

# **L- $\alpha$ -Phosphatidyl-D-*myo*-inositol 3,5-bisphosphate: total synthesis of a new inositol phospholipid via *myo*-inositol orthoacetate**

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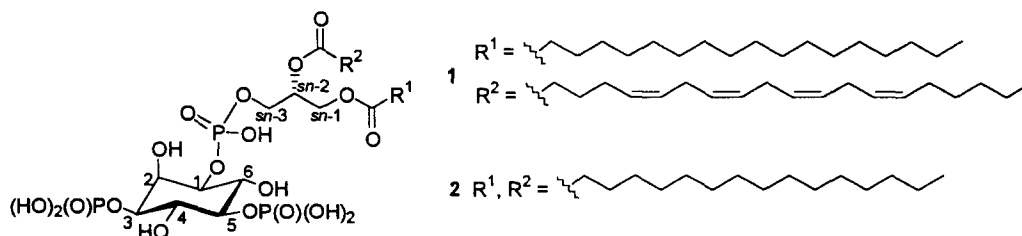
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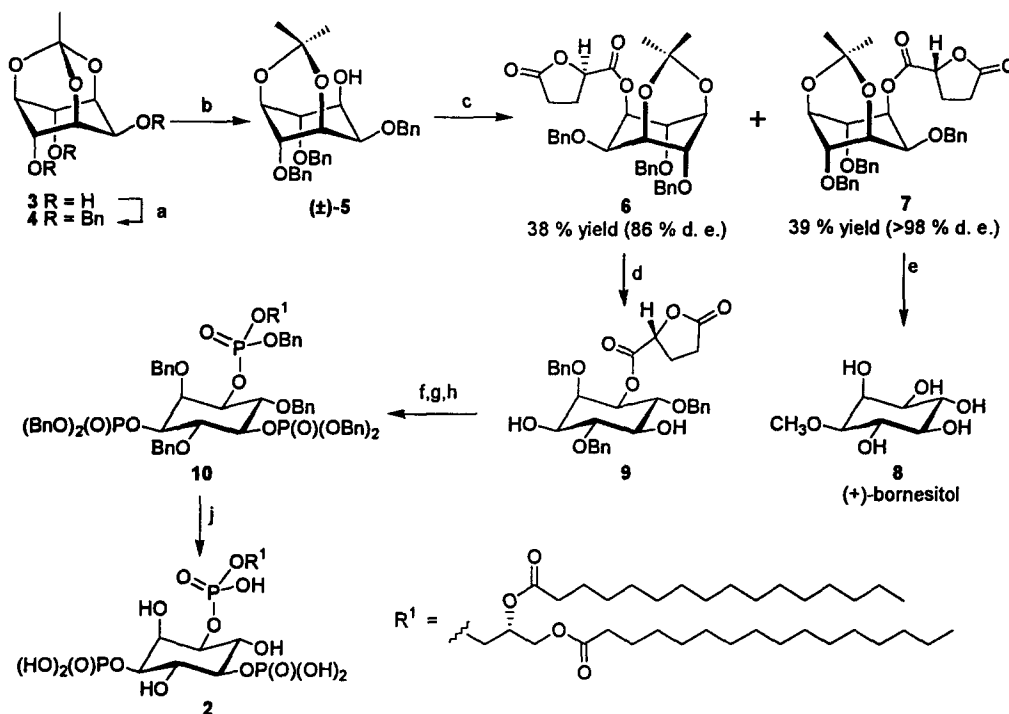
**Abstract:** The synthesis from *myo*-inositol of a newly-discovered inositol phospholipid, phosphatidylinositol 3,5-bisphosphate [PtdIns(3,5)P<sub>2</sub>], is described. The synthetic strategy, employing *inter alia*, a trimethylaluminium-mediated regioselective cleavage of a protected *myo*-inositol orthoacetate followed by an optical resolution using (*R*)-(-)-5-oxo-2-tetrahydrofuran-3-carboxylate esters, allows rapid access to dipalmitoyl PtdIns(3,5)P<sub>2</sub>. © 1998 Elsevier Science Ltd. All rights reserved.

Phosphorylation of the hydroxyl groups in phosphatidylinositol [PtdIns] at one or a combination of positions D-3, 4 and 5 of the inositol head group generates a family of phosphatidylinositol phosphates [PtdIns(P)<sub>*n*</sub>], which have been identified as components of the lipid bilayer of cell membranes. The biological functions of PtdIns(P)<sub>*n*</sub> in signal transduction, exocytosis and the regulation of membrane trafficking are currently the subject of intense interest in cell biology.<sup>1</sup> Recently, the previously unknown phosphatidylinositol 3,5-bisphosphate [PtdIns(3,5)P<sub>2</sub>, **1**] was reported to occur in mammalian cell lines<sup>2</sup> and a second study has shown that PtdIns(3,5)P<sub>2</sub> is widespread among eukaryotes.<sup>3</sup> There now exists compelling evidence that PtdIns(3,5)P<sub>2</sub> may be at the centre of a previously uncharacterised regulatory pathway,<sup>3,4</sup> but attempts to identify the cellular function of PtdIns(3,5)P<sub>2</sub> will require much larger quantities of phospholipid than can be obtained from natural sources, as well as routes adaptable to the preparation of other pharmacological probes. We therefore report here a concise and versatile synthetic route to dipalmitoyl PtdIns(3,5)P<sub>2</sub> (**2**).



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The route begins with *myo*-inositol orthoacetate (**3**).<sup>5</sup> Conventional benzylation of **3** gave the highly crystalline tri-*O*-benzyl derivative **4**<sup>6</sup> in 87 % yield without recourse to chromatography. It had previously been reported that treatment of 2,4,6-tri-*O*-benzyl-*myo*-inositol 1,3,5-orthoformate with trimethylaluminium gave ( $\pm$ )-2,4,6-tri-*O*-benzyl-1,5-*O*-ethylidene *myo*-inositol<sup>7</sup>, and we reasoned that application of a similar procedure to the orthoacetate ester **4** should give the more useful isopropylidene acetal ( $\pm$ )-**5**. The advantages of employing the orthoacetate ester are two-fold: first, the resulting isopropylidene acetal does not contain a new stereogenic centre at the bridging carbon, and second, this acetal should be more acid-labile than the corresponding ethylidene, enabling its removal under mild conditions.



**Reagents and conditions:** a) NaH, BnBr, DMF, 87%; b) Me<sub>3</sub>Al (2.5 - 3.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub>, hexane, -78°C, 91%; c) (*R*)-(-)-5-oxo-2-tetrahydrofurancarboxylic acid, DCC, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, -78°C to rt; d) 80% acetic acid, rt, then recrystallise from propan-2-ol, 78%; e) i) NaOH, MeOH, reflux; ii) NaH, MeI, DMF; iii) CH<sub>2</sub>Cl<sub>2</sub> / CF<sub>3</sub>COOH / H<sub>2</sub>O 80:19:1; iv) H<sub>2</sub>, 50 p.s.i., Pd-C, EtOH, 71% yield for 4 steps; f) (BnO)<sub>2</sub>PNPr'<sub>2</sub>, 1*H*-tetrazole, CH<sub>2</sub>Cl<sub>2</sub>; ii) *m*-CPBA, -40°C to rt, 96%; g) NH<sub>3</sub> / MeOH, rt, 91%; h) R<sup>1</sup>OP(OBn)NPr'<sub>2</sub>, 1*H*-tetrazole, CH<sub>2</sub>Cl<sub>2</sub>; ii) *m*-CPBA, -40°C to rt, 83%; j) H<sub>2</sub>, 50 p.s.i., Pd(OH)<sub>2</sub>-C, Bu'OH, 85%. Bn = benzyl.

Optimised conditions involved the use of 2.5 to 3.0 equivalents of trimethylaluminium at -78°C, followed by an alkaline work-up. Under these conditions the alcohol ( $\pm$ )-**5** was obtained in high yields on a multigram scale. Higher temperatures gave substantial amounts of the unwanted 2,4,6-tri-*O*-benzyl-*myo*-inositol. The isopropylidene acetal of ( $\pm$ )-**5** proved to be highly labile, and this property was exploited later in the synthesis.

Attempts to resolve ( $\pm$ )-5 employing the widely used (*S*)-(-)-camphanate esters were unsuccessful, as was the use of acetylmandelate esters,<sup>8</sup> but DCC-promoted esterification with (*R*)-(-)-5-oxo-2-tetrahydrofurancarboxylic [(*R*)-(-)-TOF] acid<sup>9</sup> gave the diastereoisomeric esters **6** and **7**<sup>10</sup> which were separable by flash chromatography. The less polar ester was obtained pure (as judged by <sup>1</sup>H NMR) in this way, and was converted in four steps to (+)-bornesitol<sup>11</sup> (**8**), identifying the ester as **7**. The more polar diastereoisomer **6** was obtained contaminated with some **7**. For analytical purposes, pure **6** could be isolated by further chromatography, but for the present route, it was convenient to proceed directly to the next step using partially purified **6**. Removal of the isopropylidene acetal from **6** by mild acid treatment (acetic acid at room temperature) followed by a single crystallisation from propan-2-ol gave the single diastereoisomer **9**<sup>12</sup> in 78% yield. Highly crystalline **9** could routinely be obtained on a gram scale in this way. Benzylphosphate groups were then introduced at positions 3 and 5 by phosphitylation using bis(benzyloxy)(*N,N*-diisopropylamino)phosphine<sup>13</sup> and 1*H*-tetrazole followed by *in situ* oxidation with *m*-CPBA, and the rather labile (*R*)-(-)-TOF ester was cleaved using ammonia-saturated dry methanol. Reaction at the exposed 1-OH group with benzyl 1,2-*O*-dipalmitoyl-*sn*-glyceryl *N,N*-diisopropylphosphoramidite<sup>14</sup> in the presence of 1*H*-tetrazole, gave **10** as a mixture of diastereoisomers after *m*-CPBA oxidation. Finally, deprotection by hydrogenation over palladium hydroxide on carbon in *tert*-butyl alcohol<sup>15</sup> gave dipalmitoyl PtdIns(3,5)P<sub>2</sub> (**2**).<sup>16</sup>

In conclusion, we have described an expedient route to a recently-discovered inositol phospholipid, employing a novel regioselective protecting group strategy and a chiral auxiliary new to the field of inositol chemistry. The enantiomer of **2**, potentially of use in investigating the specificity of putative target proteins for PtdIns(3,5)P<sub>2</sub>, can be synthesised either from **7** or by the use of (*S*)-(+)-5-oxo-2-tetrahydrofurancarboxylic acid. Finally, intermediate **9**, rapidly accessible in high purity from *myo*-inositol orthoacetate, provides a suitable starting point for the synthesis of other analogues of PtdIns(3,5)P<sub>2</sub>.

#### ACKNOWLEDGEMENTS:

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6. Compound **4** had  $R_f$  0.36 (EtOAc / hexane 1:3), mp 77.5 - 78.5°C (from hexane). All new compounds exhibited satisfactory spectroscopic and analytical data.
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9. Purchased from Aldrich and recrystallised once from ether / hexane before use. The enantiomer, (*S*)-(+)-5-oxo-2-tetrahydrofurancarboxylic [(*S*)-(+)-TOF] acid, also available from Aldrich, has been used as a chiral derivatising reagent for alcohols (Doolittle, R. E. and Heath, R. R. *J. Org. Chem.* **1984**, *49*, 5041-5050). (*S*)-(+)-TOF esters have also been employed for the optical resolution of ( $\pm$ )-4-benzyloxy-tetrahydrofuran-3-ol (Altenbach, H.-J. and Wolf, E. *Tetrahedron: Asymmetry* **1993**, *4*, 2155-2158).
10. Data for **6**:  $[\alpha]_D^{20}$  -21 (c 1.5, CHCl<sub>3</sub>);  $R_f$  0.40 (CHCl<sub>3</sub>/EtOAc 10:1);  $\delta_H$  (270 MHz, CDCl<sub>3</sub>, TMS) 1.38 (3 H, s, CH<sub>3</sub>), 1.43 (3 H, s, CH<sub>3</sub>), 2.07 - 2.16 (1 H, m), 2.24 - 2.40 (2 H, m), 2.42 - 2.56 (1 H, m), 4.05 (1 H, br d,  $J \sim 5.5$  Hz), 4.27-4.30 (2 H, m), 4.38 - 4.54 (5 H, m), 4.62 - 4.72 (3 H, m), 4.92 (1 H, dd,  $J$  8.4, 4.4 Hz, C-2'-H), 5.50 (1 H, dd,  $J$  7.1 5.5 Hz, C-1-H), 7.20 - 7.32 (15 H, 3  $\times$  C<sub>6</sub>H<sub>5</sub>).  
Data for **7**:  $[\alpha]_D^{16}$  -4 (c 1, CHCl<sub>3</sub>);  $R_f$  0.48 (CHCl<sub>3</sub>/EtOAc 10:1);  $\delta_H$  (400 MHz, CDCl<sub>3</sub>, TMS) 1.39 (3 H, s, CH<sub>3</sub>), 1.49 (3 H, s, CH<sub>3</sub>), 1.95 - 2.04 (1 H, m), 2.08 - 2.18 (1 H, m), 2.23 - 2.35 (2 H, m), 3.94 (1 H, br s), 4.24 (1 H, dd,  $J$  6.1 2.1 Hz), 4.38 (1 H, br s), 4.41 - 4.54 (4 H, m), 4.62 - 4.76 (4 H, m), 4.87 (1 H, dd,  $J$  8.2, 4.3 Hz, C-2'-H), 5.65 (1 H, dd,  $J$  6.1 3.1 Hz, C-1-H), 7.20 - 7.33 (15 H, m, 3  $\times$  C<sub>6</sub>H<sub>5</sub>).
11. <sup>1</sup>H NMR data for **8** agreed with those published for 1D-(-)-1-*O*-methyl *myo*-inositol, (Jaramillo, C.; Chiara, J.-L. and Martín-Lomas, M. *J. Org. Chem.* **1994**, *59*, 3135-3141) but **8** had  $[\alpha]_D^{23}$  +32 (c 2, H<sub>2</sub>O), identifying it as 1L-1-*O*-methyl *myo*-inositol [ (+)-bornesitol ]; mp 203-205°C (from MeOH / EtOH); Lit. values: + 31.9 (c 1, H<sub>2</sub>O), mp 205-207°C (Gigg, J.; Gigg, R.; Payne, S. and Conant, R. *J. Chem. Soc. Perkin Trans. 1* **1987**, 1757-1762. See also references therein).
12. Data for **9**: mp 121.5 - 123.5°C from propan-2-ol;  $[\alpha]_D^{24}$  -28° (c 1, CHCl<sub>3</sub>);  $\delta_H$  (<sup>1</sup>H - <sup>1</sup>H COSY, 400MHz, CDCl<sub>3</sub>, TMS) 1.93 - 2.05 (1 H, m, C-3'-H<sub>2A</sub>), 2.26 - 2.37 (3 H, m, C-4'-H<sub>2</sub> and C-3'-H<sub>2B</sub>), 2.39 (1 H, d,  $J$  5.2 Hz, D<sub>2</sub>O ex., C-3-OH), 2.59 (1 H, d,  $J$  2.4 Hz, D<sub>2</sub>O ex., C-5-OH), 3.58 - 3.64 (2 H, m, C-3-H and C-5-H), 3.73 (1 H, dd,  $J$  9.5, 9.3 Hz, C-4-H), 3.95 (1 H, dd,  $J$  10.1, 9.2 Hz, C-6-H), 4.06 (1 H, dd,  $J$  2.7, 2.5 Hz, C-2-H), 4.65 - 4.71 (3 H, m, OCH<sub>2</sub>Ar and C-2'-H), 4.75 - 4.90 (4 H, m, OCH<sub>2</sub>Ar), 4.87 (1 H, dd, partly buried,  $J$  10.3, 2.7 Hz C-1-H), 7.24 - 7.40 (15 H, m, 3  $\times$  C<sub>6</sub>H<sub>5</sub>).
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16. Although the free acid **2** was poorly soluble in water, and NMR spectra taken in chloroform / methanol mixtures were broad, well-resolved NMR spectra could be obtained in [<sup>2</sup>H<sub>7</sub>]DMF.  
Data for **2**:  $\delta_H$  (<sup>1</sup>H - <sup>1</sup>H COSY, 400 MHz, [<sup>2</sup>H<sub>7</sub>]DMF, TMS) 0.88 (6 H, t,  $J$  7 Hz, palmitoyl CH<sub>3</sub>), 1.22 - 1.38 (48 H, m, palmitoyl CH<sub>2</sub>), 1.54 - 1.64 (4 H, m, palmitoyl  $\beta$ -CH<sub>2</sub>), 2.32 (2 H, t,  $J$  7.6 Hz, palmitoyl  $\alpha$ -CH<sub>2</sub>), 2.34 (2 H, t,  $J$  7.6 Hz, palmitoyl  $\alpha$ -CH<sub>2</sub>), 3.98 (2 H, br t,  $J$  9 Hz, inositol C-4-H, C-6-H), 4.10 (1 H, br q,  $J$  8 Hz, inositol C-5-H), 4.18 - 4.27 (5 H, m, inositol C-1-H and C-3-H, glyceryl *sn*-1-CH<sub>2A</sub>, *sn*-3-CH<sub>2</sub>), 4.42 (1 H, dd,  $J$  11.6 Hz, 2.4 Hz, glyceryl *sn*-1-CH<sub>2B</sub>), 4.52 (1 H, br s, inositol C-2-H), 5.25 (1 H, m, glyceryl *sn*-2-H);  $\delta_P$  (162 MHz, [<sup>2</sup>H<sub>7</sub>]DMF, external H<sub>3</sub>PO<sub>4</sub>) -1.70 (1P), -0.02 (1P), 1.73 (1P). Negative FABMS (cyclohexylammonium salt, *m*-NBA):  $m/z$  969.5 [(M-H)<sup>-</sup>, 100%], 647.5 [C<sub>15</sub>H<sub>31</sub>COOCH<sub>2</sub>CH(OCOC<sub>15</sub>H<sub>31</sub>)CH<sub>2</sub>OPO<sub>3</sub>H<sup>-</sup>, 80%], 255.2 [C<sub>15</sub>H<sub>31</sub>COO<sup>-</sup>, 60%], 97.0 [H<sub>2</sub>PO<sub>4</sub><sup>-</sup>, 40%].