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Synthesis, characterization and photophysical properties of a new Cu²⁺ selective phosphorescent sensor

Tianzhi Yu^{1,*}, Qianguang Yang¹, Zeyang Zhu¹, Yuling Zhao², Xiaoxiao Liu³, Chengjin Wei¹,

Hui Zhang¹

¹Key Laboratory of Opto-Electronic Technology and Intelligent Control (Ministry of Education), Lanzhou Jiaotong University, Lanzhou 730070, China

²School of Chemical and Biological Engineering, Lanzhou Jiaotong University, Lanzhou 730070, China

³School of Mechatronic Engineering, Lanzhou Jiaotong University, Lanzhou 730070, China

Abstract: A new Cu(II) selective phosphorescent sensor based on a cationic iridium(III) complex was synthesized and characterized. Compared with other commonly coexisting metal ions, the Ir(III) complex displayed significant phosphorescence quenching for detecting Cu^{2+} with a 1:1 stoichiometry in MeOH/H₂O (1:1, v/v) solution. Upon addition of Cu^{2+} , the rapid quenching of emission intensity provided a sensitive method for detection of Cu^{2+} . The corresponding detection limit was determined to be 7.6×10^{-8} M and the association constant between the Ir(III) complex and Cu^{2+} was calculated to be 0.31×10^5 M⁻¹. Luminescence imaging experiments demonstrated that this Ir(III) complex was cell permeable and suitable for monitoring intracellular Cu^{2+} in living Hela cells by confocal microscopy.

Keywords: Cationic iridium(III) complex; Phosphorescence quenching; Phosphorescent

^{*}Corresponding Author. Tel. +86-931-4956935; Fax: +86-931-4938756; e-mail: yutzh@mail.lzjtu.cn (T.Z. Yu).

chemosensor; Cu²⁺ detection; Bioimaging

1. Introduction

Recently, phosphorescent heavy-metal complexes have attracted tremendous attentions due to their efficient luminescent properties and potential applications in organic light-emitting diodes (OLEDs) ^[1-6], light-emitting electrochemical cells ^[7-10], chemosensors ^[11-14] and biological labeling ^[15,16]. Compared with the fluorescent chemosensors, the phosphorescent heavy-metal complexes have attracted more attentions and increasingly used as colorimetric or luminescent probes for biomolecules and metal ions because their relatively long phosphorescence emission is unaffected by short-lived background noise such as fluorescence and scattered light ^[17-20]. Among phosphorescent heavy-metal complexes, the phosphorescent Ir(III) complexes show some advantageous photophysical properties such as short excited-state lifetime, high quantum efficiency, color diversity and high photostability, and some Ir(III) complexes have been used as luminescent probes.

As trace element and nutrient in living organisms, the copper(\Box) is essential and plays a significant role in a basic physiological process ^[13]. However, the copper(\Box) is also one of significant metal pollutants and toxic to living cells ^[21]. Aberrant levels of copper(\Box) ions can result in neurodegenerative diseases such as Alzheimer's disease, prion disease, Menkes disease, Parkinson's disease and Wilson disease ^[22-24]. As a result, to seek a rapid sensitive sensor to detect and quantify copper(\Box) ion in environment and in living systems is very importance for biologic research and medical diagnosis. A number of fluorescent chemosensors for the highly sensitive and selective detection of Cu (II) ion have been reported in the literatures ^[25-31]. Recently, the

phosphorescent Ir(III)-based chemosensors have been successfully applied for the detection of Cu^{2+} ions ^[13,14,19,32-35]. Moreover, the phosphorescent Ir(III) complexes as luminescent probes present important advantages as their simplicity, easy visualization and high sensitivity. The incorporation of a Lewis base as a cation receptor on the ancillary ligands of the luminescent Ir(III) complexes is a very used strategy for the recognition of cations. The coordination interaction of the Cu^{2+} with an ion recognition group on the Ir(III) complex causes variations in the luminescence intensity that may be used to detect Cu^{2+} . For example, You et al. ^[19] synthesized the Ir(III) complex having two cyclometalating 2-phenylpyridine (ppy) ligands and one 2-(2'-benzo[*b*]thienyl)pyridine (btp) tethered a di(2-picolyl)amine (DPA) copper ion receptor by a methylene linker, which was used as a Cu^{2+} -sensitive probe. This Ir(III) complex has a 1:2 stoichiometry (complex : Cu^{2+}) that was demonstrated by the binding titration data, and the limit of detection for Cu^{2+} was measured to be range of 5:8 – 8.1 µM.

The di(2-picolyl)amine moiety (DPA) always exhibits high affinity and specificity for Zn^{2+} and Cu^{2+} ions over the other metal ions, which has been frequently used as turn-on sensors for Zn^{2+} and Cu^{2+} ions. In this paper we used 2-(1-naphthyl)benzothiazole (**nbt**) as cyclometalated ligand and 2,2'-dipyridine tethered DPA moiety by a methylene linker (**bp-DPA**) as the neutral auxiliary ligand to prepare a new cationic Ir(III) complex, [**Ir(nbt)**₂(**bp-DPA**)]**PF**₆. The photophysical properties and ion-recognition behaviors of the cationic Ir(III) complex with some heavy-metal ions are investigated. This cationic Ir(III) complex exhibited good sensitivity and selective on–off type response toward Cu²⁺ over other metal ions including Zn^{2+} and Hg^{2+} . Furthermore, the confocal luminescence imaging was performed to validate the ability of the Ir(III) complex to monitor intracellular Cu²⁺ ions. The synthetic routes of the neutral auxiliary ligand **bp-DPA** and the cationic Ir(III) complex were shown in Scheme 1.



Scheme 1. Synthetic routes of bp-DPA and [Ir(nbt)₂(bp-DPA)]PF₆.

2. Experimental

2.1. Materials and instruments

2-Aminothiophenol, 1-naphthaldehyde and 4,4'-dimethyl-2,2'-bipyridine were bought from Alfa Aesar. 2,2'-Dipicolylamine was obtained from Energy Chemical (China). $IrCl_3 \cdot nH_2O$ (iridium content > 60.0%) was bought from Shanxi Kaida Chemical Co. Ltd. (China) and used without further purification. All the other chemicals were analytical grade reagent. All of the organic solvents used in this study were dried over appropriate drying agents and freshly distilled prior in the related reaction.

4-Methyl-2,2'-bipyridine-4'-carboxaldehyde was synthesized as the procedure described in literature ^[36]. The cyclometalated Ir(III) μ -chlorobridged dimer of (**nbt**)₂Ir(μ -Cl)₂Ir(**nbt**)₂ was prepared as previously described ^[37].

¹H NMR spectra were recorded on Agilent Technologies DDZ 600 MHz. Mass spectrum was obtained from a Thermo Scientific Orbitrap Elite mass spectrometer. Thermogravimetric analysis

(TGA) was performed on a Perkin-Elmer Pyris system. UV-vis absorption and photoluminescent spectra were recorded on a Shimadzu UV-2550 spectrometer and on a Perkin-Elmer LS-55 spectrometer, respectively. All pH measurements were made with apH-10C digital pH meter.

2.2. Synthesis and characterization of bp-DPA and [Ir(nbt)₂(bp-DPA)]PF₆

4-Methyl-4'-(di(2-picolyl)aminomethyl)-2,2'-bipyridine (bp-DPA): Di(2-picolyl)amine (0.729 g, 3.66 mmol) and 4-Methyl-2,2'-bipyridine-4'-carboxaldehyde (0.725 g, 3.66 mmol) were dissolved in fresh MeOH (70 mL). A catalytic amount (two drops) of glacial acetic acid was added to the solution, which was refluxed for 0.5 h. The reaction mixture was cooled by using an ice bath, and then NaBH₃CN (0.460 g, 7.32 mmol) was slowly delivered to the solution. The ice bath was removed and the reaction mixture was stirred overnight at room temperature. Basic work-up using saturated aqueous sodium carbonate solution and subsequent extraction using CH₂Cl₂ was performed. The organic layer was dried over anhydrous MgSO₄ and concentrated in vacuum. The crude was purified by chromatography on silica gel using acetone/petroleum ether (1:10, v/v) to obtain a sticky brown oily-liquid, Yield (1.185 g, 84.9%). ¹H NMR (600 MHz, CDCl₃, δ , ppm): 8.50 (d, J = 7.2 Hz, 1H), 8.40 (t, J = 9.6 Hz, 3H), 8.29 (s, 1H), 8.09 (s, 1H), 7.53 (t, J = 9.0 Hz, 2H), 7.47 (d, J = 8.8 Hz, 2H), 7.35 (d, J = 7.2 Hz, 1H), 7.02-6.97 (dd, J = 9.6, 8.4 Hz, 3H), 3.74 (s, 4H), 3.69 (s, 2H), 2.26 (s, 3H).

[Ir(nbt)₂(bp-DPA)]PF₆: The chloro-bridged dimer complex $(nbt)_2Ir(\mu-Cl)_2Ir(nbt)_2$ (0.799 g, 0.534 mmol) and **bp-DPA** (0.5111 g, 1.340 mmol) were refluxed in 60 mL of dichloromethane/methanol (1:1, v/v) under nitrogen atmosphere at 75 \Box for 4 h with keeping in dark place. After cooling, an excess of KPF₆ (15 equiv) was slowly added to the reaction mixture and the solution was stirred for additional 12 h. After completion of the reaction, the solids in the

solution were removed by filtration. The residue was purified through chromatography on silica gel using CH₂Cl₂/MeOH (9:1, v/v) as the eluent to obtain a red powder (0.98 g, 74%). ¹H NMR (600 MHz, CDCl₃, δ , ppm): δ 8.66 (dd, J = 9.0, 8.4 Hz, 3H), 8.44 (t, J = 6.8 Hz, 3H), 7.95 (dd, J = 8.4, 7.8 Hz, 2H), 7.87 (d, J = 6.0 Hz, 1H), 7.82 (d, J = 6.0 Hz, 1H), 7.76-7.68 (m, 4H), 7.57 (t, J = 7.8 Hz, 2H), 7.51-7.44 (m, 5H), 7.41 (t, J = 7.2 Hz, 1H), 7.35 (t, J = 7.8 Hz, 1H), 7.25 (t, J = 7.2 Hz, 2H), 7.20 (dd, J = 8.4, 6.0 Hz, 2H), 7.03 (t, J = 6.6 Hz, 2H), 6.98 (t, J = 7.8 Hz, 1H), 6.56 (dd, J = 8.4, 8.4 Hz, 2H), 6.37 (d, J = 8.4 Hz, 1H), 6.15 (d, J = 8.4 Hz, 1H), 4.03 (s, 2H), 3.85 (s, 4H), 2.63 (s, 3H). HR EI-MS: Calcd. for [C₅₈H₄₃IrN₇S₂]⁺, 1094.27; Found, 1094.2637.

2.3. Spectroscopic experiments

Spectrophotometric titrations of the Ir(III) complex were performed in air-equilibrated CH₃OH solution. The Ir(III) complex stock solution $(1.0 \times 10^{-3} \text{ M})$ was prepared in CH₃OH. The fresh metal stock solutions (typically $1.0 \times 10^{-2} \text{ M}$) were prepared in Milli-Q water using the corresponding chloride salts (Na⁺, K⁺, Ca²⁺, Mg²⁺, Mn²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Ni²⁺, Co²⁺, Pb²⁺, Al³⁺ and Fe³⁺), nitrate salt (Ag⁺) and perchlorate salt (Hg²⁺). For emission titration experiment, 10µL of methanol solution of the Ir(III) complex ($1.0 \times 10^{-3} \text{ M}$) was taken in a quartz optical cell of 1.0 cm optical path length in 2000 µL and then stock solutions of metal ions were added gradually to it by using micropipette. Spectral data were recorded at 1 min after the addition of the metal ions for both titrations.

2.4. Cell cytotoxicity

The cell cytotoxicity of $[Ir(nbt)_2(bp-DPA)]PF_6$ was determined by MTT (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide) assays. Hela cells were placed in microassay culture plates (5 × 10⁴ cells per milliliter) and grown for 24 h at 37 °C in a

5% CO₂ incubator. Then [Ir(nbt)₂(bp-DPA)]PF₆ was added to the wells to achieve final concentrations of 1.25, 2.5, 5, 10 and 20 μ M. The plates were incubated at 37 °C in a 5% CO₂ incubator for another 24 h. Upon completion of the incubation, stock MTT dye solution (20 μ L, 1 mg/mL) was added to each well. After 4 h, dimethyl sulfoxide (150 μ L) was added to solubilize the MTT formazan.

2.5. Cell culture and confocal luminescence imaging

Vitro experiments were performed by using Hela cells. Hela cells were cultured in RPMI-1640 medium supplemented with 10% FBS (fetal bovine serum), 100 units per mL penicillin, and 100 μ g/mL streptomycin at 37 °C in a humid atmosphere containing 5% CO₂. Cells (5 × 10⁸ L⁻¹) were plated on 18 mm glass coverslips and allowed to adhere for 24 h. Then the cells were treated with [**Ir**(**nbt**)₂(**bp-DPA**)]**PF**₆ (20 μ M) and incubated for 30 min. Subsequently, the cells were treated with 100 μ M CuCl₂ • 2H₂O. The cells were incubated for 30 min and washed with PBS three times to remove free compound and ions before analysis. Hela cells cells only incubated with 20 μ M [**Ir**(**nbt**)₂(**bp-DPA**)]**PF**₆ for 30 min acted as a control. Confocal luminescence images of Hela cells cells were carried out on an Olympus FV1000 laser scanning confocal microscope and a 100x oil-immersion objective lens. Emission was collected at 550–630 nm for the Hela cells.

3. Results and discussion

3.1. Synthesis and characterization

The synthesis of the cationic Ir(III) complexes from Scheme 1 began first with preparing the ancillary ligand **bp-DPA**. The ligand **bp-DPA** was successfully obtained through two step

reactions. First, 4,4'-dimethyl-2,2'-bipyridine was selectively oxidized to be 4-Methyl-2,2'-bipyridine-4'-carboxaldehyde by SeO₂ catalyst. Subsequently, this intermediate was subjected to reductive amination with DPA. The ligand **bp-DPA** was purified by column chromatography to give the pure product. The Ir(III) complex [**Ir**(**nbt**)₂(**bp-DPA**)]**PF**₆ was synthesized by reacting the Ir(III) chloro-bridged dimer with the ancillary ligand **bp-DPA**, and followed by a counter ion-exchange reaction from Cl⁻ to PF_6^- . The Ir(III) complex was structurally characterized by ¹H NMR and the high resolution electro spray ionization mass spectra (HR ESI-MS).

3.2. Photophysical and thermal properties of [Ir(nbt)₂(bp-DPA)]PF₆

The UV-vis absorption and photoluminescence (PL) spectra of $[Ir(nbt)_2(bp-DPA)]PF_6$ in MeOH solution is shown in Fig. 1. As shown in Fig.1, the intense absorption bands before 300 nm in the absorption spectrum are assigned to the spin-allowed $(\pi-\pi^*)$ ligand-centered transition of the cyclometalated primary ligand and the ancillary ligand in the complex. The following relatively weak absorption band at 345 nm could be attributed to both spin-allowed metal-to-ligand charge-transfer (¹MLCT) and ligand-to-ligand charge-transfer (¹LLCT) transitions. Moreover, the weaker absorption band that found at 468 nm is ascribed to the spin-forbidden ³MLCT and ³LLCT ^[38-40].

Under the excitation of 468 nm, the complex $[Ir(nbt)_2(bp-DPA)]PF_6$ exhibits red emission with a maximum peak at 594 nm and a shoulder peak at 642 nm in MeOH solution at room temperature. The CIE coordinates of $[Ir(nbt)_2(bp-DPA)]PF_6$ is (0.610, 0.388). In addition, the photoluminescence quantum yield of $[Ir(nbt)_2(bp-DPA)]PF_6$ at room temperature was measured to be 5.05% in MeOH solution (ca. 10⁻⁶ mol/L) through an absolute method using the Edinburgh

Instruments spectrometer (FLS920) integrating sphere excited at 468 nm with a Xe lamp. The photoluminescence decay lifetime of $[(nbt)_2Ir(bp-DPA)](PF_6)$ in MeOH solution was measured to be 2.21 µs with a time-correlated single photon counting spectrometer using an Edinburgh Instruments spectrometer (FLS920) with a microsecond flash lamp as the excitation source (repetition rate 90 Hz) at room temperature. The observed emission lifetime is in the microsecond time scale, which indicates the phosphorescent nature of the emission. The result indicates that the Ir(III) complex possesses a relatively long lifetime to differentiate short-lived background fluorescence and enhances the signal-to-noise ratio of detection through the use of time-resolved fluorescence analysis.



Fig. 1. The UV-vis absorption and photoluminescence (PL) spectra of [Ir(nbt)₂(bp-DPA)]PF₆ in MeOH solution.

The thermal behavior of the cationic Ir(III) complex was investigated using thermogravimetric analysis (TGA) under a nitrogen atmosphere. The result of its TGA measurement is shown in Fig. 2. The complex [(nbt)₂Ir(bp-DPA)](PF₆) exhibited good thermal stability up to 256 °C. With increasing temperature, [(nbt)2Ir(bp-DPA)](PF6) undergoes two large-stage decomposition at 259 and 382 °C accompanied by two sharp weight losses in its TGA curve. It indicates that the complex meets with the thermal stability requirement of chemical

sensor application.



Fig. 2. Thermogravimetric analysis (TGA) of $[Ir(nbt)_2(bp-DPA)]PF_6$ in nitrogen atmosphere (heating rate 10 \Box /min).

3.2. Selectivity and cation-competitive experiments

The selectivity of the Ir(III) complex (10 μ M) for metal ions investigated by monitoring its photoluminescence changes upon addition of various metal ions (1 equiv) in MeOH solution is shown in Fig. 3a. The result indicated that the photoluminescence of the Ir(III) complex was quenched significantly upon addition of Cu²⁺ ion and quenched slightly upon addition Ca²⁺ or Ni²⁺ ion, whereas very weak variations in the emission spectra of the Ir(III) complex were observed upon addition of other metal ions, such as Zn²⁺, Na⁺, Mg²⁺, Cd²⁺, Fe³⁺, Mn²⁺, K⁺, Co²⁺, Hg²⁺, Al³⁺, Pb²⁺ and Ag⁺. Therefore, the Ir(III) complex displayed a turn-off phosphorescence response and high selectivity in sensing Cu²⁺ ion.

To further investigate the interference of other metal ions on the detection of Cu^{2+} ion, the cation-competitive experiments were also performed by adding Cu^{2+} ion to solutions of the Ir(III) complex in the presence of other metal ions (Fig. 3b). As shown in Fig. 3b, whether in the absence or in the presence of the other metal ions, obvious spectral changes were observed for [Ir(nbt)₂(bp-DPA)]PF₆ only upon addition of Cu^{2+} ion. The results suggests that



 $[Ir(nbt)_2(bp-DPA)]PF_6$ possesses good selectivity for Cu²⁺ ion.

Fig. 3. (a) Photoluminescence spectra of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 µM) upon addition of various metal ions (1 equiv) in MeOH solution; (b) Column diagrams of the PL intensity of $[Ir(nbt)_2(bp-DPA)]PF_6 + M^{n+}$. Red bars represent the addition of 10 equiv of various metal ions to the blank solution and black bars represent the subsequent addition of Cu²⁺ (1 equiv) to the aforementioned solutions. ($\lambda_{ex} = 468 \text{ nm}$, $\lambda_{ex} = 594 \text{ nm}$)

3.3. Cu²⁺ titration analysis

To further evaluate the sensing properties of the Ir(III) complex towards Cu^{2+} ion, the emission titration experiments of [Ir(nbt)₂(bp-DPA)]PF₆ with Cu^{2+} ion in varying concentrations were carried out by monitoring the emission intensity changes of [Ir(nbt)₂(bp-DPA)]PF₆. As shown in Fig. 4a, the lone Ir(III) complex exhibited strong phosphorescence emission with a maximum peak at 594 nm and a shoulder peak at 642 nm upon excitation at 468 nm, and the PL intensity of the Ir(III) complex gradually decreased with continuous addition of Cu^{2+} (0–2.0 eq.) to

the solution. The emission intensity of the Ir(III) complex drops by 97.4% as Cu^{2+} ion concentration achieves 1.0 equivalents, which indicates that there is a strong association interaction between Cu^{2+} and the Ir(III) complex.

The Job's plot experiment was performed by monitoring the PL intensity change of $[Ir(nbt)_2(bp-DPA)]PF_6$ with the variation of the concentration of both $[Ir(nbt)_2(bp-DPA)]PF_6$ and Cu²⁺ ion (Fig. 4b), indicating a 1:1 stoichiometry between the Ir(III) complex and Cu²⁺ ions.



Fig. 4. (a) Photoluminescence spectra of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 µM) upon titration with Cu^{2+} (0–2.0 eq.) in MeOH solution; (b) Job's plot for the complex formation between $[Ir(nbt)_2(bp-DPA)]PF_6$ and Cu^{2+} ion in MeOH solution. ($\lambda_{ex} = 468 \text{ nm}$, $\lambda_{ex} = 594 \text{ nm}$, $X_{Cu}^{2+} = [Cu^{2+}]/[[Ir(nbt)_2(bp-DPA)]PF_6]+[Cu^{2+}])$

From the titration experiments, plotting emission intensity versus Cu^{2+} concentration (0–1.0 equiv.) afforded a good linear relationship (R = 0.9897) (Fig. 5a). The detection limit of

[Ir(nbt)₂(bp-DPA)]PF₆ was calculated from the equation DL = 3S/k, where S is the standard deviation of blank measurement, k is the slope between intensity versus sample concentration ^[33,41,42]. The limit of detection was calculated to be 7.6×10^{-8} M. The association constant was determined by the Benesi-Hildebrand equation and calculated to be 0.31×10^5 M⁻¹ (Fig. 5b). Compared with the similar Ir(III) complex ^[19], our synthesized Ir(III) complex exhibited lower detection limit for Cu²⁺ ion.



Fig. 5. (a) The linear response of emission intensity at 594 nm of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 μ M) to the Cu²⁺ concentrations changes (0–1.0 equiv.) in MeOH solution; (b) Benesi-Hildebrand plot of $[Ir(nbt)_2(bp-DPA)]PF_6$ at different Cu²⁺ concentrations in MeOH solution. ($\lambda_{ex} = 468$ nm)

Reversibility of the probe is an important consideration in practical application. The reversibility of interaction between $[Ir(nbt)_2(bp-DPA)]PF_6$ and Cu^{2+} ion was performed by addition of an aqueous of EDTA disodium (1 eq.) to the mixture of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 μ M) and Cu^{2+} ion (10 μ M) in MeOH/H₂O (1:1, v/v) solution (Fig. 6). As shown in Fig. 6, after

adding of Na₂EDTA (1 eq.) to the mixture of the Ir(III) complex and Cu^{2+} ion, the emission intensity at 594 nm was almost completely recovered. The result indicated that the interaction between [Ir(nbt)₂(bp-DPA)]PF₆ and Cu²⁺ ion is chemically reversible.



Fig. 6. The emission spectra of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 μ M), $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 μ M) + Cu²⁺ (10 μ M) and $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 μ M) + Cu²⁺ (10 μ M) + Na₂EDTA (10 μ M) in MeOH/H₂O (1:1, v/v) solution. ($\lambda_{ex} = 468 \text{ nm}, \lambda_{ex} = 594 \text{ nm}$)



Fig. 7. The emission intensity of $[Ir(nbt)_2(bp-DPA)]PF_6$ (10 µM) with and without Cu²⁺ (10 µM) as a function of pH in MeOH/H₂O (1:1, v/v) solution. ($\lambda_{ex} = 468 \text{ nm}$, $\lambda_{ex} = 594 \text{ nm}$)

The effect of pH on the photoluminescence response of $[Ir(nbt)_2(bp-DPA)]PF_6$ to Cu^{2+} ions was also investigated in MeOH/H₂O (1:1, v/v) solution (Fig. 7). As shown in Fig. 7, the PL intensity of $[Ir(nbt)_2(bp-DPA)]PF_6$ solutions in the presence and absence of Cu^{2+} were both no dramatic change over a pH range of 2–10. However, pH>10, with the coordinated Cu^{2+} ions gradually dissociating from the Ir(III) complex– Cu^{2+} system, the emission of free the Ir(III) complex recovered gradually.

From the above results, the possible mechanisms of detection and the structures of $[Ir(nbt)_2(bp-DPA)]PF_6$, both with and without the addition of Cu^{2+} , are shown in Fig. 8. In the absence of Cu^{2+} , the Ir(III) complex exhibits strong red emission with a maximum peak at 594 nm when excited at 468 nm. After coordinated with Cu^{2+} , the paramagnetic effect from spin-orbit coupling of the Cu^{2+} induces the emission of the Ir(III) complex quenching. Upon the addition of Na₂EDTA into the [Ir(nbt)₂(bp-DPA)]PF₆-Cu²⁺ system, the emission of the Ir(III) complex was restored due to dissociating from the Ir(III) complex-Cu²⁺ system by EDTA, which indicates the regeneration of the free [Ir(nbt)₂(bp-DPA)]PF₆.



Fig. 8. The possible sensing mechanism of [Ir(nbt)₂(bp-DPA)]PF₆ for Cu²⁺.

3.4. Cell cytotoxicity and confocal luminescence imaging

The cytotoxicity of $[Ir(nbt)_2(bp-DPA)]PF_6$ was investigated in Hela cells using an MTT assay (Fig. 9). As shown in Fig. 9, the Ir(III) complex had some cytotoxicity to Hela cells. When the concentration of $[Ir(nbt)_2(bp-DPA)]PF_6$ was greater than 5 µM, the viability of the cells kept changeless and the cell viability remained above 40%. Herein, we selected 20 µM $[Ir(nbt)_2(bp-DPA)]PF_6$ to analyze its recognition ability for Cu²⁺ ions in living Hela cells by confocal luminescence imaging (Fig. 10). As shown in Fig. 10a, the Hela cells treated with 20 µM

[Ir(nbt)₂(bp-DPA)]PF₆ alone for 30 min at 37 °C exhibited bright red emission. However, when Cu^{2+} (100 µM) was introduced into the cells via incubation under the same condition, remarkable intracellular luminescence quenching was observed (Fig. 10d). Bright-field measurements after the treatment with [Ir(nbt)₂(bp-DPA)]PF₆ confirmed that the cells were viable throughout the imaging experiments (Figs. 10b,e). Overlays of confocal luminescence images also demonstrated that the luminescence was evident (Figs. 10c,f). Thus, the results indicated that the Ir(III) complex was cell membrane-permeable and had the ability to detect Cu²⁺ in living cells.



Fig. 9. Hela cell viability values (%) estimated by MTT proliferation test vs. incubation concentrations of $[Ir(nbt)_2(bp-DPA)]PF_6$. Cells were cultured in the presence of 0–20 μ M $[Ir(nbt)_2(bp-DPA)]PF_6$ at 37 °C for 24 h.



Fig. 10. Confocal luminescence (a and d), bright field (b and e) and overly images (c and f) of Hela cells. Hela cells were incubated with $[Ir(nbt)_2(bp-DPA)]PF_6$ (20 µM) for 30 min at 37 °C (a–c), and subsequently treated with Cu²⁺ (100 µM) for another 30 min at 37 °C (d–f).

4. Conclusion

To summarize, we have presented a new cationic Ir(III) complex as phosphorescent sensor for quantitating Cu^{2+} ion. The Ir(III) complex exhibited a rapid turn-off phosphorescent response toward Cu^{2+} ion and displayed excellent reversibility and selectivity for Cu^{2+} ion in MeOH/H₂O (1:1) solution with a broad pH range of 2–10. The detection limit and the association constant were determined to be 7.6×10^{-8} M and 0.31×10^{5} M⁻¹ for Cu²⁺, respectively. Although the Ir(III) complex had some cytotoxicity to Hela cells, it exhibited well cell-permeable and good imaging characteristics for the detection of Cu²⁺ ion in living cells.

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Highlights:

- 1. A cationic Ir(III) complex was synthesized for detection of Cu^{2+} in vitro and cells.
- 2. The detection limit for Cu^{2+} was measured to be 10^{-8} M.
- 3. The complex has well cell-permeable and imaging characteristics for Cu^{2+} in cells.