mM, pH 8.0) and incubated at room temperature for 200 min. The inactivation and reactivation cycle were repeated three times. At specified time intervals, the conjugate activity was measured by the Ellman

assay using BTC (1 mM) and DTNB (0.1 mM) as substrates. The activity of uninhibited, or activity recovered conjugates at pH 8.0 after 200 min, were used to determine the 100% activity value.

Tertiary structure-based prediction of NHS-ATRP initiator modification

The tertiary structure of BChE was downloaded from Protein Data Bank (PDB code: 6I2T). In order of influence on initiator-amino group specificity, the following properties for each lysine residue and N-termini were determined: exposed surface area (ESA), steric hindrance, pK_a, secondary structure, local charge, and H-bonding (Table S1).[76]

RESULTS AND DISCUSSION

Synthesis and Characterization of BChE-polymer-oxime conjugates

To synthesize BChE-polymer-oxime conjugates, an *N*-hydroxysuccinimide (NHS) ester modified ATRP initiator (NHS-Br) was reacted with accessible amine groups on the surface of BChE generating the macroinitiator BChE-Br. On average, 6.5 initiators were

attached to each BChE monomer.[71] To pinpoint where these modification sites were located, we employed a tertiary structure-based predictive model that we had previously developed to identify highly reactive amine groups on the surface BChE.[76] This approach enabled us to isolate the N-termini and a small group of lysine residues (K9, K60, K105, K190, and K558) as the likely sites of reaction with the ATRP initiators (Table 1). Azide-containing copolymers were grown from BChE-Br at fast-modified lysine residues and the N-termini by *in situ* ATRP with a neutral monomer, *N,N*-dimethylacrylamide (DMAA), in the presence of an azido monomer (MA-PEG_6-N_3), to generate BChE-PDMAA-N_3 conjugates, as described previously.[71] Copper catalyzed azide-alkyne cycloaddition "click" chemistry was then used to couple an alkyne-imidazolium-oxime or an alkyne-pyridinium-oxime to the azido-containing copolymers of BChE-PDMAA-N_3 conjugates (**Figure 1**). Stoichiometric alkyne-imidazolium-oxime (IO) or alkyne-pyridinium-oxime (PO) excesses of 40-fold, 80-fold or 160-fold were used to modify BChE-PDMAA-N_3 in the preparation of BChE-PDMAA-OX conjugates (BChE-PDMAA-IO and BChE-PDMAA-PO). The BChE-PDMAA-OX conjugates were purified by ultrafiltration (100 kDa cut-off membrane) and the oxime to BChE ratios were determined using the bicinchoninic acid (BCA) assay and

absorbance at 280 nm (**Figure S1**). The number of imidazolium-oximes per BChE tetramer was 28, 84 and 172 for BChE-PDMAA-IO conjugates (we refer to these conjugates as BChE-PDMAA-IO_{28}, BChE-PDMAA-IO_{84}, and BChE-PDMAA-IO_{172}, respectively). The number of pyridinium-oximes per BChE tetramer was 44, 104 and 216 for BChE-PDMAA-PO conjugates BChE-PDMAA-PO_{44}, BChE-PDMAA-PO_{104}, and BChE-PDMAA-PO_{216}, respectively. The molecular weights and purities of the BChE-PDMAA-OX conjugates were compared by gel permeation chromatography (GPC) and sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) analysis (**Table 1**, **Figure 2**). It is important to note that there are no GPC standards for these kinds of protein-polymer conjugates, so we must be cautious when interpreting observed conjugate molecular weights. That's said, GPC can be useful when interpreting the differences in apparent molecular weight between various conjugates.

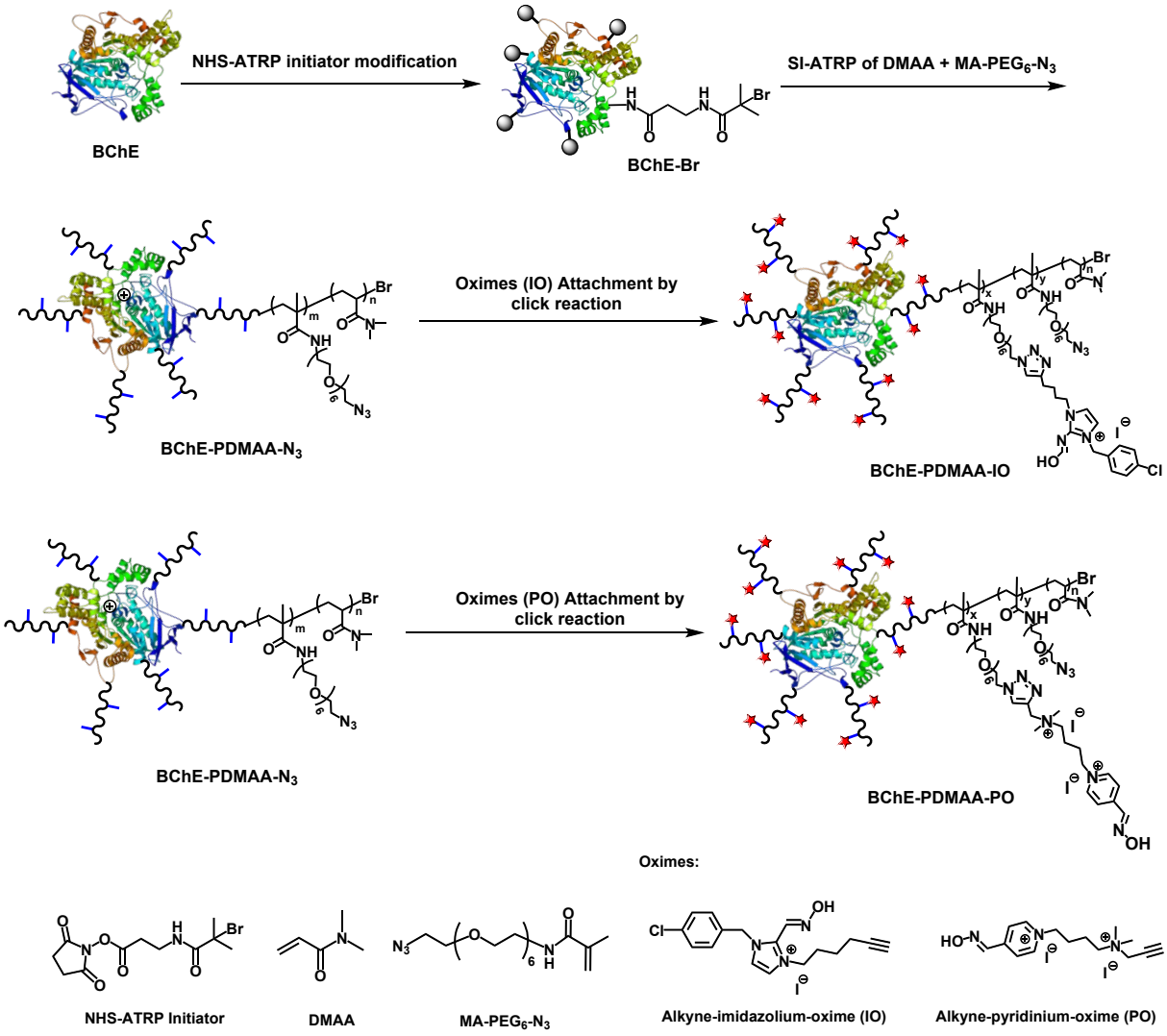

Figure 1. Synthesis of butyrylcholinesterase-polymer-oxime conjugates using atom-transfer radical polymerization (ATRP) and "click" chemistry. Additional acronyms: *N*-hydroxysuccinimide (NHS), *N,N*-dimethylacrylamide (DMAA), *N*-(20-azido-3,6,9,12,15,18-hexaoxaicosyl)methacrylamide (MA-PEG_6-N_3), butyrylcholinesterase (BChE).

We next investigated whether the BChE-PDMAA-OX conjugates retained bioactivity. Using S-butyrylthiocholine iodide (BTC) as a substrate (10 μM, pH 8.0), we compared

conjugate activity to that of native BChE. For BChE-PDMAA-IO_{28}, BChE-PDMAA-IO_{84}, and BChE-PDMAA-IO_{172}, activity was inversely proportional to oxime content (76%, 67% and 54%, respectively). The BChE-PDMAA-PO conjugates also exhibited reduced activity, but with a less pronounced inverse dependence on oxime content (63%, 48% and 48% for BChE-PDMAA-PO_{44}, BChE-PDMAA-PO_{104}, and BChE-PDMAA-PO_{216} respectively) (**Figure 3**). Previous work lead us to hypothesize that the activity loss of BChE-PDMAA-OX conjugates resulted from a combination of polymer attachment and reversible direct oxime inhibition of the enzyme.[23, 71] In addition, Cu^{2+} and Cu-ligand complexes can inhibit cholinesterase activity.[77, 78] The alkyne-imidazolium-oxime had a BChE IC_{50} of approximately 1 μM (**Figure S6**) and the alkyne-pyridinium-oxime BChE IC_{50} was > 50 μM (**Figure S7**). The BChE-PDMAA-OX conjugates reported herein were the highest activity BChE-polymer-oxime conjugates that we have studied to date. We have recently determined the Michaelis-Menten kinetics for the BChE, BChE-Br, BChE-PDMAA-N_3 starting materials.[71]

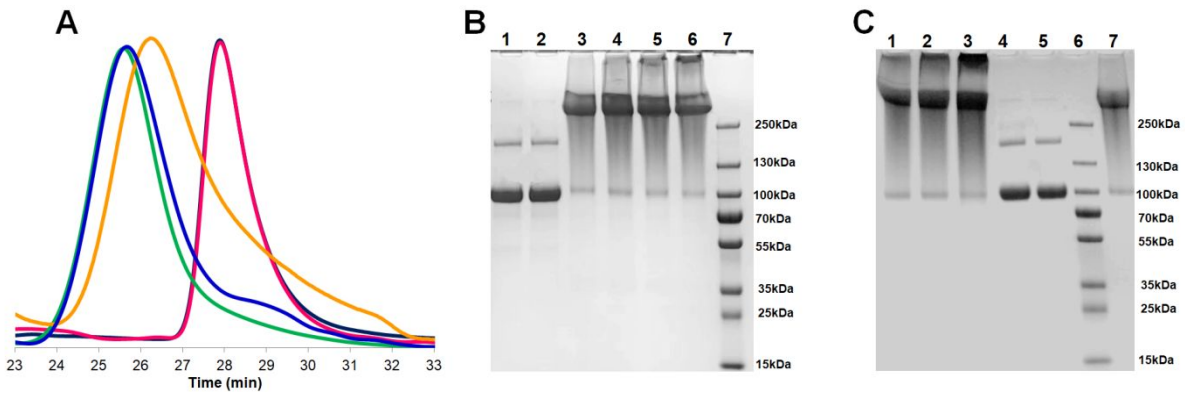

Figure 2. Characterization of BChE, BChE-Br, BChE-PDMAA-N_3 and BChE-PDMAA-OX conjugates. A) GPC analysis of BChE (black curve), BChE-Br (red curve), BChE-PDMAA-N_3 (green curve), BChE-PDMAA-IO_{84} (yellow curve) and BChE-PDMAA-PO_{104} (blue curve) conjugates. B) SDS-PAGE analysis of BChE, BChE-Br, BChE-PDMAA-N_3 and BChE-PDMAA-IO conjugates. Lane 1: BChE; Lane 2: BChE-Br; Lane 3: BChE-PDMAA-N_3; Lane 4: BChE-PDMAA-IO_{28}; Lane 5: BChE-PDMAA-IO_{84}; Lane 6: BChE-PDMAA-IO_{172}; Lane 7: Marker. C) SDS-PAGE analysis of BChE, BChE-Br, BChE-PDMAA-N_3 and BChE-PDMAA-PO conjugates. Lane 1: BChE-PDMAA-PO_{44}; Lane 2: BChE-PDMAA-PO_{104}; Lane 3: BChE-PDMAA-PO_{216}; Lane 4: BChE-Br; Lane 5: BChE; Lane 6: Marker; Lane 7: BChE-PDMAA-N_3.

Since only negatively charged oximates can reactivate BChE active sites, it was important to determine the pK_a's for the monomeric alkyne-imidazolium-oxime and alkyne-pyridinium-oxime.[76] We used a spectroscopic assay and thereby surmised that the oximes groups on BChE-PDMAA-IO and BChE-PDMAA-PO had pK_a's of 8.35 and 8.38, respectively (**Figure S8**). Thus, at pH 6.0 the conjugates would be in their fully protonated

oxime form, and therefore would be unable to undergo either inter- or intramolecular reactivation. Therefore, in order to minimize any reactivation, we inhibited the BChE-polymer-oxime conjugates by fluorogenic OPNAs (Flu-OPNA) analogues at pH 6.0 (vide infra).

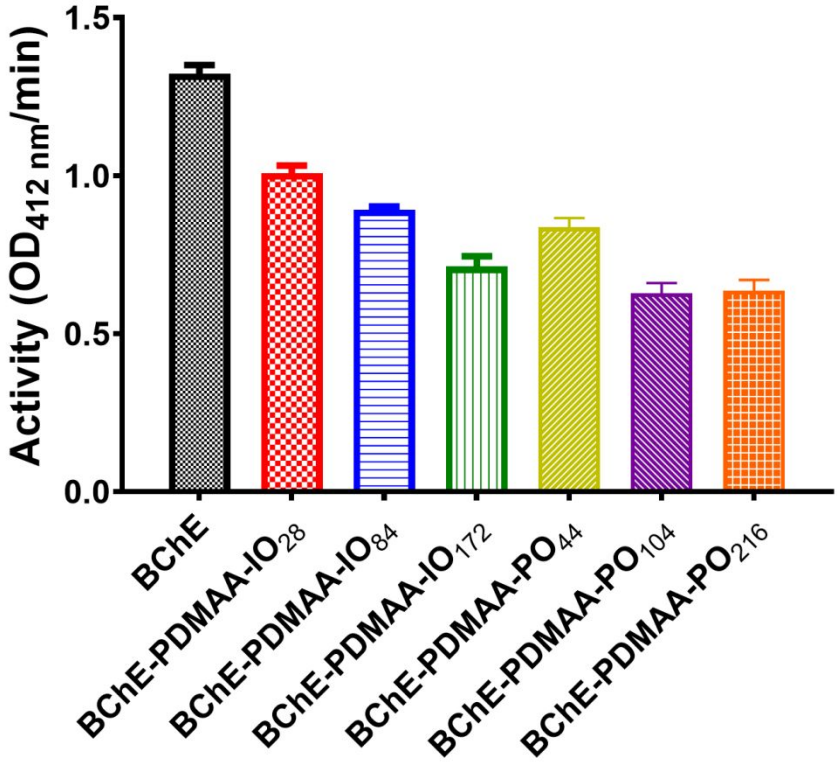

Figure 3. Enzymatic activity of BChE, BChE-PDMAA-IO and BChE-PDMAA-PO conjugates at a single concentration of BTC in PB (50 mM) buffer at pH 8.0. Results are presented as mean values ± standard deviation (n = 3).

Oxime containing copolymers protect BChE against OPs

Two racemic Flu-OPNAs analogues, which were designed to be analogues of VX or cyclosarin after reaction with BChE, were used to inhibit BChE-polymer-oxime conjugates. The Flu-OPNAs contained either an ethyl (E) or cyclohexyl (C) O-alkyl attached to a methyl-phosphoryl (MP) moiety. The fluorescent moiety, 3-cyano-4-methyl-7-hydroxy coumarin (MeCyC-OH), was released upon reaction between the BChE active site serine and Flu-OPNAs (EMP-MeCyC or CMP-MeCyC) resulting in a significant and measurable increase in fluorescence intensity.[72] BChE-PDMAA-OX conjugates (10 or 20 nM) were exposed to a 5-fold stoichiometric excess of Flu-OPNA in sodium phosphate buffer (pH 7.4, 50 mM) and the activity loss was measured over time by removing aliquots and performing an activity assay as described above. Following the addition of Flu-OPNAs, native BChE lost all activity within 10 min (**Figure S9A**). After exposure to EMP-MeCyC, BChE-PDMAA-IO_{28}, BChE-PDMAA-IO_{84}, and BChE-PDMAA-IO_{172} retained 7%, 10% and

15% activity, respectively (**Figure S9A**). The BChE-PDMAA-PO_{44}, BChE-PDMAA-PO_{104}, and BChE-PDMAA-PO_{216} conjugates retained 27%, 28% and 35% activity, respectively (**Figure S9B**). Thus, all the BChE-PDMAA-OX conjugates were all moderately resistant to inactivation by EMP-MeCyC, with the degree of protection being proportional to oxime content and related to oxime structure. Imidazolium-oxime containing copolymers also protected BChE against CMP-MeCyC (**Figure S10A**), though the highest oxime content BChE-PDMAA-PO did not resist inactivation by EMP-MeCyC (**Figure S10B**). The BChE-PDMAA-OX conjugates were all inactivated eventually by a large excess of paraoxon (POX) (**Figure S11**).

To elucidate whether the protection of oxime containing BChE-PDMAA-OX conjugates against Flu-OPNAs was consistent with intramolecular and/or intermolecular interactions, inhibition was monitored in mixtures of native BChE, BChE-PDMAA-OX conjugates and oxime containing copolymers (PDMAA-IO and PDMAA-PO) using the release of fluorescence (the leaving group is MeCyC-OH) upon covalent coupling of Flu-OPNAs to the enzyme (**Figure 4**). The total oxime content was kept constant at 172 and 216

equivalents for BChE-PDMAA-IO and BChE-PDMAA-PO respectively, delivering the oxime by polymer only or enzyme-linked oxime. The presence of free PDMAA-IO enhanced EMP-MeCyC hydrolysis. BChE-PDMAA-IO_{172} with no free PDMAA-IO had the lowest fluorescence intensity increase. The degree of fluorescence intensity increased steadily as the fraction of free PDMAA-IO increased over 20 min (**Figure 4A**). These data, when combined with control experiments, showed that free PDMAA-IO could protect BChE against EMP-MeCyC, thereby indicating that intermolecular protection accounted for the majority of the observed protection. A similar phenomenon was observed in the fluorescence monitoring assay of CMP-MeCyC mixed with BChE, PDMAA-PO and BChE-PDMAA-PO_{216} (**Figure 4B**). CMP-MeCyC did not significantly hydrolyze in PB buffer at pH 7.4 at room temperature over 40 min. PDMAA-PO addition reduced the fluorescence intensity of CMP-MeCyC (the leaving group), and the fluorescence intensity of CMP-MeCyC did not increase upon BChE addition indicating complete inactivation of the active site. Since we know that native BChE stoichiometrically reacts with OPNAs in a 1 to 1 ratio, and the fluorescence intensity of CMP-MeCyC with BChE-PDMAA-PO was higher than with BChE after 40 min, we deduced that one molecule of BChE-PDMAA-PO

could scavenge approximately 1.4 molecules of CMP-MeCyC based on the difference of fluorescence intensity between 100% BChE group and 100% BChE-PDMAA-PO_{216} group.

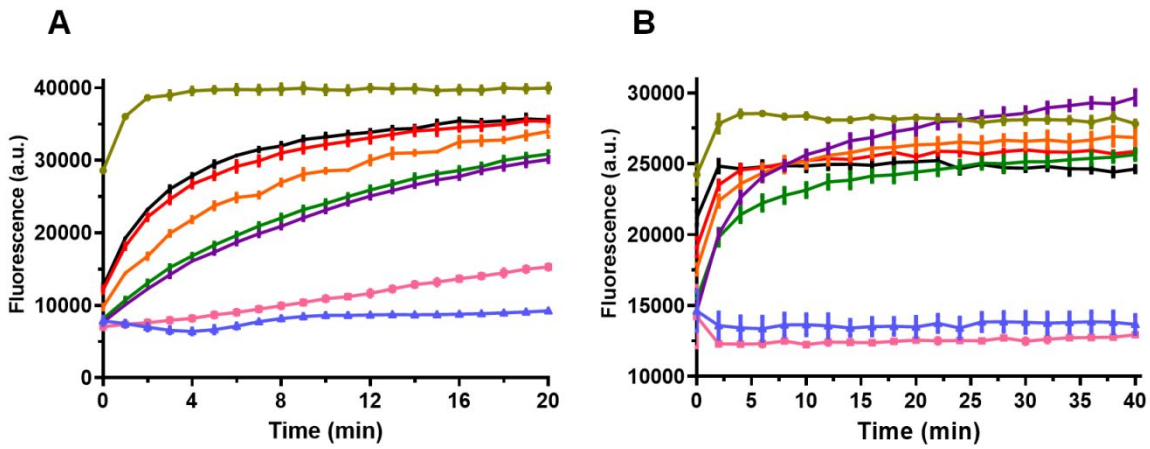

Figure 4. Time-course of Flu-OPNAs (250 nM) degradation by mixtures of BChE, polymer-oxime and BChE-PDMAA-OX conjugates in PB buffer at pH 7.4. (A) Fluorescence intensity of EMP-MeCyC mixed with BChE, PDMAA-IO and BChE-PDMAA-IO_{172} (Imidazolium-oxime/BChE tetramer = 172). (B) Fluorescence intensity of CMP-MeCyC mixed with BChE, PDMAA-PO and BChE-PDMAA-PO_{216} (Pyridinium-oxime/BChE tetramer = 216). Black curve: 100% BChE; Red curve: 80% BChE + 20% BChE-PDMAA-IO_{172} (or BChE-PDMAA-PO_{216}); Orange curve: 50% BChE + 50% BChE-PDMAA-IO_{172} (or BChE-PDMAA-PO_{216}); Green curve: 20% BChE + 80% BChE-PDMAA-IO_{172} (or BChE-PDMAA-PO_{216}); Purple curve: 100% BChE-PDMAA-IO_{172} (or BChE-PDMAA-PO_{216}); Yellow curve: only BChE; Pink curve: only PDMAA-IO; Blue curve: PB buffer. Results are presented as mean values ± standard deviation (n = 3).

To discover whether or not oxime side chains reacted directly with EMP-MeCyC and/or CMP-MeCyC, we monitored the fluorescence intensity of Flu-OPNAs after the addition of free imidazolium- or pyridinium-oximes. Alkyne-imidazolium-oxime addition enhanced the fluorescence of the MeCyC-OH moiety released from EMP-MeCyC and CMP-MeCyC (**Figure S12**). We observed a reduction in the fluorescence of EMP-MeCyC and CMP-MeCyC after the addition of alkyne-pyridinium-oxime (**Figure S12B**). EMP-MeCyC spontaneously hydrolyzed in PB buffer at pH 7.4 at room temperature (**Figure S12A**). The decreased inhibition rates of the conjugates by EMP-MeCyC and CMP-MeCyC could have been caused by a direct reaction between the imidazolium-oxime and Flu-OPNAs. We were able to rule out, however, that the protection by pyridinium-oxime containing copolymer on the surface of BChE-polymer-oxime conjugates was related to a direct reaction between pyridinium-oxime and CMP-MeCyC. The reversible inhibition of the BChE active site by pyridinium-oxime apparently masked the active site toward inhibition by CMP-MeCyC.

Reactivation of Flu-OPNAs-inhibited BChE-PDMAA-OX conjugates

To determine whether free alkyne-imidazolium-oxime and alkyne-pyridinium-oxime were effective reactivators of BChE-EMP and BChE-CMP covalent complexes, Flu-OPNA-inhibited BChE was treated with a 25,000-fold stoichiometric excess of free oxime (0.5 mM) (**Figure S13**). More than 40% of EMP-MeCyC- and CMP-MeCyC- inhibited BChE activity was recovered after the addition of alkyne-pyridinium-oxime within 24 h (**Figure S13**). Addition of 2-PAM (0.5 mM), a positive control reagent, restored more than 65% activity over the same time period. There was no significant reactivation after the addition of the same amount of alkyne-imidazolium-oxime. Alkyne-imidazolium-oxime had an IC_{50} of 1 μM (**Figure S6**). The active site of BChE, inhibited by Flu-OPNA, may be reactivated by the nucleophilic attack of alkyne-imidazolium-oxime, but reversibly inhibited by excess oxime. To determine whether alkyne-imidazolium-oxime could restore the activity of Flu-OPNAs inhibited BChE, EMP-MeCyC-inhibited BChE was treated with different concentrations of alkyne-imidazolium-oxime. Lower concentrations of alkyne-imidazolium-oxime induced the reactivation of EMP-MeCyC inhibited BChE. More than 25% of enzyme activity was recovered 5 hours after the addition of 2 μM alkyne-

imidazolium-oxime, whereas a higher concentration of alkyne-imidazolium-oxime reactivated less than 25% of the activity due to reversible inhibition of BChE (**Figure S14**).

The degree to which the polymer-bound oxime could reactivate Flu-OPNA-inhibited BChE-PDMAA-OX conjugates was determined by first inhibiting the enzyme at low pH (6.0) and then tracking recovery of activity at high pH (8.0).[71] At pH 6.0, more than 2 units below the pK_a of alkyne-imidazolium-oxime and alkyne-pyridinium-oxime, we were able to inhibit the BChE active site of the BChE-PDMAA-OX conjugates without reactivation or interference by the polymer oxime side chains. BChE-PDMAA-OX conjugates and native BChE lost all activity after being mixed with a 10-fold molar excess of Flu-OPNAs at pH 6.0 within 10 min. The EMP-BChE-PDMAA-OX or CMP-BChE-PDMAA-OX conjugates were purified by removing excess Flu-OPNAs by ultrafiltration. Fluorescence intensity analysis showed that more than 99% of free Flu-OPNAs were removed after three cycles of ultrafiltration (**Figure S15**). The EMP-BChE-PDMAA-OX or CMP-BChE-PDMAA-OX conjugates were then diluted 50-fold in PB buffer at pH 8.0 and reactivation was measured using the Ellman assay with BTC as the substrate at pH 8.0 (**Figure 5**).

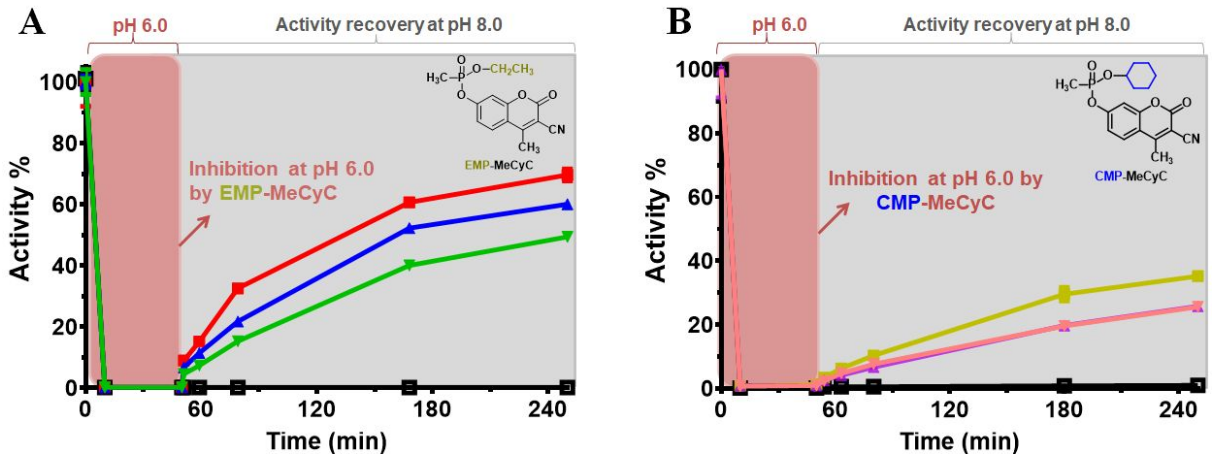

Figure 5. Reactivation assay of BChE-PDMAA-IO conjugates inhibited by EMP-MeCyC (A) and BChE-PDMAA-PO conjugates inhibited by CMP-MeCyC (B). 1 μM BChE-PDMAA-IO, BChE-PDMAA-PO or free BChE were inhibited by 10-fold excess of Flu-OPNAs in PB buffer at pH 6.0 for 10 min, remove the excess of Flu-OPNAs, then diluted 50-fold in PB buffer at pH 8.0. Black curve: BChE; Green curve: BChE-PDMAA-IO_{28}; Blue curve: BChE-PDMAA-IO_{84}; Red curve: BChE-PDMAA-IO_{172}; Pink curve: BChE-PDMAA-PO_{44}; Purple curve: BChE-PDMAA-PO_{104}; Yellow curve: BChE-PDMAA-PO_{216}. Results are presented as mean values ± standard deviation (n = 3).

The degree of reactivation was determined relative to the respective initial activity of native BChE or a given BChE-PDMAA-OX conjugate in the assay. After complete inactivation by EMP-MeCyC, the combination of intra- and intermolecular reactivation was proportional to the oxime content of the polymer for the BChE-PDMAA-IO conjugates (**Figure 5A**). BChE-PDMAA-PO conjugates also recovered significant activity, but with a less pronounced dependence on oxime content (**Figure 5B**). Prolonging the reactivation

time to 24 hours at pH 8.0, for the highest oxime content conjugates, resulted in dramatic levels of reactivation (84% for BChE-PDMAA-IO and 55% for BChE-PDMAA-PO) (**Figure S16**).

We next challenged the performance of imidazolium-oxime containing BChE-PDMAA-IO conjugates as catalytic bioscavengers against VX using the fluorogenic analogue EMP-MeCyC. The enzyme-polymer conjugates were inactivated with a 10-fold molar excess of EMP-MeCyC in PB buffer at pH 6.0, then, after ultrafiltration, we followed the reactivation at pH 8.0 for 200 min. At the end of the reactivation cycle we re-adjusted the pH to 6.0 and again challenged the conjugates with another 10-fold molar excess of EMP-MeCyC. The entire process was repeated three times to determine whether the conjugates could self-reactivate after multiple challenges. In the second and third reactivation cycles, significant activity was recovered for all of the conjugates. We did observe a gradual decrease in the maximal reactivation levels at each consecutive cycle for BChE-PDMMA-IO_{172} conjugate, from 70% to 45% and 20% maximal activity recovery during the first, second and third inhibition/reactivation cycle, respectively (**Figure 6**). This

gradual decline in maximal reactivation level may stem from slow aging of inhibited conjugate active site. The distance of each oxime from the active site of the enzyme would likely influence the ability of a particular oxime to reactivate BChE.[80] Location-specific reactivation, and how it couples with cholinesterase aging, is beyond the scope of this manuscript but will be an important area for future study. The significant reactivation levels with inhibited conjugate after three inhibition/reactivation cycles were consistent, however, with catalytic reactivation of OPNA-inhibited BChE by matrix-conjugated oximes. In totality, these data demonstrated that covalent modification with imidazolium-oxime containing copolymers on the surface of BChE could catalytically decontaminate VX, even when the number of oximes per BChE tetramer molecule was as low as 28.

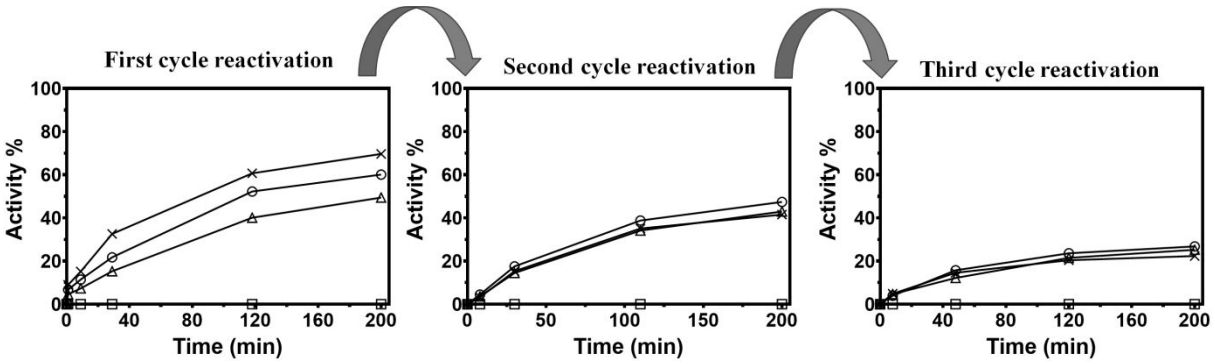

Figure 6. Inactivation and reactivation cycles of EMP-MeCyC-inhibited BChE-PDMAA-IO conjugates. Fork: BChE-PDMAA-IO_{172}; Circle: BChE-PDMAA-IO_{84}; Triangle: BChE-

PDMAA-IO_{28}; Square: BChE. Results are presented as mean values ± standard deviation (n = 3).

CONCLUSIONS

BChE-PDMAA-OX conjugates, synthesized by *in situ* ATRP and click chemistry with imidazolium-oxime and pyridinium-oxime side chains, were remarkable self-reactivators after repeated challenges with organophosphate inhibitors. The catalytic activity of these functional BChE-PDMAA-OX conjugates was dependent on the number of oximes per BChE molecule. The conjugates exhibited both inter- and intra-molecular protection against fluorescent analogues of VX and cyclosarin and functional enzyme activity in the presence of these OPs was prolonged by the conjugated polymer-oximes. After complete inactivation by EMP-MeCyC and CMP-MeCyC at a low pH (6.0), the conjugates recovered up to 84% and 55% activity after self-reactivation pH (8.0) within 5-6 hours. Imidazolium-oxime containing conjugates achieved three cycles of inhibition and auto-reactivation, demonstrating high-performance decontamination bio-scavenging.

ASSOCIATED CONTENT

Supporting Information for this article can be found online.

Complete dataset, working curves, synthesis schemes, UV-VIS absorbance spectrum, additional GPC data, Fluorescence assay, activity assay, inhibition assay and Reactivation assay

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Supervision, Writing - review & editing. Alan J. Russell: Supervision, Funding acquisition, Writing - review & editing.

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Conflict of Interest

AR and KM have founded a company that is commercializing protein-ATRP.

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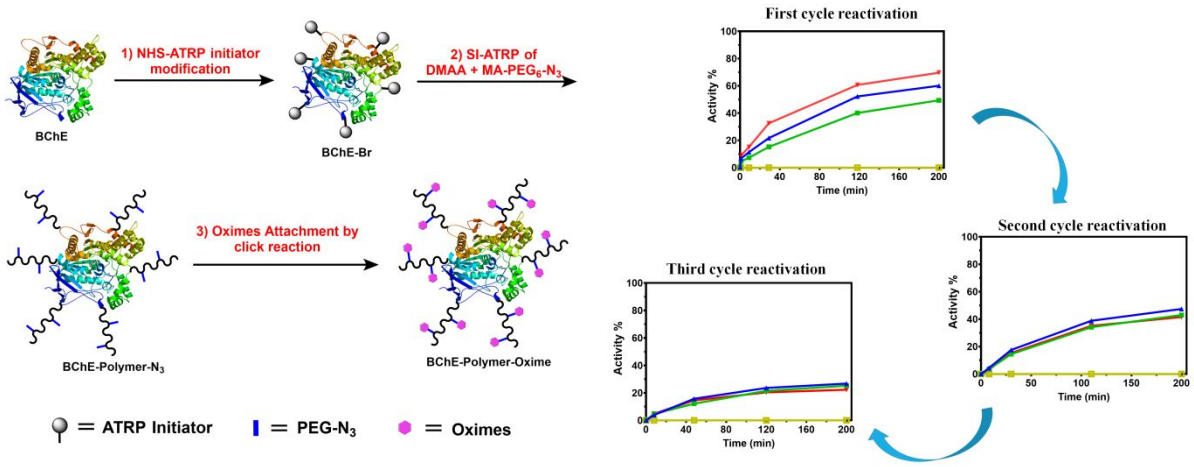

Butyrylcholinesterase (BChE)-polymer-oxime conjugates were designed and synthesized by in-situ growth of azide containing copolymers and azide-alkyne "click" chemistry. BChE-polymer-oxime conjugates self-reactivate at up to 85%.