

# Sinulamide: an H,K-ATPase Inhibitor from a Soft Coral *Sinularia* sp.<sup>1</sup>

Noriko U. Sata, Michihiro Sugano, Shigeki Matsunaga,  
and Nobuhiro Fusetani\*

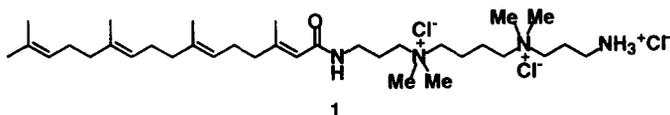
Laboratory of Aquatic Natural Products Chemistry, Graduate School of Agricultural and Life Sciences, The University of Tokyo,  
Bunkyo-ku, Tokyo 113-8657, Japan

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**Abstract:** Sinulamide (1), a new tetraprenylated spermine derivative, has been isolated from a soft coral *Sinularia* sp. as an H,K-ATPase inhibitor. The structure was assigned on the basis of spectroscopic data and confirmed by a total synthesis. © 1999 Elsevier Science Ltd. All rights reserved.

**Keywords:** Coelenterates; Enzyme inhibitors; Biologically active compounds; Natural products

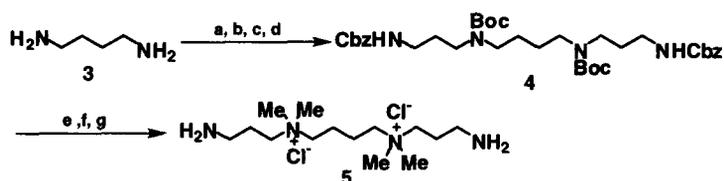
Polyamine metabolites are reported in increasing numbers from marine sources [2]. Soft corals of the genus *Sinularia* often contain acylated spermidine derivatives [3-5]. In our screening of Japanese marine invertebrates for inhibitors of gastric H,K-ATPase [6-8], the hydrophilic extract of *Sinularia* sp. collected in western Japan exhibited potent activity. Bioassay-guided isolation afforded an active substance named sinulamide, which was initially assigned structure 1 based on spectral data [9]. We now have succeeded in refining its structure by a combination of spectral analysis and total synthesis. This paper describes isolation and structure elucidation.



The water-solubles of the 70% EtOH extract of *Sinularia* sp. (500 g wet weight) collected off Tatsukushi on Shikoku Island were chromatographed on TSK G3000S gel with H<sub>2</sub>O, MeOH/H<sub>2</sub>O (1:1), MeOH/H<sub>2</sub>O (7:3), MeOH, and acetone. The three active late fractions were combined and gel-filtered on Toyopearl HW-40 with MeOH followed by ODS HPLC with MeOH/H<sub>2</sub>O/AcOH (45:55:0.1) to yield 120 mg of sinulamide (1) as a colorless gum. Sinulamide (1) showed ion peaks at  $m/z$  581 (M-Cl)<sup>+</sup> and  $m/z$  651 (M+Cl)<sup>-</sup> in the positive and

negative mode FAB mass spectra, respectively. Considering  $^{13}\text{C}$  NMR data as well as distribution of isotope peaks in the mass spectra, a molecular formula of  $\text{C}_{34}\text{H}_{66}\text{Cl}_2\text{N}_4\text{O}$  was established. The IR spectrum revealed the presence of conjugated amide ( $1660$  and  $1640\text{ cm}^{-1}$ ) and amine ( $3350$ , and  $3250\text{ cm}^{-1}$ ) functions. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra exhibited characteristic signals for a linear diterpenoid unit, e.g. five olefinic methyls [ $\delta_{\text{H}}$  1.77, 1.62, 1.55 and 1.53 (6H);  $\delta_{\text{C}}$  26.0, 25.3, 18.5, 16.2 (2C)], four tri-substituted olefins [ $\delta_{\text{H}}$  5.72, 5.12, 5.08 (2H);  $\delta_{\text{C}}$  154.1s, 134.4s, 133.8s, 129.9s, 123.4d (2C), 123.2d, 117.6d], and allylic methylenes ( $\delta_{\text{H}}$  1.90-2.12). One of the olefins was vicinal to a polar group ( $\delta_{\text{C}}$  154.1 and 117.6), thereby suggesting the terpenoid portion terminated in a carbonyl function. This was supported by fragment ions at  $m/z$  388, 319, 251, 184, and 103 in the positive ion FABMS. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra also contained two exchangeable protons [ $\delta$  8.62 (br, 2H) and 8.27 (br, 1H)], four *N*-methyls [ $\delta_{\text{H}}$  3.06 (6H, s) and 3.08 (6H, s);  $\delta_{\text{C}}$  49.7 (4C)], six nitrogen-bearing methylenes [ $\delta_{\text{C}}$  64.4 (2C), 63.6, 62.0, 36.9 and 34.1], and four additional methylenes [ $\delta_{\text{C}}$  23.9, 22.1 and 20.6 (2C)], which indicated that a  $N^2,N^2,N^3,N^3$ -tetramethylspermine unit was linked to the diterpenoid unit through an amide bond. The downfield shifts for four nitrogen-bearing methylene carbons indicated that the two middle nitrogens were both *N,N*-dimethylated to form ammonium salts. The presence of a primary amide was confirmed by formation of a monoacetamide [ $m/z$  623 (M-Cl) $^+$ ;  $\delta_{\text{H}}$  1.81;  $\delta_{\text{C}}$  22.5 and 167.4] upon treatment with  $\text{Ac}_2\text{O}$ /pyridine. The  $^{13}\text{C}$  NMR chemical shift ( $\delta_{\text{C}}$  26.0) for one of the olefinic methyls suggested the presence of a *Z*-olefin. Unfortunately, further spectral studies were hampered due to decomposition during storage in an NMR tube.

Scheme 1

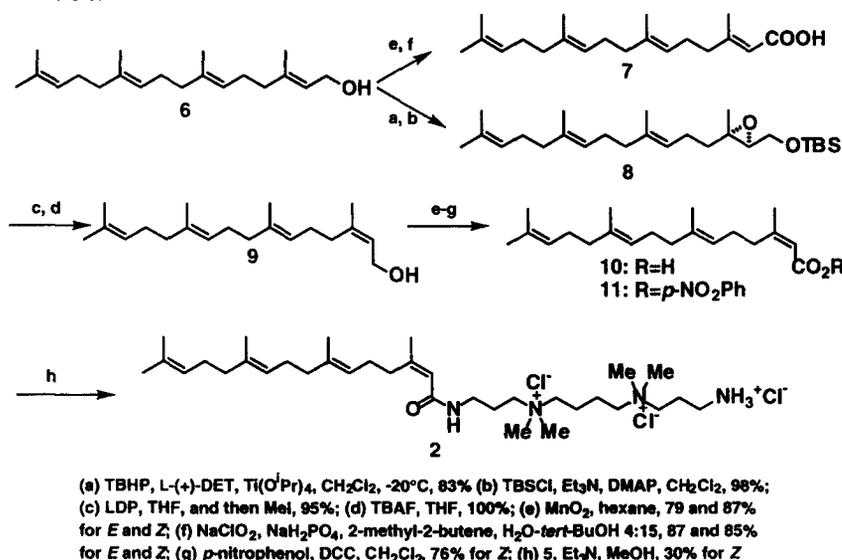


(a)  $\text{CH}_2=\text{CHCN}$ , MeOH, 92%; (b)  $\text{Boc}_2\text{O}$ ,  $\text{Et}_3\text{N}$ , 98%; (c)  $\text{NaBH}_4$  /  $\text{CoCl}_2$ , MeOH, 80%;  
 (d)  $\text{CbzCl}$ , 1N  $\text{NaOH}\cdot\text{Et}_2\text{O}$ , 58%; (e) TFA,  $\text{CH}_2\text{Cl}_2$ , 91%; (f) MeI,  $\text{KHCO}_3$ , MeOH, 80%;  
 (g)  $\text{HBr}\cdot\text{AcOH}$  1:3, 100%

In order to confirm the structure of sinulamide we attempted to synthesize the all-*E* isomer 1. Synthesis of  $N^2,N^2,N^3,N^3$ -tetramethylspermine (5) was begun with dicyanoethylation [10] of putrescine (3) which was followed by Boc protection, reduction with  $\text{NaBH}_4/\text{CoCl}_2$  [11], and Cbz protection to furnish 4. Fully protected spermine 4 was treated with TFA to generate two secondary amines which were methylated with MeI/ $\text{KOH}\cdot\text{K}_2\text{CO}_3$  [12]. The deprotection of the Cbz group with  $\text{HBr}/\text{AcOH}$  afforded the desired compound 5

(Scheme 1). Geranylgeranoic acid (**7**) was prepared from geranylgeraniol (**6**) by two oxidations with  $\text{MnO}_2$  [13] and then with  $\text{NaClO}_2$  [14]. Coupling of **5** and **7** was catalyzed by DCC (Scheme 2). Although the major product was the digeranylgeranoyl amide, we were able to isolate the desired product **1** [15] by HPLC. Synthetic **1** showed NMR data almost superimposable on those of the natural product except for the chemical shift of the olefinic methyl in the conjugated olefin (natural product:  $\delta_{\text{H}}$  1.77,  $\delta_{\text{C}}$  26.0; synthetic product:  $\delta_{\text{H}}$  2.11,  $\delta_{\text{C}}$  18.6). Therefore, it was concluded that sinulamide had *Z*-geometry of the pertinent olefin, *i.e.* geranylneroylamide of  $N^2, N^2, N^3, N^3$ -tetramethylspermine.

Scheme 2



We then tried to prepare geranylneroyl acid (**10**), which could be prepared from **6** by inversion of the terminal olefin and oxidation of the primary alcohol. Geranylgeraniol was subjected to Sharpless epoxidation [16] followed by protection with TBS to furnish **8** which was treated with lithium diphenylphosphide and then with MeI; deprotection of TBS group afforded geranylnerol (**9**) [17]. Oxidation of **9** as described above furnished **10** which was converted to the *p*-nitrophenyl ester **11** which was coupled with **5** to yield a compound indistinguishable from natural sinulamide. Therefore, the structure of sinulamide is **2** [18].

Natural sinulamide (**2**) not only inhibits H,K-ATPase with an  $\text{IC}_{50}$  value of  $5.5 \mu\text{M}$ , but also is cytotoxic against L1210 and P388 with  $\text{IC}_{50}$  values of 3.1 and  $4.5 \mu\text{g/mL}$ , respectively. It is the first acylated spermine derivative of soft coral origin; however, acylated spermidines are known from soft corals of the same genus [3-5].

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## References and Notes

- [1] Bioactive marine metabolites. 88. Part 87: Sata NU, Wada S, Matsunaga S, Watabe S, van Soest RWM, Fusetani N. submitted.
- [2] Faulkner DJ. *Nat. Prod. Rep.* 1998;113-158 and previous compilations.
- [3] Kazlauskas R, Murphy PT, Ravi BN, Sanders RL, Wells R. J. *Aust. J. Chem.* 1982;35:69-75.
- [4] Schmitz FJ, Hollenbeak, KH, Prasad RS. *Tetrahedron Lett.* 1979;36:3387-3390.
- [5] Choi Y, Schmitz FJ. *J. Nat. Prod.* 1997;60:495-496.
- [6] Fusetani N, Sugano M, Matsunaga S, Hashimoto K. *Tetrahedron Lett.* 1987;28:4311-4312.
- [7] Fusetani N, Sugano M, Matsunaga S, Hashimoto K, Shikama H, Ohta A, Nagano H. *Experientia*, 1987;43:1233-1234.
- [8] Fusetani N, Sugano M, Matsunaga S, Hashimoto K. *Experientia*, 1987;43:1234-1235.
- [9] Fusetani N. *New J. Chem.* 1990;14:721-728.
- [10] Israel M, Rosenfield JS, Modest EJ. *J. Med. Chem.* 1964;1:710-717.
- [11] Buhleier E, Wehner W, Vögtle F. *Synthesis* 1978:155-158.
- [12] Chen FCM, Benoiton NL. *Can. J. Chem.* 1976;54:3310-3311.
- [13] Fatiadi AJ. *Synthesis* 1976:65-104 and 133-167.
- [14] Isobe M, Ichikawa Y, Bai DL, Masaki H, Goto T. *Tetrahedron* 1987;43: 4767-4776.
- [15] 1: Colorless powder; TLC on cellulose,  $R_f$  0.20 [*n*-BuOH/AcOH/H<sub>2</sub>O (4:1:2)], positive to Dragendorff and ninhydrin reagents]; <sup>1</sup>H NMR (CD<sub>3</sub>OD, 500 MHz)  $\delta_H$  5.69 (s, 1H), 5.12 (t, 1H,  $J=7.0$  Hz), 5.09 (t, 1H,  $J=7.0$ ), 5.08 (t, 1H,  $J=6.5$ ), 3.40 (m, 4H), 3.14 (m, 8H), 3.10 (m, 8H), 3.05 (m, 4H), 2.17 (m, 2H), 2.14 (m, 2H), 2.11 (s, 3H), 2.07 (m, 6H), 1.98 (m, 6H), 1.85 (m, 4H), 1.65 (s, 3H), 1.61 (s, 3H), 1.58 (s, 6H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 125MHz)  $\delta_C$  170.2s, 155.9s, 137.0s, 136.0s, 132.1s, 125.4d (2C), 124.5d, 118.9d, 64.8t, 64.5t, 63.7t, 62.2t, 51.5q (4C), 41.9t, 40.8t (2C), 37.6t, 37.0t, 27.8t, 27.6t, 27.2t, 25.9q, 24.1t, 22.1t, 20.6t (2C), 18.6q, 17.7q, 16.1q (2C); FABMS (glycerol) *m/z* (rel int.) 581 (5), 531 (0.5), 479 (1.5), 443 (3), 389 (2), 344 (37), 193 (28), 157 (18), 143 (17), 100 (100), 58 (68), 41 (28).
- [16] Katsuki T, Sharpless KB. *J. Am. Chem. Soc.* 1980;102:5974-5976.
- [17] Vedejs E, Fuchs PL. *J. Am. Chem. Soc.* 1973;95: 822-825.
- [18] 2: Colorless powder; TLC on cellulose,  $R_f$  0.20 [*n*-BuOH/AcOH/H<sub>2</sub>O (4:1:2)]; IR (KBr)  $\nu_{max}$  3350 (br), 3250, 2900, 2800, 1660, 1640, 1525, 1480, 1440, 1370, 1250, 1170, 980, 895, 850, 750 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta_H$  8.63 (brs, 2H), 8.22 (brt, 1H, NH), 5.70 (s, 1H), 5.11 (t, 1H,  $J=7.2$  Hz), 5.06 (t, 1H,  $J=7.2$ ), 5.05 (t, 1H,  $J=7.2$ ), 3.53 (m, 2H), 3.39 (m, 4H), 3.34 (m, 2H), 3.13 (dt, 2H,  $J=5.8, 6.2$ ), 3.07 (s, 6H, NMe<sub>2</sub>), 3.04 (s, 6H, NMe<sub>2</sub>), 2.85 (m, 2H), 2.58 (dd, 2H,  $J=7.7, 8.1$  Hz), 2.12 (m, 2H), 2.06 (q, 2H,  $J=7.3$ ), 2.03 (q, 2H,  $J=7.3$ ), 2.01 (q, 2H,  $J=7.3$ ), 1.91 (t, 2H,  $J=5.8$ ), 1.90 (t, 2H,  $J=6.5$ ), 1.85 (m, 2H), 1.77 (s, 3H), 1.75 (m, 4H), 1.62 (s, 3H), 1.55 (s, 3H), 1.53 (s, 6H); <sup>13</sup>C NMR (CD<sub>3</sub>OD)  $\delta_C$  167.2s, 154.1s, 134.4s, 133.8s, 129.9s, 123.4d (2C), 123.2d, 117.6d, 64.4t (2C), 63.6t, 62.0t, 49.7q (4C), 40.8t (2C), 37.7t, 36.9t, 34.1t, 27.9t, 27.7t, 27.6t, 26.0q, 25.3q, 23.9t, 22.1t, 20.6t (2C), 18.5q, 16.2q (2C) (DMSO-*d*<sub>6</sub>, 150 MHz)  $\delta_C$  165.8s, 152.3s, 134.6s, 134.3s, 130.6s, 124.1d, 123.9d (2C), 119.2d, 62.2t (2C), 61.5t, 60.0t, 50.4s (2C), 50.1s (2C), 39.1t (2C), 35.9t, 35.2t, 32.3t, 26.4t, 26.1t, 26.0t, 25.4q, 24.7q, 22.4t, 20.2t, 19.0t(2C), 17.5q, 15.7q (2C); FABMS (glycerol) *m/z* (rel int.) 581 (23), 488 (2), 443 (4), 389 (1), 344 (10), 100 (84), 69 (100), 58 (100), 41 (48), 30 (23); HRFABMS [C<sub>14</sub>H<sub>16</sub>, <sup>35</sup>ClN<sub>4</sub>O (M-Cl)<sup>+</sup> obsd. *m/z* 581.4930,  $\Delta$  +0.5 mmu].