

Steroidal guanidinium receptors for the enantioselective recognition of *N*-acyl α -amino acids

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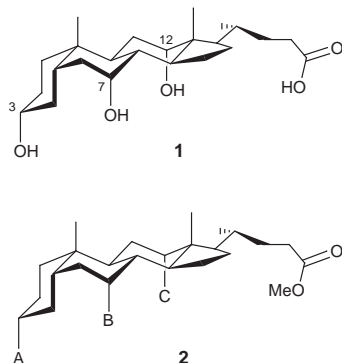
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Guanidinium cations **4 and **5** extract *N*-acetyl α -amino acids into CHCl_3 from an aqueous medium with enantiomeric excesses of up to 80%.**

Enantioselective recognition is of continuing interest in supramolecular chemistry,¹ especially where relevant to the large-scale resolution of racemates. Enantioselective phase transfer is particularly attractive, raising the possibility of 'catalytic' resolutions based on the transport of substrates through otherwise impermeable barriers.² Carboxylates/carboxylic acids are suitable targets for this approach, because of their ability to partition between aqueous and organic phases. Amino acids, and their *N*-acyl derivatives, are attractive substrates because of their biological significance and practical importance.³

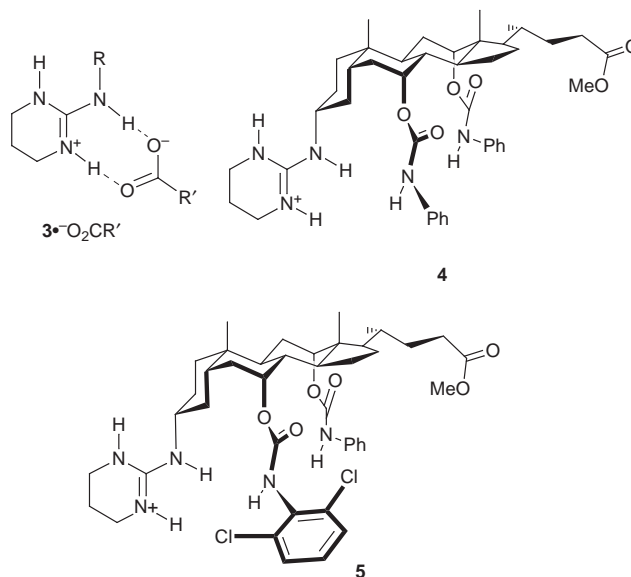
Herein we describe a new strategy for the enantioselective extraction of chiral carboxylates from aqueous into organic media, and its realisation in the form of receptors which show significant enantiodiscrimination in the case of *N*-acetyl α -amino acids. In common with previous work on carbohydrate⁴ and anion recognition,⁵ our design exploits cholic acid **1** as a starting material. The secondary hydroxy groups in **1** can be independently modified to give differentiated, co-directed substituents as in **2**.⁶ Groups A–C are spaced so as to allow, in



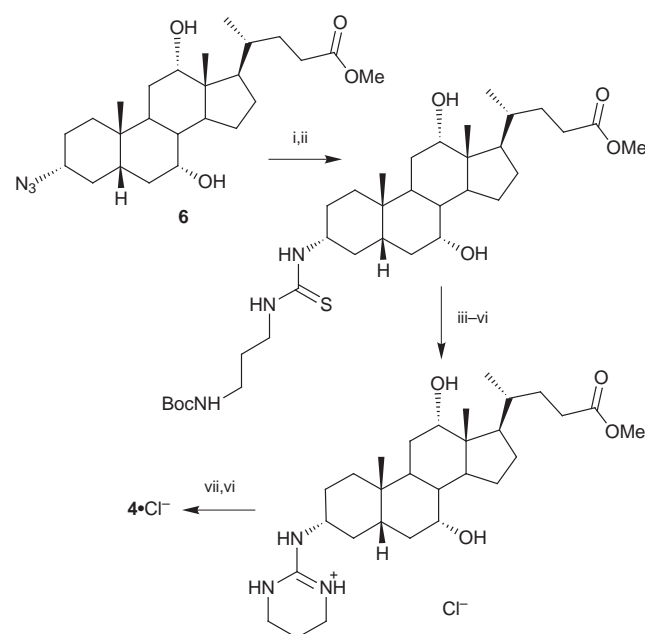
a typical case, cooperative effects on a substrate with minimum interference from intramolecular interactions. The design is suggestive of 'three-point contact', as required for the classical model of enantioselective recognition.⁷

For carboxylate extraction, it is useful that one of groups A–C should form a specific, electroneutral complex with the anionic centre. To serve this purpose we have chosen the guanidinium unit **3**, capable of binding carboxylates as shown.⁸ Of the possible variations on our general theme, we report the synthesis and properties of two initial examples; the bis-phenylcarbamate **4** and its asymmetrically-substituted relative **5**. Receptor **4** was accessible from 3 α -azide **6**⁹ as shown in Scheme 1, while receptor **5** was prepared *via* a longer sequence with alcohol **7** as the penultimate intermediate.

As anticipated, solutions of **4**·Cl[−] and **5**·Cl[−] in CHCl_3 were capable of extracting carboxylic acids from neutral or basic aqueous solutions, presumably through exchange of carboxylate for chloride. In the case of *N*-acetyl α -amino acids the ¹H NMR spectra of the complexed substrates were enantiomer-



dependent, allowing the determination of enantioselectivities, as well as extraction efficiencies, by simple integration. The results are shown in Table 1. Extraction efficiencies were moderate to good for substrates with non-polar side-chains, although neither receptor was effective with the polar asparagine derivative. Receptor **4** proved remarkably consistent in its



Scheme 1 Reagents and conditions: i, Zn dust, AcOH, room temp., 24 h; ii, $\text{SCN}(\text{CH}_2)_3\text{NHBoc}$, Pr_2NEt , dry CH_2Cl_2 , room temp., 72 h; iii, MeI, MeOH, reflux; iv, TFA, CH_2Cl_2 , room temp.; v, Pr_2NEt , MeOH, room temp., 24 h; vi, aq. NaOH then aq. HCl; vii, PhNCO , conc. aq. HCl (cat.), $\text{CH}_2\text{ClCH}_2\text{Cl}$, reflux, 72 h.

Table 1 Extractions by **4**-Cl[−] and **5**-Cl[−] of racemic *N*-acetyl α -amino acids from aqueous buffer (pH 7.4) into CHCl₃^a

Substrate	Receptor 4		Receptor 5	
	Extraction efficiency (mol%) ^b	Enantio-selectivity (L:D) ^c	Extraction efficiency (mol%) ^b	Enantio-selectivity (L:D) ^c
<i>N</i> -Ac-alanine	52	7:1	76	6:1
<i>N</i> -Ac-phenylalanine	87	7:1	93	9:1
<i>N</i> -Ac-tryptophan	83	7:1	92	6:1
<i>N</i> -Ac-valine	71	7:1	89	9:1
<i>N</i> -Ac- <i>tert</i> -leucine	<i>d</i>	<i>d</i>	82	5:2
<i>N</i> -Ac-methionine	<i>d</i>	<i>d</i>	93	9:1
<i>N</i> -Ac-proline	<i>d</i>	<i>d</i>	74	4:1
<i>N</i> -Ac-asparagine	0	—	0	—

^a Solutions of receptor in CHCl₃ (6 mM, 1 ml), and substrate in phosphate buffer (7–8 mM, 5 ml), were stirred vigorously for 2 h. The organic phases were isolated, dried by passage through hydrophobic filter paper, then evaporated. The residues were dissolved in CDCl₃ (0.6 ml) and analysed by ¹H NMR spectroscopy. ^b Determined by ¹H NMR integration of substrate α -CH and NH vs. 7/12 β -H of receptor. ^c Determined by ¹H NMR integration of α -CH and NH signals for enantiomers of substrates. Assignments confirmed through control experiments with enantiopure substrates. ^d Not determined.

ability to differentiate between enantiomers, irrespective of side-chain bulk. Receptor **5** showed generally higher extraction abilities, possibly due to the greater acidity of the dichlorophenylcarbamoyl NH, and was more sensitive to side-chain structure. Perhaps surprisingly, the substrate with the most sterically hindered asymmetric centre (*N*-Ac-*tert*-leucine) gave the lowest selectivity.

¹H NMR spectroscopy and molecular modelling combined to suggest plausible models for the binding geometries. A Monte Carlo Molecular Mechanics (MCM) search† on the complex between **5** and *N*-acetyl-L-valinate **8** yielded the configuration shown in Fig. 1. The carboxylate accepts H-bonds from the 7-carbamoyl and two guanidinium NH groups, while the acetyl oxygen is bound to the 12-carbamoyl NH. In support of this structure, the receptor carbamate and 2 of the 3 guanidinium NH signals moved downfield on complex formation, while a weak

intermolecular NOE was observed from the α -CH in **L-8** to the *ortho* protons of **5**-NHPH (consistent with Fig. 1, allowing for some rotational freedom about the N–Ph bond‡). A similar MCM search† on **5** + **D-8** yielded a higher-energy structure in which the acetyl O...HN interaction is missing, the 12-carbamoyl NH forming a fourth (apparently strained) hydrogen bond to the carboxylate.

Viewed as forerunners of an extended family of receptors, **4** and **5** show encouraging levels of enantioselectivity. Many variants are within easy reach, a majority with much greater differentiation between the three substituents. We hope to report examples with improved performance in the foreseeable future.

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Notes and references

† Calculations employed MacroModel V5.5 (ref. 10), the Amber* force field, CHCl₃ GB/SA solvation, and 5000 steps of MCM. Six and three separate searches were conducted for the L and D substrates respectively, all from widely differing starting geometries and all yielding essentially similar final structures.

‡ Rotation about N–Ph allows an *ortho* proton to make van der Waals contact with the substrate α -CH.

- 1 T. H. Webb and C. S. Wilcox, *Chem. Soc. Rev.*, 1993, **22**, 383.
- 2 M. Newcomb, J. L. Toner, R. C. Hegelson and D. J. Cram, *J. Am. Chem. Soc.*, 1979, **101**, 6294.
- 3 Enantioselective extraction/transport of amino acids or *N*-acyl amino acids : (a) J. L. Sessler and A. Andrievsky, *Chem. Eur. J.*, 1998, **4**, 159; (b) N. Voyer and B. Guerin, *Chem. Commun.*, 1997, 2329; (c) J. Y. Zheng, K. Konishi and T. Aida, *Tetrahedron*, 1997, **53**, 9115; (d) K. Konishi, K. Yahara, H. Toshihige, T. Aida and S. Inoue, *J. Am. Chem. Soc.*, 1994, **116**, 1337; (e) A. Metzger, K. Gloe, H. Stephan and F. P. Schmidtchen, *J. Org. Chem.*, 1996, **61**, 2051; (f) H. Tsukube, J. Uenishi, T. Kanatani, H. Itoh and O. Yonemitsu, *Chem. Commun.*, 1996, 477; (g) G. J. Pernia, J. D. Kilburn and M. Rowley, *J. Chem. Soc., Chem. Commun.*, 1995, 305; (h) A. Galán, D. Andreu, A. M. Echavarren, P. Prados and J. de Mendoza, *J. Am. Chem. Soc.*, 1992, **114**, 1511; (i) J. de Mendoza and F. Gago, in *Computational approaches in supramolecular chemistry*, ed. G. Wipff, Kluwer Academic Publishers, 1994, p. 79. Ref. 3(a) serves as a leading reference to amino acid recognition in general.
- 4 A. P. Davis, *Chem. Soc. Rev.*, 1993, **22**, 243; A. P. Davis, R. P. Bonar-Law and J. K. M. Sanders, in *Comprehensive Supramolecular Chemistry*, ed. Y. Murakami, Pergamon, Oxford, 1996, vol. 4 (*Supramolecular Reactivity and Transport: Bioorganic Systems*), p. 257.
- 5 A. P. Davis, J. F. Gilmer and J. J. Perry, *Angew. Chem., Int. Ed. Engl.*, 1996, **35**, 1312; A. P. Davis, J. J. Perry and R. P. Williams, *J. Am. Chem. Soc.*, 1997, **119**, 1793.
- 6 Podand-type receptors derived from the bile acids have been reported by other groups. See for example: R. Boyce, G. Li, H. P. Nestler, T. Suenaga and W. C. Still, *J. Am. Chem. Soc.*, 1994, **116**, 7955; Y. A. Cheng, T. Suenaga and W. C. Still, *J. Am. Chem. Soc.*, 1996, **118**, 1813; L. J. D'Souza and U. Maitra, *J. Org. Chem.*, 1996, **61**, 9494. However, none of these systems possesses three differentiated α -substituents as implied for **2**.
- 7 C. Dalglish, *J. Chem. Soc.*, 1952, 3940.
- 8 The analogous five-membered ring has been widely used in carboxylate and phosphate receptors. For examples, see: E. Fan, S. A. V. Arman, S. Kincaid and A. D. Hamilton, *J. Am. Chem. Soc.*, 1993, **115**, 369; M. S. Muche and M. W. Göbel, *Angew. Chem., Int. Ed. Engl.*, 1996, **35**, 2126; A. Metzger, V. M. Lynch and E. V. Anslyn, *Angew. Chem., Int. Ed. Engl.*, 1997, **36**, 862. The six-membered **3** was chosen for the present work because of its greater stability and lipophilicity, and because it was expected to hold a substrate closer to the α -face of the steroid.
- 9 For a two-step synthesis of **6** from methyl cholate, see: A. P. Davis, S. Dresen and L. J. Lawless, *Tetrahedron Lett.*, 1997, **38**, 4305.
- 10 F. Mohamadi, N. G. J. Richards, W. C. Guida, R. Liskamp, C. Caufield, G. Chang, T. Hendrickson and W. C. Still, *J. Comput. Chem.*, 1990, **11**, 440.

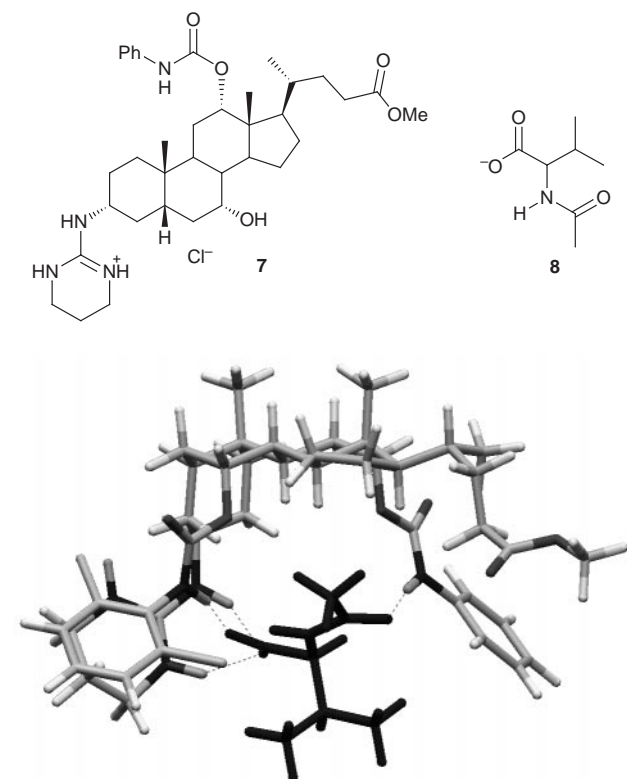


Fig. 1 Structure of **5** + **L-8** (black) derived from computer-based molecular modelling. Intermolecular hydrogen bonds are shown as broken lines.

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