

SYNTHESIS OF A BRANCHED D-GLUCOTETRAOSE, THE REPEATING UNIT OF THE EXTRACELLULAR POLYSACCHARIDES OF *Grifola umbellata*, *Sclerotinia libertiana*, *Porodisculus pendulus*, AND *Schizophyllum commune* FRIES*

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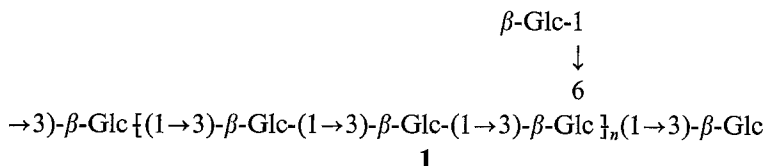
(Received October 14th, 1981; accepted for publication, November 21st, 1981)

ABSTRACT

The synthesis of D-glucotetraose, 3-O-[3-O-β-D-glucopyranosyl-β-D-glucopyranosyl]-6-O-β-D-glucopyranosyl-α (and β)-D-glucopyranose, the repeating unit of the extracellular polysaccharides of *Grifora umbellata*, *Sclerotinia libertiana*, *Porodisculus pendulus*, and *Schizophyllum commune* Fries, is described.

INTRODUCTION

Recently, it was proposed that the structure of the extracellular polysaccharides of *Grifola umbellata*², *Sclerotinia libertiana*³, and *Porodisculus pendulus*⁴ is **1**. Water-soluble D-glucan **1** from *Grifora umbellata* had been reported⁵ to inhibit the growth of subcutaneously implanted Sarcoma 180 in mice. Quite recently, schizophyllan, a water-soluble β-D-glucan elaborated by *Schizophyllum commune* Fries, was partially depolymerized by ultrasonic irradiation to a low-molecular-weight polysaccharide⁶ that exhibited the same antitumor activity against Sarcoma-180 as the native schizophyllan, and the structure of which was confirmed to be **1**, as proposed earlier for the structure of the native D-glucan⁷.

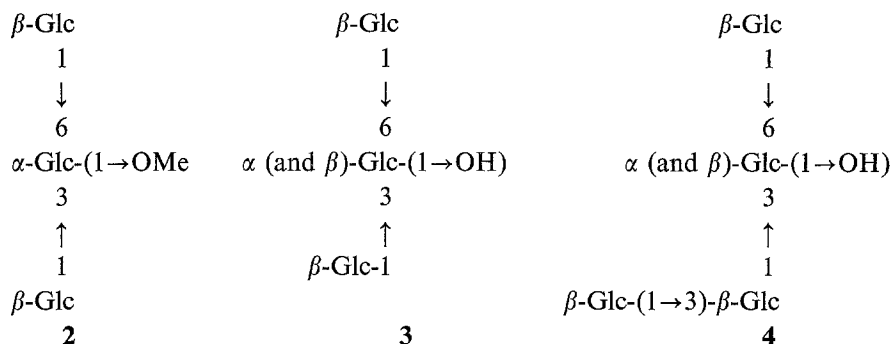


A similar (1→3)-β-D-glucan structure was also observed for the extracellular polysaccharides of *Rhizobium japonicum*⁸, which may play a role in the specificity of symbioses between *Rhizobium* species and their plant hosts⁹. A family of β-D-

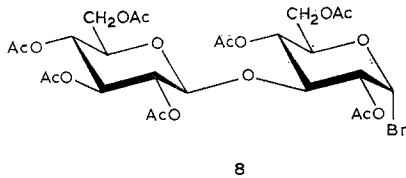
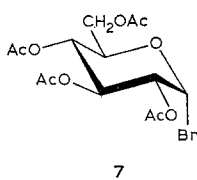
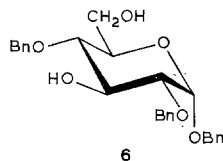
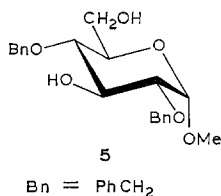
*Glucan Synthesis, Part II. For Part I, see ref. 1.

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glucans of very similar structure had been isolated from the soybean pathogen *Phytophthora megasperma* var. *sojae*, and they were demonstrated to elicit the formation of phytoalexins from soybeans and potatoes¹⁰.



In view of the biological significance, the repeating unit (**4**) of the β -D-glucan structure **1**, as well as simpler structures **2** and **3**, were chosen as the synthetic targets for the present study. Retrosynthetic examination of these target molecules demonstrated the necessary synthons **5** and **6** as two glycosyl acceptors, and **7** and **8** as two glycosyl donors.

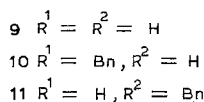
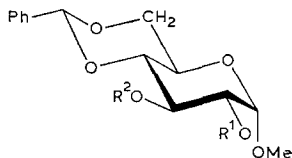


RESULTS AND DISCUSSION

Synthesis of methyl 3,6-di-O-β-D-glucopyranosyl-β-D-glucopyranoside (2). — Until 1979, 3,6-di-O-β-D-glucopyranosyl-D-glucopyranose derivatives had, to the best of our knowledge, been synthesized¹¹ in low yield as byproducts of the Koenigs-Knorr reactions between 2,3,4,6-tetra-O-acetyl-α-D-glucopyranosyl bromide (**7**)

and alkyl 4,6-*O*-benzylidene- β -D-glucopyranosides. In 1980, Eby and Schuerch¹² reported a regioselective synthesis of 2-(4-aminophenyl)ethyl 3,6-di-*O*- β -D-glucopyranosyl- α -D-glucopyranoside that employed, as the key glycosyl acceptor, 2-[4-(*p*-toluenesulfonamido)phenyl]ethyl 2,4-di-*O*-benzyl- α -D-glucopyranoside, which, in turn, was prepared *via* 1,6-anhydro-2,4-di-*O*-benzyl- β -D-glucopyranose¹³. Quite recently, Koto *et al.*¹⁴ reported a regioselective synthesis of 3,6-di-*O*- β -D-glucopyranosyl- β -D-glucopyranose (**3**) that used, as the glycosyl acceptor, benzyl 3-*O*-acetyl-2,4-di-*O*-benzyl- α -D-glucopyranoside, which, in turn, was prepared *via* partial benzylation of benzyl 6-*O*-trityl- α -D-glucopyranoside.

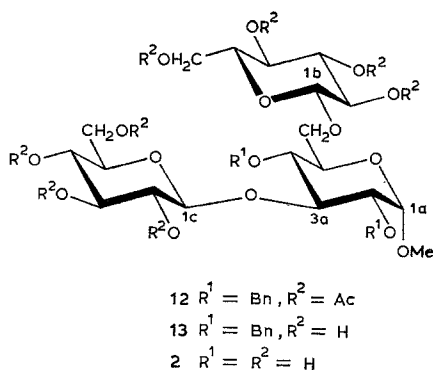
Our synthetic approach to the synthesis of glycosyl acceptors **5** and **6** depends on the regioselective hydrogenolysis of the benzylidene group of **10** and **17** according to the method of Lipták *et al.*¹⁵. In order to synthesize monobenzyl ether **10**, regioselective benzylation of diol **9** was studied by using three different methods. First, diol **9** was converted into the stannylidene derivative¹⁶ by treatment with Bu_2SnO , and subsequent reaction with benzyl bromide in HCONMe_2 gave 2-benzyl ether^{17,18} **10** and 3-benzyl ether¹⁷⁻¹⁹ **11** in 69.8 and 20.2 % yield, respectively. Second, tributylstannylation²⁰ of **9**, and subsequent alkylation with benzyl bromide, gave monobenzyl ethers **10** and **11** in 59.1 and 31.9 % yield, respectively. Finally, monobenzylation of **9**, using a phase-transfer catalyst according to Garegg *et al.*¹⁸, gave **10** and **11** in 40 and 20 % yield, respectively. Therefore, in our hands, the stannylidene approach was found to give the highest yield of the desired monobenzyl ether **10**.



Reductive cleavage¹⁵ of the benzylidene group in **10** was achieved with LiAlH_4 - AlCl_3 , to give crystalline dibenzyl ether¹⁷ **5** in 71 % yield. The assigned structure **5** was confirmed by ^{13}C -n.m.r. data, which showed two deshielded signals for C-2 and C-4 at δ 79.6 and 77.1, due to benzylation at O-2 and O-4, and a shielded signal for C-1 at δ 97.5, due to the β -effect of alkylation at O-2. Absence of the signal¹ at δ ~82.5–82.8, and the presence of a signal at δ 61.9 for C-6, proved that the two benzyl groups were linked neither to O-3 nor O-6. Furthermore, signals for methylene carbon atoms of 2-*O*-benzyl and 4-*O*-benzyl groups appeared at δ 73.1 and 74.5, in agreement with a previous observation¹.

Glycosylation of **5** with 3 equivalents of the D-glucosyl donor **7**, in the presence²¹ of HgBr_2 -powdered molecular sieve 4 A at 45°, gave protected trisaccharide **12** in 59.3 % yield. Zemplén deacetylation of **12**, to give **13** in 63 % yield, and hydrogenolysis

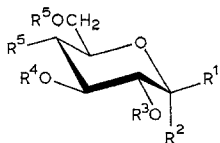
of **13**, afforded D-glucotrioside **2** in 97% yield. The configurations of the newly introduced anomeric centers were assignable as β from the ^1H - and ^{13}C -n.m.r. data. Three anomeric protons were observed as three doublets at δ 4.71, with J 4.0 Hz for H-1a, and δ 4.54 and 4.38, both with J 8.0 Hz, for H-1c and H-1b, respectively.



Two signals at δ 103.15, with $^1J_{\text{CH}}$ 160.2 Hz, and δ 99.64, with $^1J_{\text{CH}}$ 169.9 Hz, were observed in the ^{13}C -n.m.r. spectrum in the ratio of 2:1, and were assigned to C-1b and C-1c, and C-1a, respectively, from the value²² of $^1J_{\text{CH}}$.

Synthesis of glucotrioside 3 and glucotetraoside 4. — The key glycosyl acceptor **6** was prepared from D-glucose in 4 steps, as follows. D-Glucose was successively treated with (i) benzyl alcohol-HCl, and (ii) $\text{PhCH}(\text{OMe})_2\text{-TsOH} \cdot \text{H}_2\text{O}$, to give crystalline benzyl 4,6-O-benzylidene- α -D-glucopyranoside²³ (**15**) in 10.6% yield without use of chromatography. Selective monobenylation of **15** by the stannylidene method afforded a 77% yield of crystalline 2-benzyl ether **16**. The site of benzylation was proved to be O-2 by the transformation of **16** into monoacetate **17**, whose ^1H -n.m.r. data revealed a deshielded signal for H-3 as a triplet at δ 5.60 with J 10 Hz, along with a signal for the methyl group of acetate as a singlet at δ 2.03.

Selective cleavage¹⁵ of the benzylidene group of **16** afforded the desired glycosyl acceptor **6** in 53% yield. The substitution pattern of **6** was expected to be at O-2 and O-4 (from the synthetic sequence), and this was confirmed by the ^{13}C -n.m.r.

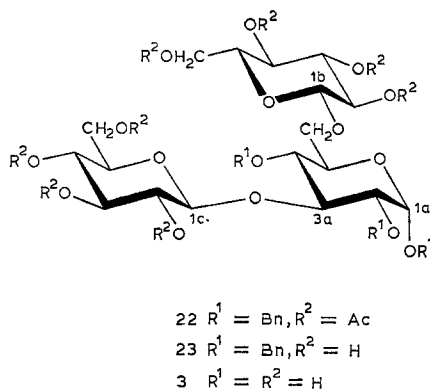
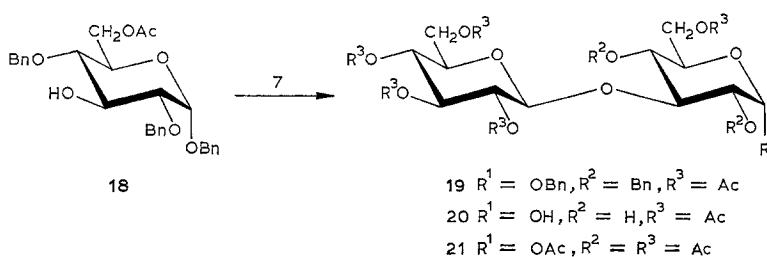


- $14 \text{ } R^1 (\text{or } R^2) = \text{OBn}, R^2 (\text{or } R^1) = \text{H}$
 $R^3 = R^4 = R^5 = \text{H}$
 $15 \text{ } R^1 = R^3 = R^4 = \text{H}, R^2 = \text{OBn}, R^5, R^5 = \text{CHPh}$
 $16 \text{ } R^1 = R^4 = \text{H}, R^2 = \text{OBn}, R^3 = \text{Bn}, R^5, R^5 = \text{CHPh}$
 $17 \text{ } R^1 = \text{H}, R^2 = \text{OBn}, R^3 = \text{Bn}, R^4 = \text{Ac}, R^5, R^5 = \text{CHPh}$

spectrum, which contained two deshielded signals, for C-2 and C-4, at δ 79.53 and 77.15, respectively, and the signal for C-6 remained shielded at δ 61.83.

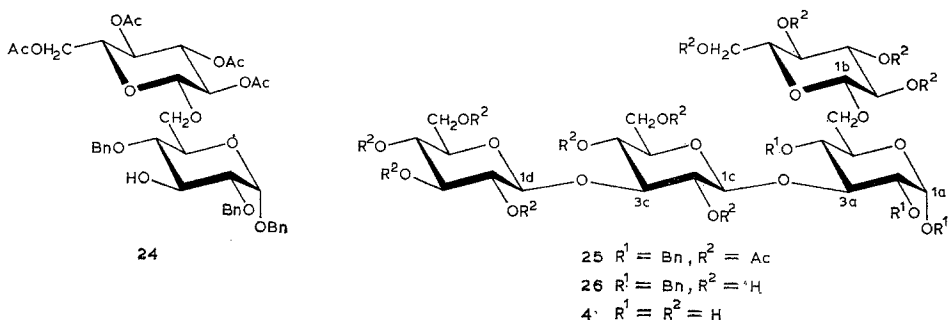
The practical synthesis of laminarabiosyl donor **8** was recently reported, either by use²⁴ of a partially protected α -D-glucopyranoside, or by acetolysis²⁵ of laminaran. We have developed an alternative route to **8** by use of partially benzylated α -D-glucopyranoside **6**.

Selective acetylation of the primary hydroxyl group of **6** gave an 82% yield of monoacetate **18**. Reaction of **18** with the D-glucosyl donor **7**, in the presence of HgBr_2 and powdered molecular sieve 4 A, led to the isolation of protected laminarabioside **19** in 73% yield. Hydrogenolysis of **19**, to give **20** in 57.5% yield, and acetylation of **20**, afforded the octaacetate **21** as a mixture of the α and β anomer in the ratio of 1:1. Oily **21** was converted into crystalline **8** in 81% yield. The ^1H -n.m.r. data for **8** confirmed the structure assigned.



Having prepared the key glycosyl acceptor **6**, and the two glycosyl donors **7** and **8**, syntheses of D-glucotriose (**3**) and D-glucotetraose (**4**) were examined. Glycosylation of **6** with 1.2 equivalents of the D-glucosyl donor **7** according to Sinay *et al.*²¹ led to the isolation of properly protected glucobioside **24** in 26% yield, and a 50% recovery of starting diol **6**. On the other hand, glycosylation of **6** with 3.6 equivalents of **7**, using the same Lewis acid and acid captor, afforded a 47% yield of protected D-glucotriose **22**. Deacetylation of **22**, to give **23**, and hydrogenolysis of **23**, gave

the desired D-glucotriose **3**. The ^{13}C -n.m.r. data confirmed the regio- and stereo-structure of **3**, which showed a signal for C-1b and C-1c at δ 103.04, with $^1J_{\text{CH}}$ 162.1 Hz, and two signals, for C-1a α and C-1a β , in the ratio of 1:1, at δ 96.02 with $^1J_{\text{CH}}$ 161.1 Hz, and at δ 92.36 with $^1J_{\text{CH}}$ 169.9 Hz, respectively. Two deshielded signals for C-3a β and C-3a α were also observed, at δ 84.72 and 82.46.



Properly protected D-glucobioside **24** was now submitted to the glycosylation reaction with **8**, in the presence of $\text{AgOSO}_2\text{CF}_3$ and powdered molecular sieve 4 A, to give an 81% yield of the protected D-glucotetraoside **25**. Deacylation of **25** to give tribenzyl ether **26** in 76% yield, and hydrogenolysis of **26**, afforded the target D-glucotetraose **4**. The regiochemistry of **4** was evident from the synthetic sequence, and the configurations of the anomeric centers newly introduced were determined to be all β by ^{13}C -n.m.r. data, which contained a signal for C-1b, C-1c, and C-1d at δ 103.08, with $^1J_{\text{CH}}$ 163.1 Hz, and two signals, for C-1a β and C-1a α in the ratio of 1:1, at δ 96.06 with $^1J_{\text{CH}}$ 157.6 Hz, and at δ 92.36, with $^1J_{\text{CH}}$ 169.9 Hz, respectively. Two deshielded signals for C-3c and C-3a β , and for C-3a α , were observed at δ 84.56 and 82.30, respectively, due to the α -effect of glucosylation.

In conclusion, the first synthesis of D-glucotetraose **4**, the repeating unit of the extracellular D-glucans of *Grifola umbellata*, *Sclerotinia libertiana*, *Porodisculus pendulus*, and *Schizophyllum commune* Fries was achieved in a regio- and stereo-controlled way by using the properly protected D-glucobioside **24** as the key intermediate.

EXPERIMENTAL

General. — Melting points were determined with a Yanagimoto micro melting-point apparatus and are uncorrected. Optical rotations were determined with a Perkin-Elmer Model 141 polarimeter for solutions in CHCl_3 at 25° , unless otherwise noted. I.r. spectra were recorded with an EPI-G2 Hitachi Spectrophotometer, using KBr discs for the crystalline samples, and neat films for the liquid samples. ^1H -N.m.r. spectra were recorded with a Varian HA-100 n.m.r. spectrometer, using tetramethylsilane as the internal standard. ^{13}C -N.m.r. spectra were recorded with a JNM-FX 100FT n.m.r. spectrometer operated at 25.05 MHz. The values of δ_{C} and

δ_{H} are expressed in p.p.m. downwards from the internal standard, for solutions in CDCl_3 , unless otherwise noted. Flash column-chromatography²⁶ was performed on columns of Wakogel C-300 (Wako Pure Chemical Industries, Ltd.). Thin-layer chromatography was performed on precoated plates (layer thickness, 0.25 mm) of Silica Gel 60 F₂₅₄ (E. Merck, Darmstadt, Germany).

Methyl 2-O-benzyl-4,6-O-benzylidene- α -D-glucopyranoside (10) and methyl 3-O-benzyl-4,6-O-benzylidene- α -D-glucopyranoside (11). — (A) A mixture of **9** (843 mg, 3 mmol) and Bu_2SnO (903 mg, 3.6 mmol) in 10:1 benzene–MeOH (20 mL) was boiled under reflux for 2 h, cooled, and evaporated *in vacuo*. A mixture of the residue with DMF (5 mL)–benzyl bromide (5 mL) was stirred for 2 h at 90–110°, cooled, evaporated *in vacuo*, and the residual oil chromatographed on SiO_2 (100 g) with 7:2 toluene–EtOAc, to give **10** (776 mg, 69.8%), m.p. 126–127° (EtOH), $[\alpha]_{\text{D}} + 34.7^\circ$ (c 0.95); R_{F} 0.40 in 7:2 toluene–EtOAc; δ_{H} : 5.48 (s, 1 H, CHPh), 4.59 (d, 1 H, J 4 Hz, H-1), and 3.35 (s, 3 H, OMe).

Anal. Calc. for $\text{C}_{21}\text{H}_{24}\text{O}_6$: C, 67.73; H, 6.50. Found: C, 67.85; H, 6.51.

Further elution gave **11** (225 mg, 20.2%), m.p. 188–189° (EtOH), $[\alpha]_{\text{D}} + 59.3^\circ$ (c 1.00); R_{F} 0.29 in 7:2 toluene–EtOAc; δ_{H} : 5.54 (s, 1 H, CHPh), 4.78 (d, 1 H, J 4 Hz, H-1), and 3.42 (s, 3 H, OMe).

Anal. Calc. for $\text{C}_{21}\text{H}_{24}\text{O}_6$: C, 67.73; H, 6.50. Found: C, 67.51; H, 6.46.

(B) A mixture of **9** (11.28 g, 40 mmol) and $(\text{Bu}_3\text{Sn})_2\text{O}$ (17.88 g, 30 mmol) in toluene (100 mL) was boiled under reflux for 15 h, with continuous azeotropic removal of water, and evaporated *in vacuo*. A mixture of the residual oil with benzyl bromide (100 mL) was stirred for 3 days at 80–83° under argon, cooled, and evaporated *in vacuo*; the residue was mixed with EtOAc (100 mL), the mixture treated with²⁷ aq. KF, and the suspension filtered. The filtrate was washed with H_2O , dried (MgSO_4), and evaporated, and the residue was chromatographed on SiO_2 (1 kg) with 3:1 toluene–EtOAc, to give **10** (7.03 g, 59.1%) and **11** (3.75 g, 31.9%).

Methyl 2,4-di-O-benzyl- α -D-glucopyranoside (5). — To a stirred solution of **10** (500 mg, 1.34 mmol) in 1:1 Et_2O – CH_2Cl_2 (10 mL) was added, portionwise, LiAlH_4 (170 mg, 4.48 mmol). To this stirred mixture was added, dropwise, a solution of AlCl_3 (500 mg, 3.75 mmol) in Et_2O (5 mL) during 30 min under reflux. After refluxing for a further 1 h, the excess of LiAlH_4 was decomposed by adding EtOAc and H_2O . The mixture was diluted with EtOAc (100 mL), washed with H_2O , dried (MgSO_4), and evaporated *in vacuo*. The residue was chromatographed on SiO_2 (45 g) with 1:1 toluene–EtOAc, to give **5** (355 mg, 71%), m.p. 71–72° (EtOH–pentane), $[\alpha]_{\text{D}} + 98.3^\circ$ (c 1.00); R_{F} 0.51 in 1:1 toluene–EtOAc; δ_{H} : 7.4–7.2 (m, 10 H, aromatic), 4.90 and 4.66 (ABq, 2 H, J 11 Hz, $\text{CH}_2\text{-Ph}$), 4.65 (s, 2 H, CH_2Ph), 4.58 (d, 1 H, J 4 Hz, H-1), 4.08 (t, 1 H, J 9 Hz, H-3), and 3.29 (s, 3 H, OMe); δ_{C} : 97.5 ($^1\text{J}_{\text{CH}}$ 167.0 Hz, C-1), 79.6 (C-2), 77.1 (C-4), 74.5 (4- $\text{O-CH}_2\text{Ph}$), 73.4 (C-3), 73.1 (2- $\text{O-CH}_2\text{Ph}$), 70.2 (C-5), 61.9 (C-6), and 55.1 (OMe).

Anal. Calc. for $\text{C}_{21}\text{H}_{26}\text{O}_6$: C, 67.36; H, 7.00. Found: C, 67.37; H, 7.00.

Methyl 2,4-di-O-benzyl-3,6-di-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (12). — To a stirred mixture of powdered molecular sieve 4 A

(1 g), HgBr_2 (108 mg, 0.3 mmol), and **5** (190 mg, 0.5 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (5 mL) was added dropwise a solution of **7** (617 mg, 1.5 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (5 mL). The mixture was stirred for 4 h at 15–20° and then for 4 h at 45°, Et_3N (2 mL) was added, the mixture was evaporated *in vacuo*, and the residue was chromatographed on SiO_2 C-300 (70 g) with 2:1 toluene–EtOAc, to give **12** (313 mg, 59.3%); R_F 0.23 in 2:1 toluene–EtOAc.

Methyl 2,4-di-O-benzyl-3,6-di-O-β-D-glucopyranosyl-α-D-glucopyranoside (13). — A solution of **12** (313 mg, 0.30 mmol) in MeOH (5 mL)–*m* NaOMe (0.3 mL) was stirred for 15 h at 15–20°, the base neutralized with Amberlist 15 (H^+), and the suspension filtered. The filtrate was evaporated *in vacuo*, and the residue was chromatographed on SiO_2 C-300 (10 g) with 15:2 CH_2Cl_2 –MeOH, to give **13** (134 mg, 63.0%); R_F 0.18 in 15:2 CH_2Cl_2 –MeOH.

Methyl 3,6-di-O-β-D-glucopyranosyl-α-D-glucopyranoside (2). — A mixture of **13** (134 mg, 0.19 mmol) and 10% Pd–C (50 mg) in AcOH (2 mL) was stirred under H_2 for 24 h at 15–20°. Processing afforded an oily product that still carried some benzyl groups (as judged by its ^1H -n.m.r. spectrum). A mixture of this oil and 10% Pd–C (100 mg) in 1:1 EtOH– H_2O (2 mL) was stirred under H_2 for 2 days; processing then afforded amorphous **2** (98.6 mg, 97%) $[\alpha]_D + 28.5^\circ$ (*c* 0.50, MeOH); R_F 0.19 in 10:5:4 1-BuOH–*i*-PrOH– H_2O ; δ_H (D_2O , 75°, 400 MHz): 4.708 (d, 1 H, *J* 4.0 Hz, H-1a), 4.538 (d, 1 H, *J* 8.0 Hz, H-1c), and 4.381 (d, 1 H, *J* 8.0 Hz, H-1b); δ_C (D_2O , 75°): 103.15 ($^1J_{\text{CH}}$ 160.2 Hz, C-1b,1c), 99.64 ($^1J_{\text{CH}}$ 169.9 Hz, C-1a), 82.53 (C-3a), 76.25 (C-3b*,3c*), 75.94 (C-5b*,5c*), 73.80 (C-2b**), 73.48 (C-2c**), 71.07 (C-5a), 70.83 (C-2a), 69.94 (C-4b,4c), 68.85 (C-6a), 68.14 (C-4a), 61.09 (C-6b,6c), and 55.63 (OMe).

Anal. Calc. for $\text{C}_{19}\text{H}_{34}\text{O}_{16} \cdot \text{H}_2\text{O}$: C, 42.53; H, 6.76. Found: C, 42.62; H, 6.87.

Benzyl 4,6-O-benzylidene-α-D-glucopyranoside²³ (15). — To benzyl alcohol (500 mL) was added AcCl (50 mL) dropwise, and the mixture was stirred for 15 min at 20°. To the solution was added D-glucose (100 g), and the mixture was stirred for 3 days at 15–20° and for 1 day at 50°, the acid neutralized with Amberlist **21** (OH^-), and the suspension filtered. The filtrate was evaporated *in vacuo*, the residual oil dissolved in DMF (100 mL), $\text{PhCH}(\text{OMe})_2$ (244 mL) and $\text{TsOH} \cdot \text{H}_2\text{O}$ (13 g) were added, the mixture was stirred for 2 days at 40–45°, and the acid was neutralized with Et_3N . The solution was evaporated *in vacuo*, the residue was partitioned between EtOAc and H_2O , the organic layer was dried (MgSO_4), and evaporated *in vacuo*, and the residue was triturated with EtOH, to give crystalline **15** (38.1 g, 10.6%); m.p. 158–159° (CH_2Cl_2 – Et_2O), $[\alpha]_D + 131.3^\circ$ (*c* 1.22); R_F 0.28 in 1:1 toluene–EtOAc; δ_H : 5.50 (s, 1 H, *CHPh*), 4.97 (d, 1 H, *J* 4 Hz, H-1), 4.77 (d, 1 H, *J* 11 Hz, *CH}_2\text{Ph}*), and 4.53 (d, 1 H, *J* 11 Hz, *CH}_2\text{Ph}*).

Anal. Calc. for $\text{C}_{20}\text{H}_{22}\text{O}_6$: C, 67.02; H, 6.19. Found: C, 66.99; H, 6.21.

Benzyl 2-O-benzyl-4,6-O-benzylidene-α-D-glucopyranoside (16). — A mixture of **15** (15 g, 42 mmol) and Bu_2SnO (10.5 g, 42 mmol) in 10:1 benzene–MeOH (132

*The assignments with asterisks may have to be interchanged.

mL) was boiled, and stirred, under reflux for 2.5 h, cooled, and evaporated; the residue was dissolved in DMF (70 mL), benzyl bromide (70 mL) was added, and the mixture was stirred for 3.5 h at 100–120°, cooled, and evaporated *in vacuo*. A solution of the residue in EtOAc (400 mL) was stirred with aq. KF for 3 h, and the suspension was filtered, to remove precipitated Bu_2SnF_2 . The filtrate was dried (MgSO_4), and evaporated, to afford an oily product which was triturated with EtOH, to give crystalline **16** (14.3 g, 77%), m.p. 133–134° (EtOH), $[\alpha]_D +110.6^\circ$ (*c* 1.1); R_F 0.52 in 3:1 toluene–EtOAc; δ_H : 7.5–7.3 (m, 15 H, aromatic) and 5.54 (s, 1 H, *CHPh*).

The 3-acetate of **17** was prepared in the usual way; δ_H 5.60 (t, 1 H, *J* 10 Hz, H-3) and 2.03 (s, 3 H, Ac).

Benzyl 2,4-di-O-benzyl- α -D-glucopyranoside (6). — To a stirred mixture of **16** (5.0 g, 14.5 mmol) and LiAlH_4 (2.2 g, 58.0 mmol) in 1:1 CH_2Cl_2 – Et_2O (100 mL), was added, dropwise, a solution of AlCl_3 (7.7 g, 58.0 mmol) in Et_2O (50 mL) during 30 min under reflux, and the mixture was stirred for 2 h under reflux. Processing as described for **5**, and purification by flash chromatography on SiO_2 C-300 (300 g) with 3:1 toluene–EtOAc, gave **6** (2.66 g, 53%), m.p. 77–78° (EtOH–pentane), $[\alpha]_D +157.6^\circ$ (*c* 1.3); R_F 0.27 in 3:1 toluene–EtOAc; δ_C : 95.16 ($^1J_{\text{CH}}$ 167.0 Hz, C-1), 79.53 (C-2), 77.15 (C-4), 74.58 (4-*O-CH}_2\text{Ph}*), 73.33 (C-3), 72.63 (2-*O-CH}_2\text{Ph}*), 70.68 (C-5), 69.31 (1-*O-CH}_2\text{Ph}*), and 61.83 (C-6).

Anal. Calc. for $\text{C}_{27}\text{H}_{30}\text{O}_6$: C, 72.00; H, 6.71. Found: C, 71.72; H, 6.75.

Benzyl 6-O-acetyl-2,4-di-O-benzyl- α -D-glucopyranoside (18). — To a stirred solution of **6** (3.0 g, 6.76 mmol) in CH_2Cl_2 (10 mL)–pyridine (1.09 mL, 13.5 mmol) was added AcCl (0.53 mL, 7.44 mmol) dropwise at -10 to -15° ; the mixture was stirred for 2 h, further AcCl (0.03 mL) was added (in order to consume **6** completely), and extractive processing then afforded an oily product which was chromatographed on SiO_2 C-300 (300 g) with 10:1 toluene–EtOAc, to give **18** (2.68 g, 82%), $[\alpha]_D +157.2^\circ$ (*c* 1.3); R_F 0.51 in 3:1 toluene–EtOAc; δ_H : 2.0 (s, 3 H, Ac).

Anal. Calc. for $\text{C}_{29}\text{H}_{32}\text{O}_7$: C, 70.71; H, 6.55. Found: C, 70.75; H, 6.60.

Benzyl 6-O-acetyl-2,4-di-O-benzyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (19). — To a stirred mixture of **18** (3.5 g, 7.1 mmol), powdered molecular sieve 4 A (5.0 g), and HgBr_2 (1.024 g, 2.84 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (5 mL) was added dropwise a solution of **7** (4.5 g, 10.9 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (15 mL). The mixture was stirred for 2.5 h at 20° , filtered through Celite, and the filtrate successively washed with 3% aq. AgNO_3 and aq. NaHCO_3 , dried (MgSO_4), and evaporated *in vacuo*. The residue was chromatographed on SiO_2 C-300 (200 g) with 8:1 toluene–EtOAc, to give **19** (4.3 g, 73.0%), $[\alpha]_D +65.6^\circ$ (*c* 1.5); R_F 0.47 in 8:1 toluene–EtOAc; δ_H : 7.38 (bs, 15 H, aromatic), 2.10 (s, 3 H, Ac), 2.04 (s, 9 H, 3 Ac), and 1.98 (s, 3 H, Ac).

Anal. Calc. for $\text{C}_{43}\text{H}_{50}\text{O}_{16}$: C, 62.77; H, 6.12. Found: C, 62.30; H, 6.11.

6-O-Acetyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-D-glucopyranose (20). — A mixture of **19** (1.84 g, 2.24 mmol) and 10% Pd–C (500 mg) in AcOH (15 mL) was stirred for 2 days at 40° under H_2 , and then filtered through Celite. The filtrate was evaporated *in vacuo*, and the residue was chromatographed on SiO_2

C-300 (30 g) with 1:5 toluene–EtOAc, to give **20** (710 mg, 57.5%), m.p. 170–171° (CH₂Cl₂–hexane); R_F 0.24 in 1:5 toluene–EtOAc; δ_H : 2.12 (s, 9 H, 3 Ac), 2.06 (s, 3 H, Ac), and 2.04 (s, 3 H, Ac).

Anal. Calc. for C₂₂H₃₂O₁₆: C, 47.83; H, 5.84. Found: C, 47.56; H, 5.80.

1,2,4,6-Tetra-O-acetyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α - and - β -D-glucopyranose (21). — Compound **20** (540 mg, 0.98 mmol) was acetylated with AcCl and pyridine, to give **21** (as a mixture of the α and β anomers in the ratio of 1:1); R_F 0.5 in 1:3 toluene–EtOAc; δ_H : 6.22 (d, 0.5 H, J 4 Hz, H-1 α), and 5.61 (d, 0.5 H, J 8 Hz, H-1 β). Mixture **21** was used directly for the next step.

2,4,6-Tri-O-acetyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranosyl bromide (8). — A solution of **21** (504 mg) in 30% HBr–AcOH (5 mL) was stirred for 1.5 h at 15–20°. The mixture was co-evaporated several times with toluene *in vacuo*, and the residue was chromatographed on SiO₂ C-300 (50 g) with 1:1 toluene–EtOAc, to give **8** (689 mg, 81%), m.p. 182–183° (EtOAc–i-Pr₂O), $[\alpha]_D$ +98.8° (*c* 1.1); δ_H : 6.50 (d, 1 H, J 4 Hz, H-1), 2.20 (s, 3 H, Ac), 2.11 (s, 6 H, 2 Ac), 2.08 (s, 3 H, Ac), 2.02 (s, 3 H, Ac), 2.00 (s, 3 H, Ac), and 1.97 (s, 3 H, Ac).

Anal. Calc. for C₂₆H₃₅BrO₁₀: C, 44.64; H, 5.04; Br, 11.42. Found: C, 44.62; H, 5.05; Br, 11.51.

Benzyl 2,4-di-O-benzyl-6-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (24). — To a stirred mixture of **6** (2.66 g, 5.91 mmol), powdered molecular sieve 4 A (10 g), and HgBr₂ (510 mg, 1.42 mmol) in Cl(CH₂)₂Cl (15 mL) was added dropwise a solution of **7** (2.915 g, 7.09 mmol) in Cl(CH₂)₂Cl (15 mL). The mixture was stirred for 3.5 h at 20°, and filtered (Celite). The filtrate was diluted with EtOAc, successively washed with aq. NaHCO₃ and H₂O, dried (MgSO₄), and evaporated *in vacuo*. The residue was chromatographed on SiO₂ C-300 (200 g) with 2:1 i-Pr₂O–Et₂O, to give recovered **6** (1.3 g, 50%) and **24** (1.2 g, 26%), $[\alpha]_D$ +67.8° (*c* 1.07); R_F 0.23 in 2:1 i-Pr₂O–Et₂O; δ_H : 2.07 (s, 3 H, Ac), 2.04 (s, 3 H, Ac), 2.02 (s, 3 H, Ac), and 1.97 (s, 3 H, Ac).

Anal. Calc. for C₄₁H₄₈O₁₅: C, 63.10; H, 6.20. Found: C, 62.77; H, 6.21.

Benzyl 2,4-di-O-benzyl-3,6-di-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (22). — To a stirred mixture of **6** (374 mg, 0.83 mmol), powdered molecular sieve 4 A (4 g), and HgBr₂ (216 mg, 0.6 mmol) in Cl(CH₂)₂Cl (5 mL) was added dropwise a solution of **7** (1.233 g, 3 mmol) in Cl(CH₂)₂Cl (5 mL). The mixture was stirred for 24 h at 45°, diluted with Cl(CH₂)₂Cl (30 mL), and filtered (Celite). The filtrate was successively washed with aq. NaHCO₃ and H₂O, dried (MgSO₄), and evaporated *in vacuo*. The residue was chromatographed on SiO₂ C-300 (100 g) with 3:1 toluene–EtOAc, to give **22** (440 mg, 47%), $[\alpha]_D$ +12.8° (*c* 0.58); R_F 0.15 in 2:1 i-Pr₂O–Et₂O; δ_H : 7.38 (bs, 15 H, aromatic), 2.09 (s, 3 H, Ac), 2.08 (s, 3 H, Ac), 2.06 (s, 6 H, 2 Ac), 2.04 (s, 9 H, 3 Ac), and 1.94 (s, 3 H, Ac).

Anal. Calc. for C₅₅H₆₆O₂₄: C, 59.45; H, 5.99. Found: C, 59.19; H, 6.02.

Benzyl 2,4-di-O-benzyl-3,6-di-O- β -D-glucopyranosyl- α -D-glucopyranoside (23). — A solution of **22** (251 mg) in MeOH (5 mL)–*m* NaOMe (0.1 mL) was stirred for

15 h at 20°. The usual processing afforded **23** (167 mg, 95%) as a white powder, R_F 0.86 in 5:1 CH_2Cl_2 –MeOH, which was used directly for the next step.

3,6-Di-O- β -D-glucopyranosyl- α - and - β -D-glucopyranose (3). — A mixture of **23** (167 mg, 0.22 mmol) and 10% Pd–C (100 mg) in 1:1 EtOH– H_2O (10 mL) was stirred under H_2 for 3 days at 20°. Processing gave **3** as a white powder (48 mg, 44%), $[\alpha]_D -13.0$ (30 min) $\rightarrow +6.8^\circ$ (3 days) (c 0.53, H_2O); R_F 0.17 in 10:5:4 1-BuOH–i-PrOH– H_2O ; δ_C (D_2O): 103.04 ($^1J_{\text{CH}}$ 162.1 Hz, C-1b,1c), 96.02 ($^1J_{\text{CH}}$ 161.1 Hz, C-1 $\alpha\beta$), 92.36 ($^1J_{\text{CH}}$ 169.9 Hz, C-1a α), 84.72 (C-3a β), and 82.46 (C-3a α).

Anal. Calc. for $\text{C}_{18}\text{H}_{32}\text{O}_{16} \cdot 1.5 \text{H}_2\text{O}$: C, 40.68; H, 6.64. Found: C, 40.94; H, 6.78.

Benzyl 2,4-di-O-benzyl-6-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-3-O-[2,4,6-tri-O-acetyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosyl]- α -D-glucopyranoside (25). — To a stirred mixture of **24** (359 mg, 0.46 mmol), powdered molecular sieve 4 A (4 g), and $\text{AgOSO}_2\text{CF}_3$ (150 mg, 0.59 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (5 mL) was added dropwise a solution of **8** (270 mg, 0.39 mmol) in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ (5 mL) at -10 to -15° . The mixture was stirred for 3 h at -10 to -15° , and processed as usual. The crude product was chromatographed on SiO_2 C-300 (30 g) with 3:2 toluene–EtOAc, to give **25** (438 mg, 81%), $[\alpha]_D +1.8^\circ$ (c 0.53); R_F 0.23 in 3:2 toluene–EtOAc; δ_H : 7.32 (bs, 15 H, aromatic), 2.08 (s, 3 H, Ac), 2.06 (s, 3 H, Ac), 2.02 (s, 21 H, 3 Ac), 1.97 (s, 3 H, Ac), and 1.92 (s, 3 H, Ac).

Anal. Calc. for $\text{C}_{67}\text{H}_{82}\text{O}_{32}$: C, 57.51; H, 5.91. Found: C, 57.16; H, 5.90.

Benzyl 2,4-di-O-benzyl-6-O- β -D-glucopyranosyl-3-O-(3-O- β -D-glucopyranosyl- β -D-glucopyranosyl)- α -D-glucopyranoside (26). — A solution of **25** (552 mg, 0.39 mmol) in MeOH (5 mL)–M NaOMe (0.18 mL) was stirred for 15 h at 15° . Neutralization of the base with Amberlist **15** (H^+), and the usual processing, gave **26** (284 mg, 76%), m.p. 187–188° (MeOH), $[\alpha]_D +16.0^\circ$ (c 0.5, MeOH); R_F 0.76 in 3:3:2:2 1-BuOH–i-PrOH–EtOH– H_2O ; δ_H (MeSO- d_6): 7.40 (bs, 15 H, aromatic).

Anal. Calc. for $\text{C}_{45}\text{H}_{60}\text{O}_{21} \cdot \text{MeOH}$: C, 57.01; H, 6.66. Found: C, 56.87; H, 6.37.

6-O- β -D-Glucopyranosyl-3-O-(3-O- β -D-glucopyranosyl- β -D-glucopyranosyl)- α - and - β -D-glucopyranose (4). — A mixture of **26** (219 mg, 0.23 mmol) and 10% Pd–C (200 mg) in 1:1 EtOH– H_2O (10 mL) was stirred under H_2 for 2 days at 30° . Filtration through Celite, and the usual processing, afforded **4** (148 mg, 90%), $[\alpha]_D -0.3^\circ$ (c 0.50, H_2O); R_F 0.27 in 3:3:2:2 1-BuOH–i-PrOH–EtOH– H_2O ; δ_C : 103.08 ($^1J_{\text{CH}}$ 163.1 Hz, H-1b,1c,1d), 96.06 ($^1J_{\text{CH}}$ 157.6 Hz, H-1a β), 92.36 ($^1J_{\text{CH}}$ 169.9 Hz, H-1a α), 84.56 (H-3c,3a β), and 82.30 (H-3a α).

Anal. Calc. for $\text{C}_{24}\text{H}_{42}\text{O}_{21} \cdot 1.5 \text{H}_2\text{O}$: C, 41.56; H, 6.54. Found: C, 41.62; H, 6.55.

ACKNOWLEDGMENTS

We thank Dr. J. Uzawa and Mrs. T. Chijimatsu for recording and measuring the n.m.r. spectra, and Dr. H. Homma and his staff for the elemental analyses. We also thank Miss A. Sone for her technical assistance.

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