

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters



journal homepage: www.elsevier.com/locate/bmcl

Hit to lead studies on (hetero)arylpyrimidines—Agonists of the canonical Wnt-β-catenin cellular messaging system

Adam M. Gilbert ^{a,*}, Matthew G. Bursavich ^a, Nippa Alon ^a, Bheem M. Bhat ^c, Frederick J. Bex ^c, Michael Cain ^d, Valerie Coleburn ^c, Virginia Gironda ^c, Paula Green ^c, Diane B. Hauze ^b, Yogendra Kharode ^c, Girija Krishnamurthy ^f, Matthew Kirisits ^a, Ho-Sun Lam ^c, Yao-Bin Liu ^c, Sabrina Lombardi ^a, Jeanne Matteo ^c, Richard Murrills ^c, John A. Robinson ^c, Sally Selim ^c, Michael Sharp ^c, Raymond Unwalla ^b, Usha Varadarajan ^e, Weiguang Zhao ^c, Paul J. Yaworsky ^d

^a Chemical Sciences, Wyeth Research, 401 North Middletown Road, Pearl River, NY 10965, United States

^b Chemical Sciences, Wyeth Research, 500 Arcola Road, Collegeville, PA 19426, United States

^c Tissue Repair, Wyeth Research, 500 Arcola Road, Collegeville, PA 19426, United States

^d Tissue Repair, Wyeth Research, 200 Cambridge Park Drive, Cambridge, MA 02140, United States

^e Biological Technologies, Wyeth Research, 87 Cambridge Park Dr Cambridge, MA 02140, United States

f Screening Sciences, Wyeth Research, 401 North Middletown Road, Pearl River, NY 10965, United States

ARTICLE INFO

Article history: Received 17 September 2009 Revised 20 October 2009 Accepted 21 October 2009 Available online 25 October 2009

Keywords: Wnt-β-catenin agonist (Hetero)arylpyrimidines Osteoporosis Calvaria

ABSTRACT

A series of (hetero)arylpyrimidines agonists of the Wnt- β -catenin cellular messaging system have been prepared. These compounds show activity in U2OS cells transfected with Wnt-3a, TCF-luciferase, Dkk-1 and tk-Renilla. Selected compounds show minimal GSK-3 β inhibition indicating that the Wnt- β -catenin agonism activity most likely comes from interaction at Wnt-3a/Dkk-1. Two examples **1** and **25** show in vivo osteogenic activity in a mouse calvaria model. One example **1** is shown to activate non-phosphorylated β -catenin formation in bone.

© 2009 Elsevier Ltd. All rights reserved.

Activation of the Wnt- β -catenin signaling pathway has been shown to play important roles in development, tissue regeneration, stem cell control, tumor progression and metastases.^{1,2} This pathway has also been implicated in the regulation of bone homeostasis. The Wnt co-receptor LRP5 is central to this function where both gain- and loss-of-function mutations have been described in humans resulting in high bone mass or an osteoporosis pseudoglioma syndrome, respectively.^{3–5} Murine models of both genetic conditions have been successfully generated.^{6,7} Pharmacologic inhibition of either Dickkopf-1 (Dkk-1) or Sclerostin, two LRP5binding negative regulators of Wnt- β -catenin signaling, results in increased bone mineral density in rodents.^{8,9} These data provide additional evidence that agonists of Wnt- β -catenin activity could yield an osteogenic agent.

Signaling by the Wnt-β-catenin pathway has been extensively studied in many systems.^{1,2,10} Briefly, secreted Wnt ligands bind to cells via Frizzled receptors and the LRP5 or LRP6 co-receptors. This ligand-receptor interaction activates the cytoplasmic protein

* Corresponding author. E-mail address: gilbera@wyeth.com (A.M. Gilbert). Disheveled which in turn inactivates a protein complex comprising Axin, Adenomatous Polyposis Coli, and Glycogen Synthase Kinase- 3β (GSK- 3β). The resulting repression of GSK- 3β activity leads to an accumulation of unphosphorylated β -catenin enabling it to translocate to the nucleus and form a transcriptional co-activator complex with T-cell factor/lymphoid enhancer-binding factor (TCF/LEF). Extracellularly, this mechanism can be antagonized by several secreted molecules including Dkk-1, Sclerostin and the Secreted Frizzled Related Proteins.

Both pharmacologic and genetic data support that the Wnt- β catenin pathway is a key mediator of the normal adaptive response to mechanical loading in bone.^{11–13} As a result, agents capable of mimicking the beneficial effects of Wnt- β -catenin activation on the skeleton would represent a novel approach for the treatment of osteoporosis and other bone disorders. In this Letter, we report on the hit to lead studies on a class of (hetero)arylpyrimidines: agonists of the Wnt- β -catenin pathway and their use as potential anabolic agents to increase bone mass starting from **1**, a (hetero)arylaminopyrimdine found from high-throughput screening.

ero)arylaminopyrimdine found from high-throughput screening. Compounds **4** are generally prepared according to Scheme 1.¹⁴ Thus 2-chloropyrimidine **2** when reacted with aryl/heteroaryl



Scheme 1. Reagents and conditions: (a) (1) Ar^1Li , Et_2O , -78 to 0 °C; (2) DDQ, THF, 23 °C; (b) $H_2N(CHR)_nAr^2$, NMP, 90 °C or $H_2N(CHR)_nAr^2$, NaH, DMSO, 80 °C.

lithiums in Et₂O produces the corresponding 4-aryl/heteroaryl compounds.^{15,16} Target compounds **4** are prepared by reacting **3** with amines in hot NMP or by first deprotonating the amine with NaH in DMSO and then heating with **3** in hot DMSO.



Wnt-3a/Dkk-1/TCF-Luci EC $_{50}$: 4.2 μM (2.9 Fl @ 20 μM) GSK-3 β IC $_{50}$: 51 μM

Compounds were assayed using U2OS cells transfected with Wnt-3a, TCF-luciferase, Dkk-1 and tk-Renilla as a signal normalizer compared to control U2OS cells transfected with Wnt-3a, TCF-Luciferase and tk-Renilla.¹⁷ The signal from the TCF-luciferase is reduced in the presence of the inhibitor Dkk-1. Compounds that modulate Dkk-1 activity or act downstream of the Wnt-3a/Dkk-1 complex (i.e., GSK-3 β inhibition) cause an increase in the luciferase signal. For all compounds in this manuscript, EC₅₀s were determined as well as the fold-induction response at 20 μ M compared to the control U2OS cells. A [³²P]-GSK-3 β was used to assay compounds for Wnt/ β -catenin activity downstream (of, from) Wnt-3a/Dkk-1.

Compound 1 shows a robust TCF-Luciferase response at 20 μ M as well as a μ M-EC₅₀(1: TCF(20 μ M): 2.9; EC₅₀: 4.2 μ M)(Table 1). Since arylamino-pyrimidines are well-known kinase inhibitors,¹⁸ 1 was assayed for GSK-3 β inhibition but little kinase inhibition was seen (IC₅₀: 51 μ M). Modulation of the R¹ substituent to ethylpyridines (5 and 6) produces compounds of comparable efficacy/potency to 1. The replacement of R¹ with a propyl(dimethylpyrazole) group

Table 2

Wnt-β-catenin activities of compounds 1, 10-11



Compd	R ¹	R ²	TCF ^a (20 μM)	$\text{EC}_{50}{}^{b}\left(\mu M\right)$
1	CH ₂ -1 <i>H</i> -Imidazole	4-(Pyridin-4-yl)	2.9	4.2
10	CH ₂ -1H-Imidazole	4-(Pyridin-3-yl)	4.7	8.8
11	CH ₂ -1 <i>H</i> -Imidazole	4-(3-Nitrophenyl)	3.6	46.3
12	4-Pyridine	4-(Pyridin-3-yl)	3.4	20.2
13	3-(1H-Indole)	4-(Pyridin-3-yl)	10.0	4.1
14	3-(2-Methyl-1 <i>H</i> -indol-5-ol)	4-(pyridin-3-yl)	5.1	12.2
15	4-(1H-Imidazole)	4-(Pyridin-3-yl)	3.1	28.8
16	4-(1H-Imidazole)	2-(Benzo[b]thiophene)	3.7	3.3
17	4-(1H-Imidazole)	2-(Naphthyl)	1.1	12.1

^a U2OS: Wnt-3a/Dkk-1/TCF-Luci fold induction at 20 μ M. Values are the average of 2 or 3 runs. Error ±20%.

 $^{\rm b}$ U2OS: Wnt-3a/Dkk-1/TCF-Luci EC_{50}. Values are the average of 2 or 3 runs. Error ±20%.

Table 1

Wnt-β-catenin activities of compounds 1, 5-9



Compd	R ¹	TCF ^a (20 μM)	ЕС ₅₀ ^в (µМ)	
1	N-(3-(1H-Imidazol-1-yl)propane)	2.9	4.2	
5	N-(2-(Pyridin-4-yl)ethane)	4.3	6.8	
6	N-(2-(Pyridin-3-yl)ethane)	2.8	4.6	
7	N-(3-(3,5-Dimethyl-1H-pyrazol-1-yl)propyl)	2.1	29.6	
8	N-(2-(1H-Indol-3-yl)ethane)	7.5	2.7	
9	N-(S)-3-(1H-Indol-3-yl)-2-propan-1-ol amine	2.2	FTC ^c	

 a U2OS: Wnt-3a/Dkk-1/TCF-Luci fold induction at 20 $\mu M.$ Values are the average of 2 or 3 runs. Error ±20%.

 $^{\rm b}$ U2OS: Wnt-3a/Dkk-1/TCF-Luci EC_{50}. Values are the average of 2 or 3 runs. Error $\pm 20\%$

^c FTC: EC₅₀ determination failed to converge.

produces a less potent compound (**7**) while the ethylindole derivative (**8**) gives a 2.5-fold more efficacious analog of **1** (**8**: TCF (20 μ M): 7.5; EC₅₀: 2.7 μ M). The more complex **9** containing a propanolindole substituent possesses similar efficacy to **1** but the EC₅₀ fails to converge.

Table 2 outlines the effects of varying the 4-pyrimidine position on compounds similar to **1**. Changing R^2 from a pyridine-4-yl to a pyridine-3-yl results in a slight increase in efficacy (**10**: TCF (20 µM): 4.7) while a 3-NO₂-phenyl group gives a much less potent compound (**11**: EC₅₀: 46.3 µM). Compound **13** where $R^1 = 3-(1H$ indole) shows µM-potency with large fold-induction (**13**: TCF (20 µM): 10.0; EC₅₀: 4.1 µM).

Of the 4-(1*H*-imidazole) indole compounds **15–17**, the 2-(benzo[*b*]thiophene) analog **16** possesses the best combination of potency and efficacy (EC₅₀: 3.3 μ M, TCF (20 μ M): 3.7). Several phenethyl analogs were also prepared, but they showed poor Wnt- β -catenin agonism.

To follow-up compound **16** which shows μ M potency and moderate efficacy in the Wnt-3a/Dkk-1/TCF-Luci assay, 2-(naphthyl)-2-(benzo[*b*]thiophene)-derivatives were prepared according to the



Scheme 2. Reagents and conditions: (a) SOCl₂, MeOH, 60 °C; (b) Trit–Cl, TEA, MeCN, 23 °C; (c) LiAlH₄. THF 0 to 23 °C; (d) NaH, R¹X, DMF, 50 °C; (e) HCl, THF, reflux; (f) Scheme 1, step b.

chemistry outlined in Schemes 2 and 3. Histidine **18** may be converted to methyl ester **19** using standard conditions. After being bis(tritylated) to produce **20**, reduction with LiAlH₄ produces the primary alcohol **21**. Alkylation with various alkyl and aryl bromides produces **22**. Deprotection with HCl yields the derivatized histidines **23** which were incorporated into products **4** using the procedure of Scheme 1, Step b. Analogs where the imidazole nitrogen is substituted are prepared by reacting compounds like **25**



Scheme 3. Reagents and conditions: (a) NaH, R¹X, THF/DMSO, 23 °C.

Table 3

Wnt-β-catenin activities of compounds 25-29



Compd	R ¹	R ²	TCF ^a (20 μM)	EC ₅₀ ^b (μΜ)
25	Н	2-(Naphthyl)	4.9	6.8
26	Ethyl	2-(Naphthyl)	1.9	14.4
27	Methylenecyclohexyl	2-(Naphthyl)	2.2	20.8
28	(2-Fluoro-3-	2-(Naphthyl)	1.6	10.9
29	(trifluoromethyl)benzyl Prop-2-ynyl	2-(Benzo[b]thiophene)	3.5	FTC

^a Values are the average of 2 or 3 runs. Error ±20%.

^b FTC: EC₅₀ failed to converge.

with NaH followed by the addition of an alkylating agent to produce **30**.

O-Substituted derivatives in Table 3 show moderate Wnt-3a/ Dkk-1/TCF-Luci efficacy at 20 μ M, but most of the compounds were not as potent as and efficacious as **16**. This was true for alkyl, aryl and alkynyl derivatives **25–29**. The exception was hydroxyl analog **25**. Analogs of **16** where the imidazole nitrogen is alkylated show similar to greater efficacy compared to the O-substituted analogs as a number of substituents are tolerated (**31**, **32**: benzyl, **33**: propynyl, **34**: acetamide, **35**, **36**: 3-propanitrile), but comparatively weaker potency to **25** (see Table 4).

Example **25** represents a compound with good properties of a chemical lead based on **1**. It has robust Wnt-3a/Dkk-1/TCF-Luci efficacy @ 20 μ M (FI 4.9), good potency (EC₅₀: 6.8 μ M) and an acceptable ligand efficiency. The calculated physical properties are in line for a molecule with good drug-like properties. Good aqueous solubility, moderate PAMPA permeability and good rat liver microsome stability is also seen.





In order to establish the osteogenic activity of this series, **1** was evaluated in vivo via local subcutaneous injection over the right side of the calvaria of wild type C57BL/6 mice.¹¹ As a positive control, an inhibitor of GSK-3 β (GSKi), 3-(3-chloro-4-hydroxyphenylamino)-4-(2-nitrophenyl)-1*H*-pyrole-2,5-dione),¹⁹ was administered at 1 mg/kg/day as previously described.¹¹ Vehicle alone was used as the negative control. Using standard dynamic histomorphometry measurements, the mineral apposition rate of new bone formation was

 Table 4

 Wnt-β-catenin activities of compounds 31–38



Compd	R ¹	R ²	$\text{TCF}^{a}\left(20\;\mu\text{M}\right)$	$\text{EC}_{50}{}^{\text{b}}\left(\mu M\right)$
31	3,5-Difluorobenzyl	2-(Benzo[b]thiophene)	1.7	16.0
32	3,5-Difluorobenzyl	2-(Naphthyl)	2.0	FTC
33	Prop-2-ynyl	2-(Benzo[b]thiophene)	3.0	14.6
34	2-Acetamide	2-(Naphthyl)	3.9	FTC
35	3-Propanitrile	2-(Benzo[b]thiophene)	4.1	FTC
36	3-Propanitrile	2-(Naphthyl)	5.5	FTC

^a Values are the average of 2 or 3 runs. Error ±20%.

^b FTC: EC₅₀ determination failed to converge.





0.1

Compound 25 (mg/kg/day)

Figure 1. Compounds 1 and 25 are osteogenic in vivo. (A) Quantitative dynamic histomorphometry was performed on murine calvaria following 7 days of 1 treatment. Mineral apposition rates were calculated and are presented as an index of anabolic, osteoblast activity. Veh, vehicle; GSKi, Glycogen synthase kinase-3ß inhibitor,¹⁹ p < 0.01 versus vehicle. (B–D) Immunohistochemical detection of nonphosphorylated β-catenin is enhanced in compound-treated calvaria suggesting that Wnt-\beta-catenin signaling is activated. Positive cells are identified by the pink staining as indicated by the arrowheads. Vehicle (B), GSKi (C), 0.3 mg/kg/day 1 (D), 1.0 mg/kg/day 1 (E). (F) Compound 25 is osteogenic in the same assay showing significantly elevated mineral apposition rates versus vehicle.

calculated exactly as previously described.¹¹ All doses of **1** showed a significant increase in anabolic bone activity over vehicle with levels equivalent to that attained by the GSKi positive control in all but the lowest tested dose of 1 (Fig. 1A). Mechanistically, this anabolic bone activity could be ascribed to activated Wnt- β -catenin signaling as visualized by the immunohistochemical detection of non-phosphorylated β-catenin in the osteoblastic cells lining the periosteal surface of the calvaria (Fig. 1B-E). Furthermore, using the same in vivo model, 25 was also shown to be an anabolic bone agent (Fig. 1F). Together these data support that **1** and **25** are both Wnt- β -catenin pathway agonists and efficacious osteogenic agents in vivo.

In conclusion, we have disclosed a series of (hetero)arylpyrimidines that act as agonists of the Wnt- β -catenin pathway. While we currently don't know their exact mechanism of action, these compounds don't function via inhibition of GSK-3B. Moreover, examples of this equity show osteogenic in vivo activity in mouse calvaria as well as immunohistochemical data indicating that Wnt-β-catenin activity is activated in bone.

Acknowledgments

We thank Dr. John Ellingboe for support of this work and the Chemical Sciences Compound Properties group for providing physicochemical profiling data on all analogs disclosed in this work.

References and notes

- Klaus, A.; Birchmeier, W. Nat. Rev. Cancer 2008, 8, 387.
- Nusse, R. Cell Res. 2008, 18, 523. 2.
- Boyden, L. M.; Mao, J.; Belsky, J.; Mitzner, L.; Farhi, A.; Mitnick, M. A.; Wu, D.; 3. Insogna, K.; Lifton, R. P. N. Eng. J. Med. 2002, 346, 1513.
- Little, R. D.; Carulli, J. P.; Del Mastro, R. G.; Dupuis, J.; Osborne, M.; Folz, C.; Manning, S. P.; Swain, P. M.; Zhao, S. C.; Eustace, B.; Lappe, M. M.; Spitzer, L.; Zweier, S.; Braunschweiger, K.; Benchekroun, Y.; Hu, X.; Adair, R.; Chee, L.; FitzGerald, M. G.; Tulig, C.; Caruso, A.; Tzellas, N.; Bawa, A.; Franklin, B.; McGuire, S.; Nogues, X.; Gong, G.; Allen, K. M.; Anisowicz, A.; Morales, A. J.; Lomedico, P. T.; Recker, S. M.; Van Eerdewegh, P.; Recker, R. R.; Johnson, M. L. Am. J. Hum. Genet. 2002, 70, 11.
- Gong, Y.; Slee, R. B.; Fukai, N.; Rawadi, G.; Roman-Roman, S.; Reginato, A. M.; Wang, H.; Cundy, T.; Glorieux, F. H.; Lev, D.; Zacharin, M.; Oexle, K.; Marcelino, J.; Suwairi, W.; Heeger, S.; Sabatakos, G.; Apte, S.; Adkins, W. N.; Allgrove, J.; Arslan-Kirchner, M.; Batch, J. A.; Beighton, P.; Black, G. C.; Boles, R. G.; Boon, L. M.; Borrone, C.; Brunner, H. G.; Carle, G. F.; Dallapiccola, B.; De Paepe, A.; Floege, B.; Halfhide, M. L.; Hall, B.; Hennekam, R. C.; Hirose, T.; Jans, A.; Juppner, H.; Kim, C. A.; Keppler-Noreuil, K.; Kohlschuetter, A.; LaCombe, D.; Lambert, M.; Lemyre, E.; Letteboer, T.; Peltonen, L.; Ramesar, R. S.; Romanengo, M.; Somer, H.; Steichen-Gersdorf, E.; Steinmann, B.; Sullivan, B.; Superti-Furga, A.; Swoboda, W.; van den Boogaard, M. J.; Van Hul, W.; Vikkula, M.; Votruba, M.; Zabel, B.; Garcia, T.; Baron, R.; Olsen, B. R.; Warman, M. L. Cell 2001, 107, 513.
- Babij, P.; Zhao, W.; Small, C.; Kharode, Y.; Yaworsky, P. J.; Bouxsein, M. L.; 6 Reddy, P. S.; Bodine, P. V.; Robinson, J. A.; Bhat, B.; Marzolf, J.; Moran, R. A.; Bex, F. I. Bone Miner. Res. 2003, 18, 960.
- 7 Kato, M.; Patel, M. S.; Levasseur, R.; Lobov, I.; Chang, B. H.; Glass, D. A., 2nd; Hartmann, C.; Li, L.; Hwang, T. H.; Brayton, C. F.; Lang, R. A.; Karsenty, G.; Chan, L. J. Cell Biol. 2002, 157, 303.
- 8 Glantschnig, H.; Hampton, R.; Wei, N.; Scott, K.; Nantermet, P.; Zhao, J.; Chen, F.; Fisher, J.; Su, Q.; Pennypacker, B.; Cusick, T.; Sandhu, P.; Reszka, A.; Strohl, W.; Flores, O.; Wang, F.; Kimmel, D.; An, Z. J. Bone Miner. Res. 2008, 23, S60.
- Li, X.; Ominsky, M. S.; Warmington, K. S.; Morony, S.; Gong, J.; Cao, J.; Gao, Y.; 9. Shalhoub, V.; Tipton, B.; Haldankar, R.; Chen, Q.; Winters, A.; Boone, T.; Geng, Z.: Niu, O. T.: Ke, H. Z.: Kostenuik, P. I.: Simonet, W. S.: Lacev, D. L.: Pasztv, C. J. Bone Miner. Res. 2009, 24, 578.
- 10. Angers, S.: Moon, R. T. Nat. Rev. Mol. Cell Biol. 2009, 10, 468.
- Robinson, J. A.; Chatterjee-Kishore, M.; Yaworsky, P. J.; Cullen, D. M.; Zhao, W.; Li, 11 C.; Kharode, Y.; Sauter, L.; Babij, P.; Brown, E. L.; Hill, A. A.; Akhter, M. P.; Johnson, M. L.; Recker, R. R.; Komm, B. S.; Bex, F. J. J. Biol. Chem. 2006, 281, 31720.
- Armstrong, V. J.; Muzylak, M.; Sunters, A.; Zaman, G.; Saxon, L. K.; Price, J. S.; 12. Lanyon, L. E. J. Biol. Chem. 2007, 282, 20715.
- Sawakami, K.; Robling, A. G.; Ai, M.; Pitner, N. D.; Liu, D.; Warden, S. J.; Li, J.; 13 Maye, P.; Rowe, D. W.; Duncan, R. L.; Warman, M. L.; Turner, C. H. J. Biol. Chem. 2006, 281, 23698.
- All newly prepared compounds were characterized by reversed phases-HPLC/ 14. MS spectroscopy. A selected number of compounds were also characterized by ¹H NMR.
- 15. Harden, D. B.; Mokrosz, M. J.; Strekowski, L. J. Org. Chem. 1988, 53, 4137.

- Strekowski, L.; Harden, M. J.; Grubb, W. B., III; Patterson, S. E.; Czarny, A.; Mokrosz, M. J.; Cegla, M. T.; Wydra, R. L *J. Heterocycl. Chem.* **1990**, *27*, 1393.
 Bhat, B. M.; Allen, K. M.; Liu, W.; Graham, J.; Morales, A.; Anisowicz, A.; Lam, H.
- Bhat, B. M.; Allen, K. M.; Liu, W.; Graham, J.; Morales, A.; Anisowicz, A.; Lam, H. S.; McCauley, C.; Coleburn, V.; Cain, M.; Fortier, E.; Bhat, R. A.; Bex, F. J.; Yaworsky, P. J. *Gene* **2007**, *391*, 103.
- Rewcastle, G. W.; Denny, W. A.; Showalter, H. D. H. *Curr. Org. Chem.* 2000, 4, 679.
 Coghlan, M. P.; Culbert, A. A.; Cross, D. A.; Corcoran, S. L.; Yates, J. W.; Pearce, N. J.; Rausch, O. L.; Murphy, G. J.; Carter, P. S.; Roxbee Cox, L.; Mills, D.; Brown, M.
- J.; Rausch, O. L.; Murphy, G. J.; Carter, P. S.; Roxbee Cox, L.; Mills, D.; Brown, M. J.; Haigh, D.; Ward, R. W.; Smith, D. G.; Murray, K. J.; Reith, A. D.; Holder, J. C. *Chem. Biol.* **2000**, *7*, 793.