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Article

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Structure-Activity Relationship of Pyrrolyl Diketo Acid Derivatives as Dual Inhibitors of HIV-1 Integrase and Reverse Transcriptase Ribonuclease H Domain

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KEYWORDS: HIV, integrase, reverse transcriptase, ribonuclease H, dual inhibitors, pyrrole, diketoacid.

ABSTRACT. The development of HIV-1 dual inhibitors is a highly innovative approach aimed to reduce drug toxic side effects as well as therapeutic costs. HIV-1 integrase (IN) and reverse transcriptase-associated ribonuclease H (RNase H) are both selective targets for HIV-1 chemotherapy and the identification of dual IN/RNase H inhibitors is an attractive strategy for new drug development. We synthesized newly pyrrolyl derivatives that exhibited good potency against IN and a moderate inhibition of the RNase H function of RT, confirming the possibility to develop dual HIV-1 IN/RNase H inhibitors and obtaining new information for the further development of more effective dual HIV-1 inhibitors.

INTRODUCTION

The development of dual-action drugs is a prosiming approach to ameliorate drug-drug interactions, reduce toxic side effects still suppressing viral resistance selection.^{1,2,3,4} Among dual-action drugs, dual inhibitors are single compounds which are able to inhibit two enzyme activities. Several reports have shown that dual inhibitors may have a role in the treatment of different diseases such as Alzheimer,⁵ Parkinson,⁶ inflammation⁷ and cancer.^{1,8,9} This approach had been attempted also in the virological arena aiming to inhibit rhinovirus replication.¹⁰ Recently, tropolones,^{11,12,13} madurahydroxylactone¹⁴ and 2-hydroxyisoquinolin-1,3(2*H*,4*H*)-diones^{15,16} have been reported to act as dual inhibitors against HIV-1, targeting viral integrase (IN) and reverse transcriptase (RT)- ribonuclease H (RNase H) activities.

HIV-1 IN is the viral enzyme responsible for the integration of the proviral dsDNA into the cell host chromosome through two coordinated enzyme functions, both accomplished by the same active site.¹⁷ In the first reaction, termed 3'-end processing (3'-P), IN removes the two terminal nucleotides (GT) from each 3'-end of the dsDNA.¹⁷ In the second reaction, termed strand-transfer (ST), IN catalyzes a nucleophilic attack by the free 3'-OH of the viral processed DNA to the target chromosomal DNA, resulting in covalent joining of the two molecules. Several classes of integrase inhibitors have been identified;¹⁸ among these the diketoacids (DKAs) showed greatest promise and the first DKA bioisoster, raltegravir (1), has been approved in 2008 for HIV-1 therapy.¹⁹

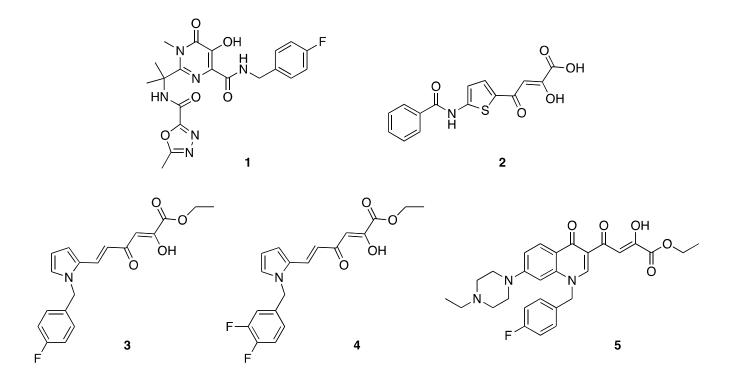
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HIV-1 IN belongs to the functionally diverse superfamily of DDE(D) nucleotidyltransferases, whose other notable members include RNaseH and MuA, Tn5, Mos1 transposases as well.¹⁷ The active sites of these enzymes typically contain three essential carboxylates that coordinate a pair of divalent metal cations (usually Mg²⁺). Thus, chelating inhibitors can be active across several classes of viral metal-dependent enzymes and chelation has been successfully used in drug design, also of dual inhibitors.²⁰ In particular, DKAs have been reported to chelate the divalent cations in the IN active site^{17,18} and notably, DKAs originally developed against HIV-1 IN have been also reported to inhibit the HIV-1 RNase H.^{21,22} The HIV-1 RT-associated RNase H function hydrolizes the RNA strand of the replicative intermediate RNA:DNA hybrid and, hence, is essential for viral replication.²³ Even though several compounds have been approved for HIV therapy. Therefore, this viral function is a very attractive target for drug development.

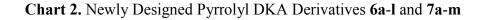
The first DKA IN inhibitor later described also as RNase H inhibitor was the 4-[5-(benzoylamino)thien-2-yl]-2,4-dioxobutanoic acid (BTDBA, **2**) discovered by Merck.²¹ Recently we reported that 6-[1-(4-fluorophenyl)methyl-1*H*pyrrol-2-yl)]-2,4-dioxo-5-hexenoic acid ethyl ester (RDS 1643, **3**) inhibited the HIV-1 RT-associated RNase H function in biochemical assays with an IC₅₀ value of 8 μ M, the HIV-1 IN ST (the IC₅₀ value was 98 μ M) and blocked the HIV-1 replication in cell based assays with an EC₅₀ value of <0.2 μ M (Chart 1).²² Starting from these observations, more recently we designed a small library of **3** analogues with the aim to obtain dual IN/RNase H HIV-1 inhibitors and found compounds active at micromolar concetration against RNase H and low nanomolar IC₅₀ values against IN in recombinant assays.³¹ The best dual inhibition was found for compounds showing a diketo ester group and fluorine atoms as substituents on the benzyl portion, as exemplified by compound **4** (Chart 1, IC_{50,IN} = 0.6 μ M, IC_{50,RH} = 3.0 μ M EC₅₀ = 2 μ M). Further, we obtained similar results with a quinolonyl diketo acid series, in which less marked dual activity was found, having in the best case IC_{50,IN} = 26.2 μ M, IC_{50,RH} = 2.4 μ M and EC₅₀ = 3.6 μ M (compound **5**).³²

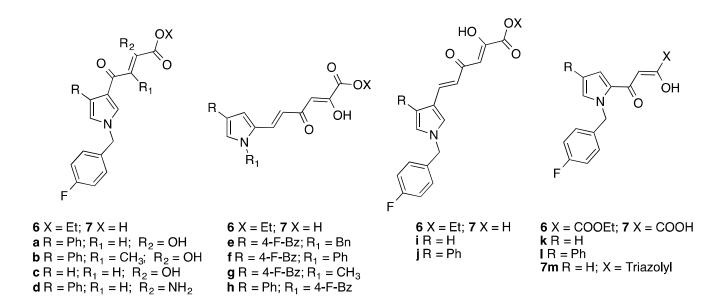
Herein we present the design, synthesis and biological evaluation of new compounds related to **3**, to better define the structure-activity relationship (SAR) within the most interesting pyrrolyl diketo acid series. All the newly designed pyrrolyl DKA both ester **6** and acid derivatives **7** are shown in Chart 2. Basically, starting from **3** we fixed the pyrrole ring and the DKA chain while wider tranformations included one or more of the following modifications: i) introduction of aromatic substituent in position 4 of the pyrrole ring; ii) shift of the diketo hexenoic chain from 2 to 3 position of the pyrrole moiety; iii) shortening of the diketo hexenoic branch into a diketo butanoic group; iv) introduction of alkyl or aryl group replacing the fluorobenzyl moiety; v) replacement of carboxylic function with a triazole ring; vi) introduction of alkyl group within the DKA branch; vii) replacement of keto group of DKA moiety with NH₂ function. Notably, among the compounds described in this paper, **7k** has been the first DKA derivative reported by Merck as selective ST IN inhibitor.³³ We decided to include this compound in this study to define its properties as dual inhibitor of IN and RNase H.

Chart 1. Selective Inhibitor of IN Enzyme (1), first Described Dual IN/RNase H Inhibitor (2), and Recently Discovered Dual IN/RNase H Inhibitors 3-5



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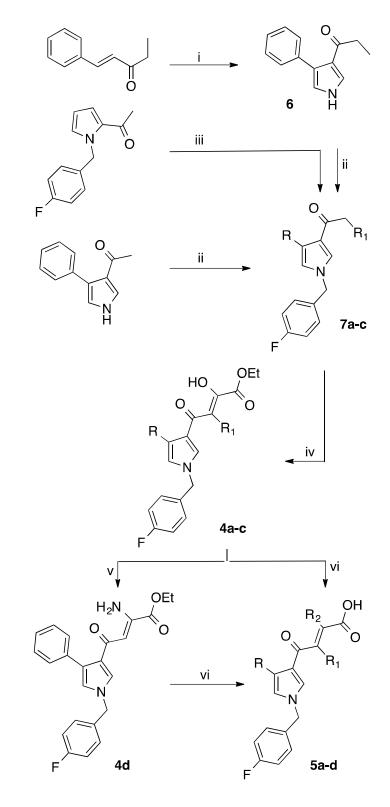




RESULT AND DISCUSSION

Chemistry. The synthesis of derivatives **6a-1** and **7a-m** is outlined in Schemes 1-4. Derivatives **6a-d** and **7a-d** were synthesized according to the pathway described in the Scheme 1. The acetyl or propionyl pyrrole intermediates **9a-c** were obtained by two different procedures: 1) the alkylation with 4-fluorobenzyl bromide in alkaline medium (K_2CO_3) of 1-(4-phenyl-1H-pyrrol-3-yl)ethanone³⁴ or derivatives **8**, achieved *via* toluene-4-sulphonylmethyl isocyanide (TosMIC) reaction from (E)-1-phenylpent-1-en-3-one,³⁵ 2) termal transposition of the acetyl chain of 2-acetyl-1-[(4-fluorophenyl)methyl]pyrrole³⁶ from 2- to 3- position of the pyrrole ring in the presence of CF₃COOH. Derivatives **9a-c** were condensed in turn with diethyl oxalate in the presence of sodium ethoxide to provide the diketobutanoic ethyl esters **6a-c**. Compound **6a** was used as substrate to provide i) **6d**, obtained by reacting the enol **6a** with ammonium acetate in acid medium (CH₃COOH) following a known procedure reported for DKA derivarives³⁷, and ii) **7a**, achieved by hydrolysis of ester **6a** in the presence of 6 N NaOH. The last conditions have been also used to obtain **7b-c** starting from **6b-c**, respectively. A sligthly different condition has been used to obtain **7d**, as previously decribed.³⁷

Scheme 1. Synthetic Route to Pyrrolyl DKAs 6a-d and 7a-d^a

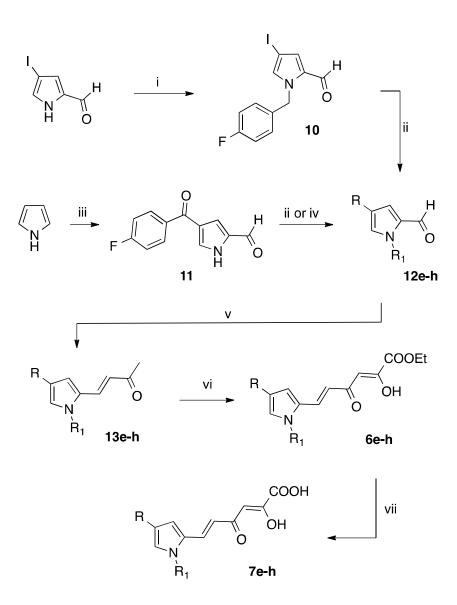


^{*a*} Reagents and conditions: (i) TosMIC, NaH, Et₂O/DMSO, room temp, 1 h; (ii) Trifluoroacetic acid, 80 °C, 24 h; (iii) 4-F-benzyl bromide, K₂CO₃, DMF, 100 °C, 24 h; (iv) diethyl oxalate, C₂H₅ONa, THF, room temp, 2 h; (v) CH₃COONH₄, benzene, glacial acetic acid, reflux, 20 h; (vi) 1 N NaOH, THF/CH₃OH, room temp, 1 h.

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Derivatives **6e-h** and **7e-h** were obtained according to Scheme 2. The pyrroles **10** and **11** were used as starting materials to obtain the key intermediates **12e-h**. Compound **10** was obtained by alkylation of commercially available 4-iodo-2-formylpyrrole in alkaline medium (K₂CO₃) with 4-F-benzyl bromide.

Scheme 2. Synthetic Route to Pyrrolyl DKAs 6e-h and 7e- h^a



^{*a*} Reagents and conditions: (i) alkylating agent, K_2CO_3 , DMF, 100 °C, 24 h; (ii) phenylboronic acid, Cs_2CO_3 , $P(t-But)_3$, $Pd_2(dba)_3$, dioxane, 80 °C, 24 h; (iii) 1) DMF, 1,2-dichloroethane dry, oxalyl chloride, 0 °C, 15 min, room temp, 15 min; 2) 4-F-benzoyl chloride, AlCl₃, room temp, 4 h; (iv) phenylboronic acid, copper (II) acetate anhydrous, pyridine/NMP 1:1, MW: 60 Watt, 120 °C, 6 min; (v) acetone, 4 N NaOH, room temp, 24 h; (vi) diethyl oxalate, C_2H_5ONa , THF, room temp, 2 h; (vii) 1 N NaOH, THF/CH₃OH, room temp, 1 h.

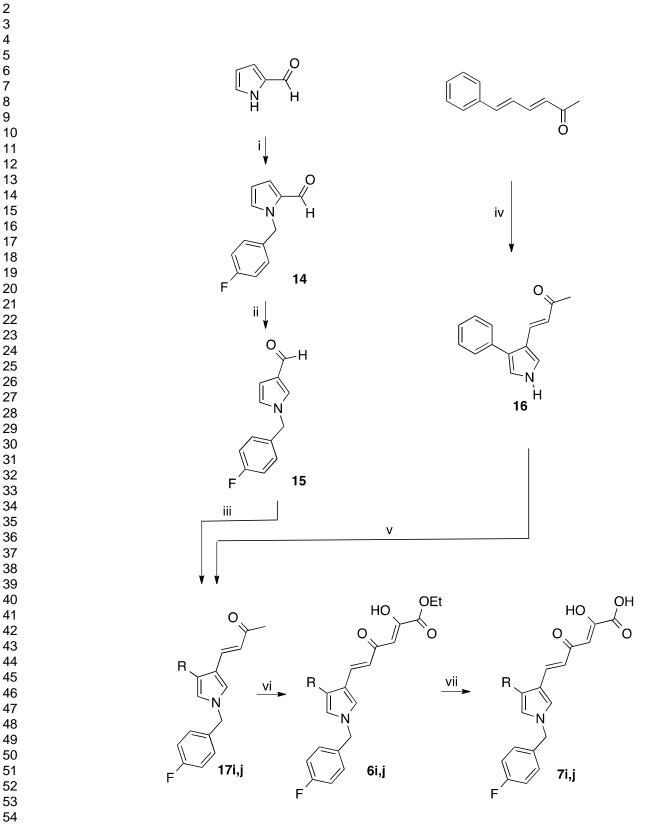
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Derivative **11** was obtained starting form pyrrole that underwent to two step one-pot reaction comprising i) formylation in the presence of oxalylchloride and DMF, and ii) Friedel-Craft reaction with 4-F-benzoyl chloride. Thus, **10** and **11** were alkylated with the appropriate alkyl halide to furnish **12e** and **12g**, or arylated on nitrogen atom of pyrrole ring to give **12f**, or finally coupled by a Suzuki reaction to obtain **12h**, respectively. The formylpyrroles **12e-h** were condensed with acetone in alkaline medium (4 N NaOH) to obtain α,β -insaturated ketones **13e-h**. The last compounds were in turn condensed with diethyl oxalate in the presence of sodium ethoxide to provide the diketobutanoic ethyl esters **6e-h** that were finally hydrolyzed by reaction with 1 N NaOH to give the corresponding carboxylic acids **7e-h**.

The synthetic pathway to obtain derivatives **6i,j** and **7i,j** is outlined in Scheme 3. The enones **17i,j** were obtained starting from **15** and **16**, respectively. Pyrrole **15** has been achieved starting form pyrrole-2-carboxaldehyde that was alkylated in alkaline medium (K_2CO_3) to obtain **14**. The termal transposition of the formyl chain from position 2 to 3 of the pyrrole ring in the presence of trifluoro acetic acid, led to derivative **15**. In a parallel pathway (3E,5E)-6-phenylhexa-3,5-dien-2-one³⁸ underwent to ring closure by reacting with TosMIC, giving the pyrrole derivatives **16**. Interestingly, the attack of the TosMIC reagent was specific on the 5,6 double bond of the starting dienone. Afterwards, derivative **15** was condensed with acetone affording **17i**; conversely, compound **16** was converted into **17j** by reaction of **4**-F-benzyl bromide in alkaline medium (K_2CO_3). Finally, intermediates **17i,j** were converted into esters **6i,j** by condensation with diethyl oxalate and then converted to acids **7i,j** by alkaline hydrolysis.

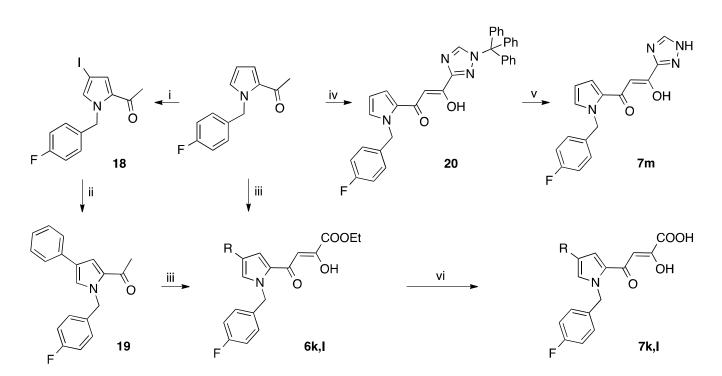
Derivatives **6k,l** and **7k,l** were obtained according to the pathway described in Scheme 4. The iodination of 2-acetyl-1-[(4-fluorophenyl)methyl]pyrrole in presence of *N*-iodosuccinimide (NIS) afforded derivative **18**, which underwent to a Suzuki coupling reaction to give **19**. Intermediates **19** and 2-acetyl-1-[(4-fluorophenyl)methyl]pyrrole were converted into the diketobutanoic acid derivatives **7k,l** through the ethyl esters **6k,l**, according to the condensation/hydrolysis procedure described above.

Scheme 3. Synthetic Route to Pyrrolyl DKAs 6i,j and $7i,j^a$



^{*a*} Reagents and conditions: (i) 4-F-benzyl bromide, K_2CO_3 , DMF, 100 °C, 24 h; (ii) Trifluoroacetic acid, 80 °C, 24 h; (iii) acetone, 4 N NaOH, room temp, 24 h; (iv) Et₂O/DMSO, NaH, TosMIC, room temp, 1 h; (v) 4-F-benzyl bromide, K_2CO_3 , DMF, 100 °C, 24 h; (vi) diethyl oxalate, C_2H_5ONa , THF, room temp, 2 h; (vii) 1 N NaOH, THF/CH₃OH, room temp, 1 h.

Scheme 4. Synthetic Route to Pyrrolyl DKAs 6k,l and 7k,l^a



^{*a*} Reagents and conditions: (i) NIS, acetone, -78 °C, 96 h; (ii) phenylboronic acid, Cs_2CO_3 , $P(t-But)_3$, $Pd_2(dba)_3$, dioxane, 80 °C, 24 h; (iii) diethyl oxalate, C_2H_5ONa , THF, room temp, 2 h; (iv) 1-trityl-1H-[1,2,4]triazole-3-carboxylic acid ethyl ester,³⁹ *n*-BuLi, THF, from -78 °C to 0 °C, 3.5 h; (v) 3 M HCl sol, 1,4-dioxane, 60 °C, 30 min; (vi) 1 N NaOH, THF/CH₃OH, room temp, 1 h.

Finally, 2-acetyl-1-[(4-fluorophenyl)methyl]pyrrole was subjected to a condensation reaction in presence of 1-trityl-1H-[1,2,4]triazole-3-carboxylic acid ethyl ester³⁹ to afford the triazole-protected derivative **20**, which was deprotected by hydrolysis in the presence of 3M HCl to obtain the triazole derivative **7m**.

Evaluation of biological activities. All newly synthesized compounds **6a-1** and **7a-m** were tested *in vitro* in enzyme assays against both recombinant RNase H and IN. The IC₅₀ values obtained for each compound in the inhibition of both the IN ST reaction and HIV-1 RT-associated RNase H function were plotted against each other in correlation plots (Figure 1). In these plots, single dots correspond to single compounds. The compounds are distributed around two perpendicular axes crossing the IN IC₅₀ axis (X axis) at 1 μ M and the RNase H IC₅₀ axis (Y axis) at 10 μ M (bolded crosshair in the center of each graph, Figure 1).

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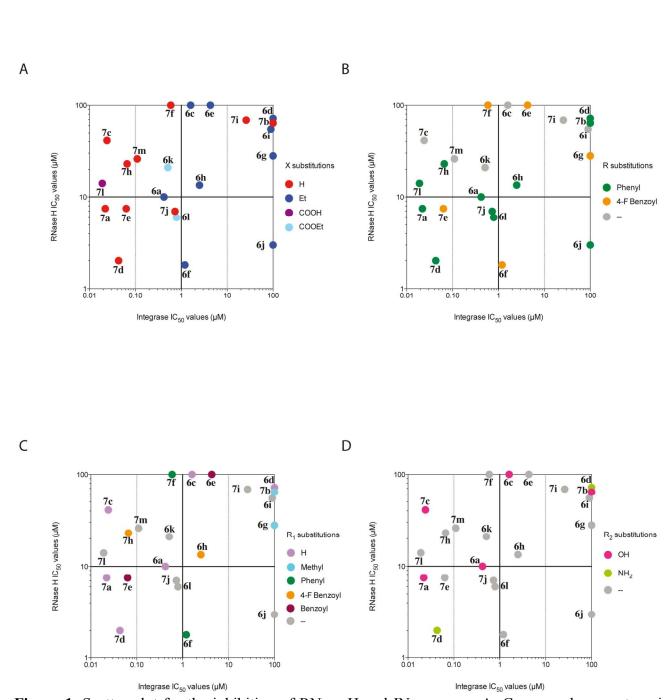


Figure 1. Scatter plot for the inhibition of RNase H and IN enzymes. A. Compounds are categorized according to their acidic or ester function. B. Compounds are categorized according to the nature of their R substitution. C. Compounds are categorized according to the nature of their R_1 substitution. D. Compounds are categorized according to the nature of their R_2 substitution. Compounds with one IC_{50} value missing such as **6d** have been left out of the plot and compounds with IC_{50} values above 111 μ M have been arbitrary positioned at the 100 μ M value.

These two axes splice the graph into four quarts corresponding to RNase H/IN dual inhibitors (lower left quarter), RNase H-selective inhibitors (lower right quarter), IN selective inhibitors (upper left quarter) and inhibitors of lower potency (upper right quarter). As seen on Figure 1, these graphs do not show any particular correlation between RNase H and IN inhibition.

The newly synthesized pyrrolyl derivatives **6a-1** and **7a-m** exhibited good potency of inhibition when tested on the HIV-1 IN ST (Table 1). In general, as reported previously,^{31,32} the acid derivatives **7a-m** were endowed of the highest potency showing IC₅₀ values in the range of 26-0.019 μ M. In our assay Merck compound **7k** was confirmed as potent IN ST inhibitor showing IC₅₀ = 0.057 μ M (literature data IC₅₀ = 0.08 μ M).³³ As seen on Figure 1A, 80% of the acid compounds (red dots) are distributed in the left half of the graph, suggesting the acid function is critical for IN inhibition but not critical for RNAse H inhibition. Among them, 7 compounds (**7a,c,d,e,h,k,l**) proven sub-micromolar activity (IC₅₀ value were in the range of 66 - 19 nM), while two derivatives (**7b,i**) can be considered less active to almost inactive (IC₅₀ values were 111 and 26 μ M, respectively). The most active compound was the 4phenylpyrrole derivative **7l** with an IC₅₀ value of 19 nM, three time less active than **1** in inhibiting of the ST reaction of IN. Interestingly, even though not all the synthetized analogues were tested on the HIV-1 3'-P activity, results showed that the newly synthesized acids were selective inhibitors of the ST step, with the IC₅₀ values on the 3'-P step 2-3 orders of magnitude higher with respect to the ones obtained on ST, thus confirming that the DKAs are selective ST inhibitors (data not shown).

The newly synthesized pyrrolyl DKA derivatives **6a-I** and **7a-m** can be divided into two classes: the diketobutanoic (**6a-d,k,l** and **7a-d,k,l**) and the diketobexenoic (**6e-j** and **7e-j**) derivatives. In the diketobutanoic structure of derivatives **7a-d**, **7k-l**, two main differences involving the substitution of the pyrrole ring emerged: the diketobutanoic chain can be linked on the pyrrole ring into two different positions (2- or 3-position), along with a phenyl ring, that can be substituted in position 4 of the pyrrole ring. Both two variables did not influence the IN inhibitory activity. Only the phenyl substitution at position 4 of the pyrrole ring (R substituent) seems to favor slightly IN vs. RNaseH inhibition with a

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majority of compounds bearing this substitution distributed in the two left quarters of the correlation graphs (green dots, Figure 1B). In fact, from a comparison of the inhibition data of diketobutanoic/diketohexenoic 7a/7l and 7c/7k, which are characterized respectively by the presence of the phenyl ring and by its absence, a correspondence in the orders of magnitude of their IC₅₀ values was observed. Moreover, 7a and 7c linked the diketobutanoic chain in position 3 of the pyrrole ring, while 7k and 7l in 2-position of the same ring. All these compounds were characterized by similar potencies (IC₅₀ values of 22 nM, 24 nM, 57 nM and 19 nM, respectively). The 4-fluoro-benzoyl substitution at position 4 of the pyrrole ring (R substituent) does not seem to favor the inhibition of either enzymes (orange dots, Figure 1B).

Other three modifications involved exclusively the diketobutanoic chain: (1) the substitution of the enolic OH with a NH₂ (**7d**), (2) the introduction of a methyl group in 3-position (**7b**), and (3) the substitution of the carboxylic acid function with its bioisoster triazole ring (**7m**). The NH₂ (**7d**) and triazole (**7m**) derivatives were 2-fold less active with respect to their OH (**7a**) and COOH (**7h**) counterparts, respectively (IC₅₀ values were 43 nM, 110 nM, 22 nM and 57 nM, respectively). On the contrary, the 3-methylbutenoate derivative (**7b**) completely lost its ability to inhibit IN enzyme (IC₅₀ values of > 111 μ M). It appears that IN seems to tolerate a wide variety of substitutions at the R₁ position, including the absence of substituent, with a majority of compounds with such substitutions distributed in the two left quarters of the correlation graphs (Figure 1C). No preferential inhibition pattern can be observed for compounds with substitutions at the R₂ position on the correlation plot presented in Figure 1 D.

When the two series of diketobutanoic and the diketohexenoic esters and acids were tested on the HIV-1 RT-associated RNase H function in biochemical assays, the most active derivatives were the diketohexenoic ester **6f** and the diketobutanoic acid **7d**, with IC_{50} value of 1.8 μ M, and 2 μ M, respectively. Interestingly, both compounds were more active then the references **3** and **4**. Noteworty, the structure of **6f** is related to the references **3** since both are pyrrolyl diketohexenoic derivatives, but **6f** bring a phenyl ring on nitrogen replacing the 4-F-benzyl group of **3** and a 4-F-benzyl moiety linked **ACS Paragon Plus Environment**

in 4 position of the pyrrole ring. In the matter of compound **7d**, it is a 3-pyrrolyl-diketobutanoic acid derivative characterized by the presence of an amine function that replaces the enole OH in position 3 of the diketobutanoic chain. Since **7d** is a carboxylic acid, its stronger inhibition towards IN than RNase H function was expected; conversely, the ester **6f** is the best dual inhibitor IN/RNase H of this series. From a first analysis of the results we can state that i) as known, the acid function confer a better inhibitory activity on IN, ii) the ester function is amenable for inhibition of RNase H function of RT, confirming the results recently reported by as by the means of docking studies and mutagenesis experiments,⁴⁰ and iii) contemporaneously, the ester function is necessary for a dual inhibition IN/RNase H.

Among the ester derivatives (**6a-l**), the diketohexenoic compounds (**6e-j**, IC₅₀ values in the range of 1.8 - 55 μ M) were able to inhibit the HIV-1 RNase H function with a slightly higher potency then the diketobutanoic counterpart (**6a-d,k,l**, IC₅₀ values in the range of 6 - 72 μ M). Differently, when the acid derivatives (**7a-m**) were tested this difference was not observed.

In both the diketohexenoic and the diketobutanoic ester and acid derivatives, the presence of a phenyl substituent in position 4 of the pyrrole ring influenced the inhibitory activity. In fact, the 4-phenyl substituted diketobutanoic ester and acid **6a**/**7a** (IC₅₀ values of 10 μ M, 7.5 μ M, respectively) and diketohexenoic **6j**/**7j** (IC₅₀ values of 3.0 μ M, 7.0 μ M, respectively) were consistently more active then the unsubstituted counterparts **6c**/**7c** and **6i**/**7i** (IC₅₀ values of > 100 μ M, 41 μ M, 55 μ M and 69 μ M respectively).

In general, when the DKA chain was shifted from 2- to 3-position, a 2-4 fold increase in potency of inhibition was observed (compare **6h**, **7h** and **7l** with **6j**, **7j** and **7a**: IC₅₀ values of 13.4 μ M and 23 μ M 14 μ M 3 μ M, 7 μ M, 7.5 μ M, respectively), with the sole exception of **6a** and **6l**, that showed comparable potency (IC₅₀ values of 10 μ M and 6 μ M, respectively).

The substitution of the enolic OH on the diketobutanoic chain (**6a**/**7a**) with a NH₂ group (**6d**/**7d**) in the ester series, lead to a 7-fold decrease of the potency of RNase H inhibition (**6d**, IC₅₀ values of 72 μ M) with respect to the unmodified counterpart (**6a**, IC₅₀ value of 10 μ M); while within the acid series,

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the NH₂ derivative improved its potency of inhibition (7d, IC₅₀ value of 2 μ M; 7a, IC₅₀ value of 7.5 μ M). The introduction of a methyl group in position 3 of the diketobutanoic chain (7b) reduced the inhibition (7b and 7a IC₅₀ values of 64 μ M and 7.5 μ M, respectively) likewise the substitution of the carboxylic acid function with its bioisoster triazole (7a and 7m, IC₅₀ values of 7.5 μ M and 26 μ M, respectively).

Among all the newly synthesized derivatives, the 2-pyrrolyldiketohexenoic ester **6f**, athough not very potent, emerged as dual inhibitor showing similar IC₅₀ values against both IN enzyme and RNase H function of RT (1.2 μ M and 1.8 μ M, respectively). This compound retains the ester function that is demonstrated necessary for the dual inhibiton³¹ and is characterized by a phenyl ring on nitrogen replacing the 4-F-benzyl group of **3** and by a 4-F-benzoyl moiety linked in 4 position of the pyrrole ring.

Cell-based assays. Among the newly synthesized pyrrolyl DKA derivatives **6a-1** and **7a-m**, seven derivatives (**6a,c,l** and **7a,c-d,l**) were characterized by a good anti-HIV activity showing a EC₅₀ values in the sub-micromolar concentration (EC₅₀ in the range of $0.56 - 0.9 \mu$ M) and a good selectivity index (SI). Three compounds, the **6k-j** and **7k** derivatives, showed an anti-HIV activity in the range of the micromolar concentration (EC₅₀ in the range of $1 - 4.3 \mu$ M), while 10 compounds were less asctive or inactive (**6d-i** and **7b,e-f,i,m**, EC₅₀ in the range of $17.2 - 50 \mu$ M). In general, all the compounds had a low citotoxicity index (CC₅₀ > 50 μ M). Compound **7c**, characterized by the diketobutanoic chain in 3-position of the pyrrole ring, showed the best antiviral efficacy on HIV-1 infected cell (EC₅₀ = 0.58 μ M) and a low citotoxicity (CC₅₀ > 50 μ M, SI > 86). The best dual inhibitor derivative **6f** that showed activity at micromolar concentration in biochemical assays, was 20 times less potent in cell-based assays.

Table 1.	Cytotoxicity,	Enzymatic, a	and Antiviral	Activities of C	Compounds 6a-l and 7a-m
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					activity enzyme assay,]	e	antiviral activity		
Cpd	R	R ₁	R ₂	Х	RH^{b}	ST ^c	EC_{50}^{d}	CC_{50}^{e}	Sľ
6a	Ph	Н	OH	Et	10	0.42	0.56	11	19.6
6b	Ph	Me	ОН	Et	-	-	-	-	-
6c	Н	Н	ОН	Et	>100	1.6	0.90	>50	>55
6d	Ph	Н	NH_2	Et	72	NT	19	>50	-
6e	4-FBz ^g	Bn^h	-	Et	>100	4.3	19	>50	-
6f	4-FBz	Ph	-	Et	1.8	1.2	20	>50	-
6g	4-FBz	Me	-	Et	28	>333	48	>50	-
6h	Ph	4-FBn	-	Et	13.4	2.5	-	-	-
6i	Н	-	-	Et	55	90	50	>50	-
6j	Ph	-	-	Et	3.0	>21	4.3	26.9	6.3
6k	Н	-	-	COOEt	21	0.51	1.2	33	27
61	Ph	-	-	COOEt	6.0	0.79	0.70	3.9	6
7a	Ph	Н	ОН	Н	7.5	0.022	0.66	>50	>75
7b	Ph	CH_3	ОН	Н	64	>111	>50	>50	-
7c	Н	Н	ОН	Н	41	0.024	0.58	>50	>86
7d	Ph	Н	NH_2	Н	2.0	0.043	0.63	>50	>79
7e	4-FBz	Bn	-	Н	7.5	0.063	>50	-	-
7f	4-FBz	Ph	-	Н	100	0.59	>50	-	-
7g	4-FBz	Me	-	Н	20	-	-	-	-
7h	Ph	4-FBn	-	Н	23	0.066	-	-	-
7i	Н	-	-	Н	69	26	>50	>50	-
7j	Ph	-	-	Н	7.0	0.73	17.2	>50	>2.9
7k ⁱ	Н	-	-	СООН	54	0.057	1.0	28	28
71	Ph	-	-	СООН	14	0.019	0.7	>50	>72

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7m	Н	-	-	Tr ^j	26	0.11	20.4	>50	-
1						0.007	0.016	>250	>15000
2					3.2	1.9	>50		
3					8	98	< 0.2	>50	>250
4					3	0.60	2	>50	>25
5					26.2	2.4	3.6	>50	>13.8

^{*a*}Inhibitory concentration 50% (μ M) determined from dose response curves. ^{*b*}Experiments performed against HIV-1 RT-associated RNase H activity. ^{*c*}Experiments performed against HIV-1 IN ST activity. ^{*d*}Effective concentration 50% (μ M). ^{*e*}Cytotoxic concentration 50% (μ M). ^{*f*}SI = CC₅₀/EC₅₀. ^{*g*}Bz, benzoyl. ^{*h*}Bn, benzyl. ^{*i*}See also reference 33. ^{*j*}Tr, triazolyl.

CONCLUSION

The development of HIV-1 dual inhibitors is a highly innovative approach aimed to reduce drug toxic side effects and therapeutic costs as well. Since HIV-1 IN and RNase H are both selective targets for HIV-1 chemotherapy, the identification of dual IN/RNase H inhibitors is an attractive strategy for new drug development. The newly synthesized pyrrolyl derivatives 6a-l and 7a-m exhibited good potency against IN and a moderate inhibition of the RNase H function of RT. In general, comparing the inhibition data among the ester and the acid derivatives a different behaviour was observed. As expected, the acid derivatives showed an higher potency of IN inhibition with respect to the corresponding esters, while the latter have been often found more potent than the corresponding acids in inhibiting the RNase H function of RT enzyme. Notably, compound 6f, although was not very potent on HIV-infected cell, showed a good correlation between HIV-1 IN and RNase H inhibition. It is characterized by a diketoester function, a phenyl ring on nitrogen and a 4-F-benzoyl moiety linked in 4 position of the pyrrole ring. We can state that although the acid function confer a better inhibitory activity on IN an ester function is amenable for inhibition of RNase H function of RT and as the conseguence the ester function is necessary for a dual inhibition IN/RNase H. This basic chemical features should be considered for development of further more potent dual inhibitors. Overally, the data reported in this work confirm the possibility to develop dual HIV-1 IN/RNase H inhibitors and give new information for the further development of effective dual HIV-1 inhibitors.

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EXPERIMENTAL SECTION

Chemistry. General. Melting points were determined on a Bobby Stuart Scientific SMP1 melting point apparatus and are uncorrected. Compounds purity were always >95% determined by high pressure liquid chromatography (HPLC). HPLC analysis were carried out with a Shimadzu LC-10AD VP CTO-10AC VP. Column used was generally Discovery Bio Wide Pore C18 (10 cm X 4.6 mm, 3 µm). Infrared (IR) spectra were recorded on a Perkin-Elmer Spectrum-one spectrophotometer. 1H NMR spectra were recorded at 400 MHz on a Broker AC 400 Ultrashield 10 spectrophotometer (400 MHz). Dimethylsulfoxide-d6 99.9% (code 44,139-2) and deuterochloroform 98.8% (code 41,675-4) of isotopic purity (Aldrich) were used. Column chromatographies were performed on silica gel (Merck; 70-230 mesh) column or alluminium oxide (Sigma-aldrich; 150 mesh) column. All compounds were routinely checked on TLC by using aluminium-baked silica gel plates (Fluka DC-Alufolien Kieselgel 60 F254) or alluminium oxide (Fluka DC-Alufolien Aluminium oxide). Developed plates were visualized by UV light. Solvents were reagent grade and, when necessary, were purified and dried by standard methods. Concentration of solutions after reactions and extractions involved the use of rotary evaporator (Büchi) operating at a reduced pressure (ca. 20 Torr). Organic solutions were dried over anhydrous sodium sulfate (Merck). All reactions were carried out under nitrogen; all solvents were freshly distilled under nitrogen and stored over molecular sieves for at least 3 h prior to use.

Microwave Irradiation Experiments. Microwave reactions were conduced using a CEM Discover system unit (CEM. Corp., Matthews, NC). The machine consists of a continuous focused microwave-power delivery system with operator selectable power output from 0 to 300 W. The temperature of the contents of the vessel was monitored using a calibrated infrared temperature control mounted under the reaction vessel. All experiments were performed using a stirring option whereby the contents of the vessel are stirred by means of a rotating magnetic plate located below the floor of the microwave cavity and a Teflon-coated magnetic stir bar in the vessel.

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General procedure A (GP-A): Synthesis of pyrrole nucleus. A solution of α , β -unsaturated ketone (5.42 mmol) and toluene-4-sulfonylmethylisocyanide (1.16 g, 5.96 mmol, 1.1 eq) dissolved in a mixture of anhydrous dimethyl sulfoxide/ethyl ether (14:30 mL) was added dropwise into a well-stirred suspension of sodium hydride (60% in paraffine oil; 0.48 g, 11.93 mmol, 2.2 eq) in dry ethyl ether (30 mL) under argon atmosphere. After the addition the mixture was stirred at room temperature for 1 hour. The reaction was treated with water and extracted with ethyl acetate. The organic layer was washed with brine, dried over anhydrous sodium sulfate and concentrated under vacuum. The crude product was purified by chromatography on aluminum oxide (chloroform as eluent) to afford the pure product. Yield (%), melting point (°C), recrystallization solvent, IR, ¹H NMR are reported for each compound.

General procedure B (GP-B): Alkylation of the pyrrole nitrogen. A mixture of the appropriate pyrrole (1.1 mmol), alkylating agent (3.3 mmol), and anhydrous K_2CO_3 (210 mg, 1.5 mmol) in dry DMF (10 mL) was stirred at 100 °C for 2 h. Then the mixture was cooled, treated with water (40 mL) and extracted with ethyl acetate. The organic layer was washed with brine, dried over anhydrous sodium sulfate and concentrated under vacuum. The crude product was purified by chromatography on silica gel to afford the pure product. Chromatography eluent, yield (%), melting point (°C), recrystallization solvent, IR, ¹H NMR are reported for each compound.

General Procedure C (GP-C): Condensation of pyrrole carboxaldehyde with acetone. The proper pyrrole carboxaldehyde (0.075mol) was dissolved in 250 mL of acetone. To this mixture was added 4 N NaOH (110 mL) and the reaction was stirred at room temperature for 24 h. After this period water (300 mL) and ethyl acetate (250 mL) were added. The organic layer was separated, washed with water (2 x 100 mL), dried over sodium sulfate, filtered and evaporated under reduced pressure. The crude product was purified by column chromatography on silica gel to obtain pure products. Chromatography eluent, yield (%), melting point (°C), recrystallization solvent, IR, ¹H NMR are reported for each compound.

General procedure (GP-D): Acetyl Transposition. A mixture of opportune α acetyl-substituted pyrrole (1.23 mmol) in trifluoroacetic acid (5 mL), was heated at 80 °C for 20 h. After this period the reaction was quenched with water (30 mL) and extracted with ethyl acetate (2 x 50 mL). The organic ACS Paragon Plus Environment layers were collected, dried over sodium sulfate, filtered and evaporated under vacuum. The crude product was purified by chromatography on silica gel (chloroform as eluent) to afford pure product as a brown oils. Yield (%), melting point (°C), recrystallization solvent, IR, ¹H NMR are reported for each compound.

General Procedure e (GP-E): Suzuki reaction. $Pd_2(dba)_3$ (0.1 g, 1.7 mmol) was added ino a well stirred mixture of appropriate 4-iodopyrrole (1.7 mmol), phenylboronic acid (0.85 g, 7.0 mmol), Cs_2CO_3 (0.665 g, 2.0 mmol) and $P(t-But)_3$ in dioxane (20 mL). The reaction was stirred at 80 °C for 24 h under argon atmosphere. Then the reaction was cooled to room temperature, filtered and washed with dioxane. The organic layer was evaporated under vacuum. The raw material was extracted with water (50 mL) and ethyl acetate (50 mL). The organic phase was separated, dried over sodium sulfate, filtered and evaporated under vacuum. The raw material was purified by silica gel chromatography. Chromatography eluent, yield (%), melting point (°C), recrystallization solvent, IR, ¹H NMR are reported for each compound.

General Procedure F (GP-F): Synthesis of Diketo Esters. Freshly prepared sodium ethoxide (390 mg, 5.5 mmol) was added into a well-stirred mixture of the appropriate acetyl derivative (2.7 mmol) and diethyl oxalate (790 mg, 5.4 mmol) in anhydrous THF (2.7 mL) under nitrogen atmosphere. The mixture was stirred at room temperature for 2 h, and then was poured into *n*-hexane (50 mL). The collected precipitate was vigorously stirred for 30 min in 1 N HCl (50 mL). The yellow solid that formed was filtered, washed with water, and dried under IR lamp to afford the pure diketo esters. Yield (%), melting point (°C), IR, ¹H NMR are reported for each compound.

General Procedure G (GP-G): Synthesis of diketo acids. A mixture of 1 N NaOH (6.5 mL) and the appropriate ester (1.3 mmol) in 1:1 THF/methanol (12 mL) was stirred at room temperature for 40 min and then poured onto crushed ice. The aqueous layer was treated with 1 N HCl until pH 3 (pH 7 for 1d), and the solid that formed was collected by filtration, then washed with water and dryed under warming lamp to afford pure acids. Yield (%), melting point (°C), IR, ¹H NMR are reported for each compound.

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1-(4-Phenyl-1H-pyrrol-3-yl)propan-1-one (8).⁴¹ Compound **8** was prepared from (E)-1-phenylpent-1-en-3-one³⁵ by means GP-A. 79% as a yellow solid; 169-170 °C; toluene. Anal. (C₁₃H₁₃NO) C, H, N.

1-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)ethanone (9a). Compound **9a** was prepared from 1-(4-phenyl-1H-pyrrol-3-yl)ethanone³⁴ by means GP-B, using 4-fluorobenzyl bromide as alkylatig agent. Chloroform; 100% as brown oil; IR *v* 1705 (C=O ketone) cm⁻¹; NMR (DMSO d_6) δ 2.26 (s, 3H, *CH*₃), 5.04 (s, 2H, CH₂), 6.63 (s, 1H, pyrrole α-proton), 7.06 (m, 2H, benzyl H), 7.19 (m, 2H, benzyl H), 7.25-7.41 (m, 6H, benzene H and pyrrole α-proton). Anal. (C₁₉H₁₆FNO) C, H, N, F.

1-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)propan-1-one (9b). Compound 9b was prepared from 8 by means GP-B, using 4-fluorobenzyl bromide as alkylating agent. Chloroform/acetate 50:1; 66% as brown oil.; IR v 1656 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 1.04 (t, 3H, J = 8 Hz, CH₂CH₃), 2.75 (q, 2H, J = 8 Hz, CH₂CH₃), 5.18 (s, 2H, CH₂), 7.07 (d, 1H, J = 2.2 Hz, pyrrole C5-H), 7.2-7.3 (m, 3H, benzene H), 7.32 (t, 2H, benzyl H), 7.4 (m, 2H, benzene H), 7.47 (m, 2H, benzyl H), 7.87 (d, 1H, J = 2 Hz, pyrrole C2-H). Anal. (C₂₀H₁₈FNO) C, H, N, F.

1-(1-(4-Fluorobenzyl)-1H-pyrrol-3-yl)ethanone (9c). Compound 9c was prepared from 1-(1-(4-fluorobenzyl)-1H-pyrrol-2-yl)ethanone³⁶ by means GP-D. 50% as brown oil; IR *v* 1655 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.37 (s, 3H, CH₃), 5.03 (s, 2H, CH₂), 6.60-6.63 (m, 2H, pyrrole C4-H and C5-H), 7.03 (t, 2H,benzene H), 7.12 (m, 2H, benzene H), 7.28 (t, 1H, *J* = 2.0 Hz, pyrrole C2-H,). Anal. (C₁₃H₁₄FNO) C, H, N, F.

1-(4-Fluorobenzyl)-4-iodo-1H-pyrrole-2-carboxaldehyde (10). Compound 10 was prepared from commercially available 4-iodopyrrole-2-carboxaldehyde by means GP-B, using 4-fluorobenzyl bromide as alkylatig agent. Chloroform/*n*-hexane 4:1; 39% as brown oil; IR v 1651 (C=O) cm⁻¹. ¹H NMR (CDCl₃) δ 5.47 (s, 2H), 6.9-7.0 (m, 4H, pyrrole α-proton, pyrrole β-proton and benzene H), 7.02 (m, 2H, benzene H), 9.48 (s, 1H, CHO). Anal. (C₁₂H₉FINO) C, H, N, F, I.

4-(4-Fluorobenzoyl)-1*H***-pyrrole-2-carboxaldehyde (11).** To a well stirred solution of DMF (3.9 mL, 50 mmol) in 1,2-dichloroethane (100 mL) refrigerated in ice bath was added drop wise a solution of

oxalyl chloride (6.35 g, 50 mmol) in 1,2 dichloroethane (100 mL) in a period of 15 minutes. After addiction, the suspension was stirred at room temperature for 15 minutes. After this time the reaction mixture was refrigerated in ice bath, and treated with a solution of pyrrole (49.9 mmol) in 1,2dichloroethane (100 mL). The mixture was stirred for 15 minutes at room temperature and then treated with AlCl₃ (14.6 g, 109 mmol) and 4-fluoro benzoyl chloride (50 mmol). The reaction was maintained at room temperature for 4 h. After this period the reaction was quenched with ice and water, and extracted with ethyl acetate, dried over sodium sulfate, filtered and evaporated under vacuum. The crude product was purified with chromatography on aluminum oxide (1:1 ethyl acetate-chloroform as eluent) to afford **5** as yellow solid. 85%; 110-111°C; benzene/cyclohexane; IR v 2900 (enole), 1660 (C=O ketone) 1640 (C=O) cm⁻¹. ¹H NMR: (CDCl₃) δ 7.1-7.4 (m, 3H, benzene H and pyrrole β-proton), 7.45 (s, 1H, pyrrole α-proton), 7.8-7.9 (m, 2H, benzene H), 9.60 (s, 1H, CHO), 12 (sb, 1H, NH). Anal. (C₁₂H₈FNO₂) C, H, N, F.

1-Benzyl-4-(4-fluorobenzoyl)-1H-pyrrole-2-carbaldehyde (12e). Compound **12e** was prepared from **11** by means GP-B, using benzyl bromide as alkylatig agent. Acetate/*n*-hexane 1:2; 34% as brown oil; IR v 1672 (C=O aldehyde) 1638 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 5.62 (s, 2H, CH₂), 7.1-7.2 (m, 4H, benzyl H and pyrrole β -proton), 7.3-7.4 (m, 4H, benzyl H and benzoyl H), 7.58 (s, 1H, pyrrole α -proton), 7.8-7.9 (m, 2H, benzoyl H), 9.62 (s, 1H, CHO). Anal. (C₁₉H₁₄FNO₂) C, H, N, F.

4-(4-Fluorobenzoyl)-1-phenyl-1H-pyrrole-2-carbaldehyde (12f). Compound 11, phenylboronic acid, pyridine, and copper (II) acetate anhydrous were dissolved in NMP (2.4ml) in a MW reactor tube and leave to react at 60 Watt, 120 °C for 6 min. After this period the reaction was quenched with water, and extracted with ethyl acetate (5 x 20ml), washed with water (5 x 20ml), dried over sodium sulfate, filtered and evaporated under vacuum. The crude product was purified with chromatography on silica gel (1:1 ethyl acetate-hexane as eluent) to afford **12f** as brown solid (63% yield). 130-131°C. benzene/cyclohexane; IR v 1680 (C=O aldehyde) 1632 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 7.21 (t, 2H, benzoyl H), 7.4-7.5 (m, 2H, benzene H), 7.5-7.6 (m, 4H, benzene H and pyrrole β–proton), 7.67 (d,

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1H, J = 2 Hz, pyrrole α -proton), 7.9-8.0 (m, 2H, benzoyl H), 9.68 (s, 1H, CHO). Anal. (C₁₈H₁₂FNO₂) C, H, N, F.

1-Methyl-4-(4-fluorobenzoyl)-1H-pyrrole-2-carboxaldehyde (12g). Compound 12g was prepared from 11 by means GP-B, using iodo methane as alkylatig agent. Chloroform; 90% as grey solid; 115-116°C; benzene/cyclohexane; IR v 1660 (C=O aldehyde) 1640 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 4.03 (s, 3H, N-CH₃), 7.2 (m, 2H, benzoyl H), 7.4 (d, 1H, pyrrole β–proton), 7.5 (s, 1H, pyrrole α–proton), 7.8-7.9 (m, 2H, benzoyl H), 9.60 (s, 1H, CHO). Anal. (C₁₃H₁₀FNO₂) C, H, N, F.

1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrole-2-carbaldehyde (12h). Compound 12h was prepared from 10 by means GP-E. 1:1 ethyl acetate-hexane as eluent; 95% as brown oil; IR v 1642 (C=O aldehyde) cm⁻¹. ¹H NMR (CDCl₃) δ 5.54 (s, 2H, CH₂), 6.82 (d, 2H, J = 7.0 Hz, benzene H), 6.91 (t, 1H, J = 7.0 Hz, benzene H), 6.99 (t, 2H, benzyl H), 7.16-7.24 (m, 4H, pyrrole β–proton, pyrrole α–proton, benzyl H), 7.48 (d, 2H, J = 7 Hz, benzene H), 9.58 (s, 1H, CHO). Anal. (C₁₈H₁₄FNO) C, H, N, F.

4-(1-Benzyl-4-(4-fluorobenzoyl)-1H-pyrrol-2-yl)but-3-en-2-one (13e). Compound 13e was prepared from 12e by means GP-C. 71% as yellow solid; 94-95 °C; benzene/cyclohexane; IR v 1675 (C=O ketone) 1637 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.26 (s, 3H, CH₃), 5.29 (s, 2H, CH₂), 6.60 (d, 1H, J = 16 Hz, butenone C3-H), 7.1-7.2 (m, 5H, butenone C4-H, benzyl H and pyrrole β-proton), 7.3-7.4 (m, 4H, benzyl H and benzoyl H), 7.46 (d, 1H, J = 2 Hz, pyrrole α-proton), 7.90 (m, 2H, benzoyl H). Anal. (C₂₂H₁₈FNO₂) C, H, N, F.

4-(1-Phenyl-4-(4-fluorobenzoyl)-1H-pyrrol-2-yl)but-3-en-2-one (13f). Compound 13f was prepared from 12f by means GP-C. 30% as yellow solid; 145-146 °C; benzene/cyclohexane; IR v 1680 (C=O ketone) 1634 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.24 (s, 3H, CH₃), 6.56 (d, 1H, J = 16 Hz, butenone C3-H), 7.19 (t, 2H, benzoyl H), 7.24 (d, 1H, J = 16 Hz, butenone C4-H), 7.35 (d, 1H, J = 2Hz, pyrrole β–proton), 7.38 (d, 1H, J = 2 Hz, pyrrole α-proton), 7.5-7.6 (m, 5H, benzene H), 7.94 (m, 2H, benzoyl H). Anal. (C₂₁H₁₆FNO₂) C, H, N, F. 4-(1-Methyl-4-(4-fluorobenzoyl)-1H-pyrrol-2-yl)but-3-en-2-one (13g). Compound 13g was prepared from 12g by means GP-C. 70 % as yellow solid; 117-118 °C; benzene/cyclohexane; IR v 1660 (C=O ketone) 1640 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.32 (s, 3H, CH₃), 3.78 (s, 3H, N-CH₃), 6.62 (d, 1H, J = 16 Hz, butenone C3-H), 7.1-7.2 (m, 3H, benzene H and pyrrole β–proton), 7.35 (d, 1H, J = 3.7 Hz, pyrrole α-proton), 7.43 (d, 1H, J = 16 Hz, butenone C4-H), 7.8-7.9 (m, 2H, benzene H). Anal. (C₁₆H₁₄FNO₂) C, H, N, F.

4-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)but-3-en-2-one (13h). Compound 13h was prepared from 12h by means GP-C. 15% as yellow solid; 160-161°C; benzene/cyclohexane; IR v 1604 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.28 (s, 3H, CH₃), 5.24 (s, 2H, CH₂), 6.57 (d, 1H, J = 16 Hz, butenone C3-H), 7.0-7.1 (m, 5H, pyrrole β-proton and benzyl H), 7.18 (d, 1H, J = 2 Hz, pyrrole α-proton,), 7.24 (t, 1H, J = 7 Hz, benzene H), 7.3-7.4 (m, 3H, butenone C4-H and benzene H), 7.5-7.6 (m, 2H, benzene H). Anal. (C₂₁H₁₈FNO) C, H, N, F.

1-(4-Fluorobenzyl)-1H-pyrrole-2-carboxaldehyde (14). Compound 14 was prepared from commercially available pyrrole-2-carboxaldehyde by means GP-B, using 4-fluorobenzyl bromide as alkylating agent. Chloroform; 80% as brown oil; IR v 1640 (C=O) cm⁻¹. ¹H NMR (CDCl₃) δ 5.54 (s, 2H, CH₂), 6.30 (t, 1H, *J* = 4 Hz, pyrrole C4-H), 6.9-7.0 (m, 4H, benzene H), 7.15 (d, 1H, *J* = 4 Hz, pyrrole C3-H), 7.17 (d, 1H, *J* = 4 Hz, pyrrole C5-H), 9.57 (s, 1H, CHO). Anal. (C₁₂H₁₀FNO) C, H, N, F.

1-(4-Fluorobenzyl)-1H-pyrrole-3-carboxaldehyde (10). Compound 10 was prepared from 9 by means GP-D. 55% as brown oil; IR v 1640 (C=O aldehyde) cm⁻¹. ¹H NMR (CDCl₃) δ 5.08 (s, 2H, CH₂), 6.6 (s, 1H, pyrrole C4-H), 6.7 (s, 1H, pyrrole C-2H), 7.0-7.2 (m, 4H, benzene H), 7.31 (s, 1H, pyrrole C2-H), 9.75 (s, 1H, CHO). Anal. (C₁₂H₁₀FNO) C, H, N, F.

4-(4-Phenyl-1H-pyrrol-3-yl)but-3-en-2-one (16).⁴² Compound **16** was prepared from (3E,5E)-6-phenylhexa-3,5-dien-2-one³⁸ by means GP-A. 56% as brown solid; toluene; IR v 1640 (C=O ketone) cm⁻¹. Anal. (C₁₄H₁₃NO) C, H, N.

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4-(1-(4-Fluorobenzyl)-1H-pyrrol-3-yl)but-3-en-2-one (17i). Compound 17i was prepared from 15 by means GP-C. 70% as brown oil; IR v 1655 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.34 (s, 3H, CH₃), 5.07 (s, 2H, CH₂), 6.43-6.48 (m, 2H, pyrrole C4-H and butenone C3-H), 6.71 (s, 1H, pyrrole C2-H), 7.0 (s, 1H, pyrrole C5-H), 7.06-7.12 (m, 2H, benzene H), 7.15-7.18 (m, 2H, benzene H), 7.49 (d, 1H, butenone C4-H, J = 16 Hz). Anal. (C₁₅H₁₄FNO) C, H, N, F.

4-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)but-3-en-2-one (17j). Compound **17j** was prepared from **16** by means GP-B, using 4-fluorobenzyl bromide as alkylating agent. Chloroform; 63% as brown oil; IR v 1640 (C=O) cm⁻¹. ¹H NMR (CDCl₃) δ 2.44 (s, 3H, CH₃), 5.07 (s, 2H, CH₂), 6.82 (s, 1H, *J* = 16.3 Hz, butenone C3-H), 6.90 (s, 1H,), 6.99-7.54 (m, 11H, pyrrole C2-H, pyrrole C5-H, benzene H and benzyl H), 7.80 (s, 1H, *J* = 16.3 Hz, butenone C4-H). Anal. (C₂₁H₁₈FNO) C, H, N, F.

1-(1-(4-Fluorobenzyl)-4-iodo-1H-pyrrol-2-yl)ethanone (18). A mixture of 1-(1-(4-fluorobenzyl)-1H-pyrrol-2-yl)ethanone³⁶ (4g, 18.4 mmol) in acetone dry (100 mL) was cooled at -78 °C, NIS (4.98 g, 22.1 mmol) was added. The reaction was stirred and the temperature was slowly increased to 25 °C in a period of 96 h. After this period the mixture was evaporated, ethyl acetate (50 mL) and NaHCO_{3(aq)} (50 mL) were added. The organic phase was separated, dried over sodium sulfate, filtered and evaporated under vacuum. The raw material was purified with a column chromatography on silica gel (1:10 ethyl acetate-hexane as eluent) to afford **18** as white solid with a yield of 40%. 81-82 °C; *n*-hexane; IR v 1640 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.40 (s, 3H, CH₃), 5.50 (s, 2H, CH₂), 6.92 (d, 1H, *J* = 2.0 Hz, pyrrole C3-H), 7.00 (t, 2H, benzene H), 7.07 (d, 1H, *J* = 2.0 Hz, pyrrole C5-H), 7.1-7.2 (m, 2H, benzene H). Anal. (C₁₃H₁₁FINO) C, H, N, F, I.

1-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)ethanone (19). Compound 19 was prepared from 18 by means GP-E. 1:7 ethyl acetate-hexane as eluent. 43% as colourless oil; IR v 1650 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 2.48 (s, 3H, CH₃), 5.58 (s, 2H, CH₂), 6.97-7.02 (t, 2H, benzyl H), 7.14-7.27 (m, 5H, benzene H, pyrrole C3-H and pyrrole C5-H), 7.35-7.39 (t, 2H, benzyl H), 7.51 (d, 2H,benzene H). Anal. (C₁₈H₂₁FNO) C, H, N, F.

 1-(1-(4-Fluorobenzyl)-1H-pyrrol-2-yl)-3-hydroxy-3-(1-trityl-1H-1,2,4-triazol-3-yl)prop-2-en-1one (20). A solution of 1-[1-(4-fluorobenzyl)-1H-pyrrol-2-yl]ethanone³⁶ (1 g, 4.6 mmol) in THF anhydrus (5 ml) was thermostated at -32 °C, LHMDS (9.2 mL) was added and the reaction was stirred at the some temperature for 2 h. A solution of 1-trityl-1H-[1,2,4]triazole-3-carboxylic acid ethyl ester³⁹ (2 g, 5.3 mmol) in THF anhydrus (18 mL) was added dropwise to the solution thermostated at -32 °C. After the addiction, the reaction mixture was stirred for 1.5 h at room temperature. The reaction was pored into 1 N HCl (100 mL), and extract with ethyl acetate. The organic phase was separated, washed with water, dried over sodium sulfate, filtered and evaporated under vacuum obtaining 2.6 g of crude product as light yellow solid. The raw material was purified by recristallization from benzene/cyclohexane obtaining 1.84 g of pure **20**. 54%; 110-112 °C; benzene/cyclohexane; IR v 2954 (OH enole), 1626 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 5.62 (s, 2H, CH₂), 6.3 (t, 1H, pyrrole C4-H), 6.9-7.0 (m, 4H, butenoate C3-H, pyrrole C3-H, benzyl H), 7.1-7.2 (m, 9H, benzyl H, pyrrole C5-H and benzene H), 7.3-7.4 (m, 9H, benzene H), 8.01 (s, 1H, triazole H), 15 (br s, 1H, OH enole). Anal. (C₃₅H₂₇FN₄O₂) C, H, N, F.

Ethyl 4-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-4-oxobut-2-enoate (6a). Compound 6a was prepared from 9a by means GP-F. 88%; 111-112°C; benzene; IR v 2900 (OH enole), 1720 (C=O ester), 1620 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 1.29 (t, 3H, *CH*₃CH₂), 4.25 (q, 2H, *CH*₂CH₃), 5.06 (s, 2H, CH₂ benzyl), 6.48 (s, 1H, butenoate C3-H), 6.67 (d, 1H, *J* = 1.5 Hz, pyrrole C5-H), 7.02-7.46 (m, 10H, pyrrole C2-H, benzene H and benzyl H), 15 (br s, 1H, OH enole). Anal. (C₂₃H₂₀FNO₄) C, H, N, F.

Ethyl 4-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-3-methyl-4-oxobut-2-enoate (6b). Compound 6b was prepared from 9b by means GP-F. 6b was extract with ethyl acetate. The organic phase was separated, washed with brine, dried over sodium sulfate, filtered and evaporated under vacuum obtaining a crude product that was purified with a column chromatography on silica gel (ethyl acetate/*n*-hexane 1:2); 30% as yellow oil; IR v 1730 (C=O ester), 1650 (C=O ketone) cm⁻¹. ¹H

NMR (DMSO d_6) δ 1.14 (d, 3H, J = 7 Hz, CH₃), 1.34 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 4.33 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 5.25 (s, 2H, CH₂), 7.14 (d, 1H, J = 1.9 Hz, pyrrole C2-H) 7.22-7.51 (m, 9H, benzene H and benzyl H), 8.08 (s, 1H, J = 1.9 Hz, pyrrole C5-H), 14 (br s, 1H, enole). Anal. (C₂₄H₂₂FNO₄) C, H, N, F.

Ethyl 4-(1-(4-fluorobenzyl)-1H-pyrrol-3-yl)-2-hydroxy-4-oxobut-2-enoate (6c). Compound 6c was prepared from 9c by means GP-F. 93% as yellow solid; 63-65 °C; ligroine; IR v 3500-2500 (OH enole), 1726 (C=O ester), 1633 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 1.28 (t, 3H, J = 7 Hz, CH_3CH_2), 4.27 (q, 2H, J = 7 Hz, CH_2CH_3), 5.17 (s, 2H, CH₂), 6.62 (t, 1H, pyrrole C4-H), 6.73 (s, 1H, butenoate C3-H), 7.02 (t, 1H, pyrrole C5-H), 7.18 (t, 2H, benzene H), 7.36 (dd, 2H, benzene H), 8.07 (s, 1H, pyrrole C2-H), 15 (br s, 1H, enole). Anal. (C₁₇H₁₈FNO₄) C, H, N, F.

Ethyl 2-amino-4-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-4-oxobut-2-enoate (6d). To a stirred mixture of 6a (1 g, 2.5 mmol), ammonium acetate (0.22 g, 2.9 mmol) in benzene (30 mL), was added acetic acid glacial (0.2 ml, 3.9 mmol). The reaction was stirred at reflux for 20 h with a Dean-Stark apparatus. After this period the reaction was cooled to room temp and washed with a saturated solution of NaHCO₃ (50 mL). The organic layer was separated, dried over sodium sulfate, filtered and evaporated under vacuum. The raw material was purified with chromatography on aluminum oxide (chloroform as eluent) to afford 6d as yellow solid with an yield of 50%. 135-136°C. benzene; IR v 3500 (NH₂), 1720 (C=O ester), 1620 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 1.27 (t, 3H, *CH*₃CH₂), 4.25 (q, 2H, CH₃*CH*₂), 5.08 (s, 2H, CH₂), 6.13 (s, 1H, butenoate C3-H), 6.67 (d, 1H, *J* = 1.5 Hz, pyrrole C5-H), 7.0-7.6 (m, 10H, benzene H, benzyl H and pyrrole C2-H), 9 (br s, 2H, NH₂). Anal. (C₂₃H₂₁FN₂O₂) C, H, N, F.

Ethyl 6-(1-benzyl-4-(4-fluorobenzoyl)-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6e). Compound 6e was prepared from 13e by means GP-F. 62% as yellow solid; 154-155°C; benzene/cyclohexane; IR v 1730 (C=O ester), 1636 (C=O ketone) cm⁻¹. ¹H NMR (Acetone d₆) δ 1.34 (t, 3H, *CH*₃CH₂), 4.32 (q, 2H, *CH*₂CH₃), 5.60 (s, 2H, CH₂), 6.86 (d, 1H, *J* = 15.6 Hz, hexanoate C5-H), 7.2-7.5 (m, 9H, J = 1.6 Hz, benzene H, pyrrole β -proton, benzoyl H and hexanoate C3-H), 7.72 (d, 1H, J = 15.6 Hz, hexanoate C6-H), 7.90 (d, 1H, J = 1.6 Hz, pyrrole α -proton), 7.9-8.0 (m, 2H, benzoyl H), 14 (bs, 1H, enole). Anal. (C₂₆H₂₂FNO₅) C, H, N, F.

Ethyl 6-(4-(4-fluorobenzoyl)-1-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6f). Compound 6f was prepared from 13f by means GP-F. 80% as yellow solid; 129-130°C; benzene/cyclohexane; IR v 3500 (OH enole), 1720 (C=O ester), 159 (C=O ketone) cm⁻¹. ¹H NMR (CD₃OD) δ 1.36 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 4.31 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 7.26-7.48 (m, 3H, benzoyl H and hexanoate C5-H), 7.39 (d, 1H, J = 15.6 Hz, hexanoate C6-H), 7.43-7.48 (m, 3H, benzene H and hexanoate C3-H), 7.58-7.62 (m, 4H, benzene H and pyrrole β-proton), 7.72 (s, 1H, pyrrole αproton), 7.97-8.01 (m, 2H, benzoyl H), 14 (bs, 1H, OH enole). Anal. (C₂₅H₂₀FNO₅) C, H, N, F.

Ethyl 6-(4-(4-fluorobenzoyl)-1-methyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6g). Compound 6g was prepared from 13g by means GP-F. 83% as yellow solid; 166-167°C; benzene/cyclohexane; IR v⁻¹ 2900 (OH enole), 1720 (C=O ester), 1650 (C=O ketone) cm^{-. 1}H NMR (CDCl₃) δ 1.39 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 3.81 (s, 3H, N-CH₃), 4.39 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 6.45 (s, 1H, hexanoate C3-H), 6.50 (d, 1H, J = 15.4 Hz, hexanoate C5-H), 7.12-7.20 (m, 3H, benzene H and pyrrole β-proton), 7.36-39 (m, 1H, pyrrole α-proton), 7.63 (d, 1H, J = 15.4 Hz, hexanoate C6-H), 7.83-7.91 (m, 2H, benzene H), 15 (bs, 1H, enole). Anal. (C₂₀H₁₈FNO₅) C, H, N, F.

Ethyl 6-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6h). Compound 6h was prepared from 13h by means GP-F. 92% as red solid; 169-170 °C; benzene; IR v 2900 (OH enole), 1723 (C=O ester), 1602 (C=O ketone) cm⁻¹. ¹H NMR (Acetone d_6) δ 1.35 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 4.32 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 5.53 (s, 2H, CH₂), 6.46 (s, 1H, hexanoate C3-H), 6.74 (d, 1H, J = 16 Hz, hexanoate C5-H), 7.13-7.24 (m, 3H, benzyl H and benzene H), 7.30-39 (m, 4H, benzyl H, hexanoate C6-H and pyrrole β-proton), 7.43 (s, 1H, pyrrole α-proton), 7.65 (d, 2H, benzene H), 7.7-7.8 (m, 2H, benzene H), 15 (bs, 1H, enole). Anal. (C₂₅H₂₂FNO₄) C, H, N, F.

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Ester 6-(1-(4-fluorobenzyl)-1H-pyrrol-3-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6i). Compound 6i was prepared from 17i by means GP-F. 41% as yellow solid; 88-90 °C; ligroine; IR v 3400 (OH enole), 1725 (C=O ester), 1625 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 1.44 (t, 3H, *J* = 7 Hz, *CH*₃CH₂), 4.41 (q, 2H, *J* = 7 Hz, *CH*₂CH₃), 5.08 (s, 2H, CH₂), 6.39 (s, 1H, *J* = 16 Hz, hexanoate C5-H), 6.5-6.6 (m, 2H, pyrrole C4-H and hexanoate C3-H), 6.7 (t, 1H, pyrrole C5-H), 7.0 (t, 1H, pyrrole C2-H), 7.08-7.20 (m, 4H, benzene H), 7.75 (s, 1H, *J* = 16 Hz, hexanoate C6-H), 14 (br s, 1H, enole). Anal. (C₁₉H₁₈FNO₄) C, H, N, F.

Ethyl 6-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-4-oxohexa-2,5-dienoate (6j). Compound 6j was prepared from 17j by means GP-F. 56% as yellow solid; 121-122°C; cyclohexane; IR v 2900 (OH enole), 1720 (C=O ester), 1620 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 1.43 (t, 3H, *CH*₃CH₂), 4.40 (q, 2H, *CH*₂CH₃), 5.09 (s, 2H, CH₂), 6.73 (s, 1H, hexanoate C3-H) 6.87 (d, 1H, *J* = 16.6 Hz, hexanoate C5-H), 7.01 (s, 1H, pyrrole C5-H), 7.08-7.54 (m, 10H, pyrrole C2-H, benzene H and benzyl H), 7.74 (d, 1H, hexanoate C6-H), 15 (bs, 1H, enole). Anal. (C₂₅H₂₁FNO₄) C, H, N, F.

Ethyl 4-(1-(4-fluorobenzyl)-1H-pyrrol-2-yl)-2-hydroxy-4-oxobut-2-enoate (6k). Compound 6k was prepared from 1-(1-(4-fluorobenzyl)-1H-pyrrol-2-yl)ethanone³⁶ by means GP-F. 41% as yellow solid; 88-89°C; ligroine; IR v 2900 (OH enole), 1720 (C=O ester), 1620 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 1.26 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 4.25 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 5.59 (s, 2H, CH₂), 6.32 (dd, 1H, J = 2.5 Hz, J = 3.5 Hz, pyrrole C4-H), 6.84 (s, 1H, butenoate C3-H), 7.0-7.2 (m, 4H, benzene H), 7.43 (d, 1H, J = 3.5 Hz, pyrrole C3-H), 7.53 (s, 1H, pyrrole C5-H), 14 (br s, 1H, enole). Anal. (C₁₇H₁₆FNO₄) C, H, N, F.

Ethyl 4-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxobut-2-enoate (61). Compound 6l was prepared from 19 by means GP-F. 95% as yellow solid; 102-103 °C; benzene/cyclohexane; IR v 3400 (OH enole), 1720 (C=O ester), 1620 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 1.34 (t, 3H, J = 7.5 Hz, CH_3 CH₂), 4.33 (q, 2H, J = 7.5 Hz, CH_2 CH₃), 5.68 (s, 2H, CH₂), 7.06 (s, 1H, butenoate C3-H), 7.17-7.31 (m, 5H, benzene H and benzyl H), 7.40-7.44 (m, 2H, benzene H), 7.75 (d, 2H, benzene H), 8.01 (s, 1H, pyrrole C5-H), 8.13 (s, 1H, pyrrole C3-H), 14 (bs, 1H, enole). Anal. (C₂₂H₂₀FNO₄) C, H, N, F.

4-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-4-oxobut-2-enoic acid (7a). Compound 7a was prepared from 6a by means GP-G. 98% as brown solid; 109-110°C; toluene; IR v 3500-2000 (OH acid and enole), 1740 (C=O acid), 1620 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 5.06 (s, 2H, CH₂), 6.48 (s, 1H, butenoate C3-H), 6.67 (d, 1H, J = 1.5 Hz, pyrrole C5-H), 7.06-7.10 (t, 2H, benzyl H), 7.18-7.25 (m, 2H, benzyl H), 7.30-7.39 (m, 5H, benzene H), 7.49 (s, 1H, pyrrole C2-H), 14 (br s, 2H, OH acid and enole). Anal. (C₂₁H₁₆FNO₄) C, H, N, F.

4-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-3-methyl-4-oxobut-2-enoic acid (7b). Compound 7b was prepared from 6b by means GP-G. 38% as white solid; 233-234°C; benzene; IR v 3500-2500 (OH acid and enole), 1700 (C=O acid), 1640 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 1.14 (d, 3H, CH₃, J = 7 Hz), 5.13 (s, 2H, CH₂), 7.1-7.2 (m, 3H, benzyl H and pyrrole C5-H), 7.34 (t, 2H, benzyl H), 7.47 (s, 1H, pyrrole C2-H), 7.51 (m, 3H, benzene H), 7.9-8.0 (m, 2H, benzene H), 14 (br s, 2H, enole and acid). Anal. (C₂₂H₁₈FNO₄) C, H, N, F.

4-(1-(4-Fluorobenzyl)-1H-pyrrol-3-yl)-2-hydroxy-4-oxobut-2-enoic acid (7c). Compound **7c** was prepared from **6c** by means GP-G. 57% as yellow solid; 146-147°C; toluene; IR v 3500-2500 (OH acid and enole), 1727 (C=O acid), 1621 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 5.16 (s, 2H, CH₂), 6.60 (dd, 1H, $J_1 = 1.5$ Hz, $J_2 = 3.0$ Hz, pyrrole C4-H), 6.72 (s, 1H, butenoate C3-H), 7.00 (dd, 1H, $J_1 = 1.5$ Hz, $J_2 = 3$ Hz, pyrrole C5-H), 7.18 (t, 2H, benzene H), 7.36 (dd, 2H, benzene H), 8.04 (s, 1H, pyrrole C2-H), 14 (br s, 1H, enole), 15 (br s, 1H, COOH). Anal. (C₁₅H₁₂FNO₄) C, H, N, F.

6-(1-Benzyl-4-(4-fluorobenzoyl)-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7e). Compound 7e was prepared from 6e by means GP-G. 87% as orange solid; 165-166°C; benzene; IR v 3500-2500 (OH acid and enole), 1727 (C=O acid), 1630 (C=O ketone) cm⁻¹. ¹H NMR (CD₃OD) δ 5.22 (s, 2H, CH₂), 6.69 (s, 1H, *J* = 15.2 Hz, hexanoate C5-H), 7.15-7.44 (m, 9H, hexanoate C3-H, benzyl H,

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benzoyl H and pyrrole β-proton), 7.65 (s, 1H, J = 15.2 Hz, hexanoate C6-H), 7.79 (s, 1H, pyrrole αproton), 7.92-7.97 (m, 2H, benzoyl H). Anal. (C₂₄H₁₈FNO₅) C, H, N, F.

6-(4-(4-Fluorobenzoyl)-1-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7f). Compound 7f was prepared from 6f by means GP-G. 85% as yellow solid; 183-184°C; benzene; IR v 3500-2500 (OH acid and enole), 1724 (C=O acid), 1596 (C=O ketone) cm⁻¹. ¹H NMR (CD₃OD) δ 6.69 (s, 1H, J = 16 Hz, hexanoate C5-H), 7.26 (t, 2H, benzoyl H), 7.41 (d, 1H, hexanoate C6-H), 7.47-7.49 (m, 2H, pyrrole β-proton and hexanoate C3-H), 7.73 (s, 1H, pyrrole α-proton), 7.98-8.01 (m, 2H, benzoyl H). Anal. (C₂₃H₁₆FNO₅) C, H, N, F.

6-(4-(4-Fluorobenzoyl)-1-methyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7g). Compound 7g was prepared from 6g by means GP-G. 95% as orange solid; 162-163°C; ethanol; IR v 3500-2500 (OH acid and enole), 1700 (C=O acid), 1600 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 3.32 (s, 3H, N-CH₃), 6.52 (s, 1H, hexanoate C3-H), 6.93 (d, 1H, hexanoate C5-H), 7.32-7.86 (m, 7H, benzene H, pyrrole β-proton, pyrrole α-proton and hexanoate C6-H), 14 (br s, 2H, OH enole and acid). Anal. (C₁₈H₁₄FNO₅) C, H, N, F.

6-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7h). Compound 7h was prepared from 6h by means GP-G. 73% as red solid; decompose; benzene; IR v 3500-2500 (OH acid and enole), 1707 (C=O acid), 1571 (C=O ketone) cm⁻¹. ¹H NMR (Acetone d_6) δ 5.51 (s, 2H, CH₂), 6.49 (s, 1H, hexanoate C3-H), 6.7, (d, 1H, hexanoate C5-H), 7.07-7.42 (m, 7H, benzyl H, pyrrole β-proton, pyrrole α-proton and hexanoate C6-H), 7.6-7.8 (m, 5H, benzene H), 14 (br s, 2H, enole and acid). Anal. (C₂₃H₁₈FNO₄) C, H, N, F.

6-(1-(4-Fluorobenzyl)-1H-pyrrol-3-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7i). Compound 7i was prepared from 6i by means GP-G. 92% as yellow solid; >300 °C; DMF/H₂O; IR v 3500-2500 (OH acid and enole), 1720 (C=O acid), 1630 (C=O ketone) cm⁻¹. ¹H NMR (DMF d_7) δ 5.34 (s, 2H, benzyl), 6.39 (s, 1H, hexanoate C3-H), 6.54 (d, 1H, hexanoate C5-H), 6.7 (bs, 1H, pyrrole C4-H), 7.38-7.67 (m,

7H, pyrrole C2-H, pyrrole C5-H, benzene H and hexanoate C6-H), 14 (br s, 2H, OH enole and acid). Anal. (C₁₇H₁₆FNO₄) C, H, N, F.

6-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-2-hydroxy-4-oxohexa-2,5-dienoic acid (7j). Compound 7j was prepared from 6j by means GP-G. 75% as yellow solid; 128-129°C; benzene; IR v 3500-2500 (OH acid and enole), 1700 (C=O acid), 1600 (C=O ketone) cm⁻¹. ¹H NMR (CDCl₃) δ 5.20 (s, 2H, CH₂), 6.75 (bs, 1H, hexanoate C3-H), 6.80 (d, 1H, hexanoate C5-H), 7.1-7.6 (m, 12H, pyrrole C2-H, pyrrole C5-H, benzene H, benzyl H and hexanoate C6-H), 14 (br s, 2H, enole and acid). Anal. (C₂₃H₁₇FNO₄) C, H, N, F.

4-(1-(4-Fluorobenzyl)-1H-pyrrol-2-yl)-2-hydroxy-4-oxobut-2-enoic acid (7k). Compound **7k** was prepared from **6k** by means GP-G 80% as yellow solid; 156-157°C; benzene; IR v 3500-2500 (OH acid and enole), 1700 (C=O acid), 1620 (C=O ketone) cm⁻¹. ¹H NMR (DMSO d_6) δ 5.59 (s, 2H, CH₂), 6.30 (dd, 1H, J = 2.5 Hz, J = 3.5 Hz, pyrrole C4-H), 6.81 (s, 1H, butenoate C3-H), 7.0-7.09-7.16 (m, 4H, benzene H), 7.39 (d, 1H, J = 3.5 Hz, pyrrole C3-H), 7.51 (s, 1H, pyrrole C5-H) 14 (br s, 2H, OH enole and acid). Anal. (C₁₅H₁₂FNO₄) C, H, N, F.

4-(1-(4-Fluorobenzyl)-4-phenyl-1H-pyrrol-2-yl)-2-hydroxy-4-oxobut-2-enoic acid (71). Compound 7l was prepared from 6l by means GP-G. 82% as yellow solid; 195-196 °C; toluene; IR v 3500-2000 (OH acid and enole), 1740 (C=O acid), 1620 (C=O ketone) cm⁻¹. ¹H NMR: (DMSO d_6) δ 5.68 (s, 2H, CH₂ benzyl), 7.05 (s, 1H, butenoate C3-H), 7.1-7.3 (m, 5H, benzyl H and benzene H), 7.41 (t, 2H, benzene H), 7.76 (d, 2H, benzene H), 8.01 (s, 1H, pyrrole C3-H), 8.13 (s, 1H, pyrrole C5-H) 14 (br s, 1H, OH enole), 15 (bs, 2H, OH acid). Anal. (C₂₀H₁₆FNO₄) C, H, N, F.

(E/Z) 2-Amino-4-(1-(4-fluorobenzyl)-4-phenyl-1H-pyrrol-3-yl)-4-oxobut-2-enoic acid (7d). Compound 6d (340 mg, 0.9 mmol) was dissolved in anhydrous THF (4.5 mL) under argon atmosphere and cooled to 0 °C. To this was added dropwise a solution of 0.5 N KOH (2 mL, 1.0 mmol) and the mixture allowed to stir at room temperature overnight. The reaction mixture was concentrated in vacuo and the residue partitioned between water and ethyl acetate. The aqueous layer was cooled on ice and

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acidified with 1 N HCl. After chilling at 4 °C for 2 h the resulting precipitate was extracted with ethyl acetate. The organic layer was washed with brine, dried over Na₂SO₄ and concentrated under vacuum to yield the acid **7d** (250 mg). 76% yellow solid; 86-88°C; toluene; IR v 3000-2500 (OH acid and enole), 1700 (C=O acid) cm⁻¹. ¹H NMR (CDCl₃) δ 4.95 (m, 2H, CH₂ E/Z form), 6.70 (s, 1H, butenoate C3-H), 6.8-7.43 (m, 11H, pyrrole C2-H, pyrrole C5-H, benzene H and benzyl H), 14 (br s, 2H, acid and enole). Anal. (C₂₁H₁₇FN₂O₃) C, H, N, F.

1-(1-(4-Fluorobenzyl)-1H-pyrrol-2-yl)-3-hydroxy-3-(1H-1,2,4-triazol-3-yl)prop-2-en-1-one (7m). 20 (1.84 g, 1.8 mmol) was suspended in 12 mL of dioxane and treated with 4.4 mL of 1 N HCl. The reaction mixture was stirred at 70 °C for 4 h. After cooling the miture was pured into 4.4 mL of 1.5 N NaOH, the formed precipitate was filtered and portioned between ethyl acetate and 1 N NaOH. The water phase was separated and acidify untyl pH = 4 with concentrated HCl. The formed solid was filtered, washed with water and recrystallized from absolute ethanol obtaining 430 mg of pure **7m.** 64%; 188-189 °C; ethanol; IR v 3200-2400 (NH, OH enole and acid), 1712 (C=O ketone) cm⁻¹. ¹H NMR (DMSO *d*₆) δ 4.50 (s, 1H, butanoate C3-H), 5.47 and 5.62 (s, 2H, CH₂ keto and enole form), 6.21 and 6.29 (t, 1H, pyrrole C4-H), 6.91 (s, 1H, butenoate C3-H), 7.06-7.16 (m, 6H, benzene H keto and enole form), 7.22 and 7.29 (m, 1H, pyrrole C3-H), 7.36 and 7.45 (s, 1H, pyrrole C2-H keto and enole form), 8.59 (bs, 1H, NH), 14 (br s, 1H, OH enole and acid). Anal. (C₁₆H₁₃FN₄O₂) C, H, N, F.

Biological methods. *RT expression and purification.* The recombinant HIV-1 RT protein, whose coding gene was subcloned in the p6HRT_prot plasmid, was expressed in *E. coli* strain M15.^{43,44} The bacteria cells were grown up to an OD₆₀₀ of 0.8 and induced with 1.7 mM IPTG for 5 hrs. HIV-1 RT purification was carried out as described. Briefly, cell pellets were resuspended in Lyses Buffer (20 mM Hepes pH 7.5, 0.5 M NaCl, 5 mM β -mercaptoethanol, 5 mM imidazole, 0.4 mg/mL lysozime), incubated on ice for 20 min, sonicated and centrifuged at 30,000 x *g* for 1 hr. The supernatant was applied to a His-binding resin column and washed thoroughly with Wash Buffer (20 mM Hepes pH 7.5, 0.3 M NaCl, 5 mM β -mercaptoethanol, 60 mM imidazole, 10% glycerol). The RT protein was eluted

with Elute Buffer and the enzyme-containing fractions were pooled, dialyzed and aliquots were stored at -80 °C.

HIV-1 RT RNase H Inhibition. The RT-associated RNase H function was measured in a polymeraseindependent cleavage assay, in which the Poly(dC)-[³H]Poly(rG) hybrid was used as reaction substrate as previously described.⁴³

HIV-1 IN Inhibition. HIV-1 IN gel-based assays were carried out as previously published.⁴⁵

HIV-1 Replication Inhibition. Compounds antiviral activity was determined in a cell-based assay according to the procedure described previously⁴⁶ and modified as follows. HeLa-CD4-LTR-β-gal cells were maintained in DMEM with 10% serum and 0.5 mg/mL G418. The day prior experimentation, 96-well plates were prepared to contain 10000 cells per well in 100 µL of Dulbecco's Modified Eagle's Medium (DMEM) medium complemented with 10% serum. On day one, each drug was serial diluted directly on cells following a 3-fold dilution over 6 points and each well was then filled to 200 µL with either fresh medium or concentrated viral supernatant (HIV-1(IIIB), Advanced Biotechnologies Inc.). The highest compound concentration tested was 50 µM. On day two, cells were washed three times with PBS before adding 200 µL of a solution containing 50 mM Tris-HCl pH 7.5, 100 mM β-mercaptoethanol, 0.05% Triton X100, and 5 mM 4-methyl-umbelliferyl-β-D-galactopyranoside (4-MUG, Sigma). On day three, sealed plates were read in a SpectraMax GEMINI-XS (Molecular Devices) with λex/em = 360/460 nm.

Cellular Toxicity. Similarly to the antiviral assays, plates were prepared with 10,000 HeLa-CD4-LTR- β -gal cells per well and a serial dilution of compounds in 100 µL. After 24 h of culture, 100 µL of ATPlite reagent (PerkinElmer) was added to each well. After 5 min at room temperature, plates' luminescence was quantified using an EnVision multilabel reader (PerkinElmer) according to manufacturer's instructions.

ASSOCIATED CONTENT

Supporting Information

Analyses of compounds **6a-l** and **7a-m**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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ABBREVIATIONS USED

IN, integrase; RT, reverse transcriptase; RNase H, ribonuclease H; 3'-P, 3'-processing; ST, strand transfer; DKA, diketo acid; BTDBA, 4-[5-(benzoylamino)thien-2-yl]-2,4-dioxobutanoic acid; SI, selectivity index; TosMIC, toluene-4-sulphonylmethyl isocyanide; NIS, N-iodosuccinimide; GP, general procedure; DMEM, Dulbecco's Modified Eagle's Medium.

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Table of Contents graphic \cap ЮH ЮH ≓> _ [Ó N $IC_{50(RH)} = 1.2 \ \mu M$ $IC_{50(RH)} = 3.0 \,\mu M$ F $IC_{50(IN)} = 1.8 \ \mu M$ $IC_{50(IN)} = 0.6 \,\mu M$ ratio ~ 1 ratio = 5 F