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## Selective Fowler Reductions: Asymmetric Total Syntheses of Isofagomine and Other 1-Azasugars from Methyl Nicotinate

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## ABSTRACT



and other 1-azasugars

An efficient, high-yielding strategy has been developed for the asymmetric synthesis of 1-*N*-iminosugars (1-azasugars), a new class of glycosidase inhibitors with promising biomedical applications. A highly regioselective procedure for the 1,2-reduction of substituted pyridines was employed to transform methyl nicotinate into several representative 1-azasugars.

Interest continues to mount in new applications of natural and synthetic glycosidase inhibitors to basic research and medicine.<sup>1</sup> Linkage- and configuration-specific inhibitors may be useful in treating lysozomal storage diseases, diabetes, viral infection, and metastatic cancer. In the past few years, 1-*N*-iminosugars, or 1-azasugars, have emerged as a major new family of monosaccharide mimics worthy of further development. The prototype 1-azasugar **1**, named isofagomine (Figure 1), was first conceived by Bols et al. as an apparent transition state analogue mimicking equatorial glycoside cleavage.<sup>2</sup> Ichikawa et al. found that 1-*N*-iminosugars **2** and **3** inhibited glucuronidase and iduronidase,<sup>3</sup> two enzymes that promote the invasion of basement membrane components during tumor cell metastasis.<sup>4,5</sup> Since then, several other 1-*N*-iminosugars have been studied, including

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hydroxylated isofagomine derivative **4**.<sup>6</sup> Besides being potent, such 1-azasugars are anomer-selective inhibitors. For example, **1** exhibited 780-fold greater potency against  $\beta$ -glu than against the corresponding  $\alpha$ -glu.<sup>7</sup>

Here we report short, enantioselective synthetic routes to iminosugars 1, 2, and 4 in high overall yields from methyl nicotinate 5. Our synthetic strategy relied on a regioselective



Figure 1. Representative examples of 1-azasugars

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and high-yielding reduction of 5, together with efficient and stereoselective functionalizations of the resulting 1,2-dihydropyridine system. This methodology should also facilitate the synthesis of other 1-N-iminosugars having substituents and stereochemistry designed for, and tailored to, specific biological targets.

Several methods have been described for the partial reduction of pyridines using hydride sources.<sup>8</sup> In a detailed study. Sundberg et al. noted that reductions of 5 using combinations of borohydride reagents with ethyl or benzyl chloroformate afforded mixtures of the 1,2-, 1,4-, and 1,6reduction products.<sup>9</sup> No reductions of 5 using phenyl chloroformate were reported, although that reagent had been used earlier to achieve the clean reduction of pyridine to *N*-phenoxycarbonyl-1,2-dihydropyridine.<sup>10</sup> We observed that reduction of 5 with NaBH<sub>4</sub> and phenyl chloroformate afforded 2,3-dihydropyridine 6 in 90% yield (Scheme 1).



Using NaBH<sub>4</sub>-trifluoroacetic acid, 6 could further be reduced to the tetrahydropyridine 7 in virtually quantitative yield, thus providing a superior route to tetrahydropyridines of the arecoline family.<sup>11</sup>

The use of NaBH<sub>4</sub> and phenyl chloroformate proved generally applicable in the selective 1,2-reduction of a range of substituted pyridines, as indicated in Table 1. Reduction of the more sterically hindered tert-butylnicotinate 8 paralleled that of 5. In the case of 9a and 9b, smooth reduction to the corresponding 1,6-dihydropyridines 13 and 14 was observed, with regiocontrol arising from the oxygen substituent. The selective 1,2-reduction of 3-hydroxymethylpyridine 10 afforded 15 in good yield, indicating that the reaction was not limited to nicotinic esters. Acetamidonicotinate 11 was inert to reduction.

The utility of such 1,2-dihydropyridines in assembling 1-azasugars was illustrated in the following synthesis of isofagomine. Reaction of 6 with m-chloroperoxybenzoic acid (MCPBA) afforded hydroxyester 16 regioselectively and in 92% yield (Scheme 2). The regiochemistry of 16 was assigned on the basis of an <sup>1</sup>H NMR decoupling experiment.





Irradiation of the resonance at  $\delta$  5.70 corresponding to H<sub>b</sub> caused the vinylic hydrogen resonance for H<sub>a</sub> at  $\delta$  7.25 to collapse to a singlet. Since the resonances for H<sub>b</sub> and H<sub>c</sub> were broad, the relative stereochemistry in 16 could not be assigned unambiguously. Reduction of 16 with trimethylsilyl



(a) MCPBA, CH2Cl2, -70 to 0 °C, 92%; (b) TMSOTf, BH<sub>3</sub>-THF, -70 to 0 °C, 77%; (c) CrO<sub>3</sub>, acetone; (d) LiAlH<sub>4</sub>, (-)-N-Me-ephedrine, 85%; (e) 1 N HCl, reflux, 99%; (f) BH3-THF, H2O2, NaOAc, 70%; (g) LiOH, 95%.

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triflate (TMSOTf) and borane–THF complex gave rise to allylic alcohol ( $\pm$ )-17.

The asymmetric epoxidation or osmylation of 6 was explored under a variety of conditions. Although these transformations were highly regioselective for the disubstituted alkene in 6, only modest (20–25%) enantioselectivities were achieved. As an alternative, oxidation of racemic 17 using Jones reagent gave the achiral enone 18, which was immediately subjected to asymmetric LiAlH<sub>4</sub> reduction using (-)-N-methylephedrine as the chiral auxiliary<sup>12</sup> to afford optically active alcohol (+)-17 in 85% yield and 83% ee, based on Mosher ester analysis. Hydrolysis of (+)-17 afforded unsaturated hydroxyacid (+)-19 in near-quantitative yield. The hydroboration of (+)-19, following a protocol for other  $\beta$ -substituted  $\alpha,\beta$ -cyclohexenones,<sup>13</sup> was performed using BH<sub>3</sub>-THF (5 molar equiv) followed by oxidative workup, affording *trans, trans*-triol (+)-20 as the exclusive product in 65% yield. Hydrolysis of (+)-20 afforded (+)isofagomine 1 in 95% yield, thus confirming the assigned absolute configuration of (+)-17.<sup>14</sup> Overall, the synthesis of isofagomine required eight steps and gave (+)-1 in 41% yield from methyl nicotinate.

Intermediate (+)-20 also afforded ready access to glucuronidase inhibitor (+)-2 (Scheme 3). Oxidation of (+)-20



with platinum and oxygen furnished carboxylic acid (+)-21 (80%), which was deprotected in base to give (+)-2<sup>14</sup> in a total of nine steps and 33% overall yield from methyl nicotinate.

Allylic alcohol (+)-17 was also useful in synthesizing tetraol (-)-4, a hydroxylated analogue of isofagomine that is also a potent and highly selective  $\beta$ -glucosidase inhibitor.<sup>6</sup>

Osmylation of (+)-17 afforded triol (-)-22 with excellent *anti*-selectivity. Alkaline hydrolysis of the urethane and methyl ester groups in (-)-22 afforded triol-amino acid (-)-23, an hydroxylated analogue of 2 and prospective glucuronidase inhibitor (Scheme 4). All attempts to reduce the carboxyl



group in (–)-23 (or its methyl ester, not shown) proved fruitless. Therefore, to improve its poor solubility in organic solvents, (–)-23 was transformed to the moisture-sensitive persilylated ester 24, which was smoothly reduced to the desired aminotetraol (–)-4 in good yield.<sup>14</sup>

In summary, an efficient and flexible route to 1-azasugars has been developed from a readily available and inexpensive starting material.<sup>15</sup> As part of that strategy, an improved procedure has been devised for the 1,2-reduction of substituted pyridines with high regioselectivity. Taken together, the methodology reported herein should provide access to other new 1-*N*-iminosugars for use as anomer-specific glycosidase inhibitors.

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**Supporting Information Available:** Detailed experimental procedures for 16,  $(\pm)$ -17 18, and (+)-17, along with <sup>1</sup>H and <sup>13</sup>C NMR data for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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<sup>(14)</sup> Spectroscopic, chiroptical, and physical characterization data for this compound matched literature values.

<sup>(15)</sup> **Representative procedure for the Fowler reduction:** To a suspension of methyl nicotinate **5** (10 g, 72.6 mmol) and sodium borohydride (2.89 g, 72.6 mmol) in methanol (200 mL) at -78 °C was added phenyl chloroformate (9.11 mL, 72.6 mmol) in a dropwise manner, over an interval of 40 min. The mixture was stirred for 3 h and then poured onto 800 mL of distilled water to give a yellow precipitate. The precipitate was filtered, washed with distilled water (150 mL), and dried under vacuum to give **6** (18 g, 97%). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  7.42–7.35 (m, 2 H), 7.28–7.21 (m, 1 H), 7.18–7.13 (m, 3 H), 7.02 (d, 1 H, *J* = 3.0 Hz), 5.44–5.37 (m, 1 H), 4.77 (s, 1 H), 4.63 (s, 1 H), 3.77 (s, 3 H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  66.1 and 165.7, 150.9 and 150.8, 132.7, 131.2. 129.8 and 129.7, 126.5–126.4, 121.7 and 121.6, 120.4, 118.5, 105.1 and 105.0, 52.1 and 51.9, 43.4 and 43.0; IR (CH<sub>2</sub>Cl<sub>2</sub>) 2950, 1740, 1720, 1230, 1190 cm<sup>-1</sup>; EIMS *m*/z 259 (M<sup>+</sup>, 87%), 77 (100%).