Potential Antitrypanosomal Agents. 1.N²-Disubstituted 2-Amino-5-hydroxy-4-methylnaphtho[1,2-d]thiazolium Salts and Related Compounds

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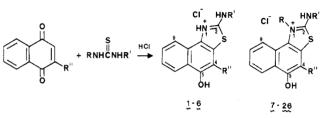
A series of 1-alkyl-2-(substituted-amino)-5-hydroxy-4-methylnaphtho[1,2-d]thiazoles having in vitro trypanocidal activity is described. Several caused complete lysis of Trypanosoma brucei organisms within 30 min at 10^{-5} M. The presence of a hydrophobic substituent on the 2-amino group was associated with high antitrypanosomal activity. Some analogues unsubstituted at the 1-position, a known class of compounds, were also active. None of the derivatives significantly prolonged the survival of T. brucei infected mice. Inhibition of activity in vitro by boyine serum albumin was observed. Because of the structural novelty of these agents in comparison with known trypanocides, their mechanism of action warrants further investigation.

African trypanosomiasis in livestock continues to be a serious constraint on livestock production in large areas of sub-Saharan Africa.¹ Epidemics of human trypanosomiasis in Africa earlier in this century have subsided to the present level of controlled endemicity largely due to chemotherapy, chemoprophylaxis, tsetse eradication efforts, and the avoidance of endemic areas.^{2,3} Nevertheless, disease foci still exist, and there is potential for new human epidemics.^{2c,d,3} Presently used trypanocides have serious drawbacks, such as toxicity or ineffectiveness in late-stage infection.^{2e-h,3,4} Virtually no new trypanocides have entered clinical or veterinary use since the introduction of melarsoprol and diminazene aceturate in the 1950's.^{2e,3} Because of the appearance of trypanosome strains resistant to currently used drug classes, trypanocides of new structural types are urgently needed.^{1b,2e,3}

Work in this laboratory has shown that certain simple quinones have in vitro activity against bloodstream forms of Trypanosoma brucei.^{5a} Others have reported that a number of cytotoxic anthraquinones show substantial activity against African trypanosomes in vitro.5b The unusual pathways of energy metabolism in these parasites appear to render them especially susceptible to the action of quinones and related agents.⁶ We have prepared and tested a variety of quinoid and quinone-derived substances in order to identify trypanocidal derivatives which lack the pharmacologically undesirable electrophilic properties of quinones. In the course of these studies some N-substituted 2-amino-5-hydroxy-4-methylnaphtho[1,2-d]thiazoles were found to have substantial rapid trypanolytic activity

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- (2) (a) de Raadt, P. Trans. R. Soc. Trop. Med. Hyg. 1976, 70, 114-116. (b) Shattuck, G. C. "Diseases of the Tropics"; Appleton Century Crofts: New York 1951; pp 108-130. (c) Scott, D. In "The African Trypanosomiases"; Mulligan, H. W., Ed.; Wiley-Interscience: New York, 1970; pp 614–44. (d) Apted, F. I. C. *Ibid.*; pp 645–660. (e) Williamson, J. *Ibid.*; pp 125–221. (f) Waddy, B. B. *Ibid.*; pp 711-725. (g) Stephen, L. E. *Ibid.*;
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- (4) Rollo, I. M. In "The Pharmacological Basis of Therapeutics; Gilman, A. G.; Goodman, L. S.; Gilman, A., Eds.; Macmillan: New York, 1980; pp 1070–1079. (a) Meshnick, S. R.; Blobstein, S. H.; Grady, R. W.; Cerami,
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Scheme I



in vitro. This paper describes the synthesis and antitrypanosomal screening of a series of these compounds (Table I).

Chemistry. Although the adducts of thioureas with quinones have been known for some time,⁷ the formation of ring-fused thiazoles from these reactants was first reported by Lau and Gompf⁸ in 1970. They prepared 1-3, 5, and other naphtho- and benzothiazoles by treating thiourea or a monosubstituted thiourea with an excess of a 1,4-naphtho- or benzoquinone in ethanolic HCl (Scheme I). In all cases described⁸ (including 3 and 5) and in 4 and 6 prepared analogously for the present study, monosubstituted thioureas afforded aminothiazole derivatives substituted on the amino group and not on the ring nitrogen. No examples using disubstituted thioureas were reported.

We found that under similar conditions a variety of N.N'-disubstituted thioureas react with 2-methyl-1,4naphthoquinone to yield 1,N²-disubstituted 2-amino-5hydroxy-4-methylnaphtho[1,2-d]thiazolium derivatives 7-24. The cyclic thioureas, imidazolidine-2-thione and hexahydropyrimidine-2-thione, afforded the tetracyclic naphthothiazolium compounds 25 and 26, respectively. In the naphthothiazole products 14-24 derived from unsymmetrical N,N'-disubstituted thioureas, selective cyclization via attack by the nitrogen bearing the sterically least-demanding group was indicated by NMR data. In 1:1 D₂O- Me_2SO-d_6 , the N²,4-dimethyl derivative 3 has methyl resonances at δ 2.25 and 3.01, while the 1,N²,4-trimethyl compound 7 has methyl resonances at δ 2.36, 3.12, and 3.95; the δ 2.25 and 2.36 resonances are assigned as 4-CH₃, the δ 3.01 and 3.12 peaks as N^2 -CH₃, and the 3.95 peak of 7 as 1-CH₃. Compounds 14-22 each display two methyl singlets which appear in the ranges δ 2.0–2.36 (4-CH₃) and 3.78-4.38 (1-CH₃). Compounds 23 and 24 are assigned as

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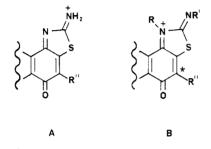
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Substituted Naphtho[1,2-d]thiazolium Salts

 Table I.
 Chemical and Antitrypanosomal Properties of 2-Amino-5-hydroxynaphtho[1,2-d]thiazoles

1-(2-hydroxyethyl) derivatives, because their N-methylene absorptions appear at ca. δ 4.87 and 4.99, respectively, compared with ca. δ 3.65 in N²-(2-hydroxyethyl) derivative 14. Inspection of space-filling molecular models suggests that substituents larger than primary alkyl in the 1-position will suffer severe steric interaction with the C-9 hydrogen and the 2-amino substituent of the naphthothiazole nucleus.

In the simpler case of 1,4-naphthoquinone, only its reaction with thiourea to form 1 was reported by Lau and Gompf.⁸ Although we were able to reproduce this result, we have so far been unable to isolate tractable products from the reaction of N,N'-disubstituted thioureas, such as N,N'-diethylthiourea or imidazolidine-2-thione, with 1,4naphthoquinone. An explanation for this failure may be found in the proposed mechanism⁸ for these reactions, which involves an oxidized intermediate of the quinone imine type, such as A in the case of cyclocondensation of



thiourea itself with a quinone. In the corresponding intermediate B from the reaction of an N,N'-disubstituted thiourea with a quinone, the nitrogen of the newly formed ring would bear an obligatory positive charge. This should render the asterisked position more susceptible to Michael attack by nucleophiles, such as other reaction intermediates, chloride, or ethanol, than would be the case in A. In the adducts derived from 2-methyl-1,4-naphthoquinone, the presence of a methyl group at R'' in B would prevent addition-rearomatization reactions from competing with the proposed intermolecular reduction⁸ which provides the final products, 7–26.

Biological Results

The in vitro activity of compounds 1–26 against T. brucei EATRO 110 was assayed by a modification of a previously described procedure.⁹ Selected concentrations in the range 5 to 1000 μ M in steps of 1.5- to 2-fold dilution were tested in order to determine a minimum lytic concentration (MLC) for each agent, defined as the lowest concentration producing complete lysis of all trypanosomes within 30 min at 37 °C. Results are shown in Table I. The known trypanocide melarsoprol had an MLC of 100 μ M in this system.

The parent compound 2-amino-5-hydroxynaphtho[1,2d]thiazolium chloride (1) does not produce complete lysis even at 500 μ M, its solubility limit in the test medium. The corresponding 4-methyl derivative 2 has significant activity, with an MLC of 100 μ M. In the 4-methyl series bearing identical N-alkyl substituents at the 1-position and the 2-amino group (7-13), activity increases approximately with increasing side-chain size up to at least four carbons, the di-n-butyl and diisobutyl derivatives 12 and 11 being lytic at 10 μ M. The di-n-hexyl analogue 13 is nonlytic at 10 μ M; its assessment at higher concentrations is precluded by partial precipitation. In the case of 25 and 26, in which N-1 and the 2-amino group are joined into a fourth ring by an ethylene or trimethylene bridge (25 and 26, respectively), the former is more active (30 vs. 100 μ M).

In the 1,4-dimethyl series (14-22), larger hydrophobic 2-amino substituents, as in 18-21, confer high activity (MLC ca. 10 μ M), but aliphatic substituents containing polar heteroatoms, as in 14-17, lead to lesser activities (MLC ca. 50-100 μ M) comparable to those of the trimethyl compound 7 and the N-unsubstituted analogue 2.

In the series of 4-methyl compounds lacking 1-substitution (2–6), activity increases with the size of the relatively nonpolar 2-amino substituents, the MLC's declining from 100 to 10 μ M.

In three pairs of compounds, no clear pattern emerges for the effect of the presence or absence of an alkyl substituent at the 1-position (7, 21, and 10 vs. 3, 5, and 4). The presence of a 1-methyl group in 7 results in poorer activity vs. 3, whereas in 21 the 1-methyl group confers greater activity compared to 5. In the case of 10 vs. 4, the presence or absence of a 1-allyl group had little effect on activity.

In comparing 18 and 12, no difference in activity is seen on changing methyl to butyl at the 1-position, when a butyl group is present at the N²-position. In contrast, for the 1-methyl derivatives 7 and 18, a large increase in activity results on changing methyl to butyl at N².

Four compounds (8, 12, 16, and 17) were also tested in the presence of 0.3% added bovine serum albumin in the suspending medium (values in parentheses in Table I). Albumin had an inhibitory effect on lytic activity; the degree of inhibition decreased with increasing polarity of the 2-alkylamino side chain.

None of the test compounds 1-26 showed significant activity against *T. brucei* infection in Swiss Webster mice at the maximum tolerated dosages (Table I).

Discussion

Of the 2-amino-5-hydroxy-4-methylnaphtho[1,2-d]thiazoles studied, those bearing hydrophobic substituents on the amino group showed rapid trypanolytic activity in vitro at concentrations in the 10-20 μ M range (ca. 4-8 ppm), but none produced significant prolongation of the lifetime of T. brucei infected mice. The tenfold inhibition of the activity of dibutyl derivative 12 in vitro by approximately one-tenth (0.3%) of the physiological concentration of serum albumin suggests that binding to host proteins is a possible explanation for the inactivity of these naphthothiazoles in vivo. A similar explanation has been advanced for the behavior of miconazole, which displays in vitro but not in vivo activity against T. brucei, the in vitro activity being subject to inhibition by serum albumin or by whole blood.¹⁰ Hydrophilic side-chain derivatives, such as 16 and 17, the trypanocidal activity of which was less strongly inhibited by albumin, were, however, intrinsically less active than the nonpolar derivatives under albumin-free conditions and displayed greater host toxicity.

Because of the structural novelty of these agents in comparison with known trypanocides, their mechanism of action warrants investigation. Preliminary data indicate that naphthothiazoles of this type may inhibit trypanosomal energy metabolism. The rate of O_2 consumption by trypanosomes in vitro, measured with an O_2 electrode in a closed cell, is inhibited by 75–80% on addition of 30 μ M 8 or 5 μ M 12. These concentrations, though they are immediately below the respective MLC's, do not cause substantial lysis or loss of motility. Lower concentrations cause less inhibition. Further research into the nature of this inhibition may provide leads for the design of intrin-

⁽⁹⁾ Meshnick, S. R.; Grady, R. W.; Blobstein, S. H.; Cerami, A. J. Pharmacol. Exp. Ther. 1978, 207, 1041–1050.

⁽¹⁰⁾ Opperdoes, F. R. Trans. R. Soc. Trop. Med. Hyg. 1980, 74, 423-424.

sically more active analogues of these compounds.

Experimental Section

Melting points were determined on a Thomas-Hoover capillary melting point apparatus and are uncorrected. Proton NMR spectra were recorded on a Varian T60A spectrometer. TLC analysis was performed on EM Laboratories silica gel 60 F-254 precoated plates. Quinones and thioureas were obtained commercially, except for those thioureas described below.

General Procedure for Unsymmetrical 1,3-Disubstituted Thioureas. A 1-2 M solution of the alkyl isothiocyanate (methyl or cyclohexyl) in ether was treated with 1 equiv of an amine. If the amine was not soluble in ether, *i*-PrOH was used as solvent. After 2-16 h, the crystalline thiourea was filtered out and washed with ether or *i*-PrOH. If the thiourea did not crystallize from the reaction mixture, the solvent was removed in vacuo and the residue was cooled and triturated with ether to induce crystallization.

N-(2-Hydroxyethyl)-N'-methylthiourea: yield 44%; mp 74-76 °C (*i*-PrOH), lit.¹¹ mp 73 °C.

N-(2-Methoxyethyl)-N-methylthiourea: yield 81%; mp 47-48 °C (Et₂O). Anal. ($C_5H_{12}N_2OS$) C, H, N.

N-[3-[Bis(2-hydroxyethyl)amino]propyl]-N'-methylthiourea: yield 62%; mp 88-90 °C (*i*-PrOH). Anal. (C₉H₂₁N₃O₂S) C, H, N.

N-[3-(4-Methyl-1-piperazinyl)propyl]-N'-methylthiourea: yield 52%; mp 80−81 °C (*i*-PrOH−Et₂O); hydrogen oxalate, mp 217 °C (*i*-PrOH−H₂O). Anal. ($C_{10}H_{22}N_4S\cdot 2C_2H_2O_4\cdot 0.25H_2O$) C, H, N.

N-Methyl-N-(2-phenylethyl)thiourea: yield 75%; mp 66–67 °C (*i*-PrOH-Et₂O), lit.¹² mp 61–63 °C.

N-(3-Acetylphenyl)-N-methylthiourea: yield 75%; mp 119-120 °C, lit.¹³ mp 120-122 °C.

N-Cyclohexyl-N² (2-hydroxyethyl)thiourea: yield 73%; mp 120-121 °C (*i*-PrOH). Anal. ($C_9H_{18}N_2OS$) C, H, N.

N-Butyl-N'-methylthiourea: yield 79%; mp 41-42 °C ($\text{Et}_2\text{O-C}_5\text{H}_{12}$), lit.¹⁴ mp 40-41 °C.

 N-Methyl-N-octylthiourea: yield 73%; mp 66–67 °C (Et₂O), lit.¹⁵ mp 58–61 °C. Anal. (C₁₀H₂₂N₂S), C, H, N. General Procedure for N-Substituted 2-Amino-5-

General Procedure for N-Substituted 2-Amino-5hydroxy-4-methylnaphtho[1,2-d]thiazoles 2-26. The method was similar to those reported⁸ for the preparation of 1-3 and 5, differing primarily in the use of more concentrated conditions. The synthesis of 8 is illustrative.

1-Ethyl-2-(ethylamino)-5-hydroxy-4-methylnaphtho[1,2d]thiazolium Chloride (8). A solution of 2-methyl-1,4naphthoquinone (13.76 g, 80 mmol) in hot absolute EtOH (100 mL) was poured into a stirred solution of N,N'-diethylthiourea (5.3 g, 40 mmol) and aqueous concentrated HCl (3.4 mL, 40 mmol) in absolute EtOH (50 mL). The solution was allowed to cool and was stirred at ambient temperature for 48 h. The tan precipitate that separated was filtered out and was largely freed of coprecipitated excess quinone and other impurities by successive washes with EtOH, i-PrOH, EtOAc, and Et₂O, affording 6.7 g (52%) of off-white powder. This was recrystallized by dissolution in hot 5:3 MeOH-CHCl₃ (40 mL) and dilution with hot EtOAc (400 mL). After the solution was cooled, the colorless crystalline solid was filtered off and dried, yielding 6.5 g (50%) of 8: mp 262-263 °C dec; NMR (D₂O) δ 1.22 (t, 3), 1.42 (t, 3), 1.96 (s, 3), 3.40 (q, 2), 3.73 (br q, 2), 7.3-8.0 (m, 4); TLC R_f 0.24 (3:2 CHCl₃-acetone). Anal. $(C_{16}H_{19}ClN_2OS)$ Ć, H. Cl, N, S.

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Compound 22 was characterized in the form of its free base, 2-[(3-acetylphenyl)imino]-1,2-dihydro-5-hydroxy-1,4-dimethylnaphtho[1,2-d]thiazole, which precipitated with loss of HCl on dilution of a methanolic solution of the crude hydrochloride with water. In the synthesis of 16 and 17, additional HCl was added to neutralize the side-chain nitrogens. The chloride form of 17, which was used for biological tests, was too hygroscopic for analysis; it was converted by treatment with $NH_4HC_2O_4$ in aqueous MeOH to the hydrogen oxalate salt: mp 206 °C (aqueous EtOH). Anal. ($C_{21}H_{28}N_4OS:3H_2C_2O_4:H_2O$) C, H, N. Further data on 1-26 are given in Table I.

Biological Methods. A. Trypanolytic Activity in Vitro. A modification of a reported procedure was used. T. brucei EATRO 110 bloodstream forms were isolated from blood of infected rats or mice as described previously.^{16,17} Suspensions of $1.5-2 \times 10^7$ trypanosomes/mL in a 1:1 mixture of PO₄-buffered saline containing glucose (PSG 6:4 of ref 17) and minimum essential medium (GIBCO Labs) were kept on ice prior to use and were used within 8 h of isolation. Aliquots in open 1-mL cuvettes maintained at 37 °C were monitored turbidometrically at 750 nm on a Beckman Acta CIII spectrophotometer. When turbidity was stable (5-10 min), the test compound in Me₂SO, aqueous Me₂SO, or H₂O was added with mixing to achieve the desired final concentration selected from the range 1000, 500, 300, 200, 100, 50, 30, 20, 10, 5 μ M. In no case did the final concentration of Me₂SO exceed 1% v/v, a level not affecting turbidity or trypanosome motility. Small aliquots removed at 5, 10, and 30 min after drug addition were examined by phase microscopy for lysis as indicated by loss of motility and loss of phase lucency. Precipitated crystals of test compound, it present, were also noted. Lysis was well correlated with loss of turbidity. The lowest concentration producing complete lysis within 30 min was designated the minimum lytic concentration (MLC). Concentrations near the MLC were replicated using two to three different passage harvests of trypanosomes.

B. Antitrypanosomal Testing in Vivo. Compounds were screened in T. brucei infected Swiss-Webster female mice weighing 20-25 g according to a reported procedure.¹⁸ Briefly, mice were infected ip with 5×10^4 T. brucei EATRO 110 organisms using freshly drawn infected mouse blood diluted with minimum essential medium. To groups of five mice, test compounds as milled suspensions in normal saline containing 0.25% methylcellulose or as solutions in normal saline were administered ip 4-6 h after infection, at which time the presence of infective trypanosomes in the host bloodstream is demonstrable by subinoculation. Untreated controls (5–10 mice per run) died 3.9 ± 0.5 days after infection. Test dosages producing mean survival times >125% of controls are retested. Dosages causing mean survival times <90% of controls or at which deaths occurred within 72 h of infection were considered toxic, and the test compound was retested at a lower dose. Dosages were selected from the range 400, 200, 100, 50, 25 mg/kg, after a preliminary toxicity screen to estimate a maximum tolerated dose. Wet films of tail blood were inspected for trypanosomes on day 3 after infection.

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