Synthesis of α,α-Difluoro-β-amino Esters or *gem*-Difluoro-β-lactams as Potential Metallocarboxypeptidase Inhibitors

Nicolas Boyer,^[a] Philippe Gloanec,^[b] Guillaume De Nanteuil,^[b] Philippe Jubault,^{*[a]} and Jean-Charles Quirion^{*[a]}

Keywords: β-Lactams / Fluorinated building blocks / 3,3-Difluoroazetidin-2-ones / α,α-Difluoro-β-amino acids / Reformatsky reaction

The synthesis of gem-difluorinated β -lactams and gem-difluorinated β -amino acids, each possessing a potential basic functional group, from ethyl bromodifluoroacetate and either imines (for β -lactams) or N-(α -aminoalkyl)benzotriazoles (for β -amino esters) was investigated. A series of these compounds were used for the design of novel metallocarboxy-

Introduction

Introduction of fluorine atoms into bioorganic and bioactive molecules often induces modifications of chemical, physical, and biological properties and as a consequence leads to the generation of novel and potent biological, pharmacological, and chemotherapeutic agents.^[1-3] The strength of the C-F bond (485.6 kJmol⁻¹) confers relative stability against metabolic transformations.^[4] Furthermore, the small nature of the steric perturbations produced in the molecule, because of the relative homology of the van der Waals radii of H (1.20 Å) and F (1.47 Å), usually allows them to enter metabolic pathways similarly to the corresponding non-fluorinated compounds. The replacement of hydrogen by fluorine does alter the properties of molecules, however, affecting the basicity or acidity of neighboring groups, lipophilicity/hydrophobicity, hydrogen bonding, dipole moment, and overall reactivity and stability. As a result, a large number of physiologically active compounds and therapeutic agents containing strategically located fluorine atoms are currently widely used or in development.^[5]

In the past decade, increasing work has been devoted to the study of nonnatural foldamers and sequence-specific oligomers that mimic various aspects of the folding, organization, and function of polypeptides or biological poly-

[b] Division D of Medicinal Chemistry, Institut de Recherches Servier, 11 rue des Moulineaux, 92150 Suresnes, France peptidase inhibitors. *N*-Alkylation and *N*-acylation of these two versatile scaffolds were carried out, leading to the expected targets in moderate to good yields.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2008)

mers.^[6] In particular, β -amino acids are recognized as pharmacologically important compounds,^[7,8] since β -peptides are able to adopt stable and well-organized conformations.^[9] Such building blocks provide useful scaffolds for the design of functional mimics of natural proteinic structures. Furthermore, β -peptides are stable to proteolytic degradation in vitro and in vivo,^[10] which represents an important advantage over natural peptides and proteins. Moreover, β amino acids also play key roles in medicinal chemistry as, for example, precursors of pharmacologically important β lactam antibiotics^[11] and components of biologically active unnatural oligopeptides.^[12]

Because of the exciting benefits of fluorine substitution and the importance of amino acids in biochemical functions, the area of fluorine-containing amino acids (FAAs) is expanding rapidly, taking a major place in the family of nonnatural amino acids.^[13] FAAs are known to show antibacterial activity, to act as proteinase inhibitors, and to exhibit promising properties as candidates for peptide modification.^[14] Given the biomedicinal and synthetic potential reported for β -amino acids, and the biological influence of fluorine substitution in the β -position relative to the amino group, it is not surprising that considerable efforts have been devoted to the development of efficient syntheses of gem-difluoromethylene compounds,^[15] and more specifically of α, α -difluoro- β -amino acids. Replacement of various functional groups by a gem-difluoromethylene group has generated potent transition-state-type inhibitors.^[16] Several α, α -difluoro- β -amino acids^[17] or gem-difluorinated peptides^[18] have been synthesized and tested as potent competitive serine protease inhibitors or [18F]-radiolabeled markers.^[19] The gem-difluoromethylene/methylene transposition in the "western" β -amino acid moiety of the naturally



 [[]a] UMR CNRS 6014, Institut de Recherche en Chimie Organique Fine, INSA et Université de Rouen,
 1 rue Tesnière, 76131 Mont-Saint-Aignan Cedex, France Fax: +33-2-35522959
 E-mail: philippe.jubault@insa-rouen.fr quirion@insa-rouen.fr.
 [h] Diricing C. functional Chamistan Institute de Bechercher

occurring antifungal cyclic tetrapeptide rhodopeptin results in improved activity and novel physical properties (Scheme 1).^[20] Incorporation of fluorinated β -amino acids in the side chain of analogues of Docetaxel (Taxotere[®]) was investigated to improve the cytotoxicity of this chemotherapy drug (Scheme 1).^[21] The introduction of a β -amino acid bearing one or two fluorine atoms in its 2-position into a β -peptide chain induces an alteration of the neighboring electronic environment without incurring steric constraints. Such modifications improve stability against enzymatic degradations and can beneficially influence the biological activity.^[22]



Scheme 1. Examples of introduction of β -amino acid moieties into bioactive compounds.

The azetidin-2-one (B-lactam) skeleton has attracted significant interest among synthetic and medicinal chemists, mainly because it is the core of natural and nonnatural antibiotics. Great efforts have been dedicated to the synthesis of new types of β -lactams to address the bacterial resistance of some of these drugs.^[23] To tackle this growing problem, the introduction of fluorine, in particular of the difluoromethylene moiety, into the azetidinone structure has been suggested because of the unique changes produced in physiochemical, and consequently biological, behavior. 3,3-Difluoroazetidin-2-ones have been already synthesized, and some of them have demonstrated potential biological activities^[18a,24] (Scheme 2) or have turned out to be versatile synthetic intermediates and useful building blocks.^[25,26] The growing interest in 3.3-difluoroazetidines^[27] is illustrated by their potential therapeutic inhibitory activity towards dipeptidyl peptidase-IV for the antihyperglycaemic treatment of type 2 diabetes.^[28]



Scheme 2. Examples of bioactive 3,3-difluoroazetidin-2-ones.

The importance and usefulness of β -lactams as versatile synthetic intermediates has also been widely recognized with the development of the " β -lactam synthon method"^[29] for the preparation of various amino acids, peptides, peptidomimetics, human cytomegalovirus protease inhibitors based on activated carbonyl groups,^[30] cholesterol absorption inhibitors,^[31] and other types of compounds of biological and medicinal interest.

In the course of a medicinal chemistry program devoted to the synthesis of basic metallocarboxypeptidase inhibitors, we were interested in small and original molecules. In this study we report the synthesis and functionalization of α,α -difluoro- β -amino acid and 3,3-difluoroazetidin-2-one templates. Our strategy was based on the salt bridge interactions between the C-terminal carboxylate of the substrate and the basic residues of the active site of the carboxypeptidase. 3,3-Difluoroazetidin-2-ones can be regarded as the masked forms of gem-difluoroacetyl functional groups, and have turned out to be prodrugs for the corresponding β amino acids. Since a metalloprotease substrate requires a zinc-binding group (ZBG) for coordination with the catalytic zinc atom of the active site, it was decided to use the neutral thiol moiety and the carboxylic acid function as ZBG. We were therefore interested in molecules of small size and high water solubility possessing a carboxylate or a thiol as zinc-chelating group and a basic functional group. β-Substituted α,α -difluoro-β-amino acids and 3,3-difluoroazetidin-2-ones are attractive scaffolds for the design of novel metallocarboxypeptidase inhibitors (Scheme 3).

$$R^1$$
 O R^1 NH O $R^1 = Zn^{2+}$ binding group
 R^2 F R^2 OH $R^2 =$ basic functional group

Scheme 3. Design of potent metallocarboxypeptidase inhibitors.

Several general methodologies for the syntheses of the 3,3-difluoroazetidin-2-one and α, α -difluoro- β -amino ester skeletons have been developed. Among them, Reformatskytype reactions^[32] with ethyl halodifluoroacetate are usually applied either with carbonyl compounds or with imines, thus leading to β-hydroxy esters and β-amino ester/β-lactam mixtures,^[33] respectively (Scheme 4). In the first case, the pathways mainly involve either a two-step strategy with conversion of the formed 2,2-difluoro-3-hydroxy esters into the desired 2,2-difluoro-3-hydroxypropionamides^[21a,26,34] or hydroxamates,^[18a] followed by internal N1-C4 cyclization or the replacement of a hydroxy group by different nucleophiles^[21a,35] under Mitsunobu conditions. In the second case, condensation of the appropriate Reformatsky reagents derived from ethyl bromodifluoroacetate with aldimines or synthetic equivalents can lead, depending on the



Scheme 4. General strategies directed towards the production of 3,3-difluoroazetidin-2-ones and α,α -difluoro- β -amino esters through Gilman–Speeter and Reformatsky reactions.

reaction conditions and/or the substitution of the substrate, either to the β -amino ester or 3,3-difluoroazetidin-2-one derivatives or to mixtures of these two products (these are difficult to separate in most cases).^[18b,34c,36] The Gilman– Speeter reaction, also called ester-imine condensation, has emerged as a powerful approach for the synthesis of *gem*difluoro- β -amino carbonyl compounds, mainly in their racemic forms, but also in the enantiomerically pure series.^[25b-25d,37]

We recently disclosed the stereoselective and chemoselective preparation of 3,3-difluoroazetidin-2-ones and α,α -difluoro- β -amino acids.^[38] The key transformation is the addition, with high levels of stereoselectivity (up to 98%), of the organozinc reagent derived from ethyl bromodifluoroacetate to chiral 1,3-oxazolidines or aldimines derived from aliphatic and aromatic aldehydes, and either (*R*)-phenylglycinol or (*R*)-methoxyphenylglycinol.

To extend the usefulness of this type of reaction in the synthesis of biologically active compounds, we now report our results relating to the straightforward and chemoselective synthesis of β -lactams and β -amino esters derived from several aldehydes bearing basic functional groups. We have also investigated the introduction of thiol and carboxylate zinc-binding groups in good to excellent yields by *N*-alkylation of β -lactams and *N*-acylation of β -amino esters.

Results and Discussion

To develop original compounds as novel metallocarboxypeptidase inhibitors, we prepared a series of β -lactams and β -amino esters substituted (R², Scheme 3) with protected or unprotected basic groups [namely pyridin-3-yl, pyridin-4yl, piperidin-4-yl, and 4-(aminomethyl)benzyl groups]. Such functional groups can be introduced by use of the corresponding aldehydes.

Preparation of Starting Aldehydes

The Cbz-protected piperidine-4-carbaldehyde **4b** was prepared by the synthetic route outlined in Scheme 5. Reduction of commercially available methyl isonipecotate (1) with LiAlH₄ in THF gave **2** in 90% yield.^[39] The protection of (piperidin-4-yl)methanol as its Cbz derivative followed by Swern oxidation afforded aldehyde **4b** in 58% overall yield.^[40]



Scheme 5. (a) LiAlH₄, THF, room temp.; (b) CbzCl, Na₂CO₃, CH₂Cl₂/H₂O, room temp.; (c) (COCl)₂, DMSO, DIEA, CH₂Cl₂, $-60 \text{ }^{\circ}\text{C} \rightarrow \text{room temp.}$

Cbz-protected 4-(aminomethyl)benzaldehyde derivative **4c** was prepared from 4-cyanobenzaldehyde (**5**) in a fourstep sequence as shown in Scheme 6. Formation of dioxolane-type acetal **6** was followed by the reduction of the cyano group.^[41] The primary amine **7** was converted into the corresponding *N*-Cbz-protected amine and was then subjected to mild acidic conditions to remove the acetal in 93% yield.^[42]



Scheme 6. (a) Ethylene glycol, PTSA, toluene, reflux; (b) LiAlH₄, THF, 0 °C \rightarrow 65 °C; (c) CbzCl, Na₂CO₃, CH₂Cl₂/H₂O, room temp.; (d) AcOH, H₂O, room temp.

Synthesis of gem-Difluoro-β-lactams and -β-amino Esters

Racemic *N*-deprotected 3,3-difluoroazetidin-2-ones can be efficiently prepared in a three-step route based on Gilman–Speeter reactions between the Reformatsky reagent derived from ethyl bromodifluoroacetate and the appropriate aldimines (Scheme 7). (*p*-Methoxybenzyl)amine was used, because the PMB protecting group can be easily removed under mild conditions. The imines were easily prepared by condensation of the appropriate arene-, heterocycle-, and alkanecarbaldehydes **4a–e** and (*p*-methoxybenzyl)amine in CH_2Cl_2 in the presence of MgSO₄, and were isolated quantitatively without further purification. Compounds **9a–e** were treated with ethyl bromodifluoroacetate (2 equiv.) in the presence of freshly activated Zn dust in



Scheme 7. Synthesis of 3,3-difluoroazetidin-2-ones. (a) MgSO₄, *p*-MeOC₆H₄CH₂NH₂, CH₂Cl₂, room temp.; (b) BrZnCF₂CO₂Et, THF, reflux; (c) CAN, CH₃CN, H₂O, 0 °C.

THF at reflux over 2 h, leading to the expected products in high yields. In situ activation of zinc, performed with dibromoethane and chlorotrimethylsilane at room temp., and preparation of the organozinc reagent prior to the addition of the aldimine led to higher yields under milder conditions.^[43] The results are summarized in Table 1. The difluoroazetidin-2-ones 10 were the major or the only isolated products except in the case of 9c, in which a particular result was observed (Table 1, Entry 3): a 1:1 β -lactam 10c/ β amino ester 11c ratio was obtained in good overall yield. As previously described by our group,^[38b] the presence of a primary carbamate greatly disfavored the cyclization step. Pyridine derivatives **9d** and **9e** each gave 10% yields of α . α difluoro- β -amino esters **11d** and **11e**, which could not be eliminated after purification on silica gel by flash chromatography. Nevertheless, this noncyclized byproduct was degraded during the oxidative debenzylation step. Deprotection was then carried out by a classical procedure with 3 equiv. of ceric ammonium nitrate (CAN) in a mixture of acetonitrile and water in a 9:1 ratio, leading to racemic azetidin-2-ones 12a-e in moderate to good yields.[44]

Table 1. Preparation of azetidin-2-ones 10 and 12.

Entry	Starting aldehyde	Yield $10 + 11$ [%] ^[a]	Ratio 10/11 [%] ^[b]	Yield 12 [%] ^[c]
1	4 a	74	100:0	83 ^[d]
2	4 b	70	100:0	38
3	4c	72	52:48 ^[e]	43
4	4d	85 ^[f]	89:11 ^[g]	62 ^[h]
5	4e	86 ^[f]	90:10 ^[g]	49 ^[h]

[a] Global isolated yield. [b] Ratios determined by ¹⁹F NMR spectroscopy. [c] Isolated yield from β -lactam **10**. [d] See ref.^[38a] for full characterization. [e] The two products can be separated on silica gel. [f] See ref.^[37b] for full characterization of this product. [g] Not separable by silica gel flash chromatography. [h] See ref.^[38b] for full characterization.

As we were looking for an efficient and short way to prepare β -amino ester derivatives in a selective manner, we decided to investigate Katritzky's procedure,^[36c] using the ethyl bromodifluoroacetate zinc derivative and *N*-(α -amino-alkyl)benzotriazoles for the preparation of α , α -difluoro- β -amino esters.

N-(α -Aminoalkyl)benzotriazoles **13** (Scheme 8), easily prepared from dibenzylamine, benzotriazole, and aldehydes **4b**-e, were used without purification as iminium salt precur-

sors in Katritzky's methodology.^[45] To our delight, benzotriazole-mediated aminoalkylations provided protected α , α difluorinated β -amino esters **14b**–e in high yields through Reformatsky reactions with ethyl bromofluoroacetate, activated zinc, and trimethylsilyl chloride in THF at reflux.

We then turned our attention to the conversion of dibenzylated amines into primary amines by catalytic hydrogenolysis (Scheme 9). Selective removal of benzyl protecting groups from 14b was achieved by a two-step route. Hydrogenolysis of the Cbz group followed by in situ Boc protection gave 14f in quantitative yield. Debenzylation of the tertiary amine by hydrogenolysis in the presence of Pearlman's catalyst in ethanol or in a mixture of ethanol and ethyl acetate was inefficient, and a complex mixture of products was obtained, due to transcarbamoylation. Other conditions such as catalytic transfer hydrogenation with cyclohexene turned out to be inefficient in our case.^[46] Hydrogenolysis of the two benzyl protecting groups was finally carried out with the following combination: H_2 (1 bar)/Pearlman's catalyst/EtOH/MeOH/room temp. to afford the β-amino ester 15b in 17% yield.



Scheme 9. (a) H_2 (1 bar), Boc_2O , Pd/C, EtOH, 40 °C; (b) H_2 , Pd(OH)₂/C, EtOH/MeOH, 40 °C; (c) H_2 (1 bar), Pd/C, EtOH, Ac-OEt, HCl/*i*PrOH (6 N), 40 °C; (d) Boc_2O (1 equiv.), Et₃N, CH₂Cl₂, room temp.; (e) H_2 (1 bar), Pd/C, EtOH, HCl/*i*PrOH (6 N), room temp.



Scheme 8. Synthesis of β -branched ethyl 3-(dibenzylamino)-2,2-difluoropropanoates. (a) Bn₂NH, BtH, EtOH, 78 °C; (b) BrZnCF₂CO₂Et, TMSCl, THF, 67 °C.



Removal of the benzyl functional groups in 14c was achieved by hydrogenolysis [Pearlman's catalyst, H_2 (1 bar), EtOH, AcOEt, HCl/*i*PrOH 6 N, 40 °C] to afford diamine 16 as a monochloride salt (72%). Selective protection with Boc₂O (1 equiv.) was tentatively carried out in a conventional manner but was not selective enough, so 15c (and its methyl analog 15c' obtained by transesterification of 15c during the purification process due the presence of methanol in the eluent composition) was isolated in only 19% yield. Cleavage of the benzyl groups in 14d and 14e was best achieved by hydrogenolysis in the presence of Pd/C catalyst under 1 bar of hydrogen in EtOH and HCl/*i*PrOH (6 N) to afford 15d and 15e in excellent yields.

N-Alkylation of gem-Difluorinated β-Lactams

N-Alkylation of β -lactams **12a**–e, as required for the preparation of potential metallocarboxypeptidase inhibitors, was then investigated. A review of the literature revealed that N-alkylation of N-unsubstituted β-lactams was not consistently efficient.^[47] Strong bases (such as NaH,^[48] NaNH₂, tBuOK,^[49] nBuLi,^[50] or LiHMDS^[51]) have been employed with basic scaffolds, but such basic conditions induce either polymerization of azetidin-2-ones or epimerization of chiral centers (especially at C-3). Alkylation under less basic conditions have been described either with organic non-nucleophilic bases such as Et₃N or DIEA or with inorganic bases (K2CO3 in acetone,^[52] Cs2CO3 in MeCN,^[53] CsF in DMF,^[54] Ag₂O in MeCN^[55]). These milder conditions were compatible with various functionalities and prevented potential epimerization. However, only highly reactive electrophiles gave satisfactory levels of conversion and yields, probably because of the low nucleophilicity of the nitrogen atom. Good yields can be obtained by phase-transfer catalysis methodology (KOH/nBu₄NBr/THF) as described by Reuschling et al.,^[56] despite the risks of side reactions or nucleophilic opening of the β -lactam ring.

In order to find the best conditions for alkylation, we first turned our attention to the introduction of the carbox-

ylate moiety with bromoacetate esters (17a: R = methyl, 17b: R = benzyl). The required side chain can be introduced by base-mediated alkylation of the nitrogen atom in a 1unsubstituted 3,3-difluoroazetidin-2-one. To the best of our knowledge, only one example of such a strategy has been reported. De Kimpe^[57] described the synthesis of *N*-substituted *gem*-difluorinated azetidinones under phase-transfer conditions.

The N-alkylation of model β -lactam 12a was therefore carefully optimized by varying the electrophile, the base, and experimental conditions (Scheme 10, Table 2). Extensive experimentation showed that numerous sets of conditions were either unsuccessful or gave inconsistent results. Treatment with strong bases such as NaH or LDA proved to be inefficient. Attempts with KOH in homogeneous or heterogeneous systems induced complete β-lactam degradation. Functionalization of 12a under the phase-transfer conditions described by De Kimpe^[57] turned out to be inefficient in our case. Whereas Hünig's base caused rapid degradation and triethylamine gave low levels of conversion, treatment of β -lactam 12a with methyl or benzyl bromoacetates, and tetramethylguanidine in the presence of TBAI (tetrabutylammonium iodide) provided the N-substituted azetidin-2-ones 18a and 18b in excellent yields (Table 2, Entries 5–6).

In order to introduce a sulfhydryl group as a Zn-chelating function, we then applied these conditions to *S*-(3-bromopropyl)thioacetate.^[58] Unfortunately, none of these experimental conditions allowed access to the corresponding *N*-substituted β -lactam, due to the poor electrophilicity of this alkylating agent. We next examined deprotection of **18a** and **18b** in order to obtain functionalized *N*-substituted lactams. Compound **18a** was subjected to various sets of acidic or basic conditions in order to carry out saponification. In most cases, we observed rapid degradation of the β -lactam skeleton. However, when benzyl ester **18b** was subjected to hydrogenolysis conditions (H₂, Pd/C), we were pleased to observe the formation of 2-oxoazetidin-1-ylacetic acid **19** in 91% yield.



Scheme 10. Synthesis of N-substituted azetidin-2-ones.

Table 2.	Optimization	of the	N-alkylation	step

Entry	Base [equiv.]	BrCH ₂ CO ₂ R [equiv.]	Solvent	Temperature	Time [h]	18, yield [%]
1	tBuOK (1.6)	17a (1.5)	THF	$0 ^{\circ}\mathrm{C} \rightarrow \mathrm{room} \mathrm{temp}.$	20	18a , 37 ^[a]
2 ^[b]	KOH (2)	17a (1.5)	THF	room temp.	20	18a, 50 ^[c]
3	CsF (10)	17a (3)	DMF	room temp.	72	18a , 10 ^[c]
4	$Et_{3}N(1)$	17a (10)	THF	room temp.	14	18a, 20 ^[c]
5 ^[b]	$TMG^{[d]}(2)$	17a (10)	THF	room temp.	6	18a, quantitative
6 ^[b]	$TMG^{[d]}(2)$	17b (10)	THF	room temp.	18	18b , 91 ^[a]

[a] Isolated yields. [b] Addition of 0.5 equiv. of TBAI. [c] Conversion determined by ¹⁹F NMR relative to **12a**. [d] 1,1,3,3-Tetramethyl-guanidine.

This synthetic strategy was then applied to several highly functionalized 3,3-difluoroazetidin-2-ones (Scheme 11). With this goal, we performed alkylation of β -lactams **12b**-



Scheme 11. (a) BrCH₂CO₂Bn, TMG, TBAI, MeCN, room temp.; (b) H₂, Pd/C, EtOH, room temp.; (c) BrCH₂CO₂tBu, TMG, TBAI, MeCN, room temp.; (d) TFA, CH₂Cl₂, 0 °C.

Table 3. Syntheses of functionalized 3,3-difluoroazetidinones.

Entry	Substrate	Alkylation agent [equiv.]	R	20 or 22 , yield [%] ^[a]	21, yield[%] ^[b]
1	12b	17b (37)	Bn	20b , 30	21b , 50
2	12c	17b (11)	Bn	20c , 95	21c, 25
3	12d	17b (1.1)	Bn	20d , 29	n.r. ^[c]
4	12e	17b (1.1)	Bn	20e , 50	n.r. ^[c]
5	12d	17c (1.1) ^[d]	tBu	22d , 47	21d , 44 ^[e]
6	12e	17c (1.1) ^[d]	<i>t</i> Bu	22e , 50	21e , 56 ^[e]

[a] Isolated yield. [b] Isolated yield after purification by gel permeation chromatography. [c] No reaction under atmospheric pressure of hydrogen, and higher pressure induced the slow formation of unidentified compounds. [d] Use of NaI (1 equiv.) instead of TBAI. [e] Partial degradation: formation of β -amino acids produced by cleavage of the N–C(O) bond in the β -lactam intermediate by water during purification. e with benzyl bromoacetate **17b** under previously optimized conditions, to provide **20b–e** in moderate to excellent yields (Table 3).

In the case of pyridinyl derivatives (12d and 12e), 1 equiv. of benzyl bromoacetate was used in order to minimize alkylation of the pyridine ring. Treatment of 20b and 20c with hydrogen in the presence of Pd/C resulted in removal of the *N*-Cbz protecting group and the labile benzyl ester to afford amino acids 21b and 21c (Scheme 11). Unexpected partial degradation occurred during hydrogenolysis of 20c, leading to the cleavage of the N1-C4 bond of the β-lactam ring and the isolation of the corresponding amido acid. Disappointingly, removal of the benzyl ester protecting groups from compounds 20d and 20e with Pd/C hydrogenation systems turned out to be ineffective, whereas higher pressures of hydrogen induced side reactions. To circumvent this problem, we investigated the synthesis of tert-butyl esters that could be used to facilitate deprotection of the carboxylate. Subsequent treatment of azetidin-2-ones 12d and 12e with tertbutyl bromoacetate (17c) and TMG in MeCN afforded Nalkylated products (22d and 22e) in 50% yield. Acidic deprotection (50% TFA/CH₂Cl₂) of these compounds afforded the acids, which were purified by gel permeation chromatography to recover β -lactams **21d** and **21e** in 44% and 56% yields, respectively.

N-Acylation of gem-Difluorinated β-Amino Acids

In an effort to synthesize novel potential metallocarboxypeptidase inhibitors, we decided to develop a family of sulfhydryl acids containing basic groups. We envisioned the preparation of these targets by N-acylation of the β -amino



Scheme 12. (a) DIEA, DMAP, CH_2Cl_2 , room temp.; (b) 50% TFA, CH_2Cl_2 , 0 °C; (c) NaOH, H₂O, MeOH, room temp.; (d) $HCl_{(g)}$, MeOH, room temp.; (e) Et₃N, DMAP, CH_2Cl_2 , room temp.; (f) LiOH, H₂O, MeOH, THF, 0 °C.



acids derived from α, α -difluorinated β -amino esters **15b**-e, since these molecules are highly useful synthetic intermediates for the insertion of zinc-binding groups. We were interested in the introduction of thiol and carboxylate functionalities.

This strategy thus required the use of a highly reactive electrophile in order to overcome the rather low reactivity of the deactivated amine due to the inductive effect of the neighboring CF_2 moiety.

Keeping the aforementioned goals in mind, we decided to use acyl chlorides bearing masked sulfhydryl or carboxylic functions. Starting from mercaptoacetic acid, we therefore prepared (acetylsulfanyl)acetic acid (**23**; Scheme 12) by acetylation with Ac₂O and Et₃N, followed by the action of oxalyl chloride in CH₂Cl₂ at room temperature, in 43% overall yield.^[59] Commercially available methyl malonyl chloride (**24**) proved to be an excellent starting material for the introduction of a carboxyl group.

The synthesis of the series of sulfhydryl acids 27 was carried out by starting from the appropriately substituted gemdifluorinated β-amino esters 15b-e (Scheme 12). Addition of (acetylthio)acetyl chloride (23) in the presence of catalytic DMAP gave the acylated compounds 25b-e in moderate to good yields. The thioester 25b was then converted into the thiol 27b by Boc deprotection with 50% TFA/ CH₂Cl₂ followed by basic deprotection with aqueous NaOH in 59% yield. Removal of the protecting groups of 25c and 25c' with methanolic HCl followed by treatment with aqueous NaOH gave disulfide 27c in moderate yield. In addition to rapid and unavoidable oxidation of thiol functionality, the amide bond proved to be particularly weak, and, as a consequence, the corresponding β -amino acid was isolated in only 20% yield. Deprotection of 25e with aqueous LiOH afforded 27e.

With an effective sequence to hand, a similar route was used for diacid compounds **28d** and **28e**, but methyl malonyl chloride (**24**) was used instead of **23**. Treatment with triethylamine in CH_2Cl_2 resulted in acylation to give diesters **26d** and **26e** in good yields, and these were then hydrolyzed to diacids. Unfortunately, due to an arduous purification, the yields of **28d** and **28e** were only 20%.

Conclusions

We succeeded in the synthesis of the target molecules displaying specific features in the course of our medicinal program devoted to the synthesis of basic metallocarbopeptidase inhibitors. A general synthetic route, in the racemic series, of various *gem*-difluoro- β -lactams and - β -amino esters, possessing different basic groups, has been successfully developed. *N*-Alkylation of *gem*-difluoro- β -lactams and *N*acylation of *gem*-difluoro- β -amino esters were carried out, leading, in moderate to good yields, to the expected potential basic metallocarbopeptidase inhibitors.

Experimental Section

Abbreviations: AcOH = acetic acid; Boc = tert-butoxycarbonyl; BopCl = bis(2-oxo-3-oxazolidinyl)phosphinic chloride; BtH = 1H- benzotriazole; CAN = ceric ammonium nitrate; Cbz = benzyloxycarbonyl; CH₂Cl₂ = dichloromethane; CI = chemical ionization; DIEA = N,N-diisopropylethylamine; DMAP = 4-(dimethylamino)pyridine; DMF = dimethyl formamide; DMSO = dimethyl sulfoxide; EI = electron impact ionization; ESI = electrospray ionization; EtOAc = ethyl acetate; FAAs = fluorine-containing amino acids. IR = infrared; LDA = lithium diisopropylamide; LiHMDS = lithium hexamethyldisilazide; MeCN = acetonitrile; MeOH = methanol; MS = mass spectrometry; NMR = nuclear magnetic resonance; PMB = p-methoxybenzyl; PTSA = p-toluenesulfonic acid; TBAI = tetrabutylammonium iodide; TFA = trifluoroacetic acid; THF = tetrahydrofuran; TLC = thin layer chromatography; TMG = N,N,N',N'-tetramethylguanidine; TMSCI = chlorotrimethylsilane; ZBG = zinc-binding group; Zn* = activated zinc dust.

General: Unless otherwise mentioned, all the reagents were purchased from commercial sources and used as received. All glassware was dried in an oven at 100 °C prior to use. THF was distilled from sodium/benzophenone ketyl under nitrogen prior to use. Dichloromethane (CH₂Cl₂) was distilled under nitrogen from P_2O_5 prior to use. DMSO and MeCN were distilled under nitrogen from CaH₂, and DMF from BaO. NMR spectra were recorded with a Bruker DXP 300 instrument. ¹H NMR chemical shifts (300.13 MHz) are expressed in parts per million (ppm, δ) downfield from tetramethylsilane ($\delta = 0$ ppm) in CDCl₃. ¹³C NMR chemical shifts (75.47 MHz) are expressed in parts per million downfield from CDCl₃ as internal standard (δ = 77.16 ppm). ¹⁹F NMR chemical shifts (282.40 MHz) are expressed in parts per million downfield from CFCl₃ as internal standard ($\delta = 0$ ppm). Coupling constants J are reported in Hertz. Abbreviations used for peak multiplicity are: br.: broad; s: singlet; d: doublet; t: triplet; q: quadruplet; m: multiplet. Progress of the reactions was monitored by TLC on Merck silica gel plates (thickness 0.2 mm with fluorescence indicator 60F₂₅₀). Flash column chromatography purifications were carried out on 40-63 mesh silica gel 60A by "flash" methodology. Silica TLC plates were visualized under UV light, by use of a solution of phosphomolybdic acid in ethanol (10%) followed by heating, or by use of a solution of aqueous alkaline potassium permanganate (1 N) and heating. Values of R_f were measured after an elution of 6 cm. Infrared (IR) spectra were recorded with a Perkin-Elmer 1420 instrument. Absorption bands are reported in cm⁻¹. Elemental analyses were performed with a Carlo Erba 1106 instrument. Melting points are uncorrected. Mass spectra were performed with a Thermofinnigan Navigator 2.1 instrument for electrospray or with a JEOL AX500 instrument (isobutane, 200 eV) for CI. HRMS measurements were performed with a JEOL AX500 spectrometer.

Starting Materials: All substrate imines were readily synthesized by condensation of the appropriate aldehyde with (p-methoxybenzyl)amine according to the following procedure. Anhydrous MgSO4 (2 g) and the appropriate aldehyde (1 equiv.) were added at ambient temperature to a solution of the corresponding amine (10 mmol) in CH₂Cl₂ (10 mL). The mixture was stirred at room temperature for 12 h. The solids were filtered by suction, and the filtrate was washed with CH_2Cl_2 (2×20 mL). The corresponding imine was recovered quantitatively after concentration under reduced pressure, and was used without purification. Activated zinc was prepared by stirring a quantity of zinc dust in dilute hydrochloric acid for about 5 min. The zinc was then filtered off, washed neutral with deionized water, with ethanol, with acetone, and with diethyl ether, and dried at 100 °C under vacuum.^[32a] All N,N-disubstituted 1Hbenzotriazolylamines were synthesized according to the following general procedure. Dibenzylamine (3.21 g, 30 mmol) and the appropriate aldehyde (30 mmol, 1 equiv.) were successively added to

a solution of 1*H*-benzotriazole (3.57 g, 30 mmol, 1 equiv.) dissolved in a minimum amount of ethanol (ca. 5 mL). The mixture was heated at reflux for 4 h in the presence of dried molecular sieves (3 Å, 1 g), and then stirred at room temp. for 12 h. After removal of molecular sieves by suction filtration through a pad of Celite and concentration of the filtrate under vacuum, the obtained solid could be recrystallized from ethanol.

(Piperidin-4-yl)methanol (2): A solution of methyl isonipecotate (1, 10 mL, 74 mmol) in anhydrous THF (80 mL) was slowly added under argon to an ice-cooled suspension of LiAlH₄ (3.5 g, 92.2 mmol, 1.25 equiv.) in anhydrous THF (250 mL). After the visible gas evolution had ceased, the mixture was left standing at room temp. for 20 h. After treatment with water (4 mL), aqueous NaOH solution (1 N, 4 mL), and water (8 mL), and stirring with diethyl ether (200 mL) for 30 min, the slurry was filtered and washed with diethyl ether (2 \times 100 mL). The organic fraction was dried with MgSO₄, and the solvents were removed under vacuum to afford the title compound as a colorless oil; yield 7.67 g (90%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 3.42 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 3.05 (dt, ${}^{3}J_{H,H} = 3$, ${}^{2}J_{H,H} = 12$ Hz, 2 H), 2.56 (td, ${}^{3}J_{H,H} = 2.5$, ${}^{2}J_{H,H} =$ 12 Hz, 2 H), 2.15 (br. s, 2 H), 1.75-1.65 (m, 2 H), 1.65-1.50 (m, 1 H), 1.10 (ddd, ${}^{3}J_{H,H} = 4$, ${}^{3}J_{H,H} = 13$, ${}^{2}J_{H,H} = 25$ Hz, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 68.3, 46.6, 39.4, 30.2 ppm. IR (KBr): $\tilde{v}_{max} = 3368$, 1634, 1039 cm⁻¹. MS (EI+): m/z (%) = 115.0 [M]⁺. C₆H₁₃NO (115.18): calcd. C 62.57, H 11.38, N 12.16; found C 62.09, H 11.44, N 12.28.

Benzyl 4-(Hydroxymethyl)piperidine-1-carboxylate (3): Amine 2 (7.67 mmol, 66.6 mmol) was dissolved in CH₂Cl₂ (230 mL) at 0 °C. A solution of Na₂CO₃ (32 g, 302 mmol, 4.5 equiv.) in water (230 mL) was added. Benzyloxycarbonyl chloride (10 mL, 72 mmol, 1.1 equiv.) was added dropwise. After the system had been stirred at room temp. for 12 h, ice-cold water (200 mL) was added, and the mixture was extracted with CH_2Cl_2 (2×100 mL). The organic layers were combined, filtered, dried with MgSO₄, and concentrated in vacuo to give the crude material. Silica gel flash column chromatography [CH2Cl2/EtOAc (8:2 to 6:4) as eluent] gave pure 3 as a colorless oil; yield 14.78 g (89%); TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.39$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.45-7.35 (m, 5 H), 5.19 (s, 2 H), 4.35-4.20 (m, 2 H), 3.55 (d, ${}^{3}J_{\text{H,H}} = 6 \text{ Hz}, 2 \text{ H}$, 2.90–2.80 (m, 2 H), 1.85–1.75 (m, 2 H), 1.75– 1.65 (m, 1 H), 1.30–1.15 (m, 2 H) ppm. $^{13}\mathrm{C}$ NMR (CDCl_3, 75.4 MHz): δ = 155.2, 136.7, 128.4, 127.9, 127.7, 67.3, 66.9, 43.8, 38.6, 28.4 ppm. IR (KBr): $\tilde{v}_{max} = 3432$, 1697, 1439, 1247, 1215, 1038 cm^{-1} . MS (EI+): $m/z = 249 \text{ [M]}^+$, 204, 158, 142, 91. C14H19NO3 (249.31): calcd. C 67.45, H 7.68, N 5.62; found C 67.90, H 7.57, N 5.14.

1-(Benzyloxycarbonyl)piperidine-4-carboxaldehyde (4b): A stirred solution of freshly distilled DMSO (13.3 mL, 187.3 mmol, 4.5 equiv.) in anhydrous CH2Cl2 (70 mL) was cooled to -78 °C. A solution of oxalyl chloride (5.3 mL, 61.8 mmol, 1.5 equiv.) in dry CH₂Cl₂ (10 mL) was then added dropwise, and stirring was continued for 30 min. Next, a solution of alcohol 3 (10.4 g, 41.7 mmol) in anhydrous CH₂Cl₂ (150 mL) was added dropwise over 15 min, and the resultant slurry was stirred at -78 °C for 30 min. N,N-Diisopropylethylamine (36.1 mL, 217.9 mmol, 5.25 equiv.) was added, and the resultant slurry was stirred at $-60\ ^\circ\mathrm{C}$ for an additional 30 min. The mixture was allowed to warm to room temp. overnight. A solution of NaH₂PO₄ (7.5 g, 64.1 mmol, 1.5 equiv.) in water (100 mL) was added, and the mixture was extracted with CH₂Cl₂ $(2 \times 200 \text{ mL})$. The organic phases were washed with aqueous HCl (1 N, 100 mL), saturated aqueous NaHCO₃ solution (100 mL), and brine (100 mL), and were then dried with anhydrous MgSO₄. The

solvent was removed in vacuo, and the residual product was purified by chromatography on a silica gel column [cyclohexane/EtOAc (9:1 to 6:4) as eluent] to give **4b** as a colorless oil; yield 7.42 g (72%); TLC: silica gel (cyclohexane/EtOAc, 6:4), $R_f = 0.38$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 9.64$ (s, 1 H), 7.35–7.30 (m, 5 H), 5.10 (s, 2 H), 4.10–3.95 (m, 2 H), 3.00 (dt, ${}^{3}J_{H,H} = 2.5$, ${}^{2}J_{H,H} = 13$ Hz, 2 H), 2.45–2.35 (m, 1 H), 1.95–1.85 (m, 2 H), 1.56 (ddd, ${}^{3}J_{H,H} = 4$, ${}^{3}J_{H,H} = 11$, ${}^{2}J_{H,H} = 24$ Hz, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): $\delta = 202.7$, 128.5, 128.0, 127.9, 67.1, 47.7, 43.0, 25.0 ppm. IR (KBr): $\hat{v}_{max} = 1728$, 1698, 1278, 1225 cm⁻¹. C₁₄H₁₇NO₃ (247.30): calcd. C 68.00, H 6.93, N 5.66; found C 67.15, H 6.57, N 5.55.

4-(1,3-Dioxolan-2-yl)benzonitrile (6): 4-Cyanobenzaldehyde (5, 15 g, 114.4 mmol) was placed in a round flask together with ethyleneglycol (11.5 mL, 206 mmol, 1.8 equiv.), dry toluene (300 mL), and p-toluenesulfonic acid as catalyst (1.5 g, 8.7 mmol, 8% with respect to 5). The solution was heated at reflux in a Dean-Stark apparatus with water trap for 18 h. It was then neutralized with saturated aqueous NaHCO₃ solution (100 mL), and the phases were separated. The aqueous layer was extracted with toluene $(3 \times 50 \text{ mL})$, and the combined organic layers were collected, dried with MgSO₄, and filtered. The toluene solution was concentrated under reduced pressure, and the title compound 6 was obtained as a pale yellow solid; yield 20.04 g (99%); m.p. 160 °C (dec.). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.63 (d, ${}^{3}J_{H,H}$ = 8 Hz, 2 H), 7.54 (d, ${}^{3}J_{H,H}$ = 8 Hz, 2 H), 5.80 (s, 1 H), 4.10–3.95 (m, 4 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): δ = 142.9, 132.0, 127.0, 118.4, 112.7, 102.2, 65.3 ppm. IR (KBr): \tilde{v}_{max} = 2230, 1085, 942 cm⁻¹. MS (EI+): m/z = 175 [M]⁺⁻, 144, 130, 103, 73, 51. C₁₀H₉NO₂ (175.19): calcd. C 68.56, H 5.18, N 8.00; found C 68.08, H 4.88, N 7.59.

4-(1,3-Dioxolan-2-yl)benzylamine (7): A solution of nitrile 6 (10.6 g, 60.5 mmol) in anhydrous THF (40 mL) was added dropwise at 0 °C to a suspension of LiAlH₄ (5.1 g, 127.7 mmol, 2 equiv.) in anhydrous THF (200 mL). The reaction mixture was allowed to warm to room temp., heated at reflux for 4 h, and stirred at room temp. for 12 h. The mixture was then recooled in an ice bath, and diethyl ether was added (200 mL). The mixture was then carefully quenched with water (5 mL), aqueous NaOH solution (1 N, 5 mL), and a second portion of water (10 mL). After 30 min, the white suspension was filtered through a pad of Celite, and concentrated in vacuo to give amine 7 as a yellow oil; yield 10.3 g (95%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.42 (d, ³J_{H,H} = 8 Hz, 2 H), 7.30 (d, ${}^{3}J_{H,H} = 8$ Hz, 2 H), 5.77 (s, 1 H), 4.10–4.05 (m, 2 H), 4.05–3.95 (m, 2 H), 3.84 (s, 2 H), 1.58 (br. s, 2 H) ppm. $^{13}\mathrm{C}$ NMR (CDCl_3, 75.4 MHz): δ = 144.3, 136.3, 127.0, 126.6, 103.6, 65.2, 46.2 ppm. IR (KBr): $\tilde{v}_{max} = 3667$, 1616, 1081, 943 cm⁻¹. C₁₀H₁₃NO₂ (179.22): calcd. C 67.02, H 7.31, N 7.82; found C 66.84, H 7.71, N 7.59.

Benzyl [4-(1,3-Dioxolan-2-yl)benzyl]carbamate (8): A solution of sodium carbonate (19.6 g, 185 mmol, 4 equiv.) in water (60 mL) was added to a solution of amine 7 (8.14 g, 45.4 mmol) in CH₂Cl₂ (140 mL). Benzyl chloroformate (6.5 mL, 45.7 mmol, 1 equiv.) was added dropwise at 0 °C and with stirring. After having been stirred at room temp. for 12 h, the reaction mixture was extracted with CH₂Cl₂ (2×150 mL). The combined organic layers were further washed with water (100 mL), saturated aqueous NH₄Cl solution (50 mL), and brine (50 mL), and dried with anhydrous MgSO₄. After concentration, **8** was obtained as a pale yellow solid; yield 13.8 g (97%); m.p. 145 °C; TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.25$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.39$ (d, ³ $J_{H,H}$ = 14.5 Hz, 2 H), 7.35–7.30 (m, 5 H), 7.29 (d, ³ $J_{H,H} = 14.5$ Hz, 2 H), 5.78 (s, 1 H), 5.11 (s, 3 H), 4.37 (d, ³ $J_{H,H} = 6$ Hz, 2 H), 4.15– 3.95 (2×m, 4 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): $\delta = 156.4$, 139.4, 137.2, 136.4, 128.5, 128.1, 127.5, 126.7, 103.4, 66.9, 65.3, 44.8 ppm. IR (KBr): $\tilde{\nu}_{max}$ = 3280, 1681, 1554, 1260, 1077, 941 cm^{-1}. C_{18}H_{19}NO_4 (313.36): calcd. C 69.00, H 6.11, N 4.47; found C 69.14, H 6.29, N 4.41.

Benzyl (4-Formylbenzyl)carbamate (4c): A solution of acetal **8** (13.9 g, 43.9 mmol) in aqueous AcOH (50%, 140 mL) was stirred at room temp. for 12 h. After concentration, flash column chromatography [cyclohexane/EtOAc (9:1 to 6:4) as eluent] gave aldehyde **4c** as a colorless solid; yield 11.0 g (93%); m.p. 68 °C; TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.49$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 9.97$ (s, 1 H), 7.82 (d, ³J_{H,H} = 8 Hz, 2 H), 7.42 (d, ³J_{H,H} = 8 Hz, 2 H), 7.35–7.30 (m, 5 H), 5.23 (br. s, 1 H), 5.13 (s, 2 H), 4.44 (d, ³J_{H,H} = 6 Hz, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): $\delta = 191.8$, 156.4, 145.4, 136.2, 135.6, 130.1, 128.6, 128.3, 128.2, 127.5, 67.1, 44.8 ppm. IR (KBr): $\tilde{v}_{max} = 3304$, 1682, 1609, 1529, 1253, 1207, 1052 cm⁻¹. C₁₆H₁₅NO₃ (269.30): calcd. C 71.36, H 5.61, N 5.20; found C 71.09, H 5.72, N 5.14.

General Procedure for 10b-e. Benzyl 4-[3,3-Difluoro-1-(4-methoxybenzyl)-4-oxoazetidin-2-yl|piperidine-1-carboxylate (10b): A dry, 60-mL Schlenk tube was charged with a suspension of freshly acidwashed zinc dust (4.75 g, 72.7 mmol, 6 equiv.) in anhydrous THF (19 mL) under argon. Chlorotrimethylsilane (460 μ L, 5 mol-%) and 1,2-dibromoethane (315 µL, 5 mol-%) were added to the suspension. The mixture was stirred at room temp. for 10 min. Controlled addition of a solution of ethyl bromodifluoroacetate (5 g, 24.6 mmol, 2.05 equiv.) in anhydrous THF (5 mL) was performed with a syringe. A temperature of ca. 50 °C was maintained during the addition (self-heating). After the end of the addition, the reaction mixture was stirred at room temp. for 10 min. A solution of the corresponding imine 9b (12.1 mmol) in anhydrous THF (8 mL) was added. The reaction mixture was then heated to reflux for 2 h. The reaction mixture was cooled to room temp. and guenched by addition of saturated aqueous NH₄Cl solution (10 mL). After filtration, the aqueous layer was extracted with EtOAc (2×100 mL). The organic layers were combined, dried with MgSO₄, and concentrated under vacuum. The crude product was then purified by flash chromatography on silica gel [cyclohexane/EtOAc (95:5 to 70:30) as eluent] to afford β -lactam 10b as a yellow oil; yield 3.76 g (70%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.35–7.30 (m, 5 H), 7.10 (d, ${}^{3}J_{H,H}$ = 8.5 Hz, 2 H), 6.87 (d, ${}^{3}J_{H,H}$ = 8.5 Hz, 2 H), 5.09 (s, 2 H), 4.88 (d, ${}^{2}J_{H,H}$ = 15 Hz, 1 H), 4.30–4.05 (m, 2 H), 4.03 (dd, J = 2.5, ${}^{2}J_{\text{H,H}}$ = 15 Hz, 1 H), 3.78 (s, 3 H), 3.55–3.45 (m, 1 H), 2.80–2.60 (m, 2 H), 1.90–1.80 (m, 1 H), 1.70–1.60 (m, 2 H), 1.25–1.10 (m, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): $\delta = 161.4$ (t, ² $J_{C,F} =$ 30.5 Hz), 159.6, 155.0, 136.5, 129.5, 128.5, 128.0, 127.9, 125.3, 120.8 (dd, ${}^{1}J_{C,F}$ = 285.5, ${}^{1}J_{C,F}$ = 291.5 Hz), 114.5, 68.1 (dd, ${}^{2}J_{C,F}$ = 22, ${}^{2}J_{C,F}$ = 24 Hz), 67.1, 55.2, 45.7, 43.3, 36.0, 28.2 ppm. ${}^{19}F$ NMR (CDCl₃, 288.3 MHz): δ = -114.5 (d, ²J_{F,F} = 236.5 Hz), -123.6 (dd, ${}^{3}J_{\rm EH} = 73$, ${}^{2}J_{\rm EF} = 236.5$ Hz) ppm. IR (KBr): $\tilde{v}_{\rm max} =$ 1787, 1698, 1514, 1434, 1301, 1249, 1225, 1061, 699 cm⁻¹. C₂₄H₂₆F₂N₂O₄ (444.48): calcd. C 64.85, H 5.90, N 6.30; found C 64.99, H 6.03, N 6.09.

Benzyl {4-[3,3-Difluoro-1-(4-methoxybenzyl)-4-oxoazetidin-2-yl]benzyl}carbamate (10c) and Benzyl 4-{2-(Ethoxycarbonyl)-2,2-difluoro-1-[(4-methoxybenzyl)amino]ethyl}benzylcarbamate (11c): These compounds were prepared according to the above General Procedure, from aldehyde 4c (1.37 g, 5.1 mmol), (*p*-methoxybenzyl)amine (0.70 g, 5.1 mmol), activated zinc dust (2 g, 30.6 mmol), ethyl bromodifluoroacetate (2.12 g, 10.4 mmol), and anhydrous acetonitrile (16 mL). The crude product was purified by silica gel column chromatography [cyclohexane/EtOAc (85:15 to 70:30) as eluent] to give the β-lactam 10c as a colorless oil and the



 β -amino ester **11c** as a white solid; yield 0.89 g and 0.90 g (72% global yield).

β-Lactam 10c: TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.35$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.35-7.25$ (m, 7 H), 7.17 (d, ³ $J_{\text{H,H}} = 8$ Hz, 2 H), 7.00 (d, ³ $J_{\text{H,H}} = 8.5$ Hz, 2 H), 6.81 (d, ³ $J_{\text{H,H}} = 8.5$ Hz, 2 H), 5.22 (br. s, 1 H), 5.13 (s, 2 H), 4.86 (d, ² $J_{\text{H,H}} = 14$ Hz, 1 H), 4.65 (dd, ³ $J_{\text{H,F}} = 1.5$, ³ $J_{\text{H,F}} = 7$ Hz, 1 H), 4.39 (m, 2 H), 3.79 (d, ² $J_{\text{H,H}} = 14$ Hz, 1 H), 3.77 (s, 3 H) ppm. ¹³C (CDCl₃, 75.4 MHz): $\delta = 160.7$ (t, ² $J_{\text{C,F}} = 30.5$ Hz), 159.6, 156.7, 140.5, 137.1, 136.4, 130.9, 129.5, 129.2, 128.5, 128.2, 128.1, 127.7, 120.4 (t, ¹ $J_{\text{C,F}} = 292$ Hz), 114.4, 67.4 (dd, ² $J_{\text{C,F}} = 24$, ² $J_{\text{C,F}} = 26.5$ Hz), 66.9, 55.3, 44.6, 43.6 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -114.8$ (dd, ³ $J_{\text{F,H}} = 7.5$, ² $J_{\text{F,F}} = 223$ Hz), -121.9 (d, ² $J_{\text{F,F}} = 223$ Hz) ppm. IR (KBr): $\tilde{v}_{\text{max}} = 3340$, 1787, 1718, 1612, 1514, 1302, 1249, 1201, 1034 cm⁻¹. C₂₆H₂₄F₂N₂O₄ (466.49): calcd. C 66.94, H 5.19, N 6.01; found C 66.88, H 5.45, N 5.91.

β-Amino Ester 11c: M.p. 106 °C; TLC: silica gel (cyclohexane/ EtOAc, 7:3), $R_f = 0.45$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.35$ -7.25 (m, 9 H), 7.09 (d, ${}^{3}J_{\rm H,\rm H} = 7$ Hz, 2 H), 6.81 (d, ${}^{3}J_{\rm H,\rm H} = 7$ Hz, 2 H), 5.13 (br. s, 2 H), 4.40 (d, ${}^{3}J_{\rm H,\rm H} = 6$ Hz, 2 H), 4.25 (q, ${}^{3}J_{\rm H,\rm H} =$ 7 Hz, 2 H), 4.18 (dd, ${}^{3}J_{\rm H,\rm F} = 6.5$, ${}^{3}J_{\rm H,\rm H} = 13$ Hz, 1 H), 3.77 (s, 3 H), 3.69 (d, ${}^{2}J_{\rm H,\rm H} = 13$ Hz, 1 H), 3.42 (d, ${}^{2}J_{\rm H,\rm H} = 13$ Hz, 1 H), 2.07 (br. s, 1 H), 1.26 (t, ${}^{3}J_{\rm H,\rm H} = 7$ Hz, 3 H) ppm. 13 C NMR (CDCl₃, 75.4 MHz): $\delta = 163.9$ (dd, ${}^{2}J_{\rm C,\rm F} = 30.5$, ${}^{2}J_{\rm C,\rm F} = 34$ Hz), 158.8, 156.4, 139.0, 136.4, 133.4, 130.9, 129.5, 129.2, 128.5, 128.2, 128.1, 127.7, 115.1 (dd, ${}^{1}J_{\rm C,\rm F} = 253.5$, ${}^{1}J_{\rm C,\rm F} = 257$ Hz), 113.7, 66.9, 62.8, 62.5 (dd, ${}^{2}J_{\rm C,\rm F} = 21$, ${}^{2}J_{\rm C,\rm F} = 27$ Hz), 55.2, 49.9, 44.7, 13.9 ppm. 19 F NMR (CDCl₃, 288.3 MHz): $\delta = -108.6$ (dd, ${}^{3}J_{\rm F,\rm H} = 7.5$, ${}^{2}J_{\rm F,\rm F} =$ 257 Hz), -121.0 (dd, ${}^{3}J_{\rm F,\rm H} = 21.5$, ${}^{2}J_{\rm F,\rm F} = 223$ Hz) ppm. IR (KBr): $\tilde{v}_{\rm max} = 3334$ 1769, 1703, 1612, 1303, 1250, 1210, 1072, 699 cm⁻¹. C₂₈H₃₀F₂N₂O₅ (512.56): calcd. C 65.61, H 5.90, N 5.47; found C 65.13, H 5.55, N 5.38.

3,3-Difluoro-1-(4-methoxybenzyl)-4-phenylazetidin-2-one (10a): Preparation, description, and spectroscopic data for this compound have already been reported in ref.^[38a]

3,3-Difluoro-1-(4-methoxybenzyl)-4-(pyridin-4-yl)azetidin-2-one (10d) and Ethyl 2,2-Difluoro-3-[(4-methoxybenzyl)amino]-3-(pyridin-4-yl)propanoate (11d): Preparation, description, and spectroscopic data for these compounds have already been reported in ref.^[38b]

3,3-Difluoro-1-(4-methoxybenzyl)-4-(pyridin-3-yl)azetidin-2-one (10e) and Ethyl 2,2-Difluoro-3-(4-methoxybenzylamino)-3-(pyridin-3-yl)propanoate (11e): Preparation, description and spectroscopic data of these compounds have already been reported in ref.^[38b]

General Procedure for 12b-e. rac-Benzyl 4-(3,3-Difluoro-4-oxoazetidin-2-yl)piperidine-1-carboxylate (12b): CAN (7 g, 12.7 mmol, 3.6 equiv.) was added in small portions at 0 °C to N-protected βlactam 10b (1.56 g, 3.5 mmol) in a mixture of CH₃CN/H₂O (9:1, 22 mL). After 20 min at 0 °C and 6 h at room temp., the mixture was poured into water (100 mL). The aqueous layer was extracted with EtOAc $(2 \times 100 \text{ mL})$. The combined organic layers were washed with NaHCO3 (5%, 100 mL), Na2SO3 (10%, 100 mL), NaHCO₃ (5%, 100 mL), and finally brine (50 mL). After concentration and flash column chromatography [CH₂Cl₂/EtOAc (95:5 to 80:20) as eluent] the desired racemic product 12b was obtained as an orange oil; yield 0.43 g (38%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.40–7.30 (m, 5 H), 5.10 (s, 2 H), 5.02 (br. s, 1 H), 4.30–4.10 (m, 2 H), 3.62 (t, ${}^{3}J_{H,H} = 9$, ${}^{3}J_{H,F} = 9$ Hz, 1 H), 2.90–2.70 (m, 2 H), 1.70-1.50 (m, 3 H), 1.30-1.10 (m, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 161.1 (t, ² $J_{C,F}$ = 31 Hz), 155.3, 136.4, 128.5, 128.1,

127.9, 121.5 (dd, ${}^{1}J_{C,F} = 287.5$, ${}^{1}J_{C,F} = 291$ Hz), 67.4, 66.2 (dd, ${}^{2}J_{C,F} = 23$, ${}^{2}J_{C,F} = 24$ Hz), 43.3 and 43.1, 35.9, 27.9 and 27.6 ppm. 19 F NMR (CDCl₃, 288.3 MHz): $\delta = -114.2$ (d, ${}^{2}J_{F,F} = 239$ Hz), -125.4 (dd, ${}^{3}J_{F,H} = 40$, ${}^{2}J_{F,F} = 239$ Hz) ppm.

rac-Benzyl 4-(3,3-Difluoro-4-oxoazetidin-2-yl)benzylcarbamate (12c): This compound was prepared from 10c according to the same procedure as employed for 12b, from β-lactam 10c (4.3 g, 9.2 mmol) and CAN (15.2 g, 27.7 mmol, 3 equiv.) in a mixture of CH₃CN/H₂O (9:1, 170 mL). Chromatography [CH₂Cl₂/EtOAc (95:5 to 85:15) as eluent] gave pure racemic 12c as a white solid; yield 1.37 g (43%); TLC: silica gel (CH₂Cl₂/EtOAc, 9:1), *R_f*= 0.52. ¹H NMR (CD₃OD, 300.3 MHz): δ = 7.40–7.25 (m, 9 H), 5.10 (dd, ³J_{H,F} = 2.5, ³J_{H,F} = 7.5 Hz, 1 H), 5.09 (s, 2 H), 4.32 (s, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 163.4 (t, ²J_{C,F} = 30 Hz), 159.0, 141.8, 138.2, 133.0, 129.4, 129.0, 128.8, 128.6, 128.2, 122.8 (t, ¹J_{C,F} = 292 Hz), 67.5, 65.5 (dd, ²J_{C,F} = 24, ²J_{C,F} = 26 Hz), 45.1 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -116.6 (dd, ³J_{F,H} = 7.5, ²J_{F,F} = 222.5 Hz), -122.9 (d, ²J_{F,F} = 222.5 Hz) ppm.

rac-3,3-Difluoro-4-(pyridin-4-yl)azetidin-2-one (12d): This compound was prepared from the mixture of 10d and 11d according to the same procedure as employed for 12b, from β -lactam 10d (1.2 g, 3.95 mmol) and CAN (6.5 g, 11.85 mmol, 3 equiv.) in a mixture of CH₃CN/H₂O (9:1, 55 mL). Chromatography [CH₂Cl₂/MeOH/ NH₄OH (95:5:0.5 to 80:20:2) as eluent] gave pure racemic 12d as a pale beige solid; yield 0.45 g (62%); m.p. 160 °C (dec.); TLC: silica gel (CH₂Cl₂/MeOH/NH₄OH, 9:1:0.1), $R_f = 0.44$. ¹H NMR $(CDCl_3, 300.3 \text{ MHz}): \delta = 10.32 \text{ (t, } J = 12.5 \text{ Hz}, 1 \text{ H}), 8.98 \text{ (d, }^3J_{\text{H H}})$ = 6.5 Hz, 2 H), 8.01 (d, ${}^{3}J_{H,H}$ = 6.5 Hz, 2 H), 5.69 (dd, ${}^{3}J_{H,F}$ = 1.5, ${}^{3}J_{\text{H,F}} = 7 \text{ Hz}$, 1 H) ppm. ${}^{13}\text{C}$ NMR (CDCl₃, 75.4 MHz): $\delta =$ 160.3 (t, ${}^{2}J_{C,F}$ = 29.5 Hz), 151.0, 144.3, 124.9, 121.8 (t, ${}^{1}J_{C,F}$ = 294.5 Hz), 62.5 (dd, ${}^{2}J_{C,F}$ = 22.5, ${}^{2}J_{C,F}$ = 26 Hz) ppm. ${}^{19}F$ NMR (CDCl₃, 288.3 MHz): $\delta = -112.7$ (ddd, ${}^{3}J_{F,H} = 7$, ${}^{4}J_{F,H} = 13.5$, ${}^{2}J_{F,F}$ = 219.5 Hz), -118.7 (dd, ${}^{4}J_{F,H}$ = 11.5, ${}^{2}J_{F,F}$ = 219.5 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3079, 1813, 1384, 718 cm⁻¹. MS (CI+): m/z = 185 [M + H]⁺, 173, 159, 145, 93.

rac-3,3-Difluoro-4-(pyridin-3-yl)azetidin-2-one (12e): This compound was prepared from the mixture of 10e and 11e according to the same procedure as employed for 12b, from β -lactam 10e (12.6 g, 41.4 mmol) and CAN (68 g, 124 mmol, 3 equiv.) in a mixture of CH₃CN/H₂O (9:1, 550 mL). Chromatography [CH₂Cl₂/MeOH/ NH₄OH (95:5:0.5 to 80:20:2) as eluent] gave pure racemic 12e as a brown viscous oil; yield 3.74 g (49%); TLC: silica gel (CH₂Cl₂/ MeOH/NH₄OH, 95:5:0.5), $R_f = 0.25$. ¹H NMR (CD₃OD, 300.3 MHz): δ = 8.70–8.65 (m, 2 H), 8.02 (d, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 7.66 (dd, ${}^{3}J_{H,H} = 5$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 5.35 (dd, ${}^{3}J_{H,F} = 2.5$, ${}^{3}J_{H,F}$ = 7 Hz, 1 H) ppm. ¹H NMR ([D₆]DMSO, 300.3 MHz): δ = 10.04 (t, ${}^{4}J_{H,F}$ = 12 Hz, 1 H), 8.65–8.60 (m, 2 H), 7.86 (d, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 7.53 (dd, ${}^{3}J_{H,H}$ = 5, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 5.42 (dd, ${}^{3}J_{H,F}$ = 2.5, ${}^{3}J_{\rm H,F}$ = 7 Hz, 1 H) ppm. 13 C NMR (CD₃OD, 75.4 MHz): δ = 162.8 (t, ${}^{2}J_{C,F}$ = 30 Hz), 150.1, 148.3, 138.1, 131.8, 125.8, 125.6 (t, ${}^{1}J_{C,F}$ = 294.5 Hz), 63.3 (dd, ${}^{2}J_{C,F}$ = 23.5, ${}^{2}J_{C,F}$ = 26.5 Hz) ppm. ¹⁹F NMR (CD₃OD, 288.3 MHz): δ = -116.4 (dd, ³J_{F,H} = 6.5, ²J_{F,F} = 228 Hz), -122.5 (d, ${}^{2}J_{F,F}$ = 228 Hz) ppm. ${}^{19}F$ NMR ([D₆]DMSO, 288.3 MHz): $\delta = -114.2$ (ddd, ${}^{3}J_{F,H} = 7.5$, ${}^{4}J_{F,H} = 14$, ${}^{2}J_{F,F} =$ 223 Hz), -120.3 (ddd, ${}^{3}J_{\rm F,H}$ = 2, ${}^{4}J_{\rm F,H}$ = 12, ${}^{2}J_{\rm F,F}$ = 223 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3079, 1813, 1384, 718 cm⁻¹. MS (CI+): *m*/*z* = 185 $[M + H]^+$, 165, 142, 100, 91.

General Procedure for 14b–e. *rac*-Benzyl 4-[1-(Dibenzylamino)-2-(ethoxycarbonyl)-2,2-difluoroethyl]piperidine-1-carboxylate (14b): The *N*,*N*-disubstituted 1*H*-benzotriazolylamine 13b was prepared according to the above General Procedure; dibenzylamine (0.79 g, 4 mmol) and aldehyde 4b (1 g, 4.05 mmol, 1 equiv.) were successively added to a solution of 1H-benzotriazole (0.48 g, 4 mmol, 1 equiv.) dissolved in dry ethanol (1 mL). The mixture was heated at reflux for 4 h in the presence of dried molecular sieves (3 Å, 0.5 g) and was then stirred at room temp. for 12 h. After removal of molecular sieves by suction filtration and concentration of the filtrate under vacuum, the obtained solid could be recrystallized from ethanol. Chlorotrimethylsilane (1.1 mL, 8.6 mmol, 2.2 equiv.) was added to a suspension of activated zinc dust (0.79 g, 12 mmol, 3 equiv.) in anhydrous THF (10 mL), stirred in a dry Schlenk tube under argon, followed 10 min later by ethyl bromodifluoroacetate (1.83 g, 9 mmol, 2.2 equiv.). After 10 min, a solution of 13b (4 mmol) in anhydrous THF (4 mL) was added dropwise. After 3 h at 60 °C, the mixture was cooled, poured into saturated aqueous NaHCO₃ solution (20 mL), and then filtered through Celite. The layers were separated, and the aqueous phase was extracted with EtOAc (2×60 mL). The organic layers were combined and washed with brine (40 mL), and were then dried with MgSO₄. After filtration and evaporation of the solvent, the crude product was chromatographed [cyclohexane/EtOAc (95:5 to 80:20) as eluent] to give pure ester 14b as a yellow oil; yield 1.88 g (83%; 2 steps). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.25–7.10 (m, 15 H), 4.96 (br. s, 2 H), 4.11 (q, ${}^{3}J_{H,H}$ = 7 Hz, 2 H), 4.10–3.90 (m, 2 H), 3.79 (d, ${}^{2}J_{H,H}$ = 13.5 Hz, 2 H), 3.72 (d, ${}^{2}J_{H,H}$ = 13.5 Hz, 2 H), 3.25–3.05 (m, 1 H), 2.65-2.50 (m, 2 H), 1.95-1.85 (m, 1 H), 1.85-1.70 (m, 1 H), 1.50-1.40 (m, 1 H), 1.17 (t, ${}^{3}J_{H,H} = 7$ Hz, 3 H), 1.10–1.00 (m, 1 H), 0.90–0.85 (m, 1 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 164.3 (t, ${}^{2}J_{C,F}$ = 32 Hz), 155.0, 139.0, 136.8, 129.4, 128.4, 128.2, 127.9, 127.8, 127.2, 119.0 (t, ${}^{1}J_{C,F}$ = 260 Hz), 67.0, 62.9, 62.2 (t, ${}^{2}J_{C,F}$ = 20 Hz), 55.0, 44.2 and 43.8, 35.3, 30.4 and 28.8, 13.8 ppm. $^{19}\mathrm{F}$ NMR (CDCl₃, 288.3 MHz): δ = -102.1 (dd, ${}^{3}J_{F,H}$ = 7.5, ${}^{2}J_{F,F}$ = 259 Hz), -113.7 (dd, ${}^{3}J_{\text{F,H}} = 24.5$, ${}^{2}J_{\text{F,F}} = 259$ Hz) ppm. IR (KBr): $\tilde{v}_{max} = 1787, 1698, 1514, 1434, 1249, 1225, 1061, 699 \text{ cm}^{-1}.$ C32H36F2N2O4 (550.65): calcd. C 69.80, H 6.59, N 5.09; found C 69.22, H 6.71, N 4.97.

rac-Benzyl 4-[1-(Dibenzylamino)-2-(ethoxycarbonyl)-2,2-difluoroethyllbenzylcarbamate (14c): The N,N-disubstituted 1H-benzotriazolylamine 13c was prepared according to the same procedure as employed for 14b; dibenzylamine (5.13 g, 26 mmol) and aldehyde 4c (7 g, 26 mmol, 1 equiv.) were successively added to a solution of 1H-benzotriazole (3.1 g, 26 mmol, 1 equiv.) dissolved in dry ethanol (5 mL). The mixture was heated at reflux for 4 h in the presence of dried molecular sieves (3 Å, 2 g), and was then stirred at room temp. for 12 h. After removal of molecular sieves by suction filtration and concentration of the filtrate under vacuum, the obtained solid could be recrystallized from ethanol. Racemic β-amino ester 14c was obtained from N,N-disubstituted 1H-benzotriazolylamine 13c according to the procedure already described for the conversion of 13b into 14b, with activated zinc dust (6.3 g, 96.4 mmol, 4 equiv.), chlorotrimethylsilane (3.7 mL, 28.9 mmol, 1.2 equiv.), and ethyl bromodifluoroacetate (10.7 g, 52.9 mmol, 2.2 equiv.) in anhydrous THF (95 mL). Chromatography [cyclohexane/EtOAc (97:3 to 80:20) as eluent] gave pure ester 14c as a white solid; yield 8.93 g (60%; 2 steps); TLC: silica gel (cyclohexane/ EtOAc, 7:3), $R_f = 0.58$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.40$ – 7.10 (m, 15 H), 5.13 (s, 3 H), 4.41 (d, ${}^{3}J_{H,H} = 6$ Hz, 2 H), 4.31 (dd, ${}^{3}J_{H,F} = 8$, ${}^{3}J_{H,F} = 26$ Hz, 1 H), 4.30 (q, ${}^{3}J_{H,H} = 7$ Hz, 1 H), 4.01 $(d, {}^{2}J_{H,H} = 13.5 \text{ Hz}, 2 \text{ H}), 3.99 \text{ (q, }{}^{3}J_{H,H} = 7 \text{ Hz}, 1 \text{ H}), 3.06 \text{ (d,}$ ${}^{2}J_{H,H} = 13.5 \text{ Hz}, 2 \text{ H}), 1.12 \text{ (t, }{}^{3}J_{H,H} = 7 \text{ Hz}, 3 \text{ H}) \text{ ppm.}^{13}\text{C NMR}$ (CDCl₃, 75.4 MHz): δ = 163.9 (dd, ²J_{C,F} = 30, ²J_{C,F} = 34 Hz), 156.5, 139.0, 138.7, 136.4, 131.3, 129.1, 128.5, 128.3, 128.1, 127.3, 127.2, 117.1 (dd, ${}^{1}J_{C,F} = 253$, ${}^{1}J_{C,F} = 259.5$ Hz), 66.9, 63.3 (dd, ${}^{2}J_{C,F}$ = 19.5, ${}^{2}J_{C,F}$ = 28.5 Hz), 62.8, 54.9, 44.7, 13.6 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -102.0 (d, ²*J*_{F,F} = 262 Hz), -113.8 (d, ²*J*_{F,F} = 262 Hz) ppm.

rac-Ethyl 3-(Dibenzylamino)-2,2-difluoro-3-(pyridin-4-yl)propanoate (14d): The N,N-disubstituted 1H-benzotriazolylamine 13d was prepared according to the same procedure as employed for 14b; dibenzylamine (8.37 g, 42.4 mmol) and pyridine-4-carbaldehyde (4d, 5 g, 46.7 mmol, 1.1 equiv.) were successively added to a solution of 1Hbenzotriazole (5.06 g, 42.5 mmol, 1 equiv.) dissolved in dry ethanol (5 mL). The mixture was heated at reflux for 4 h in the presence of dried molecular sieves (3 Å, 4 g), and was then stirred at room temp. for 12 h. After removal of molecular sieves by suction filtration and concentration of the filtrate under vacuum, the obtained solid could be recrystallized from ethanol. Racemic β-amino ester 14d was obtained from N,N-disubstituted 1H-benzotriazolylamine 13d by the procedure already described for the conversion of 13b to 14b, from iminium salt 13d (9 g, 22.2 mmol), activated zinc dust (3 g, 45.9 mmol, 2 equiv.), chlorotrimethylsilane (3.5 mL, 27.4 mmol, 1.2 equiv.), and ethyl bromodifluoroacetate (6.75 g, 33.3 mmol, 1.5 equiv.) in anhydrous THF (7 mL). Chromatography [cyclohexane/EtOAc (95:5 to 70:30) as eluent] gave pure ester 14d as a pale yellow solid; yield 7.47 g (82%; 2 steps); m.p. 93 °C; TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.30$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 8.69 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 7.35–7.20 (m, 12 H), 4.45–4.30 (m, 2 H), 4.04 (d, ${}^{2}J_{H,H}$ = 13.5 Hz, 2 H), 3.10 (d, ${}^{2}J_{H,H}$ = 13.5 Hz, 2 H), 1.18 (t, ${}^{3}J_{H,H}$ = 7 Hz, 3 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): δ = 163.3 (dd, ${}^{2}J_{C,F}$ = 29.5, ${}^{2}J_{C,F}$ = 34 Hz), 149.6, 139.5, 138.0, 129.1, 128.4, 127.5, 125.7, 116.6 (dd, ${}^{1}J_{C,F} = 255$, ${}^{1}J_{C,F}$ = 261 Hz), 63.1, 62.7 (dd, ${}^{2}J_{C,F}$ = 19.5, ${}^{2}J_{C,F}$ = 28.5 Hz), 54.9, 13.6 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -102.1$ (dd, ³ $J_{\rm EH} =$ 7.5, ${}^{2}J_{\text{F,F}}$ = 259 Hz), -113.7 (dd, ${}^{3}J_{\text{F,H}}$ = 24.5, ${}^{2}J_{\text{F,F}}$ = 259 Hz) ppm. IR (KBr): \tilde{v}_{max} = 1770, 1293, 1204, 1066, 700 cm⁻¹. C₂₄H₂₄F₂N₂O₂ (410.47): calcd. C 70.23, H 5.89, N 6.82; found C 69.91, H 6.02, N 6.46.

rac-Ethyl 3-(Dibenzylamino)-2,2-difluoro-3-(pyridin-3-yl)propanoate (15e): The N,N-disubstituted 1H-benzotriazolylamine 13e was prepared according to the same procedure as employed for 14b; dibenzylamine (16.74 g, 84.9 mmol) and pyridine-3-carbaldehyde (4e, 10 g, 93.4 mmol, 1.1 equiv.) were successively added to a solution of 1H-benzotriazole (10.11 g, 84.9 mmol, 1 equiv.) dissolved in dry ethanol (15 mL). The mixture was heated at reflux for 4 h in the presence of dried molecular sieves (3 Å, 6 g), and then stirred at room temp. for 12 h. After removal of molecular sieves by suction filtration and concentration of the filtrate under vacuum, the obtained solid could be recrystallized from ethanol. Racemic β-amino ester 14e was obtained from N,N-disubstituted 1H-benzotriazolylamine 13e according to the procedure already described for the conversion of 13b into 14b, from iminium salt 13e (9 g, 22.2 mmol), activated zinc dust (3 g, 45.9 mmol, 2 equiv.), chlorotrimethylsilane (3.5 mL, 27.4 mmol, 1.2 equiv.), and ethyl bromodifluoroacetate (6.75 g, 33.3 mmol, 1.5 equiv.) in anhydrous THF (7 mL). Chromatography [cyclohexane/EtOAc (9:1 to 7:3) as eluent] gave pure ester 14e as a yellow oil; yield 7.35 g (81%); TLC: silica gel (cyclohexane/EtOAc, 7:3), $R_f = 0.42$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 8.62 (dd, ${}^{4}J_{H,H}$ = 1.5, ${}^{3}J_{H,H}$ = 5 Hz, 1 H), 8.55 (d, ${}^{4}J_{H,H} = 2$ Hz, 1 H), 7.81 (dt, ${}^{4}J_{H,H} = 1.5$, ${}^{3}J_{H,H} = 7.5$ Hz, 1 H), 7.35 (dd, ${}^{3}J_{H,H} = 5$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 7.30–7.20 (m, 10 H), 4.37 (dd, ${}^{3}J_{H,F} = 8$, ${}^{3}J_{H,F} = 9$ Hz, 1 H), 4.30 (q, ${}^{3}J_{H,H} = 7$ Hz, 2 H), 4.00 (d, ${}^{2}J_{H,H}$ = 13 Hz, 2 H), 3.03 (d, ${}^{2}J_{H,H}$ = 13 Hz, 2 H), 1.26 (t, ${}^{3}J_{\rm H,H}$ = 7 Hz, 3 H) ppm. 13 C NMR (CDCl₃, 75.4 MHz): δ = 163.5 (dd, ${}^{2}J_{C,F} = 30$, ${}^{2}J_{C,F} = 34$ Hz), 151.9, 150.0, 138.1, 137.9, 129.2, 128.4, 127.5, 126.3, 123.4, 120.3 (t, ${}^{1}J_{C,F}$ = 290 Hz), 63.0, 61.6 (dd, ${}^{2}J_{C,F}$ = 19.5, ${}^{2}J_{C,F}$ = 29.5 Hz), 54.9, 13.7 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -101.5$ (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} = 257$ Hz), -113.6



(dd, ${}^{3}J_{F,H} = 25$, ${}^{2}J_{F,F} = 257$ Hz) ppm. IR (KBr): $\hat{v}_{max} = 1770$, 1292, 1204, 1064, 700 cm⁻¹. C₂₄H₂₄F₂N₂O₂ (410.47): calcd. C 70.23, H 5.89, N 6.82; found C 69.95, H 5.83, N 6.75.

rac-tert-Butyl 4-[1-(Dibenzylamino)-2-(ethoxycarbonyl)-2,2-difluoroethyllpiperidine-1-carboxylate (14f): A solution of racemic β-amino ester 14b (10 g, 17.8 mmol) and Boc₂O (4.3 g, 19.7 mmol, 1.1 equiv.) in EtOH (80 mL) was added to a suspension of palladium on charcoal 10% (1 g) in EtOH (10 mL). The mixture was hydrogenated at atmospheric pressure and 40 °C until no starting material could be observed by TLC (ca. 48 h), filtered through Celite, and concentrated. The crude oil was crystallized from diethyl ether and, after filtration and drying, the pure β -amino ester 14f was obtained as a white solid; yield 9.19 g (quant.); m.p. 124 °C; TLC: silica gel (cyclohexane/EtOAc, 8:2), $R_f = 0.55$. ¹H NMR $(CDCl_3, 300.3 \text{ MHz}): \delta = 7.30-7.15 \text{ (m, 10 H)}, 4.25-4.05 \text{ (m, 2 H)},$ 4.05–3.90 (m, 2 H), 3.80 (d, ${}^{2}J_{H,H}$ = 12.5 Hz, 1 H), 3.73 (d, ${}^{2}J_{H,H}$ = 12.5 Hz, 1 H), 3.20 (td, ${}^{3}J_{H,H}$ = 7.5, ${}^{3}J_{H,F}$ = 7.5, ${}^{3}J_{H,F}$ = 23 Hz, 1 H), 2.60–2.45 (m, 2 H), 2.00–1.85 (m, 1 H), 1.85–1.70 (m, 1 H), 1.50–1.40 (m, 1 H), 1.24 (t, ${}^{3}J_{H,H} = 7$ Hz, 3 H), 1.15–1.00 (m, 1 H), 1.00–0.85 (m, 1 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 164.4 (t, ${}^{2}J_{C,F}$ = 32 Hz), 154.6, 139.0, 129.4, 128.2, 127.2, 119.0 (t, ${}^{1}J_{C,F}$ = 260 Hz), 79.4, 62.9, 62.3 (t, ${}^{2}J_{C,F}$ = 20 Hz), 55.1, 43.8, 35.4, 30.5 and 28.9, 28.4, 13.8 ppm. $^{19}\mathrm{F}$ NMR (CDCl_3, 288.3 MHz): δ = -106.1 (d, ${}^{2}J_{\text{EF}}$ = 250 Hz), -107.2 (dd, ${}^{3}J_{\text{EH}}$ = 11.5, ${}^{2}J_{\text{EF}}$ = 250 Hz) ppm. IR (KBr): \tilde{v}_{max} = 1773, 1695, 1295, 1210, 1176, 1063, 700 cm^{-1} .

rac-tert-Butyl 4-[1-Amino-2,2-difluoro-2-(methoxycarbonyl)ethyl]piperidine-1-carboxylate (15b): A solution of dibenzylamine 14f (7.15 g, 13.8 mmol) was added to a suspension of Pearlman's catalyst (palladium hydroxide 20% on activated charcoal, 0.5 g) in ethanol (80 mL). The reaction mixture was stirred at 40 °C under hydrogen (1 bar) for 48 h. The catalyst was filtered off and washed with ethanol $(2 \times 20 \text{ mL})$. The combined organic layers were concentrated, and the residue was chromatographed on silica gel [cyclohexane/EtOAc + 0.5% Et₃N (95:5 to 70:30) as eluent] to give 15b as a yellow oil; yield 0.76 g (17%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 4.25–4.00 (m, 2 H), 3.87 (s, 3 H), 3.08 (ddd, ${}^{3}J_{H,H}$ = 4, ${}^{3}J_{H,F}$ = 10.5, ${}^{3}J_{H,F}$ = 18 Hz, 1 H), 2.75–2.55 (m, 2 H), 1.80– 1.65 (m, 1 H), 1.42 (s, 9 H), 1.60–1.20 (m, 4 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 164.7 (d, ²J_{C,F} = 32, ²J_{C,F} = 33 Hz), 154.6, 116.7 (t, ${}^{1}J_{C,F}$ = 255 Hz), 79.4, 57.5 (t, ${}^{2}J_{C,F}$ = 23 Hz), 53.3, 43.7, 36.5, 28.4, 29.4 and 26.4 ppm. $^{19}\mathrm{F}$ NMR (CDCl₃, 288.3 MHz): δ = -111.2 (dd, J = 115, ${}^{2}J_{\text{EF}} = 263$ Hz), -117.0 (dd, ${}^{3}J_{\text{EH}} = 11$, ${}^{2}J_{\text{EF}}$ = 263 Hz) ppm.

rac-Ethyl 3-Amino-3-[4-(aminomethyl)phenyl]-2,2-difluoropropanoate Hydrochloride (16): A solution of N-protected β -amino ester 14c (10.94 g, 18.1 mmol) in EtOAc (30 mL) was added to a suspension of palladium on charcoal (10%, 0.5 g) in ethanol (30 mL), followed by the addition of HCl (5 N)/iPrOH (15 mL). The mixture was hydrogenated at atmospheric pressure and 40 °C until no starting material could be observed by TLC (ca. 3 d), filtered through a pad of Celite, washed with ethanol $(2 \times 25 \text{ mL})$, and concentrated. The hydrochloride salt of β -amino ester 16 was obtained as a hygroscopic white solid; yield 3.84 g (72%). ¹H NMR (CD₃OD, 300.3 MHz): δ = 7.60–7.40 (m, 4 H), 5.16 (dd, ${}^{3}J_{H,F}$ = 7.5, ${}^{3}J_{H,F}$ = 19 Hz, 1 H), 4.07 (q, ${}^{3}J_{H,H}$ = 7 Hz, 2 H), 4.03 (s, 2 H), 1.04 (t, ${}^{3}J_{\rm H,H}$ = 7 Hz, 3 H) ppm. 13 C NMR (CD₃OD, 75.4 MHz): δ = 161.8 (t, ${}^{2}J_{C,F} = 24$ Hz), 137.4, 131.1, 130.9, 114.2 (t, ${}^{1}J_{C,F} = 259$ Hz), 65.3, 57.3 (dd, ${}^{2}J_{C,F}$ = 22.5, ${}^{2}J_{C,F}$ = 24.5 Hz), 43.7, 14.1 ppm. ${}^{19}F$ NMR (CD₃OD, 288.3 MHz): $\delta = -108.7$ (dd, ${}^{3}J_{F,H} = 6.5$, ${}^{2}J_{F,F} =$ 260 Hz), -120.1 (dd, ${}^{3}J_{EH} = 18.5$, ${}^{2}J_{EF} = 260$ Hz) ppm. IR (KBr): $\tilde{v}_{max} = 3401, 1778, 1596-1519, 1315, 1200, 1128-1100, 1056 \text{ cm}^{-1}.$

4-[1-Amino-2-(ethoxycarbonyl)-2,2-difluoroethyl]benrac-Benzyl zylcarbamate (15c) and rac-Benzyl 4-[1-Amino-2,2-difluoro-2-(methoxycarbonyl)ethyl|benzylcarbamate (15c'): A solution of amine hydrochloride 17 (4 g, 13.6 mmol), triethylamine (4.15 mL, 30 mmol, 2.2 equiv.), Boc₂O (2.97 g, 13.6 mmol, 1 equiv.), and DMAP as catalyst (20 mg, 1 mol-%) was stirred at room temp. for 24 h. The mixture was poured into saturated aqueous NH₄Cl solution (50 mL) and extracted with CH_2Cl_2 (2×100 mL). The organic layer was dried with MgSO₄ and concentrated before purification by flash chromatography on silica gel [CH2Cl2/MeOH (98:2 to 96:4) as eluent] to afford the mixture of β -amino esters 15c and 15c' as a pale yellow oil; yield 0.91 g (19%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.35–7.15 (m, 8 H), 4.85 (br. s, 2 H), 4.43 (dd, ${}^{3}J_{H,F} = 11.5$, ${}^{3}J_{H,F}$ = 14.5 Hz, 2 H), 4.28 (d, ${}^{3}J_{H,H}$ = 6 Hz, 4 H), 4.23 (q, ${}^{3}J_{H,H}$ = 7 Hz, 2 H), 3.65 (s, 3 H), 1.76 (br. s, 2 H), 1.43 (br. s, 18 H), 1.22 (t, $^3\!J_{\rm H,H}$ = 7 Hz, 3 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 165.0 (t, ${}^{1}J_{C,F}$ = 33 Hz), 155.9, 139.6, 135.2, 128.2, 127.6, 79.6, 62.9, 58.1 (t, $^{2}J_{C,F}$ = 23.5 Hz), 53.3, 44.2, 28.5, 28.4, 13.9 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -114.4$ (dd, ${}^{3}J_{F,H} = 12$, ${}^{2}J_{F,F} = 261$ Hz), -117.8 (dd, ${}^{3}J_{F,H}$ = 16.5, ${}^{2}J_{F,F}$ = 261 Hz) ppm (ethyl β-amino ester); δ = -113.4 (dd, ${}^{3}J_{F,H}$ = 11 Hz, ${}^{2}J_{F,F}$ = 261 Hz), -117.3 (dd, ${}^{3}J_{F,H}$ = 16 Hz, ${}^{2}J_{F,F}$ = 261 Hz) ppm (methyl β-amino ester). IR (KBr): \tilde{v}_{max} = 3340, 1767, 1698, 1516, 1367, 1276, 1252, 1170, 1073, 758 cm⁻¹.

rac-Ethyl 3-Amino-2,2-difluoro-3-(pyridin-4-yl)propanoate Hydrochloride (15d): A solution of N-protected β-amino ester 14d (7.45 g, 18.1 mmol) in ethanol (30 mL) was added to a suspension of palladium on charcoal (10%, 1 g) in ethanol (15 mL), followed by the addition of HCl (5 N)/iPrOH (15 mL). The mixture was hydrogenated at atmospheric pressure and 40 °C until no starting material could be observed by TLC (ca. 3 d), filtered through a pad of Celite, washed with ethanol $(2 \times 25 \text{ mL})$, and concentrated. The hydrochloride salt of β-amino ester 16d was obtained as a hygroscopic white solid; yield 3.62 g (75%); m.p. 190 °C (dec.); TLC: silica gel $(CH_2Cl_2/MeOH/NH_4OH, 9:1:0.1), R_f = 0.70.$ ¹H NMR ([D₆]-DMSO, 300.3 MHz): δ = 8.91 (d, ${}^{3}J_{H,F}$ = 6 Hz, 2 H), 7.97 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 5.59 (dd, ${}^{3}J_{H,F}$ = 11.5, ${}^{3}J_{H,F}$ = 15 Hz, 1 H), 5.3–4.3 (br. s, 3 H), 4.28 (q, ${}^{3}J_{H,H} = 7$ Hz, 2 H), 1.18 (t, ${}^{3}J_{H,H} = 7$ Hz, 3 H) ppm. ¹³C NMR ([D₆]DMSO, 75.4 MHz): δ = 160.5 (t, ²J_{C,F} = 30.5 Hz), 146.6, 143.3, 126.0, 112.5 (t, ${}^{1}J_{C,F}$ = 258 Hz), 64.6, 54.3 $(t, {}^{2}J_{C,F} = 24.5 \text{ Hz}), 13.6 \text{ ppm}. {}^{19}\text{F} \text{ NMR} ([D_{6}]\text{DMSO}, 288.3 \text{ MHz}):$ $\delta = -109.5 \text{ (dd, } {}^{3}J_{\text{F,H}} = 11, \, {}^{2}J_{\text{F,F}} = 259 \text{ Hz}\text{)}, -112.5 \text{ (dd, } {}^{3}J_{\text{F,H}} = 15,$ ${}^{2}J_{\rm F,F}$ = 259 Hz) ppm. IR (KBr): $\tilde{v}_{\rm max}$ = 3399, 2790, 1778, 1640– 1616–1529, 1318, 1223, 1071 cm⁻¹.

rac-Ethyl 3-Amino-2,2-difluoro-3-(pyridin-4-yl)propanoate Hydrochloride (15e): This compound was prepared from N,N-dibenzyl β amino ester 14e according to the same procedure as employed for 15d, from N-protected β -amino ester 14e (2.9 g, 7.05 mmol), palladium on charcoal (10%, 0.17 g), HCl (5 N)/iPrOH (3 mL) in ethanol (10 mL). The hydrochloride salt of β-amino ester 15e was obtained as a white, viscous oil; yield 1.69 g (90%); TLC: silica gel $(CH_2Cl_2/MeOH/NH_4OH, 9:1:0.1), R_f = 0.70.$ ¹H NMR (CD₃OD, 300.3 MHz): $\delta = 9.14$ (d, ${}^{3}J_{H,H} = 6$ Hz, 1 H), 9.08 (br. s, 1 H), 8.75 (m, 1 H), 8.21 (m, 1 H), 5.80 (dd, ${}^{3}J_{H,F} = 8$, ${}^{3}J_{H,F} = 9$ Hz, 1 H), 4.42 (q, ${}^{3}J_{H,H}$ = 7 Hz, 2 H), 1.34 (t, ${}^{3}J_{H,H}$ = 7 Hz, 3 H) ppm. ${}^{13}C$ NMR (CD₃OD, 75.4 MHz): δ = 165.0 (t, ²J_{C,F} = 30.5 Hz), 147.3, 146.1, 145.6, 130.3, 128.5, 113.5 (dd, ${}^{1}J_{C,F}$ = 259.5, ${}^{1}J_{C,F}$ = 261.5 Hz), 65.9, 54.8 (t, ${}^{2}J_{C,F}$ = 24.5 Hz), 14.0 ppm. ${}^{19}F$ NMR (CD₃OD, 288.3 MHz): $\delta = -110.4$ (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} = 275$ Hz), -116.4 (dd, ${}^{3}J_{\rm F,H}$ = 16.5, ${}^{2}J_{\rm F,F}$ = 275 Hz) ppm. IR (KBr): $\tilde{v}_{\rm max}$ = 3401, 1769, 1668, 1318, 1210, 1072 cm⁻¹.

rac-Methyl 2-(3,3-Difluoro-2-oxo-4-phenylazetidin-1-yl)acetate (18a): A solution of azetidin-2-one 12a (2.3 g, 12.6 mmol), *n*Bu₄NI

(2.3 g, 6.3 mmol, 0.5 equiv.), and methyl bromoacetate (17a, 15 mL, 158 mmol, 12.5 equiv.) in anhydrous acetonitrile (17 mL) was treated dropwise with 1,1,3,3-tetramethylguanidine (2.9 g, 25.1 mmol, 2 equiv.). The mixture was stirred at room temp. for 12 h. The solvent was evaporated to dryness, and the residue was chromatographed on silica gel [cyclohexane/EtOAc (95:5 to 70:30) as eluent] to give β -lactam 18a as an opalescent oil; yield 3.21 g (quant.); TLC: silica gel (cyclohexane/EtOAc, 8:2), $R_f = 0.35$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.43 (m, 3 H), 7.28 (m, 2 H), 5.25 $(dd, {}^{3}J_{H,F} = 2.5, {}^{3}J_{H,F} = 7.5 \text{ Hz}, 1 \text{ H}), 4.49 (d, {}^{3}J_{H,H} = 18 \text{ Hz}, 1 \text{ H})$ H), 3.73 (s, 3 H), 3.61 (dd, J = 2, ${}^{2}J_{H,H} = 18$ Hz, 1 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): $\delta = 167.2$, 161.2 (t, ${}^{2}J_{CF} = 31$ Hz), 130.0, 129.6, 129.1, 128.0, 120.9 (t, ${}^{1}J_{C,F}$ = 291 Hz), 69.2 (dd, ${}^{2}J_{C,F}$ = 24, ${}^{2}J_{C,F}$ = 27 Hz), 52.7, 40.7 ppm. ${}^{19}F$ NMR (CDCl₃, 288.3 MHz): $\delta = -116.9$ (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} = 225$ Hz), -124.6 (d, ${}^{2}J_{F,F}$ = 225 Hz) ppm. IR (KBr): \tilde{v}_{max} = 1799, 1750, 1323, 1208, 1131, 1068, 703 cm⁻¹. $C_{12}H_{11}F_2NO_3$ (255.22): calcd. C 56.47, H 4.34, N 5.49; found C 56.46, H 4.75, N 5.21.

2-(3,3-Difluoro-2-oxo-4-phenylazetidin-1-yl)acetate rac-Benzvl (18b): This compound was prepared from racemic β-lactam 12a according to the procedure already described for the conversion of 12a into 18a, from azetidin-2-one 12a (2g, 10.9 mmol), nBu₄NI (2 g, 5.4 mmol, 0.5 equiv.), benzyl bromoacetate (17b, 17.35 mL, 109 mmol, 10 equiv.), and 1,1,3,3-tetramethylguanidine (2.5 g, 21.7 mmol, 2 equiv.) in anhydrous acetonitrile (10 mL). Chromatography on silica gel [cyclohexane/EtOAc (10:0 to 8:2) as eluent] gave β -lactam **18b** as a colorless oil; yield 3.28 g (91%); TLC: silica gel (cyclohexane/EtOAc, 8:2), $R_f = 0.41$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.40–7.15 (m, 10 H), 5.17 (dd, ³J_{H,F} = 2.5, ${}^{3}J_{H,F}$ = 7 Hz, 1 H), 5.11 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 5.05 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 4.46 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.58 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 166.8, 161.4 (t, ²J_{C,F} = 31 Hz), 134.7, 130.2, 129.7, 129.3, 128.9, 128.8, 128.7, 128.2, 121.0 (dd, ${}^{1}J_{C,F} = 289$, ${}^{1}J_{C,F} = 292$ Hz), 69.4 (dd, ${}^{2}J_{C,F} = 24.5$, ${}^{2}J_{C,F}$ = 27 Hz), 67.9, 41.1 (t, J = 30 Hz) ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -114.1 (d, ${}^{2}J_{\rm F,F}$ = 225.5 Hz), -121.8 (d, ${}^{2}J_{\rm F,F}$ = 225.5 Hz) ppm. IR (KBr): \tilde{v}_{max} = 1798, 1747, 1323, 1201, 1067, 701 cm⁻¹.

rac-2-(3,3-Difluoro-2-oxo-4-phenylazetidin-1-yl)acetic Acid (19): β-Lactam 18b (3.31 g, 10 mmol) was dissolved in ethanol (60 mL), and palladium on charcoal (10%, 0.2 g) was added. The mixture was first placed under nitrogen and subsequently under hydrogen (hydrogen balloon, 1 bar). After the system had been stirred at room temp. for 24 h, the hydrogen gas was removed with the aid of a nitrogen flow. The catalyst was filtered off through a pad of Celite and washed with ethanol $(2 \times 20 \text{ mL})$. The solvent was evaporated in vacuo. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/ H₂O/HCl (1 N, 500:500:10) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the title compound 20 as a white solid; yield 3.28 g (91%); m.p. 162 °C. ¹H NMR (CD₃OD, 300.3 MHz): δ = 7.50–7.45 (m, 3 H), 7.40–7.35 (m, 2 H), 5.34 (dd, ${}^{3}J_{H,F}$ = 2, ${}^{3}J_{\rm H,F}$ = 7.5 Hz, 1 H), 4.45 (d, ${}^{2}J_{\rm H,H}$ = 18 Hz, 1 H), 3.74 (dd, J = 1, ${}^{2}J_{H,H}$ = 18 Hz, 1 H) ppm. ${}^{13}C$ (CD₃OD, 75.4 MHz): δ = 170.0, 162.8 (t, ${}^{2}J_{C,F}$ = 31 Hz), 131.4, 130.9, 130.1, 129.2, 122.4 (t, ${}^{1}J_{C,F}$ = 290 Hz), 70.5 (dd, ${}^{2}J_{C,F}$ = 24, ${}^{2}J_{C,F}$ = 27 Hz), 42.1 ppm. ${}^{19}F$ NMR (CD₃OD, 288.3 MHz): $\delta = -104.2$ (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} =$ 225 Hz), -121.9 (d, ${}^{2}J_{F,F}$ = 225 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3044, 1791, 1718, 1323, 1243, 701 cm⁻¹. MS (CI+): $m/z = 242 [M + H]^+$, 198, 154, 140, 102, 93. HRMS (CI+): calcd. for C₁₁H₉F₂NO₃ 242.0629; found 242.0626. C₁₁H₉F₂NO₃ (241.20): calcd. C 54.78, H 3.76, N 5.81; found C 54.82, H 3.58, N 5.78.



rac-4-{1-[(Benzyloxycarbonyl)methyl]-3,3-difluoro-4-oxo-Benzyl azetidin-2-yl{piperidine-1-carboxylate (20b): This compound was prepared from racemic β -lactam 12a according to the procedure already described for the conversion of 12a into 18b, from azetidin-2-one 12b (0.56 g, 1.7 mmol), nBu₄NI (0.3 g, 0.8 mmol, 0.5 equiv.), benzyl bromoacetate (17b, 10 mL, 63 mmol, 37 equiv.), and 1,1,3,3tetramethylguanidine (0.39 g, 3.4 mmol, 2 equiv.) in anhydrous acetonitrile (10 mL). Chromatography on silica gel [cyclohexane/ EtOAc (95:5 to 80:20) as eluent] gave β -lactam **20b** as an orange oil; yield 0.24 g (30%); TLC: silica gel (cyclohexane/EtOAc, 8:2), $R_f = 0.40$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.40-7.25$ (m, 10 H), 5.20 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 5.15 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 5.10 (s, 2 H), 4.43 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 4.30–4.05 (m, 2 H), 3.96 (dt, ${}^{3}J_{\text{H,F}} = 2.5, {}^{3}J_{\text{H,H}} = {}^{3}J_{\text{H,F}} = 11 \text{ Hz}, 1 \text{ H}), 3.87 \text{ (d, } {}^{2}J_{\text{H,H}} = 18 \text{ Hz},$ 1 H), 2.85-2.60 (m, 2 H), 2.00-1.85 (m, 1 H), 1.75-1.65 (m, 1 H), 1.60-1.45 (m, 1 H), 1.30-1.10 (m, 2 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 166.6, 161.2 (t, ² $J_{C,F}$ = 31 Hz), 155.0, 136.5, 134.5, 128.9, 128.7, 128.6, 128.5, 128.1, 127.9, 69.8 (dd, ${}^{2}J_{C,F} = 22$, ${}^{2}J_{C,F}$ = 25 Hz), 67.8, 67.2, 43.3, 43.2 and 43.1, 35.7, 28.1 and 27.6 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -113.5 (dd, ³J_{F,H} = 8, ²J_{F,F} = 234 Hz), -114.5 (dd, ${}^{3}J_{F,H}$ = 21, ${}^{2}J_{F,F}$ = 234 Hz) ppm. IR (KBr): $\tilde{v}_{max} = 1798, 1747, 1323, 1201, 1067, 701 \text{ cm}^{-1}.$

Benzyl rac-[2-(4-{[(Benzyloxycarbonyl)amino]methyl}phenyl)-3,3-difluoro-4-oxoazetidin-1-yl]acetate (20c): This compound was prepared from racemic β -lactam 12c according to the procedure already described for conversion of 12a into 18b, from azetidin-2-one 12c (1.28 g, 3.7 mmol), *n*Bu₄NI (1.38 g, 3.7 mmol, 1 equiv.), benzyl bromoacetate (17b, 9.2 g, 40 mmol, 10.8 equiv.), and 1,1,3,3-tetramethylguanidine (0.84 g, 7.3 mmol, 2 equiv.) in anhydrous acetonitrile (80 mL). Chromatography on silica gel [CH2Cl2/EtOAc (100:0 to 98:2) as eluent] gave β -lactam **20c** as a colorless oil; yield 1.74 g (95%); TLC: silica gel (CH₂Cl₂/EtOAc, 95:5), $R_f = 0.75$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.40–7.25 (m, 12 H), 7.21 (d, ${}^{3}J_{H,H}$ = 8 Hz, 2 H), 5.21 (dd, ${}^{3}J_{H,F}$ = 2.5, ${}^{3}J_{H,F}$ = 7.5 Hz, 1 H), 5.17 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 5.12 (br. s, 3 H), 5.10 (d, ${}^{2}J_{H,H}$ = 11.5 Hz, 1 H), 4.50 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 4.38 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 3.61 (dd, J = 1.5, ${}^{2}J_{H,H} = 18$ Hz, 1 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): δ = 166.5, 161.1 (t, ²*J*_{C,F} = 31 Hz), 156.4, 140.7, 136.2, 134.5, 128.8, 128.7, 128.5, 128.4, 128.2, 128.1, 128.0, 120.8 (t, $^1\!J_{\rm C.F}$ = 291 Hz), 68.9 (dd, ${}^{2}J_{C,F}$ = 23.5, ${}^{2}J_{C,F}$ = 26.5 Hz), 67.7, 66.9, 44.5, 40.9 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -114.1 (dd, ³J_{F,H} = 7.5, ${}^{2}J_{\rm F,F}$ = 225 Hz), -121.8 (d, ${}^{2}J_{\rm F,F}$ = 225 Hz) ppm. C₂₇H₂₄F₂N₂O₅ (494.50): calcd. C 65.58, H 4.89, N 5.67; found C 65.97, H 5.06, N 5.57.

rac-Benzyl 2-[3,3-Difluoro-2-oxo-4-(pyridin-4-yl)azetidin-1-yl]acetate (20d): This compound was prepared from racemic β -lactam 12d according to the procedure already described for the conversion of 12a into 18b, from azetidin-2-one 12d (5.15 g, 28 mmol), nBu₄NI (10.3 g, 29.7 mmol, 1 equiv.), benzyl bromoacetate (17b, 7 g, 30.6 mmol, 1.1 equiv.), and 1,1,3,3-tetramethylguanidine (6.4 g, 55.6 mmol, 2 equiv.) in anhydrous acetonitrile (150 mL). Chromatography on silica gel [CH₂Cl₂/MeOH/NH₄OH (99.5:0.5: 0.05 to 98:2:0.2) as eluent] gave β -lactam **20d** as a pale yellow oil; yield 2.70 g (29%); TLC: silica gel (CH2Cl2/MeOH/NH4OH, 97:3:0.3), $R_f = 0.73$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 8.66$ (d, ${}^{3}J_{\rm H,H}$ = 6 Hz, 2 H), 7.40–7.25 (m, 5 H), 7.18 (d, ${}^{3}J_{\rm H,H}$ = 6 Hz, 2 H), 5.22 (dd, ${}^{3}J_{H,F} = 2$, ${}^{3}J_{H,F} = 7$ Hz, 1 H), 5.17 (d, ${}^{2}J_{H,H} = 12$ Hz, 1 H), 5.11 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 4.54 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.67 (dd, J = 1.5, ${}^{2}J_{H,H} = 18$ Hz, 1 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): δ = 166.3, 160.7 (t, ² $J_{C,F}$ = 31 Hz), 150.6, 138.8, 134.3, 128.9, 128.7, 128.5, 122.5, 120.7 (dd, ${}^{1}J_{C,F} = 291$, ${}^{1}J_{C,F} = 293$ Hz), 68.0 (dd, ${}^{2}J_{C,F}$ = 23.5, ${}^{2}J_{C,F}$ = 27 Hz), 67.9, 41.3 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -113.3$ (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} = 225.5$ Hz),

-120.6 (d, ${}^{2}J_{F,F} = 225.5$ Hz) ppm. MS (EI+): m/z = 332 [M]⁺⁺, 281, 232, 198, 169, 141, 91, 65.

rac-Benzyl 2-[3,3-Difluoro-2-oxo-4-(pyridin-3-yl)azetidin-1-yl]acetate (20e): This compound was prepared from racemic β-lactam 12e according to the procedure already described for the conversion of 12a into 18b, from azetidin-2-one 12e (0.5 g, 2.7 mmol), nBu₄NI (0.5 g, 3.3 mmol, 1.2 equiv.), benzyl bromoacetate (17b, 0.68 g, 3.0 mmol, 1.1 equiv.), and 1,1,3,3-tetramethylguanidine (0.62 g, 5.4 mmol, 2 equiv.) in anhydrous acetonitrile (50 mL). Chromatography on silica gel [CH₂Cl₂/MeOH/NH₄OH (99.5:0.5:0.05 to 98:2:0.2) as eluent] gave β -lactam **20e** as a pale yellow oil; yield 0.45 g (50%); TLC: silica gel (CH₂Cl₂/MeOH/NH₄OH, 98:2:0.2), $R_f = 0.48$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 8.65$ (d, ³ $J_{H,H} =$ 4 Hz, 1 H), 8.49 (s, 1 H), 7.61 (dd, ${}^{4}J_{H,H} = 1$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 7.4–7.2 (m, 6 H), 5.26 (dd, ${}^{3}J_{H,F} = 2.5$, ${}^{3}J_{H,F} = 7$ Hz, 1 H), 5.17 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 5.11 (d, ${}^{2}J_{H,H}$ = 12 Hz, 1 H), 4.49 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.63 (dd, J = 1.5, ${}^{2}J_{H,H} = 18$ Hz, 1 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): δ = 166.3, 160.8 (t, ²J_{C.F} = 30.5 Hz), 151.3, 149.6, 135.6, 134.3, 128.8, 128.7, 128.5, 124.6, 123.8, 120.8 (t, ${}^{1}J_{C,F} = 292 \text{ Hz}$), 67.9, 67.0 (dd, ${}^{2}J_{C,F} = 24$, ${}^{2}J_{C,F} = 27 \text{ Hz}$), 41.1 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -113.8$ (dd, ³ $J_{\rm EH} =$ 7.5, ${}^{2}J_{\text{F,F}}$ = 226 Hz), -121.2 (d, ${}^{2}J_{\text{F,F}}$ = 226 Hz) ppm. IR (KBr): $\tilde{v}_{max} = 1800, 1747, 1312, 1206, 1067, 699 \text{ cm}^{-1}.$

rac-tert-Butyl 2-[3,3-Difluoro-2-oxo-4-(pyridin-4-yl)azetidin-1-yl]acetate (22d): This compound was prepared from racemic β -lactam 12d according to the procedure already described for the conversion of 12a into 18b, from azetidin-2-one 12d (1.9 g, 10.3 mmol), NaI (1.8 g, 12 mmol, 1.15 equiv.), *tert*-butyl bromoacetate (17c, 2 g, 10.3 mmol, 1 equiv.), and 1,1,3,3-tetramethylguanidine (2.37 g, 20.6 mmol, 2 equiv.) in anhydrous acetonitrile (100 mL). Chromatography on silica gel [CH₂Cl₂/MeOH/NH₄OH (99:1:0.1 to 98:2:0.2) as eluent] gave β -lactam 22d as a brown oil; yield 1.44 g (47%).

rac-tert-Butyl 2-[3,3-Difluoro-2-oxo-4-(pyridin-3-yl)azetidin-1-yl]acetate (22e): This compound was prepared from racemic β -lactam 12e according to the procedure already described for the conversion of 12a into 18b, from azetidin-2-one 12e (0.65 g, 3.5 mmol), NaI (0.6 g, 4 mmol, 1.15 equiv.), tert-butyl bromoacetate (17c, 0.69 g, 3.5 mmol, 1 equiv.), and 1,1,3,3-tetramethylguanidine (0.81 g, 7 mmol, 2 equiv.) in anhydrous acetonitrile (50 mL). Chromatography on silica gel [CH2Cl2/MeOH/NH4OH (99.5:0.5:0.05 to 98:2:0.2) as eluent] gave β -lactam **22e** as a pale yellow solid; yield 0.52 g (50%); m.p. 85 °C; TLC: silica gel (CH₂Cl₂/MeOH/NH₄OH, 95:5:0.5), $R_f = 0.75$. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 8.65$ (d, ${}^{3}J_{H,H} = 5$ Hz, 1 H), 8.54 (s, 1 H), 7.63 (d, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 7.36 (dd, ${}^{3}J_{H,H} = 5$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 5.25 (dd, ${}^{3}J_{H,F} = 2.5$, ${}^{3}J_{H,F} = 3.5$ 7 Hz, 1 H), 4.33 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.47 (dd, J = 1, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 1.39 (s, 9 H) ppm. $^{13}\mathrm{C}$ NMR (CDCl₃, 75.4 MHz): δ = 165.4, 160.8 (t, ${}^{2}J_{C,F}$ = 31 Hz), 151.3, 149.6, 135.5, 125.9, 123.8, 120.8 (t, ${}^{1}J_{C,F}$ = 291.5 Hz), 83.6, 66.9 (dd, ${}^{2}J_{C,F}$ = 23.5, ${}^{2}J_{C,F}$ = 27 Hz), 41.8, 27.8 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -114.1 (dd, ${}^{3}J_{F,H}$ = 7.5, ${}^{2}J_{F,F}$ = 225.5 Hz), -121.3 (d, ${}^{2}J_{F,F}$ = 225.5 Hz) ppm. IR (KBr): $\tilde{v}_{max} = 1797, 1733, 1416, 1374, 1315, 1248, 1166,$ 1066, 720 cm⁻¹. C₁₄H₁₆F₂N₂O₃ (298.29): calcd. C 56.37, H 5.41, N 9.39; found C 56.45, H 5.45, N 9.46.

rac-2-[3,3-Difluoro-2-oxo-4-(piperidin-4-yl)azetidin-1-yl]acetic Acid Hydrochloride (21b): β -Lactam 20b (0.23 g, 0.5 mmol) was dissolved in ethanol (5 mL), and Pd/C (10%, 50 mg) was added. The mixture was first placed under nitrogen and subsequently under hydrogen (hydrogen balloon, 1 bar). After the mixture had been stirred at room temp. for 24 h, the hydrogen gas was removed with the aid of a nitrogen flow. The catalyst was filtered off through a pad of Celite, and washed with ethanol $(2 \times 10 \text{ mL})$. The solvent was evaporated in vacuo. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 100:100:2) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 21b as a pale yellow lyophilizate; yield: 70 mg (50%). ¹H NMR (D₂O, 300.3 MHz): δ = 4.13 (dt, ³J_{H,F} = 2.5, ³J_{H,F} = 9 Hz, 1 H), 4.06 (d, ${}^{2}J_{H,H}$ = 17.5 Hz, 1 H), 3.82 (d, ${}^{2}J_{H,H}$ = 17.5 Hz, 1 H), 3.45-3.35 (m, 2 H), 3.00-2.85 (m, 2 H), 2.30-2.10 (m, 1 H), 2.00–1.85 (m, 2 H), 1.65–1.45 (m, 2 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 174.0, 163.6 (t, ²J_{C,F} = 30.5 Hz), 120.5 (t, ${}^{1}J_{C,F}$ = 290 Hz), 70.5 (dd, ${}^{2}J_{C,F}$ = 22, ${}^{2}J_{C,F}$ = 24.5 Hz), 49.8, 44.2 and 44.1, 33.6, 25.5 and 24.6 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ = -115.2 (dd, ${}^{3}J_{\rm F,H}$ = 8.5, ${}^{2}J_{\rm F,F}$ = 232.5 Hz), -125.3 (d, ${}^{2}J_{\rm F,F}$ = 232.5 Hz) ppm. MS (ESI–): $m/z = 247.0 [M - H]^{-}$, 183.0.

rac-2-{2-[4-(Aminomethyl)phenyl]-3,3-difluoro-4-oxoazetidin-1-yl}acetic Acid Hydrochloride (21c): This compound was prepared from racemic β-lactam 20c according to the procedure already described for the conversion of 20b into 21b, from azetidin-2-one 20c (1.71 g, 3.4 mmol) and Pd/C (10%, 0.2 g) in methanol (60 mL). The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 500:500:10) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 21c as a white lyophilizate; yield 0.26 g (25%). ¹H NMR (D_2O , 300.3 MHz): δ = 7.65 (d, ³J_{H,H} = 8 Hz, 2 H), 7.61 (d, ³J_{H,H} = 8 Hz, 2 H), 5.58 (dd, ${}^{3}J_{H,F} = 2$, ${}^{3}J_{H,F} = 7$ Hz, 1 H), 4.58 (d, ${}^{2}J_{H,H} = 18$ Hz, 1 H), 4.32 (s, 2 H), 4.04 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H) ppm. ${}^{13}C$ NMR (D₂O, 75.4 MHz): δ = 170.9, 163.4 (t, ² $J_{C,F}$ = 31 Hz), 134.8, 130.8, 129.7, 129.4, 120.3 (t, ${}^{1}J_{C,F}$ = 288 Hz), 69.4 (dd, ${}^{2}J_{C,F}$ = 24, ${}^{2}J_{C,F}$ = 26.5 Hz), 43.0, 42.3 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): $\delta = -115.8$ (dd, ${}^{3}J_{EH} = 6.5$, ${}^{2}J_{EF} = 225.5$ Hz), -123.0 (d, ${}^{2}J_{EF} = 225.5$ Hz) ppm. IR (KBr): \tilde{v}_{max} = 3042, 1752, 1518, 1412, 1220 cm⁻¹. MS (ESI+): $m/z = 292.9 [M + Na]^+$, 270.9 $[M + H]^+$, 254.0 $[M + H - Ma]^+$ $NH_{3}]^{+}$.

2-{3-|4-(Aminomethyl)phenyl]-2,2-difluoropropanamido}acetic Acid: The hydrogenolysis deprotection step of **20c** into **21c** was accompanied by partial degradation. The corresponding hydrochloride salt was isolated from the steric exclusion chromatography as a white lyophilizate; yield 105 mg (10%). ¹H NMR (D₂O, 300.3 MHz): δ = 7.40–7.35 (m, 4 H), 4.14 (s, 2 H), 3.91 (s, 2 H), 3.43 (dt, ³J_{H,H} = 3, ³J_{H,F} = 16.5 Hz, 2 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 172.6, 166.6 (t, ²J_{C,F} = 30 Hz), 132.6, 132.3, 131.5, 129.3, 117.4 (t, ¹J_{C,F} = 252 Hz), 43.0, 41.1, 40.2 (t, ²J_{C,F} = 24.5 Hz) ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ = -106.4 (t, ²J_{F,F} = 16.5 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3330, 2038, 1697, 1551, 1407, 1230, 902, 770 cm⁻¹. MS (ESI+): *m*/*z* = 294.9 [M + Na]⁺, 273.0 [M + H]⁺, 256.0 [M + H – NH₃]⁺.

rac-2-[3,3-Difluoro-2-oxo-4-(pyridin-4-yl)azetidin-1-yl]acetic Acid Hydrochloride (21d): TFA (6 mL) was added to a solution of azetidin-2-one 22d (1.44 g, 4.85 mmol) in wet CH₂Cl₂ (8 mL). The mixture was stirred at 0 °C for 1 h. After concentration, the crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 500:500:10) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the trifluoroacetate salt of the title compound 21c as a pale yellow lyophilizate; yield 0.76 g (44%). ¹H NMR (CD₃OD, 300.3 MHz): δ = 7.65 (d, ³J_{H,H} = 8 Hz, 2 H), 7.61 (d, ³J_{H,H} = 8 Hz, 2 H), 5.58 (dd, ³J_{H,F} = 2, ³J_{H,F} = 7 Hz, 1 H), 4.58 (d, ²J_{H,H} = 18 Hz, 1 H), 4.32 (s, 2 H), 4.04 (d, ${}^{2}J_{H,H} = 18$ Hz, 1 H) ppm. ${}^{13}C$ NMR (CD₃OD, 75.4 MHz): $\delta = 169.8$, 164.1 (t, ${}^{2}J_{C,F} = 30.5$ Hz), 152.4, 144.4, 127.4, 122.3 (t, ${}^{1}J_{C,F} = 293$ Hz), 68.9 (dd, ${}^{2}J_{C,F} = 23$, ${}^{2}J_{C,F} = 27$ Hz), 43.3 ppm. ${}^{19}F$ NMR (CD₃OD, 288.3 MHz): $\delta = -77.9$ (CF₃), -115.6 (dd, ${}^{3}J_{F,H} = 7.5$, ${}^{2}J_{F,F} = 224$ Hz), -121.6 (d, ${}^{2}J_{F,F} = 224$ Hz) ppm. IR (KBr): $\tilde{v}_{max} = 3422$, 2350, 1801, 1730, 1643, 1420, 1324, 1193, 720 cm⁻¹. MS (ESI+): m/z = 242.9 [M + H]⁺.

rac-3-[(Carboxymethyl)amino]-2,2-difluoro-3-(pyridin-4-yl)propanoic Acid Hydrochloride: The trifluoroacetate form of the product of nucleophilic ring-opening was obtained during the TFA deprotection step of **22d**, after isolation from the steric exclusion chromatography as a white lyophilizate; yield 200 mg (11%). ¹H NMR (D₂O, 300.3 MHz): $\delta = 8.89$ (d, ${}^{3}J_{H,H} = 6.5$ Hz, 2 H), 8.19 (d, ${}^{3}J_{H,H} = 6$ Hz, 2 H), 5.32 (dd, ${}^{3}J_{H,F} = 8.5$, ${}^{3}J_{H,F} = 15$ Hz, 1 H), 3.80 (s, 2 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): $\delta = 170.3$, 165.8 (t, ${}^{2}J_{C,F} = 26$ Hz), 149.8, 142.6, 128.4, 114.0 (t, ${}^{1}J_{C,F} = 261$ Hz), 62.6 (dd, ${}^{2}J_{C,F} = 24$, ${}^{2}J_{C,F} = 26$ Hz), 47.5 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): $\delta = -77.9$ (CF₃), -108.3 (dd, ${}^{3}J_{F,H} = 8.5$, ${}^{2}J_{F,F} = 254.5$ Hz), -112.4 (dd, ${}^{3}J_{F,H} = 15$, ${}^{2}J_{E,F} = 254.5$ Hz) ppm. IR (KBr): $\tilde{\nu}_{max} = 3412$, 1740, 1644, 1510, 1407, 1295, 1243, 1194 cm⁻¹. MS (ESI+): m/z = 260.9 [M + H]⁺.

rac-2-[3,3-Difluoro-2-oxo-4-(pyridin-3-yl)azetidin-1-yl]acetic Acid Hydrochloride (21e): TFA (2.5 mL) was added to a solution of azetidin-2-one 22e (0.3 g, 1 mmol) in wet CH_2Cl_2 (2.5 mL). The mixture was stirred at 0 °C for 1 h. After concentration, the crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H2O/HCl (1 N, 200:200:4) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 21e as a pale yellow lyophilizate; yield 0.16 g (56%). ¹H NMR (D₂O, 300.3 MHz): δ = 8.97 (s, 1 H), 8.85 (d, ${}^{3}J_{H,H}$ = 5.5 Hz, 1 H), 8.76 (d, ${}^{2}J_{H,H}$ = 7.5 Hz, 1 H), 8.15 (t, ${}^{3}J_{H,H}$ = 6.5 Hz, 1 H), 5.74 (dd, ${}^{3}J_{H,F}$ = 2, ${}^{3}J_{H,F}$ = 6.5 Hz, 1 H), 4.45 (d, ${}^{2}J_{H,H}$ = 18.5 Hz, 1 H), 4.05 (d, ${}^{2}J_{H,H}$ = 18.5 Hz, 1 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 170.5, 166.0 $(t, {}^{2}J_{C,F} = 23.5 \text{ Hz}), 147.4, 142.6, 141.9, 131.7, 128.0, 66.6 \text{ (dd, } {}^{2}J_{C,F}$ = 23.5, ${}^{2}J_{C,F}$ = 27 Hz), 42.8 ppm. ${}^{19}F$ NMR (D₂O, 288.3 MHz): δ = -115.2 (dd, ${}^{3}J_{F,H}$ = 6.5, ${}^{2}J_{F,F}$ = 226.5 Hz), -125.5 (d, ${}^{2}J_{F,F}$ = 226.5 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3412, 1802, 1736, 1637, 1400, 1313, 1203 cm⁻¹. MS (ESI+): $m/z = 242.9 [M + H]^+$.

rac-3-[(Carboxymethyl)amino]-2,2-difluoro-3-(pyridin-4-yl)propanoic Acid Hydrochloride: The hydrochloride form of the product of nucleophilic ring-opening was obtained during the TFA deprotection step of 22e, after isolation from the steric exclusion chromatography as a pale yellow lyophilizate; yield 130 mg (44%). ¹H NMR (D₂O, 300.3 MHz): $\delta = 8.98$ (s, 1 H), 8.92 (d, ${}^{3}J_{H,H} = 6$ Hz, 1 H), 8.76 (d, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 8.18 (dd, ${}^{3}J_{H,H} = 6$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 5.36 (dd, ${}^{3}J_{H,F} = 8$, ${}^{3}J_{H,F} = 16$ Hz, 1 H), 3.84 (s, 2 H) ppm. 13 C NMR (D₂O, 75.4 MHz): $\delta = 169.9$, 165.7 (t, ${}^{2}J_{C,F} = 25$ Hz), 147.8, 143.7, 142.8, 129.9, 128.5, 114.0 (t, ${}^{1}J_{C,F} = 260$ Hz), 60.8 (dd, ${}^{2}J_{C,F} = 23.5$, ${}^{2}J_{C,F} = 26$ Hz), 47.3 ppm. 19 F NMR (D₂O, 288.3 MHz): $\delta = -108.3$ (dd, ${}^{3}J_{F,H} = 8.5$, ${}^{2}J_{F,F} = 256$ Hz), -113.0 (dd, ${}^{3}J_{F,H} = 16$, ${}^{2}J_{F,F} = 256$ Hz) ppm. IR (KBr): $\tilde{v}_{max} = 3416$, 1740, 1637, 1406, 1297, 1196 cm⁻¹. MS (ESI+): m/z = 282.9 [M + Na]⁺, 260.9 [M + H]⁺. MS (ESI-): 258.7 [M - H]⁻.

(Acetylsulfanyl)acetyl Chloride (23): Acetic anhydride (30 mL, 317 mmol, 1.1 equiv.) was added dropwise at 0 °C to a solution of mercaptoacetic acid (20 mL, 287 mmol), triethylamine (80 mL, 569 mmol, 2 equiv.), and DMAP as catalyst (100 mg) in anhydrous MeCN (200 mL). The mixture was stirred at room temp. for 18 h. The reaction was quenched with water (20 mL). The solvent was



evaporated to dryness, and the residue was taken up in CH₂Cl₂ (200 mL). The organic layer was washed with water $(2 \times 100 \text{ mL})$, and extracted with aqueous NaOH solution (1 N, 100 mL). The aqueous layer was washed with diethyl ether (100 mL), acidified to pH = 1 by the addition of aqueous HCl solution (4 N), and extracted with EtOAc (3×150 mL). The combined organic phases were washed with brine (50 mL), dried with MgSO₄, filtered, and concentrated. The residue was distilled under reduced pressure (1 Torr), and the fraction boiling between 118 and 122 °C was collected to afford pure (acetylsulfanyl)acetic acid as a pale yellow oil; yield 25.03 g (65%). ¹H NMR (CDCl₃, 300.3 MHz): δ = 10.17 (br. s, 1 H), 4.09 (s, 2 H), 2.38 (s, 3 H) ppm. ¹³C (CDCl₃, 75.4 MHz): $\delta = 193.9, 174.7, 31.3, 30.1$ ppm. IR (KBr): $\tilde{v}_{max} = 3450, 1697$ cm⁻¹. C₄H₆O₃S (134.15): calcd. C 35.81, H 4.51, S 23.90; found C 36.02, H 4.99, S 23.59. Oxalyl chloride (7.5 mL, 88.4 mmol, 1.5 equiv.) was added dropwise at 0 °C over 1 h to a solution of acid (8 g, 59.6 mmol) and DMF (0.6 mL, 7.7 mmol) in anhydrous CH₂Cl₂ (100 mL). The mixture was stirred at room temp. for 12 h, and the solvents were evaporated to dryness. The crude oil was distilled at 7 mbar to give pure acyl chloride 23 as an orange oil; yield 6.10 g (67%); b.p. 62 °C. ¹H NMR (CDCl₃, 300.3 MHz): δ = 4.15 (s, 2 H), 2.40 (s, 3 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 192.9, 169.6, 42.4, 30.4 ppm.

rac-tert-Butyl 4-[1-{[(Acetylsulfanyl)acetyl]amino}-2,2-difluoro-2-(methoxycarbonyl)ethyl]piperidine-1-carboxylate (25b): A solution of acyl chloride 23 (0.75 g, 4.9 mmol, 2.05 equiv.) in anhydrous CH₂Cl₂ (3 mL) was added under nitrogen to a solution of β-amino ester 15b (0.79 g, 2.4 mmol) in anhydrous CH₂Cl₂ (5 mL). N,N-Diisopropylethylamine (0.87 mL, 5 mmol, 2.1 equiv.) was then added dropwise to the solution at 0 °C, and the mixture was stirred at room temp. for 12 h. The reaction was quenched by the addition of saturated aqueous NaHCO₃ solution (20 mL), and the reaction mixture was extracted with CH_2Cl_2 (2 × 80 mL). The combined extracts were dried with anhydrous MgSO₄, and the solvent was evaporated to give a crude oily product. This was subjected to column chromatography [CH₂Cl₂/EtOAc (90:10 to 75:25) as eluent] to afford pure amide 25b as a yellow oil; yield 0.78 g (74%); TLC: silica gel (CH₂Cl₂/EtOAc, 7:3), $R_f = 0.78$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 6.59 (d, ${}^{3}J_{H,H}$ = 10 Hz, 1 H), 4.55–4.40 (m, 1 H), 4.15–3.95 (m, 2 H), 3.77 (s, 3 H), 3.49 (d, ${}^{2}J_{H,H}$ = 14.5 Hz, 1 H), 3.43 (d, ${}^{2}J_{H,H}$ = 14.5 Hz, 1 H), 2.70–2.50 (m, 2 H), 1.95–1.80 (m, 1 H), 1.36 (s, 9 H), 1.7-1.2 (m, 4 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 196.2, 168.5, 163.1 (dd, ${}^{2}J_{C,F}$ = 31, ${}^{2}J_{C,F}$ = 33.5 Hz), 154.4, 114.5 (t, ${}^{1}J_{C,F}$ = 257 Hz), 79.4, 53.9 (dd, ${}^{2}J_{C,F}$ = 22.5, ${}^{2}J_{C,F}$ = 27 Hz), 53.5, 43.2, 35.3, 32.6, 30.0, 29.1, 28.2, 26.6 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -109.7 (dd, ³J_{F,H} = 6, ${}^{2}J_{F,F}$ = 265 Hz), -115.6 (dd, ${}^{3}J_{F,H}$ = 18.5, ${}^{2}J_{F,F}$ = 265 Hz) ppm.

rac-Ethyl 3-{[(Acetylsulfanyl)acetyl]amino}-3-(4-{[(*tert*-butoxycarbonyl)amino]methyl}phenyl)-2,2-difluoropropionate (25c) and *rac*-Methyl 3-{[(Acetylsulfanyl)acetyl]amino}-3-(4-{[(*tert*-butoxycarbonyl)amino]methyl}phenyl)-2,2-difluoropropionate (25c'): Racemic β-amino ester 25c and its methyl ester derivative 25c' were obtained from β-amino ester 15c and its methyl ester derivative 15c' (mixture of ethyl and methyl esters) according to the procedure already described for the conversion of 15b into 25b, from 15c and 15c' (0.91 g, 2.6 mmol), DMAP (50 mg), *N*,*N*-diisopropylethylamine (0.5 mL, 2.9 mmol, 1.2 equiv.), and acyl chloride 23 (0.79 g, 5.1 mmol, 2 equiv.) in anhydrous CH₂Cl₂ (13 mL). The crude product was chromatographed on silica gel [CH₂Cl₂/EtOAc (95:5 to 90:10) as eluent] to afford 25c as a yellow oil; yield 0.42 g (34%).

Ethyl Ester 25c: 51 %. ¹H NMR (CDCl₃, 300.3 MHz): δ = 7.30–7.15 (m, 5 H), 5.58 (td, ³*J*_{H,F} = 2.5, ³*J*_{H,H} = ³*J*_{H,F} = 9.5 Hz, 1 H),

4.86 (br. s, 1 H), 4.28 (d, ${}^{3}J_{H,H}$ = 6.5 Hz, 2 H), 4.23 (2×q, ${}^{3}J_{H,H}$ = 7 Hz, 2 H), 3.53 (m, 2 H), 2.41 (s, 3 H), 1.43 (s, 9 H), 1.25 (t, ${}^{3}J_{\text{H,H}} = 7 \text{ Hz}, 3 \text{ H}$) ppm. ${}^{13}\text{C}$ (CDCl₃, 75.4 MHz): $\delta = 196.6, 167.9,$ 162.8 (t, ${}^{2}J_{C,F}$ = 25 Hz), 155.9, 140.1, 131.5, 128.5, 127.7, 117.0 (t, ${}^{2}J_{C,F}$ = 258 Hz), 79.6, 63.3, 55.0 (t, ${}^{2}J_{C,F}$ = 24 Hz), 32.8, 30.2, 28.4, 13.8 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -111.8 (dd, ³J_{F,H} = 9.5, ${}^{2}J_{F,F}$ = 258 Hz), -115.6 (dd, ${}^{3}J_{F,H}$ = 17, ${}^{2}J_{F,F}$ = 258 Hz) ppm. Methyl Ester 25c': 49%. ¹H NMR (CDCl₃, 300.3 MHz): $\delta = 7.30$ – 7.15 (m, 5 H), 5.63 (td, ${}^{3}J_{H,F} = 2.5$, ${}^{3}J_{H,H} = {}^{3}J_{H,F} = 9.5$ Hz, 1 H), 4.86 (br. s, 1 H), 4.28 (d, ${}^{3}J_{H,H}$ = 6.5 Hz, 2 H), 3.79 (s, 3 H), 3.53 (m, 2 H), 2.41 (s, 3 H), 1.43 (s, 9 H) ppm. ¹³C NMR (CDCl₃, 75.4 MHz): δ = 196.5, 167.8, 162.9 (t, ² $J_{C,F}$ = 25 Hz), 155.9, 140.1, 131.5, 128.5, 127.6, 117.0 (t, ${}^{2}J_{C,F}$ = 258 Hz), 79.6, 55.0 (t, ${}^{2}J_{C,F}$ = 24 Hz), 53.6, 32.8, 30.2, 28.4, 13.8 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -111.3 (dd, ${}^{3}J_{F,H}$ = 8.5, ${}^{2}J_{F,F}$ = 259 Hz), -115.2 (dd, ${}^{3}J_{F,H} = 17$, ${}^{2}J_{F,F} = 259$ Hz) ppm.

rac-Ethyl 3-{[(Acetylsulfanyl)acetyl]amino}-2,2-difluoro-3-(pyridin-3-yl)propionate (25e): Racemic β-amino ester 25e was obtained from β -amino ester 15e according to the procedure already described for the conversion of 15b into 25b, from 15e (1 g, 3.75 mmol), DMAP (50 mg), triethylamine (1.8 mL, 12.8 mmol, 3.4 equiv.), and acyl chloride 23 (0.73 g, 4.8 mmol, 1.25 equiv.) in anhydrous CH₂Cl₂ (10 mL). The crude product was chromatographed on silica gel [CH2Cl2/MeOH/NH4OH (98:2:0.2 to 96:4:0.4) as eluent] to afford 25e as a yellow solid; yield 0.90 g (69%); m.p. 117 °C; TLC: silica gel (CH₂Cl₂/MeOH/NH₄OH, 9:1:0.1), R_f = 0.43. ¹H NMR (CDCl₃, 300.3 MHz): δ = 8.60 (d, ³*J*_{H,H} = 1.5 Hz, 1 H), 8.58 (d, ${}^{4}J_{H,H}$ = 1.5 Hz, 1 H), 7.66 (d, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 7.40 (br. d, ${}^{3}J_{H,H}$ = 9.5 Hz, 1 H), 7.29 (dd, ${}^{3}J_{H,H}$ = 5, ${}^{3}J_{H,H}$ = 7.5 Hz, 1 H), 5.70 (m, 1 H), 4.26 (dq, J = 1.5, ${}^{3}J_{H,H} = 7$ Hz, 2 H), 3.57 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.50 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 2.40 (s, 3 H), 1.27 (t, ${}^{3}J_{H,H} = 7$ Hz, 3 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): $\delta = 196.7$, 168.0, 162.2 (dd, ${}^{2}J_{C,F} = 31$, ${}^{2}J_{C,F} =$ 32.5 Hz), 150.3, 149.6, 135.8, 128.8, 123.5, 113.2 (t, ${}^{1}J_{C,F}$ = 257.5 Hz), 63.6, 53.2 (dd, ${}^{2}J_{C,F} = 24$, ${}^{2}J_{C,F} = 27.5$ Hz), 32.7, 30.2, 13.8 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): $\delta = -110.7$ (dd, ³ $J_{\rm E,H} =$ 8.5, ${}^{2}J_{F,F}$ = 261 Hz), -115.7 (dd, ${}^{3}J_{F,H}$ = 18.5, ${}^{2}J_{F,F}$ = 261 Hz) ppm.

rac-2,2-Difluoro-3-[(mercaptoacetyl)amino]-3-(piperidin-4-yl)propionic Acid (27b): TFA (10 mL) and water (1 mL) were added to a solution of amide 25b (0.78 g, 2.4 mmol) in CH₂Cl₂ (10 mL). The mixture was stirred at 0 °C for 1 h. After concentration, the crude product was dissolved in a mixture of MeOH/H₂O (1:3, 40 mL) and treated with aqueous NaOH solution (2 N) until pH = 9 was reached. The reaction mixture was stirred at 0 °C for 12 h. Most of the solvents were evaporated, and the mixture was washed with diethyl ether (30 mL) and acidified with aqueous HCl solution (1 N) until pH = 1 was reached. The aqueous phase was washed with diethyl ether (30 mL), and lyophilized. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 500:500:10) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 27b as a yellow lyophilizate; yield 0.45 g (59%). ¹H NMR (D₂O, 300.3 MHz): δ = 4.46 (dt, ³J_{H,F} = 5.5, ${}^{3}J_{H,H}$ = ${}^{3}J_{H,F}$ = 14.5 Hz, 1 H), 3.35–3.45 (m, 2 H), 3.26 (s, 2 H), 2.90-3.05 (m, 2 H), 2.05-2.20 (m, 1 H), 1.95-2.05 (m, 2 H), 1.45–1.65 (m, 2 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 172.5, 166.0 (t, ${}^{2}J_{C,F}$ = 28 Hz), 113.8 (t, ${}^{1}J_{C,F}$ = 257 Hz), 52.3 (t, ${}^{2}J_{C,F}$ = 24.5 Hz), 41.5 and 41.4, 31.2, 25.0, 21.8 and 21.7 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ = -110.1 (dd, ${}^{3}J_{F,H}$ = 14.5, ${}^{2}J_{F,F}$ = 249.5 Hz), $-111.0 \text{ (dd, } {}^{3}J_{\text{F,H}} = 14.5, {}^{2}J_{\text{F,F}} = 249.5 \text{ Hz} \text{) ppm. IR (KBr): } \tilde{v}_{\text{max}} =$ 3390, 2496, 1756, 1666, 1545, 1422, 1202, 959, 682 cm⁻¹. MS $(ESI+): m/z = 304.9 [M + Na]^+, 283.0 [M + H]^+.$

N. Boyer, P. Gloanec, G. De Nanteuil, P. Jubault, J.-C. Quirion

rac-3-[4-(Aminomethyl)phenyl]-3-[({[({1-[4-(aminomethyl)phenyl]-2carboxy-2,2-difluoroethyl}carbamoyl)methyl|disulfanyl}acetyl)amino]-2,2-difluoropropionic Acid (27c): Anhydrous HCl was bubbled through a solution of the mixture of β -amino ester 25c and its methyl ester derivative 25c' (0.42 g, 0.9 mmol) in anhydrous MeOH for 10 min. The resulting solution was then stirred at room temp. for 3 h and concentrated. The resulting yellow oil was dissolved in a mixture of MeOH/THF (1:1, 20 mL) and treated with aqueous NaOH (2 N) solution until pH = 9 was reached. The reaction mixture was stirred at room temp. for 12 h, poured into water (30 mL), washed with diethyl ether (30 mL), and acidified with aqueous HCl (1 N) until pH = 1 was reached. The aqueous phase was washed with diethyl ether (30 mL), and lyophilized. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 500:500:10) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 27c as a pale yellow lyophilizate; yield 0.10 g (36%). ¹H NMR (D₂O, 300.3 MHz): δ (pair of diastereomers) = 7.45–7.35 (m, 8 H), 5.53 (dd, ${}^{3}J_{H,F} = 13$, ${}^{3}J_{H,F} = 15.5$ Hz, 2 H), 4.08 (s, 2 H), 4.05 (s, 2 H), 3.34 (d, ${}^{2}J_{H,H}$ = 14.5 Hz, 1 H), 3.31 (s, 2 H), 3.24 (d, ${}^{2}J_{H,H}$ = 14.5 Hz, 1 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ (pair of diastereomers) = 171.8, 167.4 (t, ${}^{2}J_{C,F}$ = 28.5 Hz), 134.3, 133.8, 129.5, 129.4, 115.2 (t, ${}^{1}J_{C,F}$ = 257 Hz), 56.0 (t, ${}^{2}J_{C,F}$ = 24.5 Hz), 42.9, 41.1 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ (pair of diastereomers) = -111.6 (dd, ${}^{3}J_{F,H}$ = 12.5, ${}^{2}J_{F,F}$ = 250.5 Hz), -111.8 (dd, ${}^{3}J_{F,H}$ = 13, ${}^{2}J_{F,F} = 250.5 \text{ Hz}$), -114.3 (dd, ${}^{3}J_{F,H} = 16$, ${}^{2}J_{F,F} = 250.5 \text{ Hz}$), -114.5 (dd, ${}^{3}J_{F,H} = 16$, ${}^{2}J_{F,F} = 250.5$ Hz) ppm. IR (KBr): $\tilde{v}_{max} =$ 3500-3000, 1753, 1656, 1543, 1420, 1190, 971, 685 cm⁻¹. MS (ESI+): $m/z = 628.9 [M + Na]^+$, 607.0 $[M + H]^+$, 589.8 $[M + H - Ma]^+$ $NH_{3}]^{+}$.

rac-3-Amino-3-[4-(aminomethyl)phenyl]-2,2-difluoropropanoic Acid Hydrochloride: The hydrochloride form of the product of amide bond cleavage was obtained during the NaOH deprotection step of 25c, after isolation by steric exclusion chromatography, as a white lyophilizate; yield 50 mg (20%). ¹H NMR (D₂O, 300.3 MHz): δ = 7.51 (s, 4 H), 5.05 (dd, ³*J*_{H,F} = 6, ³*J*_{H,F} = 9 Hz, 1 H), 4.19 (s, 2 H) ppm. ¹H NMR ([D₆]DMSO, 300.3 MHz): δ = 9.43 (br. s, 1 H), 8.70 (br. s, 3 H), 7.57 (s, 4 H), 5.09 (dd, ³*J*_{H,F} = 10, ³*J*_{H,F} = 16 Hz, 1 H), 4.01 (s, 2 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 166.5 (t, ²*J*_{C,F} = 26 Hz), 135.2, 130.0, 129.8, 129.4, 114.6 (dd, ¹*J*_{C,F} = 257.5, ¹*J*_{C,F} = 261 Hz), 57.0 (dd, ²*J*_{C,F} = 22.5, ²*J*_{C,F} = 26 Hz), 42.9 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ = -107.8 (dd, ³*J*_{F,H} = 5.5, ²*J*_{F,F} = 245 Hz), -117.4 (dd, ³*J*_{F,H} = 18.5, ²*J*_{F,F} = 245 Hz) ppm. IR (KBr): \tilde{v}_{max} = 3020, 2044, 1682, 1592, 1410, 1217, 817 cm⁻¹. MS (ESI+): *m*/*z* = 253.0 [M + Na]⁺, 231.0 [M + H]⁺, 214.0 [M + H - NH₃]⁺.

rac-2,2-Difluoro-3-[(mercaptoacetyl)amino]-3-(pyridin-3-yl)propionic Acid (27e): A solution of amide 25e (0.9 g, 2.6 mmol) in a mixture of MeOH/THF/H₂O (1:1:1, 18 mL) was treated with an aqueous LiOH solution (1 N, 7 mL). The reaction mixture was stirred at 0 °C for 12 h. Most of the solvents were evaporated, and the mixture was washed with diethyl ether (30 mL) and acidified with aqueous HCl solution (1 N) until pH = 1 was reached. The aqueous phase was washed with diethyl ether (30 mL) and lyophilized. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H₂O/HCl (1 N, 200:200:4) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 27e as a white lyophilizate; yield 0.16 g (20%). ¹H NMR (D_2O , 300.3 MHz): δ = 8.82 (s, 1 H), 8.65 (d, ${}^{3}J_{H,H}$ = 6 Hz, 1 H), 8.52 (d, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 7.95 (dd, ${}^{3}J_{H,H} = 6$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 5.70 (dd, ${}^{3}J_{H,F}$

= 11, ${}^{3}J_{\rm H,F}$ = 15.5 Hz, 1 H), 3.17 (d, ${}^{2}J_{\rm H,H}$ = 15 Hz, 1 H), 3.11 (d, ${}^{2}J_{\rm H,H}$ = 15 Hz, 1 H) ppm. 13 C NMR (D₂O, 75.4 MHz): δ = 174.1, 165.3 (dd, ${}^{2}J_{\rm C,F}$ = 26, ${}^{2}J_{\rm C,F}$ = 27 Hz), 147.0, 142.0, 141.2, 134.5, 127.9, 114.2 (dd, ${}^{1}J_{\rm C,F}$ = 256, ${}^{1}J_{\rm C,F}$ = 261 Hz), 53.4 (dd, ${}^{2}J_{\rm C,F}$ = 25.5, ${}^{2}J_{\rm C,F}$ = 28 Hz), 27.2 ppm. 19 F NMR (D₂O, 288.3 MHz): δ = -111.4 (d, ${}^{2}J_{\rm F,F}$ = 256 Hz), -112.6 (d, ${}^{2}J_{\rm F,F}$ = 256 Hz) ppm. IR (KBr): $\tilde{v}_{\rm max}$ = 3404, 2360, 1756, 1669, 1547, 1409, 1325, 1194, 682 cm⁻¹. MS (ESI–): m/z = 275.0 [M – H]⁻, 190.9, 117.0.

rac-Ethyl 2,2-Difluoro-3-[3-methoxy-3-oxopropanoyl)amino]-3-(pyridin-4-yl)propanoate (26d): A solution of methyl malonyl chloride (24, 0.7 mL, 6.05 mmol, 1.6 equiv.) in anhydrous CH₂Cl₂ (3 mL) was added under nitrogen to a solution of β -amino ester 15d (1 g, 3.75 mmol) in anhydrous CH2Cl2 (12 mL). N,N-Diisopropylethylamine (1.7 mL, 9.85 mmol, 2.6 equiv.) was then added dropwise to the solution at 0 °C, and the mixture was stirred at room temp. for 12 h. The reaction was quenched by the addition of saturated aqueous NaHCO3 solution (20 mL), and the mixture was extracted with CH_2Cl_2 (2 × 80 mL). The combined extracts were dried with anhydrous MgSO₄, and the solvent was evaporated to give a crude oily product. This was subjected to column chromatography [CH₂Cl₂/ MeOH/NH₄OH (98:2:0.2 to 96:4:0.4) as eluent] to afford pure amide 26d as a brown oil; yield 0.53 g (43%); TLC: silica gel $(CH_2Cl_2/MeOH/NH_4OH, 9:1:0.1), R_f = 0.49.$ ¹H NMR (CDCl₃, 300.3 MHz): δ = 8.61 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 8.54 (d, ${}^{3}J_{H,H}$ = 9.5 Hz, 1 H), 8.30 (d, ${}^{3}J_{H,H}$ = 6 Hz, 2 H), 5.75 (dt, ${}^{3}J_{H,F}$ = 9.5, ${}^{3}J_{\rm H,F} = 16$ Hz, 1 H), 4.26 (q, ${}^{3}J_{\rm H,H} = 7$ Hz, 2 H), 3.76 (s, 3 H), 3.42 $(d, {}^{2}J_{H,H} = 18.5 \text{ Hz}, 1 \text{ H}), 3.34 (d, {}^{2}J_{H,H} = 18.5 \text{ Hz}, 1 \text{ H}), 1.25 (t, t)$ ${}^{3}J_{\text{H,H}} = 7 \text{ Hz}, 3 \text{ H}$) ppm. ${}^{13}\text{C}$ NMR (CDCl₃, 75.4 MHz): $\delta = 169.8$, 164.8, 162.2 (t, ${}^{2}J_{C,F}$ = 31 Hz), 150.2, 141.8, 123.1, 113.1 (t, ${}^{1}J_{C,F}$ = 258 Hz), 63.6, 54.2 (dd, ${}^{2}J_{C,F}$ = 24.5, ${}^{2}J_{C,F}$ = 26 Hz), 52.7, 40.0, 13.7 ppm. ¹⁹F NMR (CDCl₃, 288.3 MHz): δ = -110.7 (dd, ³*J*_{F,H} = 8.5, ${}^{2}J_{\text{EF}} = 262 \text{ Hz}$), -114.5 (dd, ${}^{3}J_{\text{EH}} = 16$, ${}^{2}J_{\text{EF}} = 262 \text{ Hz}$) ppm.

rac-Ethyl 2,2-Difluoro-3-[3-methoxy-3-oxopropanoyl)amino]-3-(pyridin-3-yl)propanoate (26e): Racemic β-amino ester 26e was obtained from β -amino ester 15e according to the procedure already described for the conversion of 15d into 26d, with β -amino ester 15e (1.04 g, 3.9 mmol), methyl malonyl chloride (24, 0.7 mL, 6.05 mmol, 1.5 equiv.), and N,N-diisopropylethylamine (1.7 mL, 9.85 mmol, 2.5 equiv.) in anhydrous CH₂Cl₂ (10 mL). Chromatography [CH₂Cl₂/MeOH/NH₄OH (99:1:0.1 to 96:4:0.4) as eluent] gave pure amide 26e as a yellow oil; yield 0.72 g (56%). TLC: silica gel (CH₂Cl₂/MeOH/NH₄OH, 9:1:0.1), $R_f = 0.5$. ¹H NMR (CDCl₃, 300.3 MHz): δ = 8.63 (d, ${}^{4}J_{H,H}$ = 2 Hz, 1 H), 8.55 (dd, ${}^{4}J_{H,H}$ = 1.5, ${}^{3}J_{\text{H,H}} = 5$ Hz, 1 H), 8.50 (br. d, ${}^{3}J_{\text{H,H}} = 10$ Hz, 1 H), 7.72 (d, ${}^{3}J_{\text{H,H}}$ = 8 Hz, 1 H), 7.31 (dd, ${}^{3}J_{H,H}$ = 5, ${}^{3}J_{H,H}$ = 7.5 Hz, 1 H), 5.79 (dt, ${}^{3}J_{H,F} = 10, \, {}^{3}J_{H,F} = 16 \text{ Hz}, 1 \text{ H}), \, 4.26 \text{ (q, } {}^{3}J_{H,H} = 7 \text{ Hz}, 2 \text{ H}), \, 3.76$ (s, 3 H), 3.40 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 3.33 (d, ${}^{2}J_{H,H}$ = 18 Hz, 1 H), 1.26 (t, ${}^{3}J_{H,H}$ = 7 Hz, 3 H) ppm. ${}^{13}C$ NMR (CDCl₃, 75.4 MHz): $\delta = 169.9, 164.6, 162.3 \text{ (dd, } {}^{2}J_{\text{C,F}} = 31, {}^{2}J_{\text{C,F}} = 32.5 \text{ Hz}$), 150.3, 149.7, 135.8, 129.0, 123.6, 113.3 (t, ${}^{1}J_{C,F} = 257$ Hz), 63.6, 53.1 (dd, ${}^{2}J_{C,F} = 25, {}^{2}J_{C,F} = 27$ Hz), 52.7, 40.0, 13.8 ppm. ${}^{19}F$ NMR (CDCl₃, 288.3 MHz): $\delta = -110.7$ (dd, ${}^{3}J_{EH} = 8.5$, ${}^{2}J_{EF} = 262$ Hz), -114.5(dd, ${}^{3}J_{F,H} = 16$, ${}^{2}J_{F,F} = 262$ Hz) ppm. IR (KBr): $\tilde{v}_{max} = 3293$, 1770, 1748, 1671, 1550, 1330, 1292, 1203, 1133, 1073, 713 cm⁻¹.

rac-3-[(Carboxyacetyl)amino]-2,2-difluoro-3-(pyridin-4-yl)propanoic Acid (28d): A solution of amide 26d (0.54 g, 1.6 mmol) in a mixture of MeOH/H₂O (1:1, 10 mL) was treated with aqueous NaOH solution (1 N, 1.65 mL, 1 equiv.). The reaction mixture was stirred at room temp. for 2 h. Most of the solvent was evaporated, and the mixture was washed with diethyl ether (2×30 mL) and acidified with aqueous HCl solution (1 N) until pH = 1 was reached. The aqueous phase was washed with diethyl ether (30 mL) and



lyophilized. The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/ H₂O/HCl (1 N, 200:200:4) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound **28d** as a yellow lyophilizate; yield 93 mg (20%). ¹H NMR (D₂O, 300.3 MHz): δ = 8.78 (d, ³*J*_{H,H} = 6.5 Hz, 2 H), 8.10 (d, ³*J*_{H,H} = 6.5 Hz, 2 H), 5.80 (t, ³*J*_{H,F} = 13 Hz, 1 H), 3.56 (d, ²*J*_{H,H} = 16.5 Hz, 1 H), 3.47 (d, ²*J*_{H,H} = 16.5 Hz, 1 H) ppm. ¹³C NMR (D₂O, 75.4 MHz): δ = 169.0, 166.3, 163.4 (t, ²*J*_{C,F} = 31 Hz), 154.9, 141.7, 127.1, 113.7 (t, ¹*J*_{C,F} = 255.5 Hz), 56.0 (dd, ²*J*_{C,F} = 24, ²*J*_{C,F} = 27.5 Hz), 42.0 ppm. ¹⁹F NMR (D₂O, 288.3 MHz): δ = -110.9 (dd, ³*J*_{F,H} = 13, ²*J*_{F,F} = 256 Hz), −111.8 (dd, ²*J*_{F,F} = 13, ²*J*_{F,F} = 256 Hz) ppm. IR (KBr): $\tilde{v}_{max} = 3404$, 2360, 1756, 1669, 1547, 1409, 1325, 1194, 682 cm⁻¹. MS (ESI+): m/z = 288.9 [M + H]⁺.

rac-3-[(Carboxyacetyl)amino]-2,2-difluoro-3-(pyridin-3-yl)propanoic Acid (28e): Racemic β-amino acid 28e was obtained from β-amino ester 26e according to the procedure already described for the conversion of 26d into 28d, from β-amino ester 26e (0.65 g, 1.95 mmol) and an aqueous LiOH solution (1 N, 6 mL, 6 mmol, 3 equiv.) in a mixture of MeOH/THF/H2O (1:1:1, 18 mL). The crude product was purified by steric exclusion chromatography on a column of Biogel P2 (Bio-Rad) with MeCN/H2O/HCl (1 N, 200:200:4) as the solvent. Fractions containing only product were pooled, and the solvent was removed under vacuum. Lyophilization yielded the hydrochloride salt of the title compound 28e as a white lyophilizate; yield 112 mg (20%). ¹H NMR (D₂O, 300.3 MHz): δ = 8.82 (s, 1 H), 8.75 (d, ${}^{3}J_{H,H}$ = 6 Hz, 1 H), 8.62 (d, ${}^{3}J_{H,H}$ = 8.5 Hz, 1 H), 8.06 (dd, ${}^{3}J_{H,H} = 6$, ${}^{3}J_{H,H} = 8.5$ Hz, 1 H), 5.79 (t, ${}^{3}J_{H,F} = 13$ Hz, 1 H), 3.50 (d, ${}^{2}J_{H,H}$ = 16.5 Hz, 1 H), 3.41 (d, ${}^{2}J_{H,H}$ = 16.5 Hz, 1 H) ppm. ¹H NMR ([D₆]DMSO, 300.3 MHz): δ = 9.39 (d, ³J_{H,H} = 9.5 Hz, 1 H), 8.86 (s, 1 H), 8.76 (d, ${}^{3}J_{H,H}$ = 5 Hz, 1 H), 8.26 (d, ${}^{3}J_{H,H}$ = 8 Hz, 1 H), 7.78 (dd, ${}^{3}J_{H,H} = 5$, ${}^{3}J_{H,H} = 8$ Hz, 1 H), 5.84 (dt, ${}^{3}J_{H,H}$ $= {}^{3}J_{H,F} = 10, {}^{3}J_{H,F} = 18$ Hz, 1 H), 3.32 (d, ${}^{2}J_{H,H} = 15.5$ Hz, 1 H), 3.25 (d, ${}^{2}J_{H,H}$ = 15.5 Hz, 1 H) ppm. ${}^{13}C$ ([D₆]DMSO, 75.4 MHz): $\delta = 169.0, 166.3, 163.4$ (t, ${}^{2}J_{C,F} = 31$ Hz), 145.1, 144.5, 142.8, 132.9, 126.2, 113.7 (t, ${}^{1}J_{C,F}$ = 255.5 Hz), 52.0 (dd, ${}^{2}J_{C,F}$ = 24, ${}^{2}J_{C,F}$ = 27.5 Hz), 42.6 ppm. ¹⁹F NMR ([D₆]DMSO, 288.3 MHz): δ = -109.8 (dd, ${}^{3}J_{F,H} = 9.5$, ${}^{2}J_{F,F} = 254.5$ Hz), -114.6 (dd, ${}^{2}J_{F,F} = 19$, ${}^{2}J_{\text{E,F}} = 254.5 \text{ Hz}$) ppm. MS (ESI+): $m/z = 310.9 \text{ [M + Na]}^{+}$, 288.9 $[M + H]^+$. $C_{11}H_{10}F_2N_2O_5$ ·HCl (324.67): C 40.69, H 3.42, N 8.63; found C 40.64, H 3.37, N 8.66.

Acknowledgments

We gratefully acknowledge financial support (grant to N. B.) from the Institut de Recherches SERVIER. We also warmly thank Louise Chervel, Maryse Thiverny, Laëtitia Bailly and Elisabeth Roger for their initial contribution to this work.

- a) S. Purser, P. R. Moore, S. Swallow, V. Gourverneur, *Chem. Soc. Rev.* 2008, *37*, 320–330; b) J.-P. Bégué, D. Bonnet-Delpon in *Chimie bioorganique et médicinale du fluor*, CNRS Editions, Paris, 2005; c) K. Mikami, Y. Itoh, M. Yamanaka, *Chem. Rev.* 2004, *104*, 1–16.
- [2] For a general discussion of biological activity and the importance of fluorinated amino compounds, see: a) V. A. Soloshonok (Ed.) in *Enantiocontrolled Synthesis of Fluoro-Organic Compounds: Stereochemical Challenges and Biomedicinal Targets*, Wiley, Chichester, UK, **1999**; b) I. Ojima, J. R. McCarthy, J. T. Welch (Eds.) in *Biomedical Frontiers of Fluorine Chemistry*, ACS Books, American Chemical Society, Washington, DC, **1996**; c) J. T. Welch, S. Eswarakrischnan in *Fluorine in Bioorganic Chemistry*, Wiley-VCH, New York, **1991**.

- [3] For a general discussion of biological activity and the importance of fluorinated compounds, see: a) C. J. Thomas, *Curr. Top. Med. Chem.* **2006**, *6*, 1529–1543; b) K. L. Kirk, *J. Fluorine Chem.* **2006**, *127*, 1013–1029.
- [4] a) B. E. Smart, J. Fluorine Chem. 2001, 109, 3; b) M. Schlosser, Angew. Chem. Int. Ed. 1998, 37, 1496–1513; c) B. E. Smart, J. C. Tatlow in Organofluorine Chemistry – Principles and Commercial Applications, Plenum Press, New York, 1994.
- [5] a) M. Shimizu, T. Hiyama, Angew. Chem. Int. Ed. 2005, 44, 214–231; b) J.-A. Ma, D. Cahard, Chem. Rev. 2004, 104, 6119–6146; c) T. Hiyama in Organofluorine Compounds: Chemistry and Applications (Ed.: H. Yamamoto), Springer, Berlin, 2000; d) C. Dollery in Therapeutic Drugs, Churchill Livingstone, Edinburgh, U.K., 1999.
- [6] a) R. P. Cheng, S. H. Gellman, W. F. DeGrado, *Chem. Rev.* 2001, 101, 3219–3232; b) A. E. Barron, R. N. Zuckermann, *Curr. Opin. Chem. Biol.* 1999, 3, 681–687; c) K. D. Stigers, M. J. Soth, J. S. Nowick, *Curr. Opin. Chem. Biol.* 1999, 3, 714–723; d) S. H. Gellman, *Acc. Chem. Res.* 1998, 31, 173–180.
- [7] E. Juaristi, D. Quintana, J. Escalante, *Aldrichim. Acta* **1994**, *27*, 3–11.
- [8] a) T. C. Boge, G. I. Georg in *Enantioselective Synthesis of β-Amino Acids*, (Ed: E. Juaristi), Wiley-VCH, New York, **1996**, pp. 1–43; b) S. Fustero, J. F. Sanz-Cervera, V. A. Soloshonok in *Enantioselective Synthesis of β-Amino Acids*, 2nd ed. (Eds: E. Juaristi, V. A. Soloshonok), Wiley-VCH, New York, **2005**, pp. 319–350.
- [9] a) D. Seebach, D. F. Hook, A. Glättli, *Pept. Sci.* 2005, 84, 23–37; b) D. Seebach, J. L. Matthews, *Chem. Commun.* 1997, 2015–2022.
- [10] D. Seebach, S. Abele, J. V. Schreiber, B. Martinoni, A. K. Nussbaum, H. Schild, H. Schulz, H. Hennecke, R. Woessner, F. Bitsch, *Chimia* **1998**, *52*, 734–739.
- [11] M. Benaglia, M. Cinquini, F. Cozzi, Eur. J. Org. Chem. 2000, 563–572.
- [12] a) J. D. Sadowsky, J. K. Murray, Y. Tomita, S. H. Gellman, *ChemBioChem* 2007, *8*, 903–916; b) D. H. Appella, L. A. Christianson, I. L. Karle, D. R. Powell, S. H. Gellman, *J. Am. Chem. Soc.* 1996, *118*, 13071–13072.
- [13] See, for example: a) G. Shi, W. Cai, J. Org. Chem. 1995, 60, 6289-6295; b) P. D. Edwards, D. W. Andisik, C. A. Bryant, B. Ewing, B. Gomes, J. J. Lewis, D. Rakiewicz, G. Steelman, A. Strimpler, D. A. Trainor, P. A. Tuthill, R. C. Mauger, C. A. Veale, R. A. Wildonger, J. C. Williams, D. J. Wolanin, M. Zottola, J. Med. Chem. 1997, 40, 1876-1885; c) V. A. Soloshonok, I. V. Soloshonok, V. P. Kukhar, V. K. Svedas, J. Org. Chem. 1998, 63, 1878-1884; d) K. Uneyama, J. Hao, H. Amii, Tetrahedron Lett. 1998, 39, 4079-4082; e) J. M. Percy, M. E. Prime, J. Org. Chem. 1998, 63, 8049-8051; f) R. J. Cregge, S. L. Durham, R. A. Farr, S. L. Gallion, C. M. Hare, R. V. Hoffman, M. J. Janusz, H.-O. Kim, J. R. Koehl, S. Mehdi, W. A. Metz, N. P. Peet, J. T. Pelton, H. A. Schreuder, S. Sunder, C. Tardif, J. Med. Chem. 1998, 41, 2461–2480; g) A. Sutherland, C. L. Wilis, Nat. Prod. Rep. 2000, 17, 621-631; h) N. N. Sergeeva, A. S. Golubev, K. Burger, Synthesis 2001, 281-285; i) N. Fokina, A. Kornilov, I. Kulik, V. Kukhar, Synthesis 2002, 17, 2589-2596; j) X.-L. Qiu, W.-D. Meng, F.-L. Qing, Tetrahedron 2004, 60, 6711–6745; k) K. Konno, M. Kanda, T. Ishihara, H. Yamanaka, J. Fluorine Chem. 2005, 126, 1517-1523; 1) R. Smits, B. Koksch, Curr. Top. Med. Chem. 2006, 6, 1483-1498; m) F. Huguenot, T. Brigaud, J. Org. Chem. 2006, 71, 2159–2162; n) G. Li, W.A. van der Donk, Org. Lett. 2007, 9, 41-44; o) V. A. Soloshonok, A. G. Kirilenko, S. V. Galushko, V. P. Kukhar, Tetrahedron Lett. 1994, 35, 5063-5064; p) V. A. Soloshonok, A. G. Kirilenko, V. P. Kukhar, G. Resnati, Tetrahedron Lett. 1993, 34, 3621-3624; g) V. A. Soloshonok, A. G. Kirilenko, N. A. Fokina, I. P. Shishikina, S. V. Galushko, V. P. Kukhar, V. K. Svedas, E. Kozlova, Tetrahedron: Asymmetry 1994, 5, 1225-1228; r) V. A. Soloshonok, N. A. Fokina, A. V. Ryabakova, I. P. Shishkina, S. V. Galushko, A. E. Sorochinsky, V. P. Kukhar, M. V.

Savchenko, V. K. Svedas, *Tetrahedron: Asymmetry* **1995**, *6*, 1601–1610; s) V. A. Soloshonok, T. Ono, *Tetrahedron* **1996**, *52*, 14701–14712; t) V. A. Soloshonok, I. V. Soloshonok, T. Ono, V. K. Svedas, J. Org. Chem. **1997**, *62*, 7538–7539; u) V. A. Soloshonok, V. P. Kukhar, *Tetrahedron* **1997**, *53*, 8307–8314.

- [14] N. Sewald, K. Burger in *Fluorine-Containing Amino Acids Synthesis and properties* (Eds: V. P. Kukhar, V. A. Soloshonok), John Wiley and Sons Ltd., Chichester, UK, **1995**.
- [15] M. Tozer, T. Herpin, Tetrahedron 1996, 52, 8619-8683.
- [16] D. Schirlin, S. Baltzer, J. M. Altenburger, C. Tarnus, J. M. Remy, *Tetrahedron* 1996, 52, 305–318.
- [17] T. Ohba, E. Ikeda, H. Takei, *Bioorg. Med. Chem. Lett.* 1996, 6, 1875–1880.
- [18] a) S. Thaisrivongs, H. Schostarez, D. T. Pals, S. R. Turner, J. Med. Chem. 1987, 30, 1837–1842; b) M. R. Angelastro, P. Bey, S. Mehdi, N. P. Peet, Bioorg. Med. Chem. Lett. 1992, 2, 1235–1238.
- [19] a) A. Cheguillaume, S. Lacroix, J. Marchand-Brynaert, *Tetrahedron Lett.* 2003, 44, 2375–2377; b) A. Cheguillaume, J. Gillart, D. Labar, V. Grégoire, J. Marchand-Brynaert, *Bioorg. Med. Chem.* 2005, 13, 1357–1367.
- [20] a) K. Nakayama, H. C. Kawato, H. Inagaki, R. Nakajima, A. Kitamura, K. Someya, T. Ohta, *Org. Lett.* 2000, *2*, 977–980; b) H. Agematsu, H. Chiba, R. Kaneto, T. Ohta, K. Nakayama, WO9526978, 1995.
- [21] a) K. Uoto, S. Ohsuki, H. Takenoshita, T. Ishiyama, S. Iimura, Y. Hirota, I. Mitsui, H. Terasawa, T. Soga, *Chem. Pharm. Bull.* **1997**, *45*, 1793–1804; b) H. Terasawa, T. Soga, K. Uoto, JP07233159, **1995**.
- [22] D. F. Hook, F. Gessier, C. Noti, P. Kast, D. Seebach, *ChemBioChem* 2004, 5, 691–706.
- [23] a) C. Palomo, J. M. Aizpurua, I. Ganboa, M. Oiarbide, *Eur. J. Org. Chem.* **1999**, 3223–3235; b) G. S. Singh, *Tetrahedron* **2003**, 59, 7631–7649; c) S. France, A. Weatherwax, A. E. Taggi, T. Lectka, *Acc. Chem. Rev.* **2004**, *37*, 592–600; d) C. Palomo, J. M. Aizpurua, I. Ganboa, M. Oiarbide, *Curr. Med. Chem.* **2004**, *11*, 1837–1872.
- [24] a) R. Joyeau, H. Molines, R. Labia, M. Wakselman, J. Med. Chem. 1988, 31, 370–374; b) M. Wakselman, R. Joyeau, R. Kobaiter, N. Boggetto, I. Vergely, J. Maillard, V. Okochi, J. J. Montagne, M. Reboud-Ravaux, FEBS Lett. 1991, 282, 377– 381; c) I. Vergely, N. Boggetto, V. Okochi, S. Golpayegani, M. Reboud-Ravaux, R. Kobaiter, R. Joyeau, M. Wakselman, Eur. J. Med. Chem. 1995, 30, 199–208; d) S. Lacroix, A. Cheguillaume, S. Gérard, J. Marchand-Brynaert, Synthesis 2003, 16, 2483–2486.
- [25] a) J. E. Baldwin, G. P. Lynch, C. J. Schofield, *Tetrahedron* 1992, 48, 9085–9100; b) M. P. Doyle, W. Hu, *J. Org. Chem.* 2000, 65, 8839–8847; c) M. P. Doyle, I. M. Phillips, W. Hu, *J. Am. Chem. Soc.* 2001, 123, 5366–5367; d) M. P. Doyle, W. Hu, I. Phillips, WO2002045853, 2002.
- [26] A. Otaka, J. Watanabe, A. Yukimasa, Y. Sasaki, H. Watanabe, T. Kinoshita, S. Oishi, H. Tamamura, N. Fujii, J. Org. Chem. 2004, 69, 1634–1645.
- [27] a) B. Hulin, J. C. Parker, WO2003101958A2, 2003; b) W. Li, E. Oliver Jr, C. Rojas, V. Kalish, S. Belyakov, WO2004071454A2, 2004; c) J. L. Duffy, R. J. Mathvink, A. E. Weber, J. Xu, WO2004050022A2, 2004; d) V. J. Colandrea, S. D. Edmondson, R. J. Mathvink, A. Mastracchio, A. E. Weber, J. Xu, WO2004043940A1, 2004; e) J. C. Parker, B. Hulin, US2005043292A1, 2005.
- [28] A. E. Weber, J. Med. Chem. 2004, 47, 4135–4141.
- [29] a) I. Ojima, Acc. Chem. Res. 1995, 28, 383–389; b) I. Ojima, F. Delaloge, Chem. Soc. Rev. 1997, 26, 377–386; c) B. Alcaide, P. Almendros, Curr. Med. Chem. 2004, 11, 1921–1949; d) B. Alcaide, P. Almendros, C. Aragoncillo, Chem. Rev. 2007, 107, 4437–4492.
- [30] a) R. Déziel, E. Malenfant, *Bioorg. Med. Chem. Lett.* 1998, *8*, 1437–1442; b) C. Yoakim, W. Ogilvie, D. R. Cameron, C. Chabot, I. Guse, B. Haché, J. Naud, J. A. O'Meara, R. Plante,

R. Déziel, J. Med. Chem. 1998, 41, 2882–2891; c) W. Ogilvie, C. Yoakim, F. Do, B. Haché, L. Lagacé, J. Naud, J. A. O'Meara, R. Déziel, *Bioorg. Med. Chem.* 1999, 7, 1521–1531; d) P. Bonneau, F. Hasani, C. Plouffe, E. Malenfant, S. R. Laplante, I. Guse, W. W. Ogilvie, R. Plante, W. C. Davidson, J. L. Hopkins, M. M. Morelock, M. G. Cordingley, R. Déziel, J. Am. Chem. Soc. 1999, 121, 2965–2973.

- [31] a) D. A. Burnett, M. A. Caplen, M. A. Davis Jr, R. E. Burrier, J. W. Clader, J. Med. Chem. 1994, 37, 1733–1736; b) A. Zarks, S. Chackalamamil, S. Dugar, J. Org. Chem. 1996, 61, 8341–8343; c) W. D. Vaccaro, H. R. Davis, Bioorg. Med. Chem. Lett. 1998, 8, 313–318; d) R. Annunziata, M. Bengalia, M. Cinquini, F. Cozzi, Tetrahedron: Asymmetry 1999, 10, 4841–4849.
- [32] a) A. Fürstner, Synthesis 1989, 571–590; b) R. Ocampo, W. R. Dolbier Jr, Tetrahedron 2004, 60, 9325–9374.
- [33] D. J. Hart, D.-C. Ha, Chem. Rev. 1989, 89, 1447-1465.
- [34] a) b) H. Agematsu, H. Chiba, R. Kaneto, T. Ohta, K. Nakayama WO9526978, **1995**, K. Nakayama, H. C. Kawato, H. Inagaki, R. Nakajima, A. Kitamura, K. Someya, T. Ohta, *Org. Lett.* **2000**, *2*, 977–980; c) H. Amii, T. Kobayashi, K. Uneyama, *Synthesis* **2000**, *14*, 2001–2003; d) M. Bordeau, F. Frébault, M. Gobet, J.-P. Picard, *Eur. J. Org. Chem.* **2006**, 4147–4154.
- [35] N. Fokina, A. Kornilov, V. Kukhar, J. Fluorine Chem. 2001, 111, 69–76.
- [36] a) K. Iseki, Y. Kuroki, D. Asada, Y. Kobayashi, *Tetrahedron Lett.* 1997, 38, 1447–1448; b) K. Iseki, Y. Kuroki, D. Asada, M. Takahashi, S. Kishimoto, Y. Kobayashi, *Tetrahedron* 1997, 53, 10271–10280; c) A. Katritzky, D. Nichols, M. Qi, *Tetrahedron Lett.* 1998, 39, 7063–7066; d) K. Sato, A. Tarui, S. Matsuda, M. Omote, A. Ando, I. Kumadaki, *Tetrahedron Lett.* 2005, 46, 7679–7681.
- [37] a) O. Kitagawa, T. Taguchi, Y. Kobayashi, *Tetrahedron Lett.* 1988, 29, 1803–1806; b) T. Taguchi, O. Kitagawa, Y. Suda, S. Ohkawa, A. Hashimoto, Y. Iitaka, Y. Kobayashi, *Tetrahedron Lett.* 1988, 29, 5291–5294; c) A. Vidal, A. Nefzi, R. A. Houghten, J. Org. Chem. 2001, 66, 8268–8272; d) D. D. Staas, K. L. Savage, C. F. Homnick, N. N. Tsou, R. G. Ball, J. Org. Chem. 2002, 67, 8276–8279; e) V. A. Soloshonok, H. Ohkura, A. Sorochinsky, N. Voloshin, A. Markovsky, M. Belik, T. Yamazaki, *Tetrahedron Lett.* 2002, 43, 5445–5448; f) A. Sorochinsky, N. Voloshin, A. Markovsky, M. Belik, N. Yasuda, H. Uekusa, T. Ono, D. O. Berbasov, V. A. Soloshonok, J. Org. Chem. 2003, 68, 7448–7454.
- [38] a) S. Marcotte, X. Pannecoucke, C. Feasson, J.-C. Quirion, J. Org. Chem. 1999, 64, 8461–8464; b) N. Boyer, P. Gloanec, G. de Nanteuil, P. Jubault, J.-C. Quirion, *Tetrahedron* 2007, 63, 12352–12366.
- [39] M. D. Sørensen, N. M. Khalifa, E. B. Pedersen, Synthesis 1999, 11, 1937–1943.
- [40] a) P. C. Ting, J. F. Lee, J. C. Anthes, N.-Y. Shih, J. J. Piwinski, *Bioorg. Med. Chem. Lett.* **2001**, *11*, 491–494; b) Y. Yoneda, S. Kawajiri, M. Sugimura, K. Osanai, F. Kito, E. Ota, T. Mimura, *Bioorg. Med. Chem. Lett.* **2001**, *11*, 2663–2666.
- [41] O. Ouari, F. Chalier, R. Bonaly, B. Pucci, P. Tordo, J. Chem. Soc. Perkin Trans. 2 1998, 2299–2307.
- [42] a) J. Lee, J. Lee, M. Kang, M. Shin, J.-M. Kim, S.-U. Kang, J.-O. Lim, H.-K. Choi, Y.-G. Suh, H.-G. Park, U. Oh, H.-D. Kim, Y.-H. Park, H.-J. Ha, Y.-H. Kim, A. Toth, Y. Wang, R. Tran, L. V. Pearce, D. J. Lundberg, P. M. Blumberg, *J. Med. Chem.* 2003, 46, 3116–3126; b) G. Durand, A. Polidori, O. Ouari, P. Tordo, V. Geromel, P. Rustin, B. Pucci, *J. Med. Chem.* 2003, 46, 5230–5237.
- [43] A. K. Awasthi, M. L. Boys, K. J. Cain-Janicki, P.-J. Colson, W. W. Doubleday, J. E. Duran, P. N. Farid, *J. Org. Chem.* 2005, 70, 5387–5397.
- [44] H. Adams, J. C. Anderson, S. Peace, A. M. K. Pennell, J. Org. Chem. 1998, 63, 9932–9934.
- [45] a) A. R. Katritzky, B. V. Rogovoy, *Chem. Eur. J.* 2003, *9*, 4586–4593; b) A. R. Katritzky, K. Manju, S. K. Singh, N. K. Meher, *Tetrahedron* 2005, *61*, 2555–2581.



- [46] G. Brieger, T. J. Nestrick, Chem. Rev. 1974, 74, 567-580.
- [47] P. G. Mattingly, M. J. Miller, J. Org. Chem. 1981, 46, 1557– 1564.
- [48] a) G. A. Brown, S. R. Martel, R. Wisedale, J. P. H. Charmant, N. J. Hales, C. W. G. Fishwick, T. Gallagher, J. Chem. Soc. Perkin Trans. 1 2001, 1281–1289; b) E. Cesarotti, I. Rimoldi, Tetrahedron: Asymmetry 2004, 15, 3841–3845.
- [49] Y. Du, D. F. Wiemer, J. Org. Chem. 2002, 67, 5709-5717.
- [50] a) Z. Kaluza, S.-H. Park, Synlett **1996**, 895–896; b) I. N. Simpson, C. J. Urch, G. Hagen, R. Albrecht, B. Sprinkart, H. R. Pfaendler, J. Antibiot. **2003**, 56, 838–847.
- [51] a) M. W. Carland, R. L. Martin, C. H. Schiesser, *Tetrahedron Lett.* 2001, 42, 4737–4739; b) M. W. Carland, R. L. Martin, C. H. Schiesser, *Org. Biomol. Chem.* 2004, 2, 2612–2618.
- [52] G. Cainelli, P. Galletti, S. Garbisa, D. Giacomini, L. Sartor, A. Quintavalla, *Bioorg. Med. Chem.* 2003, 11, 5391–5399.
- [53] a) P. R. Guzzo, M. J. Miller, J. Org. Chem. 1994, 59, 4862–4867; b) A. Macias, A. M. Ramallal, E. Alonso, C. del Pozo, J. Gonzalez, J. Org. Chem. 2006, 71, 7721–7730.

- [54] T. B. Durham, M. J. Miller, J. Org. Chem. 2003, 68, 27-34.
- [55] Y. Takahashi, H. Yamashita, S. Kobayashi, M. Ohno, *Chem. Pharm. Bull.* **1986**, *34*, 2732–2742.
- [56] D. Reuschling, H. Pietsch, A. Linkies, *Tetrahedron Lett.* 1978, 19, 615–618.
- [57] W. Van Brabandt, N. De Kimpe, Synlett 2006, 13, 2039–2042.
- [58] This compound is readily prepared under S_N2 conditions; see:
 a) L. Bauer, K. S. Suresh, B. K. Ghosh, J. Org. Chem. 1965, 30, 949–951; b) C. Savarin, J. Srogl, L. S. Liebeskind, Org. Lett. 2000, 2, 3229–3231; c) E. Block, E. V. Dikarev, R. S. Glass, J. Jin, B. Li, X. Li, S.-Z. Zhang, J. Am. Chem. Soc. 2006, 128, 14949–14961.
- [59] D. Appleton, A. B. Duguid, S.-K. Lee, Y.-J. Ha, H.-J. Hab, F. J. Leeper, J. Chem. Soc. Perkin Trans. 1 1998, 89–101.

Received: April 9, 2008 Published Online: July 10, 2008