

## DESIGN AND SYNTHESIS OF A PYRIDONE-BASED PHOSPHOTYROSINE MIMETIC

Jian-Min Fu and Arlindo L. Castelhanos\*

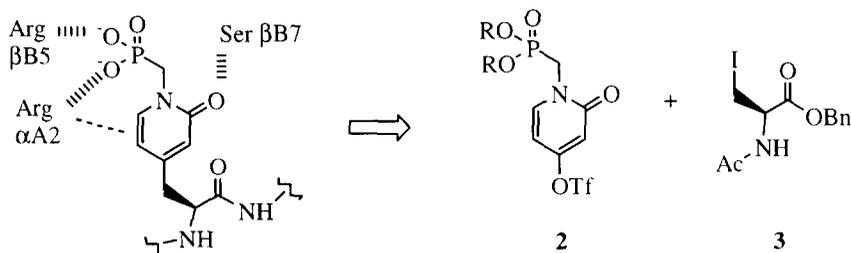
Cadus Pharmaceutical Corporation, 777 Old Saw Mill River Road, Tarrytown, NY, 10591, U.S.A.

Received 20 July 1998; accepted 1 September 1998

**Abstract:** A novel pyridone-based tyrosine analog, **6**, has been designed to mimic the binding interaction of SH2 domains with phosphotyrosine (pTyr) containing peptides. Synthesis of **6** features a key Pd catalyzed coupling of  $\beta$ -iodoalanine with phosphonomethyl 4-pyridone triflate. © 1998 Elsevier Science Ltd. All rights reserved.

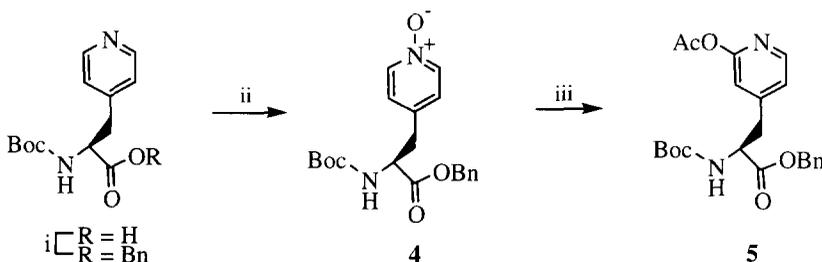
SH2 domains are phosphotyrosine-binding modules found in a variety of important signal-transducing molecules such as nonreceptor tyrosine kinases, phosphatases, and regulatory adapter proteins. Inhibitors that block SH2 domain binding have potential utility in a wide variety of therapeutic areas including metabolic diseases, cancer, inflammation and allergy.<sup>1</sup> Our interest lies with the high affinity IgE receptor, Fc $\epsilon$ RI, and associated tyrosine kinases and phosphatase PTP-1C.<sup>2</sup> Aggregation of this receptor by antigen-antibody complexes leads to the activation of *Lyn* and *Syk* with rapid phosphorylation of tyrosine residues in the  $\beta$ - and  $\gamma$ -chain cytoplasmic ITAM (immunoreceptor tyrosine-based activation motif) regions of the receptor. Association of the SH2 domain of *syk* with the phosphorylated  $\gamma$ -chain of Fc $\epsilon$ RI in basophils and mast cells leads to downstream activation signals and the allergic response.<sup>3</sup>

Structural detail provided from X-ray and NMR studies of high affinity pTyr containing peptides has guided the design of SH2-directed ligands.<sup>4</sup> Selective ligands for SH2 domains containing pTyr or phosphate-resistant pTyr analogs and pseudo-peptidic elements, have been developed for SH2 domains of pp60<sup>c-src</sup>, p85 subunit of PI-3 kinase, and other proteins.<sup>5</sup> Ligand studies with (phosphonomethyl) phenylalanine (Pmp), wherein the phosphate ester oxygen (>COPO<sub>3</sub>H<sub>2</sub>) has been replaced by a methylene unit (>CH<sub>2</sub>PO<sub>3</sub>H<sub>2</sub>) and Pmp analogs bearing fluorine or hydroxyl, indicate a pK<sub>A2</sub> requirement (pTyr pK<sub>A2</sub> = 5.7 vs. Pmp pK<sub>A2</sub> = 7.1) and an H-bond to the phosphate ester oxygen.<sup>6</sup> It occurred to us that the inductive effect of a heterocycle on phosphonate acidity (Het-CH<sub>2</sub>PO<sub>3</sub>H<sub>2</sub>) would result in a pK<sub>A2</sub> close to that of pTyr.<sup>7</sup> As indicated in Figure 1, the pyridone methylphosphonate moiety was expected to maintain ionic and H-bonding interactions observed in phosphate-based ligands.<sup>8</sup>



**Figure 1.** Modeled interactions with a SH2 domain and retrosynthesis of pyridone pTyr mimetic

The first approach in preparing the key pyridone pTyr mimetic began with commercial (4-pyridinyl)alanine. Since pyridine to pyridone conversion has been reported for simple systems,<sup>9</sup> rearrangement of  $N^\alpha$ -Boc-(4-pyridinyl-N-oxide)alanine benzyl ester to the corresponding (4-pyridone)alanine with acetic anhydride was investigated (Scheme 1). In the event, we established the presence of **5** in crude product by MS but the yield was low and pure material was elusive.



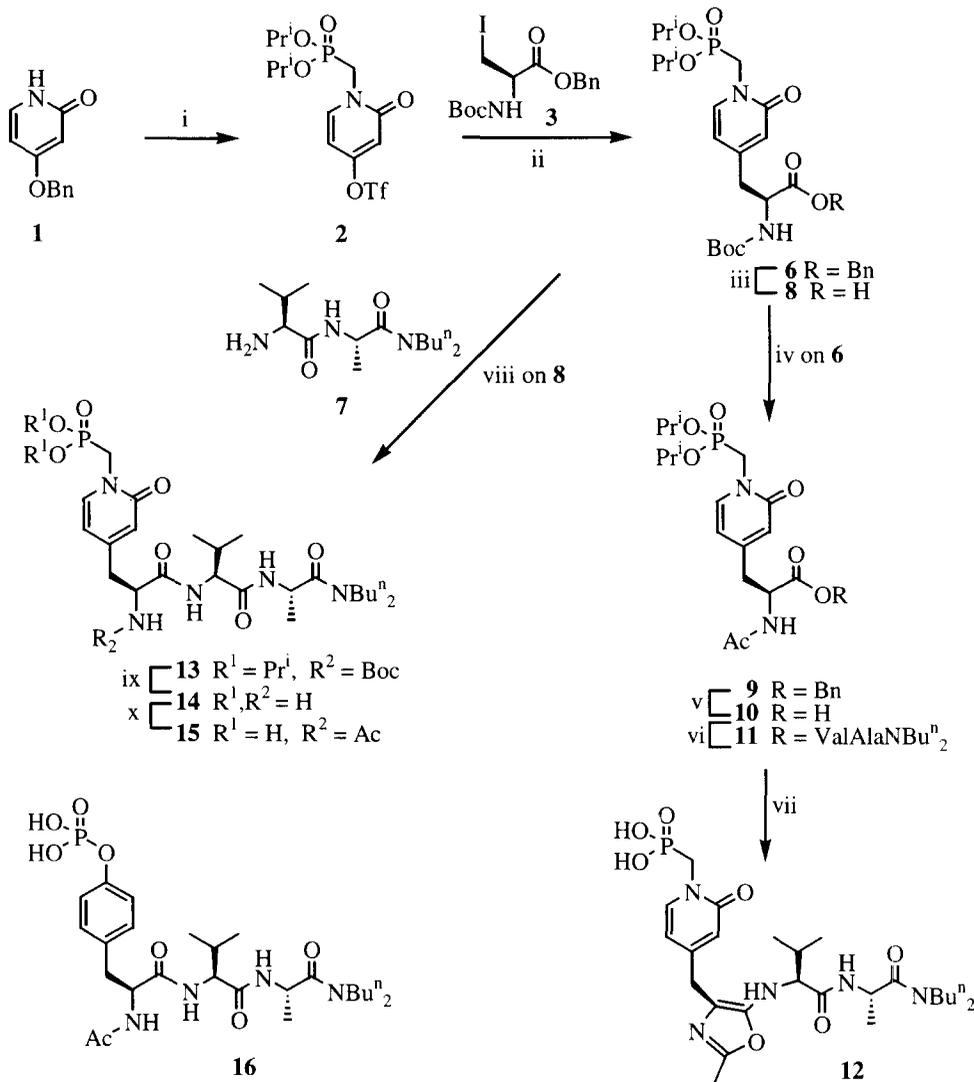
**Scheme 1.** (i)  $\text{CsCO}_3$ , DMF/ $\text{H}_2\text{O}$ , BnBr, 76% (ii) *m*-CPBA,  $\text{CH}_2\text{Cl}_2$ , 86% (iii)  $\text{Ac}_2\text{O}$ ,  $65^\circ\text{C}$ , 2.5 h

Alternatively, the palladium catalyzed cross coupling of triflate **2**, already possessing the phosphonate moiety, and  $\beta$ -iodoalanine **3** appeared to be a feasible, convergent synthesis of **6**<sup>10</sup> (Scheme 2). Starting with commercial 4-(*O*-benzyl)pyridone, alkylation with  $\text{BrCH}_2\text{P}(\text{O})(\text{O}^i\text{Pr})_2$  and  $\text{K}_2\text{CO}_3$  in acetonitrile at reflux gave *N*-alkylated product in 98% yield. The benzyl group of the phosphonomethylpyridone intermediate was then removed by hydrogenolysis in 96% isolated yield. The triflate moiety was introduced with triflic anhydride and triethylamine at  $-78^\circ\text{C}$  for 5 min in 70% isolated yield, longer reaction time led to lower yields of triflate product. Palladium catalyzed coupling of **2** with the zinc reagent of  $\beta$ -iodoalanine, prepared according to Jung,<sup>11</sup>  $\text{Pd}_2(\text{dba})_3/\text{o-tol}_3\text{P}$  at  $55^\circ\text{C}$ , provided the desired product **6** reproducibly in 43% yield.

Assembly of a pyridone-based ligand with recognition for SH2 domains involved the additional condensation of **6** with the peptidomimetic **7**, an entity developed for the P+1 to P+3 pockets,<sup>12</sup> and  $N^\alpha$ -acetylation of the *N*-terminus. Thus, treatment of **6** with TFA and acetylation with acetic anhydride proceeded in 76% yield for the two-step transformation to give **9**. Hydrogenolysis with  $\text{H}_2/\text{Pd}(\text{OH})_2/\text{EtOAc}$  gave the carboxylic acid **10** in 94% yield. Coupling of **10** with ValAla dibutyl amide **7**, afforded **11** as a single isomer revealing stereochemical integrity in the palladium coupling step. Unmasking of the phosphonate isopropyl esters with typical conditions for ethyl phosphate esters, namely iodotrimethylsilane and *N,O*-bis(trimethylsilyl) acetamide,<sup>13</sup> led to the oxazole **12** in 51% isolated yield. To avoid this intramolecular cyclization and dehydration of the acetamide moiety, the *N*-acetyl group would need to be introduced after phosphonate ester hydrolysis. This was achieved by first coupling *N*-Boc acid **8** with **7** (EDCI/HOBT) to give **13** in 85% yield. Treatment of **13** with bromotrimethylsilane in acetonitrile and subsequently aqueous acetone resulted in isopropyl ester hydrolysis and Boc removal. Acetylation of the zwitterionic intermediate **14** with  $\text{Ac}_2\text{O}$  gave the desired target compound **15** as a single isomer as determined by  $^1\text{H}$  and  $^{13}\text{C}$  NMR analysis.<sup>14</sup>

The corresponding phosphate **16** (reported<sup>12</sup> to block the association of PDGF- $\beta$  receptor with p85 C-SH2;  $\text{IC}_{50} = 0.077 \mu\text{M}$ ) was also assembled for comparative biochemical evaluation. BIAcore analysis of **15** showed 50% inhibition of binding of the p85 *N*-terminal SH2 domain to a CD19 phosphopeptide at  $50 \mu\text{M}$ .

By comparison, the canonical phosphopeptide **16** exhibited 98% inhibition at 20  $\mu\text{M}$ . This result indicates a moderate effect by the pyridone heterocyclic on phosphonate  $\text{pK}_{\text{A}2}$ .<sup>7</sup> Moreover, the Arg  $\alpha\text{A}2$ -aromatic ( $\pi$ -cation) interaction may be compromised in the pyridone case.<sup>6c,8</sup> We are continuing our studies with other SH2 domains in order to determine the potential utility of the pyridone phosphonate as a pTyr mimetic.



**Scheme 2.** (i) (a)  $\text{K}_2\text{CO}_3$ ,  $\text{CH}_3\text{CN}$ ,  $\text{BrCH}_2\text{P}(\text{O})(\text{O}^i\text{Pr})_2$ , reflux, 48 h, 98%; (b)  $\text{H}_2/\text{Pd}/\text{C}$ , MeOH, rt, 2 h, 96%; (c)  $\text{Et}_3\text{N}$ ,  $(\text{CF}_3\text{SO}_2)_2\text{O}$ ,  $\text{CH}_2\text{Cl}_2$ ,  $-78^\circ\text{C}$ , 5 min, 70% (ii) Zn dust,  $\text{Pd}_2(\text{dba})_3/o\text{-tolylP}/\text{THF-DMA}$ ,  $55^\circ\text{C}$ , 43% (iii)  $\text{H}_2$ , Pd/C, MeOH, rt, 14 h, 99% (iv) (a) TFA,  $\text{CH}_2\text{Cl}_2$ , rt, 5 min; (b)  $\text{Ac}_2\text{O}$ , NMM,  $\text{CH}_2\text{Cl}_2$ ,  $0^\circ\text{C}$  to rt, 76% for two steps; (v)  $\text{H}_2$ ,  $\text{Pd}(\text{OH})_2$ , EtOAc, 94% (vi) **7**, EDCI/HOBT, DDMF,  $0^\circ\text{C}$  to rt, 85% (vii) TMSI, BSTFA,  $\text{CH}_2\text{Cl}_2$ ,  $0^\circ\text{C}$  to rt; TFA- $\text{H}_2\text{O}$ - $\text{CH}_3\text{CN}$ , rt, 1 h, 51% (viii) **7**, EDCI, HOBT, DMF,  $0^\circ\text{C}$  to rt, 14 h, 85% (ix) TMSBr,  $\text{CH}_3\text{CN}$ , rt, 2 h;  $\text{H}_2\text{O}$ -acetone, rt, 14 h (x)  $\text{Ac}_2\text{O}$ , NMM, DMF,  $0^\circ\text{C}$  to rt, 14 h.

**Acknowledgements:** We are indebted to Dr. John Cambier and Sara Johnson, National Jewish Medical and Research Center, Denver, CO for the BIAcore analysis and to Dr. Chris Pleiman for preliminary studies.

## References and Notes

1. Plummer, M. S.; Lunney, E. A.; Para, S. K.; Vara Prasad, J. V. N.; Shahripour, A.; Singh, C. J. S.; Humblet, C.; Fergus, J. H.; Marks, J. S.; Sawyer, T. K. *Drug Design and Discovery* **1996**, *13*, 75.
2. Cambier, J. C.; Pleiman, C. M.; Clark, M. R. *Annu. Rev. Immunol.* **1994**, *12*, 457.
3. Kimura, T.; Kihara, H.; Bhattacharyya, S.; Sakamoto, H.; Appella, E.; Siraganian, R. P. *J. Biol. Chem.* **1996**, *271*, 27962.
4. Plummer, M. S.; Holland, D. R.; Shahripour, A.; Lunney, E. A.; Fergus, J. H.; Marks, J. S.; McConnell, B.; Mueller, W. T.; Sawyer, T. K. *J. Med. Chem.* **1997**, *40*, 3719.
5. (a) Domchek, S. M.; Auger, K. R.; Chatterjee, S.; Burke, Jr., T. R.; Shoelson, S. E.; *Biochemistry*, **1992**, *31*, 9865. (b) Wange, R. L.; Isakov, N.; Burke, Jr., T. R.; Otaka, A.; Roller, P. P.; Watts, J. D.; Aebersold, R.; Samelson, L. E. *J. Biol. Chem.* **1995**, *270*, 944. (c) Wange, R. L.; Isakov, N.; Burke, Jr., T. R.; Otaka, A.; Roller, P. P.; Watts, J. D.; Aebersold, R.; Samelson, L. E. *J. Biol. Chem.* **1995**, *270*, 944. (d) Plummer, M. S.; Lunney, E. A.; Para, K. S.; Shahripour, A.; Stankovic, C. J.; Humblet, C.; Fergus, J. H.; Saltiel, A.; Sawyer, T. K. *Bioorg. Med. Chem.* **1997**, *5*, 41. (e) Furet, P.; Gay, B.; G-Echeverria, C.; Rahuel, J.; Fretz, H.; Schoepfer, J.; Caravatti, G. *J. Med. Chem.* **1997**, *40*, 3551. (f) Revesz, L.; Blum, E.; Manning, U.; Demange, B. J.; Widmer, A.; Zuber, J.-F. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 2875.
6. (a) Burke, Jr., T. R.; Smyth, M. S.; Otaka, A.; Nomizu, M.; Roller, P. P.; Wolf, G.; Case, R.; Shoelson, S. E. *Biochemistry* **1994**, *33*, 6490. (b) Burke, Jr., Smyth, M. S.; Otaka, A.; Roller, P. P. *Tetrahedron Lett.* **1993**, *34*, 4125. (c) Liu, W.-Q.; Roques, B. P.; Garbay C. *Tetrahedron Lett.* **1997**, *38*, 1389. The high affinity of peptides bearing  $>CF_2PO_3H_2$  has also been attributed to a direct interaction of the fluorines with active site residues and not to lower pK<sub>a</sub> values, see: Chen, L.; Wu, L.; Otaka, A.; Smith, M. S.; Roller, P. R.; Burke, T. R.; den Hertog, J.; Zhang, Z.-Y. *Biochem. Biophys. Res. Comm.* **1995**, *216*, 976.
7. The pK<sub>A2</sub> of  $NH_2CH_2PO_3H_2$  is 5.9. Pyridone- $CH_2PO_3H_2$  and Pmp pK<sub>A2</sub> were subsequently calculated as 7.1 and 7.7 ± 0.3, respectively, using Advanced Chemistry Development, Inc., prediction software.
8. (a) Waksman, G.; Shoelson, S. E.; Pant, N.; Cowburn, D.; Kuriyan, J. *Cell* **1993**, *72*, 779. (b) Eck, M. J.; Shoelson, S. E.; Harrison, S. C. *Nature* **1993**, *362*, 87.
9. Kelly, T. R.; Bridger, G. J.; Zhao, C. *J. Am. Chem. Soc.* **1990**, *112*, 8024.
10. A similar approach to the synthesis of 4-phosphono(difluoromethyl)-L-phenylalanine and pyridylalanine regioisomers starting from 2-, 3- and 4-bromopyridine and β-iodoalanine have been described, see: (a) Smyth, M. S.; Burke, Jr., T. R. *Tetrahedron Lett.* **1994**, *35*, 551 (b) Walker, M. A.; Kaplita, K. P.; Chen, T.; King, H. D. *Synlett* **1997**, 169.
11. Jung, M. E.; Starkey, L. S. *Tetrahedron*, **1995**, *53*, 8815.
12. Eaton, S. R.; Cody, W. L.; Kent, D. R.; Paneck, R. L.; Lu, G. H.; Dahrting, T. K.; Doherty, A. M. "Peptidomimetic inhibitors of the association of platelet derived growth factor-β receptor with phosphatidylinositol 3-kinase", 25<sup>th</sup> National Medicinal Chemistry Symposium, Ann Arbor Michigan, June 18-22.
13. Szardenings, A. K.; Gordeev, M. F.; Patel, D. V. *Tetrahedron Lett.* **1996**, *37*, 3635.
14. Compound **8**: Oil; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>) δ 1.21 (s, 3H), 1.24 (s, 3H), 1.28 (s, 3H), 1.31 (s, 3H), 1.43 (s, 9H), 2.94–3.02 (m, 2H), 4.28–4.75 (m, 5H), 5.40 (br, 1H), 6.26–6.29 (d, J = 6.6 Hz, 1H), 6.56 (s, 1H), 7.43–7.46 (d, J = 6.6 Hz, ArH). MS (ES): 461 (M<sup>+</sup> + 1), 405 (M<sup>+</sup> - C(CH<sub>3</sub>)<sub>3</sub>). Compound **13**: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>) δ 0.84–0.95 (m, 12H), 1.17–1.55 (m, 32H), 2.01–2.13 (m, 2H), 2.74–3.29 (m, 6H), 3.37–3.52 (m, 2H), 4.28–4.41 (m, 4H), 4.57–4.85 (m, 4H), 5.46–5.50 (d, J = 8 Hz, 1H), 6.13–6.17 (d, J = 8 Hz, 1H), 6.43 (s, 1H), 6.92–6.96 (d, J = 8 Hz, 1H), 7.19–7.23 (d, J = 8 Hz, 1H), 7.38–7.42 (d, J = 8 Hz, 1H). MS (ES): 742.0 (M<sup>+</sup>+1). Compound **15**: MS (ES): 600.3 (M<sup>+</sup> + 1).