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Abstract: The synthesis of γ -lactam lignans from thuriferic acid via Michael addition of substituted anilines under basic conditions, followed by lactam ring closure, is described.

Key words: lignans, lactams, Michael additions, thuriferic acid, privileged structure

 γ -Lactams are important molecules with numerous pharmacological properties such as, psychotropic agents,¹ muscarinic acid agonists,² antihypertensive agents,³ peptide mimics,⁴ β-turn peptidomimetics,⁵ non-peptide mimics of somatostatin/sandostatin,⁶ serine protease inhibitors,⁷ and anti-asthma drug rolipram.⁸ Unlike β-lactams, to date γ -lactams have been found as components of only a few natural products such as fischerellin B,⁹ the antibiotic lactonamycin,¹⁰ and the alkaloid gelsemine.¹¹ Reports regarding the construction of γ -lactam libraries have been published.¹² Within this context, and in the search for ligands for a range of biological targets, we initiated a program to synthesize γ -lactam libraries based on the concept of privileged structure.¹³ The 1-aryltetralin system 1¹⁴ represents a privileged structure which remains to be exploited by combinatorial chemistry. Indeed, lignans of the type 1-aryltetralin are the constituents of plants or their synthetic derivatives and present a wide range of biological activities¹⁵ including inhibition of the polymerization of tubulin, inhibition of DNA topoisomerase II, immunosuppressive activity, and anti-HIV activity. Recently, cyclolignans as inhibitors of the insulin-like growth factor-1 receptor (IGF-1R) have been reported.¹⁶ Interestingly, the combination of 1 with γ -lactam rings was carried out by Kadow and co-workers¹⁷ with the synthesis of antitumor lactams from the anticancer drug etoposide, a glucoside derivative of the cytotoxic 1-aryltetralin lignan podophyllotoxin. We chose thuriferic acid $2^{18,19}$ a lignan isolated from Juniperus thurifera leaves,²⁰ which encompasses the 1-aryltetralin scaffold 1, and whose chemical reactivity has been the subject of a few studies^{18,19a} (Figure 1). In this paper we present the synthesis of γ -lactam lignans 3. which were synthesized by the conjugate addition of substituted anilines to the methyl ester of thuriferic acid 5, followed by intramolecular cyclization of the corresponding β -amino ketones. To our knowledge there is only one example²¹ of a Michael addition to a related substrate of **2**, 3-carboxy-2-methylene-4-(3',4',5'-trimethoxynamelv phenyl)-6,7-dimethoxy-1-tetralone, which was prepared from morpholine.



Figure 1





Scheme 1 *Reagents and conditions:* (a) *t*-BuOK, *t*-BuOH, reflux; (b) MeI, NaHCO₃, DMF, r.t.; (c) ArNH₂, Et₃N, THF, reflux; (d) *t*-BuOK (cat.), *t*-BuOH, DMF, r.t.

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Two protocols have been described^{18a,c} for the synthesis of thuriferic acid **2** from podophyllotoxone **4**. On a large scale, **2** was synthesized using the procedure of Höfert and Matusch.^{18a} To facilitate the purification step the crude acid thus obtained, after evaporation of *t*-BuOH, was converted into methyl ester **5** (Scheme 1).

The Michael addition of substituted anilines to 5 in the presence of Et₃N in THF at reflux afforded β -amino ketones 6a-i, which were characterized by ¹H NMR spectroscopy²² (Table 1). For halogenoanilines (Table 1, entries 1-4) and activated anilines (Table 1, entries 5 and 6), the reaction with 5 catalyzed by Et_3N (4 equiv) provided **6a–f** after 24 hours at reflux in THF in 78–95% yields. Intramolecular cyclization of these β -amino ketones was never observed when an excess of reactants and/or base were used. Importantly, treatment of 5 with the more basic 4-aminopyridine instead of Et₃N led to naphthoic acid methyl ester 7.23 3-Nitroaniline gave compound 6g in 43% yield after heating for 48 hours (Table 1, entry 7). 4-Trifluoroaniline provided 6h in 53% yield, regardless of the reaction time (Table 1, entry 8). Attempts to optimize the reaction with 4-nitroaniline were unsuccessful (Table 1, entries 9 and 10). The stereochemistry of 6a-i was assigned on the basis of the $J_{1,2}$ (8–11 Hz) and $J_{2,3}$ (9– 13 Hz) coupling constants which were in agreement with the 1,2 antidiaxial and 2,3 antidiaxial configurations previously reported for 11-chloro-3,11-dihydrothuriferic acid.18b,c

As expected, the β -amino ketone **6a** proved sensitive to retro-Michael reaction, for example, the conversion of the carbonyl group at C-4 into the corresponding methyloxime²⁴ was attempted under two different conditions and failed (MeONH₂·HCl, NaOAc in MeOH–THF, reflux^{25a} or MeONH₂·HCl, pyridine·HCl, EtOH, reflux^{25b}). Addition of arylhydrazines²⁶ to **6a** gave the same result. Finally, cyclization of **6a–i** into the desired γ -lactam lignans **3a–i**^{27,28} occurred under base catalysis²⁹ (*t*-BuOK) in 45–72% yields (Table 1). Surprisingly, while picropodophyllone (C-2 epimer of **4**) reacted at C-4 with *O*-methylhydroxylamine as described,³⁰ the corresponding 4-oxime derivatives of **3a** failed to form under the same reaction conditions.

In summary, the synthesis of γ -lactam lignans **3** was achieved from thuriferic acid **2** via Michael addition of substituted anilines under basic conditions followed by lactam ring closure. Extension of this work to construct diversified libraries of γ -lactam lignans for biological evaluation, using resins and parallel solution methodologies, is currently under investigation.

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LETTER

	Michael addition of anilines $5 \rightarrow 6a-i$		γ-Lactams 6a−i → 3a−i		
Entry	Aniline	Product ^a	Yield (%) ^b	Product ^a	Yield (%) ^b
1°	H ₂ N Br	6a	94	3a	63
2°	H ₂ N Br	6b	95	3b	69
3°	H ₂ N	6с	78	3c	67
4°	H ₂ N F	6d	78	3d	72
5°	H ₂ N OMe	6e	90	3e	65
6 ^c	H ₂ N Me	6f	93	3f	64
7 ^d		6g	43	3g	45
8 ^{c,d}	H ₂ N CF ₃	6h	53	3h	53
9 ^{d,e}	- NO ₂	6i	23	3i	65
10 ^f	HaN	6i	31	3i	65

^a All products were characterized by ¹H NMR, ¹³C NMR, IR, and mass spectroscopy.

^b All yields reported are isolated yields of compounds, estimated to be 95% pure by ¹H NMR spectroscopy.

^c Et₃N (4 equiv), aniline (4 equiv), 24 h.

^d Et₃N (4 equiv), aniline (4 equiv), 48 h.

^e Et_3N (4 equiv), aniline (4 equiv), 5 d.

^f Et₃N (10 equiv), aniline (10 equiv), 5 d.

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Figure 2

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(27) Synthesis of 6 and 3; General Procedure

To a solution of the methyl ester of thuriferic acid 5 (50 mg, 0.117 mmol) in anhyd THF (2 mL) were added Et₃N and the aniline at r.t. (Table 1). The reaction mixture was then heated at 65 °C for the reaction time indicated, concentrated under reduced pressure, and the crude product was purified by chromatography on silica gel (cyclohexane-EtOAc, 5:2) to give the desired β -amino ketones 6. This compound (0.165 mmol, 1 equiv) was diluted in DMF (2.5 mL) and a 1 M solution of t-BuOK in t-BuOH (1 M; 16.5 µL, 0.1 equiv) was added at r.t. The mixture was stirred for 20 min then the pH was adjusted to 7 by the addition of aq NH₄Cl. The mixture was extracted with EtOAc (3×20 mL), the combined organic phases were dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by chromatography on silica gel (cyclohexane-EtOAc, 3:1) to furnish the γ -lactam lignans **3**.

Compound **6i**: Yellow powder; yield: 31%; mp 194 °C; $[\alpha]_D^{20}$ -84 (*c* 0.19, CHCl₃). IR: 3400–3300, 2940, 1734, 1671, 1601, 1506, 1480 cm⁻¹. ¹H NMR (300 MHz, CDCl₃):

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$$\begin{split} &\delta = 8.07 \ (d, 2 \ H, J = 9.2 \ Hz, H_{3''}, H_{5''}), 7.47 \ (s, 1 \ H, H_5), 6.55 \\ &(d, 2 \ H, J = 9.2 \ Hz, H_{2''}, H_{6''}), 6.33 \ (s, 2 \ H, H_{2'}, H_{6'}), 6.29 \ (s, 1 \ H, H_8), 6.00 \ (m, 2 \ H, OCH_2O), 5.19 \ (br \ s, 1 \ H, NH), 4.36 \\ &(d, 1 \ H, J = 10.7 \ Hz, H_1), 3.86 \ (s, 3 \ H, OMe_{4'}), 3.80 \ (s, 6 \ H, OMe_{3',5'}), 3.63 \ (m, 1 \ H, H_{11a}), 3.49 \ (s, 3 \ H, CO_2Me), 3.48 \\ &(m, 1 \ H, H_{11b}), 3.24 \ (dd, 1 \ H, J = 12.9, 10.7 \ Hz, H_2), 3.19 \ (m, 1 \ H, H_{31}), ^{13}C \ NMR \ (75 \ MHz, acetone-d_6): \delta = 195.8, 174.0, 155.8, 155.5, 154.4, 149.2, 143.7, 139.6, 139.1, 138.5, 128.4, 127.7, 112.9, 110.0, 108.6, 106.7, 104.1, 61.5, 57.4, 54.0, 53.1, 50.9, 50.0, 43.7. \ MS \ (DCI, NH_3): m/z = 565 \ [M + H]^+. \end{split}$$

Compound **3i**: Yellow powder; yield: 65%; mp 235–240 °C; $[a]_D^{20} -110$ (*c* 0.34, CHCl₃). IR: 2940, 1716, 1670, 1597, 1521, 1481 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): $\delta = 8.22$ (d, J = 9.3 Hz, 2 H, H_{3"}, H_{5"}), 7.79 (d, J = 9.3 Hz, 2 H, H_{2"}, H_{6"}), 7.48 (s, 1 H, H₅), 6.73 (s, 1 H, H₈), 6.27 (s, 2 H, H_{2"}, H_{6"}), 6.04 (m, 2 H, OCH₂O), 4.81 (d, J = 1.7 Hz, 1 H, H₁), 4.38 (d, J =9.7 Hz, 1 H, H_{11a}), 4.01 (m, 1 H, H_{11b}), 3.81 (s, 3 H, OMe_{4'}), 3.76 (s, 6 H, OMe_{3',5'}), 3.40 (dd, J = 7.6, 1.7 Hz, 1 H, H₂), 3.29 (m, 1 H, H₃). ¹³C NMR (75 MHz, CDCl₃): $\delta = 193.9$, 172.3, 153.7, 153.5, 144.1, 143.6, 139.6, 138.1, 137.0, 126.9, 124.5, 118.6, 109.3, 105.8, 104.6, 102.0, 60.6, 56.0, 50.5, 43.3, 42.9, 39.5. MS (DCI, NH₃): m/z = 533 [M + H]⁺, 550 [M + NH₄]⁺.

(28) The 2,3-*cis* stereochemistry of **3a**–**i** was deduced from the $J_{1,2}$ and $J_{2,3}$ coupling constants (1.7 and 7.6 Hz, respectively for **3i**) and was confirmed from NOESY correlations of H₂/H₃, H₂/H_{2',6'}, and H₃/H_{2',6'}. Molecular modeling [Insight II,



Figure 3

Discover, MD simulations (300 K), cff 91, $\varepsilon = 4.8$ for CDCl₃] provided a unique global minimum conformation for **3i** which fitted the NOE data (Figure 3).

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