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**Chemoenzymatic Synthesis of (R) - and (S) - Atenolol and Propranolol employing
Lipase Catalyzed Enantioselective Esterification and Hydrolysis**

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Abstract : Chemoenzymatic synthesis of (R) - and (S) - atenolol and propranolol employing lipase catalyzed enantioselective esterification and hydrolysis is described.

In a drug molecule the stereochemistry plays an important role in biological activity¹. In case of 2 - propanolamines e. g. atenolol and propranolol, the β - blockers, the activity is associated with the (S) - isomer. On the contrary the opposite (R) - enantiomer may be responsible for the side effects². There have been considerable efforts done in the preparation of enantiomerically pure (S) - propanolamines by employing biocatalytic processes^{3,4}. There are several reports regarding the chiral O - acyl derivatives of aryloxypropanolamines served as prodrugs⁵⁻⁷.


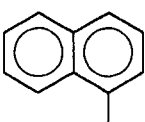
The synthesis of (R) - and (S) - atenolol have been achieved by lipase catalyzed hydrolysis³ of the intermediate O - acetyl esters. The synthesis of (R) - and (S) -

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propranolol have been achieved by using lipase catalyzed hydrolysis^{2,8} of the intermediate O - acetyl esters and also by using lipase catalyzed acylation^{9,10} of the intermediate secondary alcohols *Rhizopus arrhizus*^{11,12} have been employed for enantioselective hydrolysis of acid esters and also its lipase¹³ was employed for enantioselective hydrolysis of the secondary alcohol esters. Lipase of *Pseudomonas cepacia* (PS) was used for enantioselective O- esterification of secondary alcohol function^{2,3,8,9}. The above advantages of lipases to catalyze the enantioselective reactions (e. g. esterification, hydrolysis) and also the importance of the chiral drugs prompted us to develop a methodology for the synthesis of chirally pure atenolol and propranolol. In present methodology we report here the direct resolution of (\pm) - atenolol and (\pm) - propranolol using lipase catalysed enantioselective synthesis, by employing enantioselective esterification and hydrolysis.

The methodology consists of esterification of (\pm) - atenolol and (\pm) - propranolol with vinyl acetate [Table - 1, Scheme - 1] or succinic anhydride [Table-2, Scheme - 2] in the presence of lipase PS - D to give (S) - alcohol and (R) - ester. The products were separated by column chromatography and the intermediate (R) - ester [confirmed by co-TLC with authentic sample], was hydrolyzed [using K_2CO_3 -MeOH] to give (R) - alcohol. Further the corresponding O - acetyl ester of (\pm) - atenolol and (\pm) - propranolol were subjected to enantioselective hydrolysis [Table -3, Scheme -3] using lipase of *Rhizopus arrhizus* to produce (R) - alcohol and (S) - ester. The products were separated by column chromatography and the intermediate (S) - ester [confirmed by co -TLC with authentic sample], was hydrolyzed [using K_2CO_3 -MeOH] to give (S) - alcohol. The compounds were obtained in good optical purity and isolated yields (35 - 45 %).

Table 1 :

Substrate 1	Ar -	Product 2 : conf. ⁿ , ee, (yield [♣])	Product 3 : conf. ⁿ , ee, (yield [♣])
a		a* : (S), 94 %, (42 %) [a] _D = - 15.0 (0.5, EtOH), Lit. ⁴ [a] _D = - 16.0 (1, EtOH)	a : (R), 82 %, (40 %) [a] _D = + 13.0 (1, EtOH), Lit. ⁴ [a] _D = + 16.0 (1, EtOH)
b		b** : (S), 98 %, (45 %) [a] _D = - 10.0 (1, EtOH), Lit. ³ [a] _D = - 10.2 (1.02, EtOH)	b : (R), 88 %, (43 %) [a] _D = + 9.0 (0.5, EtOH), Lit. ³ [a] _D = + 10.2 (1.02, EtOH)

♣ The yields are isolated yields. conf.ⁿ = configuration

* ¹H NMR (DMSO-d₆) δ 1.1 (d, 6H), 2.5 (m, 1H), 2.6-2.8 (m, 2H), 3.4 (s, 2H), 3.8 (s, 2H), 3.9 (m, 1H), 5.0 (s, 4H), 6.8 (d, 2H), 7.2 (d, 2H).

** ¹H NMR (CDCl₃) : δ 1.1-1.2 (d, 6H), 2.8-3.1 (m, 3H), 4.1-4.3 (d, 3H), 6.8-8.3 (m, 7H).

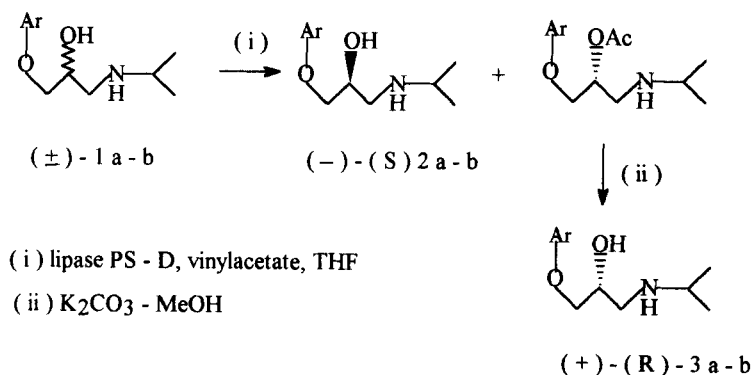
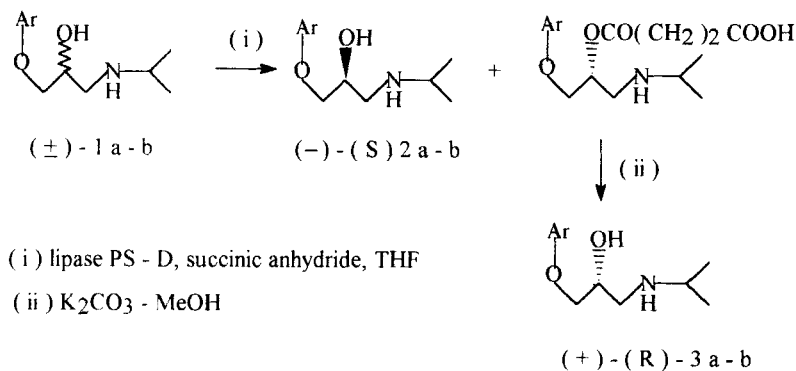
Scheme 1 :

Table 2 :

Substrate 1	Ar -	Product 2 : conf. ⁿ , ee, (yield [*])	Product 3 : conf. ⁿ , ee, (yield [*])
a		a : (S), 94 %, (40%) [a] _D = - 15.0 (1, EtOH), Lit. ⁴ [a] _D = - 16.0 (1, EtOH)	a : (R), 88 %, (38 %) [a] _D = + 14.0 (2, EtOH), Lit. ⁴ [a] _D = + 16.0 (1, EtOH)
b		b : (S), 98 %, (45 %) [a] _D = - 10.0 (1, EtOH), Lit. ³ [a] _D = - 10.2 (1.02, EtOH)	b : (R), 88 %, (38 %) [a] _D = + 9.0 (1, EtOH), Lit. ³ [a] _D = + 10.2 (1.02, EtOH)

♣ The yields are isolated yields. conf.ⁿ = configuration

Scheme 2 :

In order to optimise the reaction on larger scale, esterification of (±) - atenolol (25 mmol, 6.65 g) was carried by lipase PS - D (2.25 g) by usual procedure to give (S) - atenolol (94 % ee, 42 % yield^{*}) along with (R) - atenolol (after hydrolysis of the corresponding ester) (82 % ee, 40 % yield^{*}). This reaction was completed after 12h as

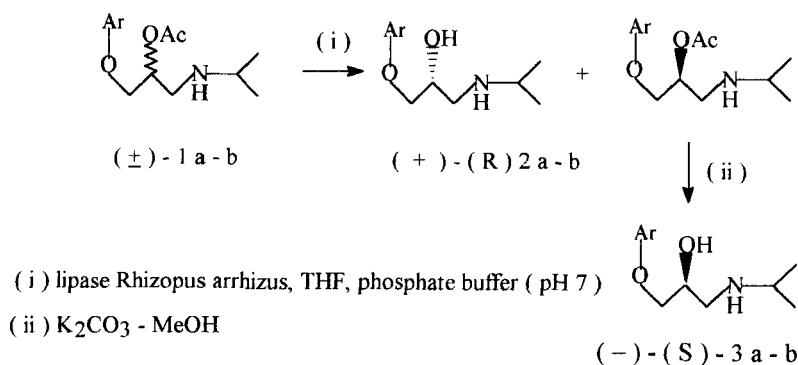
Table 3 :

Substrate I	Ar -	Product 2 : conf. ⁿ , ee, (yield [♣])	Product 3 : conf. ⁿ , ee, (yield [♣])
a		a* : (R), 82 %, (37%) [a] _D = + 13.0 (2, EtOH), Lit. ⁴ [a] _D = + 16.0 (1, EtOH)	a : (S), 69 %, (44%) [a] _D = - 11.0 (1, EtOH), Lit. ⁴ [a] _D = - 16.0 (1, EtOH)
b		b** : (R), 98 %, (35%) [a] _D = + 10.0 (0.5, EtOH), Lit. ³ [a] _D = +10.2 (1.02, EtOH)	b : (S), 78 %, (43 %) [a] _D = - 8.0 (1, EtOH), Lit. ³ [a] _D = - 10.2 (1.02, EtOH)

♣ The yields are isolated yields. conf.ⁿ = configuration

* ¹H NMR (DMSO-d₆) δ 1.1 (d, 6H), 2.5 (m, 1H), 2.6-2.8 (m, 2H), 3.4 (s, 2H), 3.8 (s, 2H), 3.9 (m, 1H), 5.0 (s, 4H), 6.8 (d, 2H), 7.2 (d, 2H).

** ¹H NMR (CDCl₃) : δ 1.1-1.2 (d, 6H), 2.8-3.1 (m, 3H), 4.1-4.3 (d, 3H), 6.8-8.3 (m, 7H).

Scheme 3 :

compared to the same reaction performed on a small scale which required 10h for completion.

Experimental :

THF was dried over sodium wire. All reactions were monitored by TLC [thin layer chromatography] on glass plates coated with 0.25 mm layer of silica gel. Compounds visualized by iodine vapours or in UV light. Column chromatography was done with silica gel 60 - 120 mesh, Merck. The substrate (\pm) - atenolol¹⁵, (\pm) - propranolol¹⁶ and their corresponding O - acetyl esters were prepared by literature procedure⁶. Lipase 'PS - D' [lipase PS immobilized on " diatomite "] was obtained from ' Amano Pharmaceutical Co., Ltd. Japan.' The *Rhizopus arrhizus* lipase was obtained from Fluka. ¹H NMR spectra were recorded on (MSL) 300 MHz spectrometer, using CDCl₃ or DMSO - d₆ as solvents. Optical rotation measurements were done on JASCO - 360 polarimeter.

General procedure for the Enantioselective Esterification of (\pm) - atenolol and (\pm) - propranolol with vinyl acetate [or succinic anhydride] :

A mixture of 5 mmol (\pm) - atenolol / (\pm) - propranolol, 6 mmol vinyl acetate/ 5 mmol succinic anhydride in 10 ml THF and 0.75 mg lipase PS - D was stirred at room temperature for 10 h. [reaction progress was monitored by TLC]. The reaction mixture was filtered through celite pad to remove the enzyme, washed with MeOH / CHCl₃. The combined organic solvent was dried over Na₂SO₄ and evaporated under reduced pressure. The products were separated by column chromatography. [atenolol - CHCl₃ : MeOH 8 : 2 / propranolol - CHCl₃ : MeOH 9.8 : 0.2] or [atenolol - Ethyl acetate : MeOH 9.5 : 0.5 / propranolol - Ethyl acetate].

General procedure for the Enantioselective Hydrolysis of (\pm) - O - acetyl ester of atenolol and propranolol :

A mixture of 1 g (\pm) - O - acetyl ester of atenolol / propranolol, 10 ml THF,

10 ml 0.1M phosphate buffer (pH 7) and 0.5 g lipase *Rhizopus arrhizus* were stirred at room temperature for 20 h. [reaction progress was monitored by TLC]. The reaction mixture was filtered through celite pad to remove the enzyme, washed with n - BuOH or CHCl₃ . Aqueous phase was extracted with n - BuOH (for atenolol) or CHCl₃ (for propranolol). The organic phase was dried over Na₂SO₄ and evaporated under reduced pressure. The products were separated by column chromatography. [atenolol - CHCl₃ : MeOH 8 : 2 / propranolol - CHCl₃ : MeOH 9.8 : 0.2].

In summary, we have developed a direct method to synthesize both the enantiomers of the two important β - blockers. As other important β - blockers like practolol, oxprenolol, metoprolol, acebutolol, moprolool, etc. are all closely related to atenolol and propranolol¹⁷, our present route should serve as a protocol for all these chiral drugs.

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References :

1. Witiak, D. T.; Inbasekaram, M. N. " Krik - Othmer Encyclopedia of Chemical Technology, 3rd Ed. " , Wiely - Interscience : New York, **1982**, 17, 311.
2. Jamali, F.; Mehvar, R.; Pasutto, F. M. *J. Pharm. Sci.*, **1989**, 78, 695.
3. Bevinakatti, H. S.; Banerji, A. A. *J. Org. Chem.*, **1991**, 56, 5372.
4. Bevinakatti, H. S.; Banerji, A. A. *J. Org. Chem.*, **1992**, 57 (22), 6003.
5. Irwin, W. J.; Belard, K. A. *Int. J. Pharm.*, **1988**, 46 (1 - 2), 57.
6. Irwin, W. J.; Belard, K. A. *Drug Dev. Ind. Pharm.*, **1987**, 13 (9 - 11), 2017.
7. Anders, B.; Hams, L.; Vincent, H. L. *Int. J. Pharm.*, **1988**, 42 (1 - 3), 51.

8. Matsuo, N.; Ohno, N. *Tetrahedron Lett.*, **1985**, 26 (45), 5533.
9. Lundell, K.; Kanerva, L. T. *Tetrahedron : Asymmetry*, **1995**, 6 (9), 2281.
10. Wen, C. T. ; Chi, C. C.; Torr, L. C.; Fu, T. D. *Bioorg. Med. Chem. Lett.*, **1997**, 7 (4), 433.
11. Salvi, N. A.; Patil, P. N.; Udupa, S. R.; Banerji, A. *Tetrahedron : Asymmetry*, **1995**, 6 (9), 2287.
12. Patil, P. N.; Salvi, N. A.; Udupa, S. R.; Banerji, A. *Biotechnology Lett.*, **1996**, 18 (2), 159.
13. Ming, G. Q.; Reddy, D. R.; Sih, C. J. *Tetrahedron Lett.*, **1986**, 27 (43), 5203.
14. Lednicer, Mitscher, L. A. " The Organic Chemistry of Drug Synthesis, John Wiley and Sons : New York, **1980**, 2, 109.
15. Capler, V., *Anal. Profiles Drugs Subt.*, **1984**, 13, 1.
16. Kaimal, T. N. B. ; Prasad, R. N. B. *Biotechnology Lett.*, **1992**, 14 (1), 21.
17. Lednicer, D. , Mitscher, L. A. " The Organic Chemistry of Drug Synthesis" , John Wiley and Sons : New York, **1980**, 2, 109.

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