SYNTHESIS OF A HEXASACCHARIDE UNIT OF A COMPLEX TYPE OF GLYCAN CHAIN OF A GLYCOPROTEIN*

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ABSTRACT

The synthesis of a branching hexasaccharide unit, methyl 3-O-[2-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- α -D-mannopyranosyl]-6-O-[2,4-di-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- α -D-mannopyranosyl]- α -D-mannopyranoside, a core structure of the complex type of glycans of glycoproteins, is described.

INTRODUCTION

Complex types of glycan chains of glycoproteins exhibit, for the glycans, unique branching modes² that have not been observed in the case of high-D-mannose types of glycan chains. Although high-D-mannose types of glycans show the 3,6-branching mode at the D-mannopyranosyl residues in the core structure, complex types of glycans exhibit the 2,4- and 2,6-branching modes, in addition to 3,6-branching, at the D-mannopyranosyl residues.

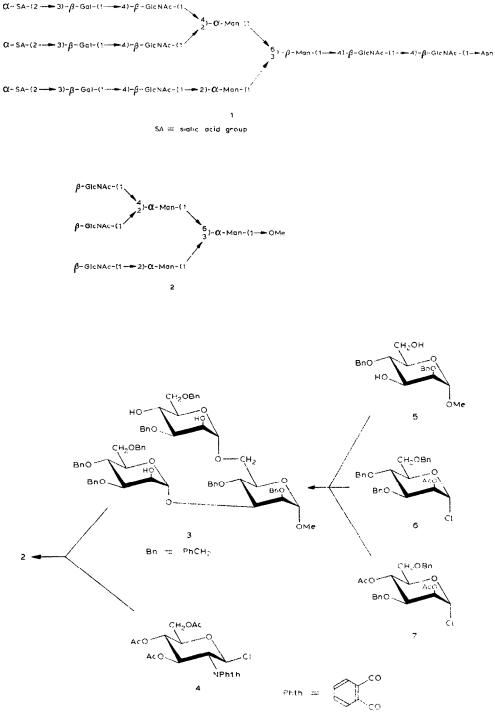
For example, in 1979, Baenziger and Fiete³ reported that the complete structure of the glycan chain of fetuin, the major glycoprotein in fetal-calf serum, is **1**, which shows both 2,4- and 3,6-branching modes. Svensson *et al.*⁴, however, proposed a similar, but isomeric, structure for this glycan. Complex types of glycans of similar structures have recently been reported for such glycoproteins as human α_1 -protease inhibitor⁵, the membrane glycoprotein of vesicular stomatitis virus⁶, α_1 -acid glycoprotein of human plasma⁷, and the membrane glycoprotein of calf-thymocyte plasma⁸.

The unique, branching structure having three antennae, as depicted in 1, and the presence of similar structures in several glycoproteins of biological importance, have stimulated our efforts directed toward their chemical synthesis. In this context, unambiguous synthesis of di- and tri-saccharides having⁹ the sequence $[\beta$ -D-Glc-NAc]_n-(1 \rightarrow X)- α -D-Man-(1 \rightarrow OMe, and synthetic routes towards lactosaminyl donors^{10,11}, were recently reported. As the next step towards the synthesis of 1, we

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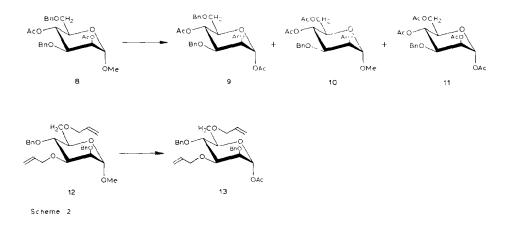


Scheme 1 (synthetic plan)

designed, as the target, compound 2, the inner-core hexasaccharide unit of 1, and developed a synthetic route¹² towards 2.

A retrosynthetic analysis of the target structure 2 indicated, as a key intermediate, the partially protected mannotriose 3, which can accept three molecules of the β -D-GlcNAc donor¹³ 4. The key intermediate 3 should, in turn, be retrosynthesized into one α -D-mannopyranosyl acceptor, 5, and two α -D-mannopyranosyl donors, 6 and 7 (see Scheme 1).

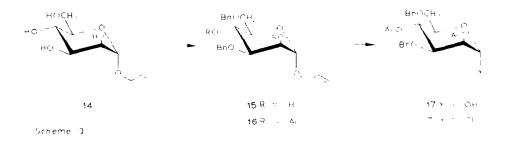
Among the four monosaccharide synthons (4, 5, 6, and 7) thus designed, three, namely, synthons 4 (ref. 13), 5 (ref. 14), and 6 (ref. 15), had already been prepared. Therefore, a synthetic route towards the monosaccharide synthon 7 was first studied. Acetolysis of methyl 2,4-di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranoside^{14a,16} (8) afforded the desired triacetate 9 in only 14.4% yield. The other two products, 10 and 11, were obtained in 36.0 and 7.4% yield, respectively, in addition to a 7.0% recovery of starting material 8. It may be noted that a parallel experiment on the acetolysis of the 3,6-di-O-allyl derivative¹⁴ 12 gave an 80% yield of triacetate 13. Thus, the relative ease of cleavage of the C-O linkages in acetolysis was found to be in the order 6-O-benzyl \geq 1-O-methyl > 6-O-allyl. The facile cleavage of the 6-O-benzyl group on α -D-Man in acetolysis had been reported by Ponpipom¹⁷ and Schuerch *et al.*¹⁸. Therefore, in order to synthesize the mannopyranosyl donor 7, the allyl group¹⁹ was chosen as the protective group for the anomeric oxygen atom, as it can be removed under mild conditions^{19,20} without causing cleavage of the 6-O-benzyl group.



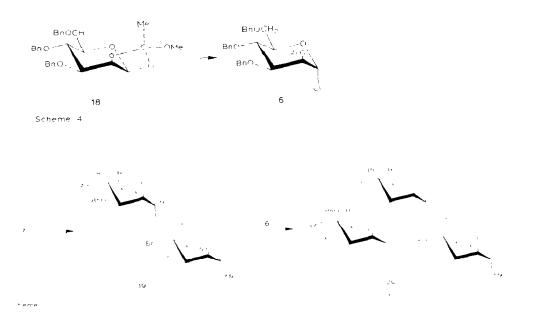
RESULTS

Allyl α -D-mannopyranoside (14) was regioselectively benzylated *via* the partially stannylated intermediate²¹, to give the 3,6-di-*O*-benzyl derivative 15 in 54% yield. Acetylation of 15 to 16, and deallylation of 16 with PdCl₂-NaOAc-aq. AcOH, afforded 17 in 64°, yield. The hydroxyl derivative 17 was quantitatively converted

into the chloride **4** by treatment with the complex²² [Me₂N⁺ = CH-OSOCI] Cl , formed *in situ* from SOCl₂ and a catalytic amount of *N*,*N*-dimethylformamide (DMF) in 1,2-dichloroethane.

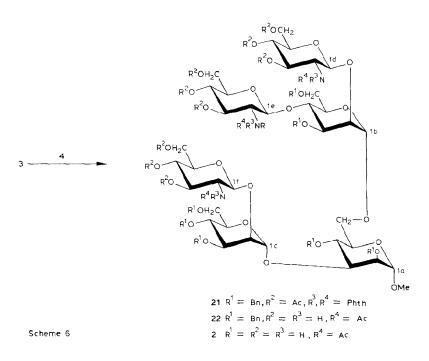


Having prepared the new α -D-mannopyranosyl donor 7, regioselective introduction of 7 at the primary hydroxyl group of 5 was now examined. Treatment of 5 with 1.7 molar equivalents of 7 in the presence of AgOSO₂CF₃-1,1,3,3-tetramethylurea, according to Hanessian and Banoub²³, afforded the desired D-mannobiosyl derivative 19 in 50 °_o yield. The stereochemistry at C-1b was expected to be α , due to be presence¹⁴ of the 2-*O*-Ac group in the D-mannosyl donor 7, and this was supported by the ¹³C-n.m.r. data for the final product (2). Introduction of another α -D-mannopyranosyl group, at O-3a of 19, was achieved, to give the D-mannotriosyl derivative 20 in 62 °_o yield, by using the α -D-mannopyranosyl donor 6, which was readily available from the orthoester 18 by a reported procedure^{14,15}. Subsequent deacetylation of 20 afforded the key intermediate 3.



228

The transformation of 3 into the target compound 2 was performed by using a method developed by Lemieux *et al.*²⁴. The reaction of 3 with 6.25 molar equivalents of the 2-amino-2-deoxy- β -D-glucosyl donor 4 in the presence of AgOSO₂CF₃collidine in 1,2-dichloroethane produced hexasaccharide 21 in 35% yield. Deacylation of 21 with²⁵ BuNH₂ in MeOH, and *N*-acetylation of 21 with Ac₂O-MeOH afforded 22 in 20% yield, and catalytic hydrogenolysis of the benzyl groups of 22 in the presence of 10% Pd-C gave the target hexasaccharide 2.



The structure of 2 was proved by the following ¹H- and ¹³C-n.m.r. data for 2. The 400-MHz, ¹H-n.m.r. spectra of 2 and the reference D-mannotrioside¹⁴ 23 are shown in Fig. 1. Because we had reported⁹ that introduction of a β -D-GlcNAc group on O-2, or O-4, or both, of an α -D-Man residue does not cause any substantial effect on the chemical shift of the anomeric proton of the α -D-Man, a close similarity between the chemical-shift values of H-1a, H-1b, and H-1c in the spectrum of 2 and in that of 23 was expected. Therefore, the assignment of each signal of 2 was made as follows. The signals for H-1a, H-1b, and H-1c were respectively assigned to three doublets (J 2 Hz) at δ 4.740, 4.936, and 5.155, in close agreement with the signals for H-1a, H-1b, and H-1c in²⁶ 23, which, respectively appeared as three doublets (J 2 Hz) at δ 4.738, 4.920, and 5.117. These observations strongly supported the 3,6-branching mode at Man-a in 2. Signals for H-1d, H-1e, and H-1f appeared at δ 4.605, 4.600, and 4.575 as three doublets, with J 8–9 Hz. Although three doublets for the anomeric protons of the GlcNAc groups could not be definitely assigned to any specific group, the observation of the vicinal coupling-constant of 8–9 Hz supported the β -D configuration for each GlcNAc group. We had also observed⁹, for the model compounds, that, when an α -D-Man residue carries a β -D-GlcNAc group at O-2, or two such, at O-2 and O-4, the signals for the anomeric carbon atoms of the α -D-Man residues are shielded by 3.2 or 3.8 p.p.m., respectively. As the signals for C-1a, C-1b, and C-1 of **23** had been reported²⁶ to appear at δ 101.8, 100.2, and 103.2, the corresponding

(1)

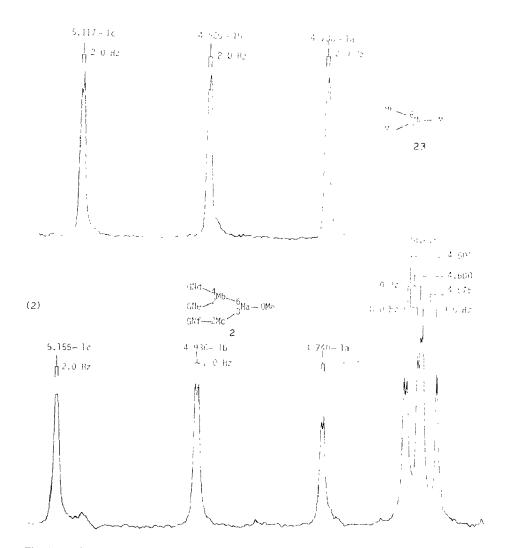


Fig. 1. (1) ¹H-N.m.r. spectrum (400 MHz) of methyl p-mannotrioside **23** in D₂O at 57°. (2) ¹H-N.m.r. spectrum (400 MHz) of hexasaccharide **2** in D₂O at 75 . [¹H-N.m.r. spectra were recorded with a JEOL JNM-FX 400 spectrometer operating at 400.5 MHz. The values of $\delta_{\rm H}$ are expressed in p.p.m. downward from the internal standard, sodium 2.2,3,3-tetradeuterio-4,4-dimethyl-4-silapentanoate. Samples were prepared by "exchanging" them with D₂O, *i.e.*, by dissolving them several times in 99.8"_o D₂O and evaporating *in vacuo*.]

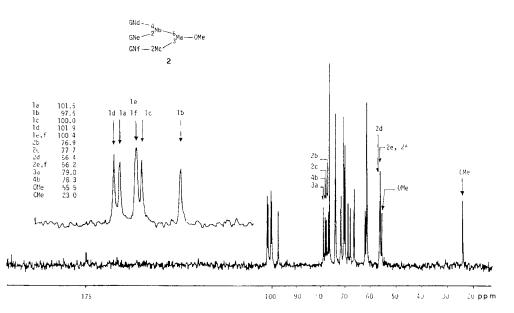


Fig. 2. ¹³C-N.m.r. spectrum of hexasaccharide 2 in D₂O at 80°. [The ¹³C-N.m.r. spectrum was recorded with a JEOL JNM-FX 100FT spectrometer operating at 25.05 MHz. The values of $\partial_{\rm C}$ are expressed in p.p.m. downward from tetramethylsilane, referenced indirectly with an internal standard of 1,4-dioxane ($\delta_{\rm C}$ 66.9).]

signals for C-1a, C-1b, and C-1c in **2** would be expected to appear at δ 101.8, 96.4 (100.2 - 3.8), and 100.0 (103.2 - 3.2). In fact, three signals, with ${}^{1}J_{CH}$ 170 Hz, were observed, at δ 101.5, 100.0, and 97.5, in the spectrum of **2** (see Fig. 2), and were readily assigned to the signals for C-1a, C-1c, and C-1b, respectively. The remaining two signals, observed at δ 101.9 and 100.4 in the ratio of 1:2, with ${}^{1}J_{CH}$ 156 Hz, were assigned as follows. As it had been reported⁹ that the anomeric carbon atoms of β -D-GlcNAc groups linked to O-2 or O-3 of α -D-Man are relatively shielded, and appear at δ 99.6–100.3, whereas those of β -D-GlcNAc groups linked to O-4 or O-6 of α -D-Man appear at δ 101.8–102.0, the signals at δ 101.9 and 100.4 were readily assigned to C-1d, and C-1e and C-1f, respectively. The 13 C- and 1 H-n.m.r. data for the anomeric carbon atoms and protons of **2** were reasonably assigned, based on our previous observations, and were found to be compatible with the target structure **2**.

In conclusion, a regio- and stereo-controlled, synthetic sequence leading to the hexasaccharide unit 2, a core structure of the complex type of glycans of glycoproteins, has now been developed. In close connection with our approach to the synthesis of 2, it may be noted that an approach to complex types of glycans having two antennae was recently reported by Arnarp and Lönngren¹¹.

EXPERIMENTAL

General. — Melting points were determined with a Yanagimoto micro melting-

point apparatus and are uncorrected. Optical rotations were determined with a Perkin–Elmer 241MC polarimeter for solutions in CHCl₃ at 25, unless noted otherwise. Column chromatography was performed on columns of Silica Gel Merck (70-230 mesh; E. Merck, Darmstadt, Germany). Thin-layer chromatography was conducted on precoated plates (layer thickness, 0.25 mm; E. Merck, Darmstadt, Germany) of Silica Gel 60 F_{254} . Flash chromatography was performed on columns of Silica Gel C-300 (200-300 mesh; Wako-Pure Chemical Industries, Osaka, Japan). I.r. spectra were recorded with an EPI-G2 Hitachi spectrophotometer, as KBr disks for the crystalline samples, and as neat films for the liquid samples. ¹H-N.m.r. spectra with a JNM-FX 100FT n.m.r. spectrometer operated at 25.05 MHz. The values of $\delta_{\rm C}$ and $\delta_{\rm H}$ are expressed in p.p.m. downward from the internal standard, tetramethyl-silane, for solutions in CDCl₃, unless noted otherwise.

Acetolysis of methyl 2,4-di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranoside (8). -- To a solution of 8 (307.7 mg), $[\alpha]_D + 56.8^+$ (c 0.48, MeOH), in Ac₂O (3 mL) was added, dropwise, 2°_{0} H₂SO₄- AcOH (3 mL) at 0-5. After stirring for 30 min at 0-5, NaOAc (0.3 g) was added, and the mixture was evaporated *in vacuo*. The residue was partitioned between EtOAc and H₂O, and the organic layer was successively washed with aq. NaHCO₃ and H₂O, dried (MgSO₄), and evaporated *in vacuo*. The residue was flash-chromatographed on SiO₂ C-300 (30 g) with 7:1 toluene– EtOAc, to give the following fractions.

(*i*) A 1:1 mixture (42.8 mg) of 8 (R_F 0.47 in 5:1 toluene-EtOAc) and 9 [(8 + 9): δ_H 6.10 (d, J 2 Hz, H-1 of 9) and 3.38 (s, OMe of 8) in the ratio of 1:3].

(*ii*) **9** (25.6 mg), $R_{\rm F}$ 0.41, $[\alpha]_{\rm D}$ +9.1° (*c* 0.96); $\delta_{\rm H}$ 6.09 (d, 1 H, J 2 Hz), 5.32 (q, 1 H, J 2.4 Hz, H-2), 5.29 (t, 1 H, J 10 Hz, H-4), 2.14 (s, 3 H, Ac), 2.10 (s, 3 H, Ac), and 1.92 (s, 3 H, Ac).

Anal. Calc. for C₂₆H₃₀O₉: C, 64.18; H, 6.22. Found: C, 64.10: H, 6.26.

(*iii*) Compound **10** (34.0 mg), $R_F 0.26$, $[\alpha]_D - 1.0^\circ$ (*c* 1.47); $\delta_H 5.36$ (q, 1 H, J 2.4 Hz, H-2), 5.24 (t, 1 H, J 10 Hz, H-4), 4.74 (d, 1 H, J 2 Hz, H-1), 3.38 (s, 3 H, OMe), 2.16 (s, 3 H, Ac), 2.11 (s, 3 H, Ac), and 2.02 (s, 3 H, Ac).

Anal. Calc. for C₂₀H₂₆O₉: C, 58.53; H, 6.39. Found: C, 58.59; H, 6.41.

(*iv*) A 3:1 mixture (86.8 mg) of 10 and 11 ($R_{\rm F}$ 0.22); (10 \vdash 11): $\delta_{\rm H}$ 6.08 (d, J 2 Hz, H-1 of 11), 4.72 (d, J 2 Hz, H-1 of 10, partially overlapped with CH_2 Ph signals), and 3.36 (s, OMe of 10). The combined yields of 9. 10, and 11 were 14.4. 36.0, and 7.4 $^{\circ}_{+0}$, respectively. A 7.0 $^{\circ}_{+0}$ recovery of 8 was also observed.

3.6-Di-O-allyl-2.4-di-O-benzyl- α -D-mannopyranosyl acetate (13). — To a solution of 12 (12.6 g) in Ac₂O (100 mL) was added, dropwise, 2°₀ H₂SO₄-AcOH (100 mL) at 0-5. After stirring the mixture for 10 min at 5. NaOAc (8.0 g) was added, to neutralize the H₂SO₄. The mixture was evaporated *in vacuo*, and the residue was partitioned between EtOAc and H₂O. The organic layer was successively washed with aq. NaHCO₃ and H₂O, dried (MgSO₄), and evaporated, to give an oil which was chromatographed on SiO₂ (700 g) with 10:1 toluene-EtOAc, to give 13 (10.6 g, 79.1 $^{\circ}_{0}$); $R_{\rm F}$ 0.54 in 5:1 toluene-EtOAc; [α]_D + 32.2° (c 1.06); $\delta_{\rm H}$ 7.5-7.3

(m, 10 H, aromatic), 6.24 (d, 1 H, J 2 Hz, H-1), 6.16–5.76 (m, 2 H, 2 $-CH=CH_2$), 5.44–5.10 (m, 4 H, 2 $-CH=CH_2$), 4.98 and 4.63 (ABq, 2 H, J 9 Hz, CH_2 Ph), 4.80 (s, 2 H, CH_2 Ph), and 2.04 (s, 3 H, Ac).

Anal. Calc. for C₂₈H₃₄O₇: C, 69.69; H, 7.10. Found: C, 69.73; H, 7.07.

Allyl 3,6-di-O-benzyl- α -D-mannopyranoside (15). — A mixture of 14 (20 g, 91 mmol) and (Bu₃Sn)₂O (77.5 g, 130 mmol) in toluene (1.000 L) was boiled and stirred under reflux for 3 h with continuous, azeotropic removal of water, cooled, and evaporated *in vacuo*. A solution of the residue in benzyl bromide (200 mL) was stirred under Ar for 2 days at 90°, cooled, evaporated *in vacuo*, a solution of the residue in EtOAc mixed with aq. KF, and the mixture stirred for 30 min. The precipitate of Bu₃SnF was filtered off, the filtrate extracted with EtOAc, and the extract washed with water, dried (MgSO₄), and evaporated *in vacuo* to an oil which was chromatographed on SiO₂ (1 kg) with 1:1 toluene–EtOAc, to give 15 (19.5 g, 53.6 %); R_F 0.44 in 1:1 toluene–EtOAc; $[\alpha]_D$ +30.8° (c 0.39); δ_H 7.30 (s, 5 H, aromatic), 7.28 (s, 5 H, aromatic), 6.1–5.65 (m, 1 H, -CH=CH₂), 5.4–5.05 (m, 2 H, CH=CH₂), and 4.88 (d, 1 H, J 1 Hz, H-1).

Anal. Calc. for C₂₃H₂₈O₆: C, 68.98; H, 7.05. Found: C, 68.96; H, 7.07.

Allyl 2,4-di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranoside (16). — To a solution of 15 (1.3 g, 3.2 mmol) in pyridine (15 mL) was added Ac₂O (7 mL) at 0-5°, and the mixture was stirred for 16 h at 20°. The usual processing, and chromatography on SiO₂ with 3:1 toluene–EtOAc afforded a quantitative yield of 16; R_F 0.59 in 3:1 toluene–EtOAc; $[\alpha]_D$ +11.1° (c 0.45); δ_H 7.28 (s, 5 H, aromatic), 7.26 (s, 5 H, aromatic), 6.0–5.64 (m, 1 H, -CH=CH₂), 4.86 (bs, 1 H, H-1), 2.11 (s, 3 H, Ac), and 1.90 (s, 3 H, Ac).

Anal. Calc. for C₂₇H₃₂O₈: C, 66.92; H, 6.66. Found: C, 66.94; H, 6.66.

2,4-Di-O-acetyl-3,6-di-O-benzyl-D-mannopyranose (17). — A mixture of 16 (500 mg, 1.03 mmol), NaOAc (200 mg, 2.4 mmol), and PdCl₂ (200 mg, 1.1 mmol) in AcOH (2 mL)-H₂O (0.1 mL) was stirred for 14 h at 20°, and evaporated *in vacuo* below 30°. A solution of the residue in EtOAc was successively washed with aq. NaHCO₃ and H₂O, dried (MgSO₄), and evaporated *in vacuo*. Chromatography of the residue on SiO₂ (30 g) with 4:1 toluene–EtOAc afforded 17 (294 mg, 64%); $R_{\rm F}$ 0.30 in 3:1 toluene–EtOAc; $[\alpha]_{\rm D}$ –18.9° (c 0.37), unchanged after 16 h; $\delta_{\rm H}$ (CDCl₃-D₂O): 7.25 (s, 5 H, aromatic), 7.24 (s, 5 H, aromatic), 5.31 (dd, 1 H, $J_{1,2}$ 2, $J_{2,3}$ 3.5 Hz, H-2), 5.14 (d, 1 H, J 2 Hz, H-1), 5.10 (t, 1 H, J 10 Hz, H-4), 3.90 (dd, 1 H, $J_{2,3}$ 3.5, $J_{3,4}$ 10 Hz, H-3), 2.08 (s, 3 H, Ac), and 1.88 (s, 3 H, Ac).

Anal. Calc. for C₂₄H₂₈O₈: C, 64.85; H, 6.35. Found: C, 64.36; H, 6.46.

2,4-Di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranosyl chloride (7). — To a solution of 17 (2.0 g, 4.5 mmol) in Cl(CH₂)₂Cl (28 mL) were added SOCl₂ (0.66 mL, 9 mmol) and DMF (0.1 mL) at 0-5°. The mixture was stirred for 14 h at 15-20°, filtered through a bed of SiO₂ (2.5 g), and the filtrate co-evaporated with toluene (three times), to give 7 (quantitative yield). An analytical sample of 7 was obtained by chromatography on SiO₂ with 3:1 toluene-EtOAc (R_F 0.52); $[\alpha]_D$ +40.0° (c 0.27); δ_H 7.26 (s, 10 H, aromatic), 6.02 (d, 1 H, J 1 Hz, H-1), 5.41 (dd, 1 H, J_{1,2} 2,

 $J_{2,3}$ 4 Hz, H-2), 5.30 (t, 1 H, J 10 Hz, H-4), 4.12 (dd, 1 H, $J_{2,3}$ 4, $J_{3,4}$ 10 Hz, H-3), 2.10 (s, 3 H, Ac), and 1.89 (s, 3 H, Ac).

Anal. Calc. for C₂₄H₂₇ClO₇: C, 62.27; H, 5.88; Cl. 7.66. Found: C, 61.69; H, 5.82; Cl, 7.72.

Methyl 2,4-di-O-benzyl-6-O-(2,4-di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside (**19**). — To a mixture of **5** (1.65 g, 4.4 mmol). AgOSO₂CF₃ (3.44 g, 13.4 mmol), and 1,1,3,3-tetramethylurea (4.2 mL, 35 mmol) in Cl(CH₂)₂Cl (20 mL) was added, dropwise, a solution of **7** (2.00 g, 4.3 mmol) in Cl(CH₂)₂Cl (5 mL) at 0–5°. After stirring for 5 h at 20°, a solution of **7** (1.5 g, 3.2 mmol) in Cl(CH₂)₂Cl (5 mL) at 0–5°. After stirring for 5 h at 20°, a solution of **7** (1.5 g, 3.2 mmol) in Cl(CH₂)₂Cl (5 mL) was added dropwise at 0–5, the mixture stirred for 16 h at 20°, diluted with Cl(CH₂)₂Cl (50 mL), filtered, and the filtrate processed, to give an oil which was ehromatographed on SiO₂ (200 g) with 4:1 toluene-EtOAc, to give **19** (1.75 g, 49.5°₀); $R_{\rm F}$ 0.38 in 3:1 toluene-EtOAc; $[\alpha]_{\rm D}$ +29.7 (*c* 0.58): $\delta_{\rm H}$ 7.32-7.20 (m, 20 H, aromatic), 5.44 (bs, 1 H, H-2b), 5.26 (t, 1 H, *J* 10 Hz, H-4b), 4.94 (bs, 1 H, H-1a or H-1b), 3.24 (s, 3 H, OMe), 2.10 (s, 3 H, Ac), and 1.86 (s, 3 H, Ac).

Anal. Cale. for C49H52O13: C, 67.49; H, 6.54. Found: C. 67.41: H, 6.55.

Methyl 3-O-(2-O-acetyl-3,4.6-tri-O-benzyl- α -D-mannopyranosyl)-2.4-di-O-benzyl-6-O-(2,4-di-O-acetyl-3,6-di-O-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside (**20**). — To a mixture of **19** (1.40 g, 1.75 mmol), AgOSO₂CF₃ (0.93 g, 3.6 mmol), and tetramethylurea (1.5 mL, 12.6 mmol) was added dropwise a solution of **6** (1.25 g, 2.45 mmol) in Cl(CH₂)₂Cl (5 mL) during 40 min at 0–5 . The mixture was stirred for 3 days at 20 , diluted with Cl(CH₂)₂Cl (100 mL), filtered (Celite), and the filtrate processed as usual. Chromatography on SiO₂ (150 g) with 4:1 toluene -EtOAc afforded **20** (1.38 g, 61.9%); R_F 0.49 in 3:1 toluene-EtOAce: $[\alpha]_D$ +35.2 (c 0.29); δ_H 7.4–7.2 (m, 35 H, aromatic), 5.54 and 5.46 (dd, 2 H, J 2.4 Hz, H-2b,2c), 5.21 (d, 1 H, J 2 Hz, H-1), 5.20 (t, 1 H, J 9 Hz, H-4b), 4.96 (d, 1 H, J 2 Hz, H-1), 3.20 (s, 3 H, OMe), 2.14 (s, 3 H, Ac), 2.10 (s, 3 H, Ac), and 1.88 (s, 3 H, Ac).

Anal. Calc. for C₇₄H₈₂O₁₉: C, 69.69; H, 6.48. Found: C. 69.42; H, 6.43.

Methyl 2,4-di-O-benzyl-6-O-(3,6-di-O-benzyl- α -D-mannopyranosyl)-3-O-(3,4,6tri-O-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside (3). A solution of 20 (638 mg, 0.5 mmol) in oxolane (10 mL)-MeOH (5 mL)-2M NaOMe (0.2 mL) was stirred for 3 h at 20°. The usual processing gave a quantitative yield of 3. A small portion of 3 was purified by chromatography on SiO₂, to give an analytical sample: $R_{\rm F}$ 0.30 in 1:1 toluene-EtOAc; $[\alpha]_{\rm D}$ +39.5° (c 0.39); $\delta_{\rm H}$ 7.4-7.1 (35 H, aromatic), 5.20 (bs, 1 H, H-1), 5.00 (bs, 1 H, H-1), and 3.22 (s. 3 H, OMe).

Anal. Cale. for C₆₈H₇₆O₁₆: C, 71.06. H, 6.67. Found: C, 70.70; H, 6.76.

Methyl 2,4-di-O-benzyl-6-O-[3,6-di-O-benzyl-2,4-di-O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)- α -D-mannopyranosyl]-3-O-[3,4,6-tri-O-benzyl-2-O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl]- α -D-mannopyranosyl]- α -D-mannopyranoside (21). – To a mixture of 3 (546 mg, 0.48 mmol), AgOSO₂CF₃ (771 mg, 3 mmol), and collidine (0.6 mL) in Cl(CH₂)₂Cl (15 mL) was added, dropwise, a solution of 4 (680 mg, 1.5 mmol) in Cl(CH₂)₂Cl (3 mL) at 0-5⁺. After stirring for 6 h at 20⁺, compound 4 (680 mg, 1.5 mmol) in Cl(CH₂)₂Cl (3 mL) was added, and the mixture was stirred for 16 h at 20°. Processing, and chromatography on SiO₂ (50 g) with 50:3 CHCl₃-acetone, afforded **21** (400 mg, 34.7%); $R_{\rm F}$ 0.70 in 25:2 CH₂Cl₂-acetone; $[\alpha]_{\rm D}$ +22.4° (*c* 0.38); $\delta_{\rm H}$ 7.8–7.0 (m, 47 H, aromatic), 3.24 (s, 3 H, OMe), and 2.04, 2.00, 1.98, 1.94, 1.92, 1.88, 1.84, 1.82, and 1.78 (9 s, 9 Ac).

Anal. Calc. for $C_{128}H_{133}N_3O_{43}$: C, 64.02; H, 5.58; N, 1.75. Found: C, 64.02; H, 5.56; N, 1.60.

Methyl 3-O-[2-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl]-2,4-di-O-benzyl-6-O-[2,4-di-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-3,6-di-O-benzyl- α -D-mannopyranosyl]- α -D- α -D (40 mL) and BuNH₂ (4.0 mL, and the suspension stirred vigorously for 1 h, and filtered. (The filtrate did not contain 22, according to t.l.c. examination.) The solid was chromatographed on SiO₂ (50 g) with 50:24:1 CHCl₃-MeOH-H₂O; [α]_D + 19.4° (c 0.18); δ _H (CD₃OD): 7.66-7.0 (m, 35 H, aromatic), 1.95 (s, 6 H, 2 NAc), and 1.86 (s, 3 H, NAc).

Anal. Calc. for $C_{92}H_{115}N_3O_{31}$: C, 62.82; H, 6.59; N, 2.39. Found: C, 62.38; H, 6.70; N, 2.29.

Methyl 3-O-[2-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- α -D-mannopyranosyl]-6-O-[2,4-di-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- α -D-mannopyranosyl]- α -D-mannopyranoside (2). A mixture of 22 (42 mg) and 10% Pd-C (70 mg) in EtOH (5 mL)-H₂O (0.7 mL) was stirred under H₂ for 8 h at 50-60°. The usual processing gave 2 quantitatively; $R_{\rm F}$ 0.20 in 10:30:3 CHCl₃-MeOH-H₂O; $[\alpha]_{\rm D}$ + 14.7° (c 0.34).

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