

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 14 (2004) 2863–2866

Potent DNA gyrase inhibitors; novel 5-vinylpyrazole analogues with Gram-positive antibacterial activity

Akihiko Tanitame,^{a,*} Yoshihiro Oyamada,^b Keiko Ofuji,^a Kenji Suzuki,^a Hideaki Ito,^b Motoji Kawasaki,^a Masaaki Wachi^c and Jun-ichi Yamagishi^b

^aChemistry Research Laboratories, Dainippon Pharmaceutical Co., Ltd, 33-94, Enoki, Suita, Osaka 564-0053, Japan ^bPharmacology and Microbiology Research Laboratories, Dainippon Pharmaceutical Co., Ltd, 33-94, Enoki, Suita, Osaka 564-0053, Japan

^cDepartment of Bioengineering, Tokyo Institute of Technology, 4259, Nagatsuda, Midori-ku, Yokohama 226-8501, Japan

Received 18 February 2004; accepted 15 March 2004

Abstract—In this study, we designed and synthesized novel 5-vinylpyrazole analogues by decreasing the lipophilicity of the parent compounds **1a**,**b**; 3-[(3-methoxycarbonyl)cyclohexylaminomethyl]indazoles while keeping the van der Waals interaction with the lipophilic area of DNA gyrase B. The selected compound **8bb** exhibited good antibacterial activity against staphylococci and enterococci, including multi-drug resistant strains.

© 2004 Elsevier Ltd. All rights reserved.

The increasing use of antibacterial agents such as β -lactams, macrolides, vancomycin or quinolones has resulted in the emergence of multi-drug resistant pathogens, especially Gram-positive bacteria.^{1,2} Over the past decade, DNA gyrase has drawn much attention as selected target for finding potent antibacterial agents against multi-drug resistant strains.³

We have previously found compounds **1a**,**b** (Fig. 1) as potent and selective inhibitors of DNA gyrase with moderate antibacterial activity against multi-drug resistant strains as well as susceptible strains (Table 1).⁴ Docking studies of **1a** with a 43 kDa fragment of DNA gyrase subunit B from *Escherichia* (*E.*) coli revealed that (1) the indazole scaffold forms postulated H-bond network with Asp73 and H₂O, (2) the methyl ester moiety is H-bonded to Arg136, (3) the benzyloxy side chain interacts—van der Waals interaction—with the lipophilic area around Ile94, (4) an additional H-bond is formed between Gly77 and the nitrogen atom on the secondary amine of **1a** (Fig. 2).



Figure 1.

In addition to our studies on compounds **1a**,**b**, we have previously reported a new class of antibacterial agents with potent inhibitory activity against bacterial type II topoisomerase using a new screening system for specific inhibitors of chromosome partitioning in *E. coli*.^{5,6} Among these compounds, the pyrazole analogues **2a**,**b** (Fig. 1) showed potent Gram-positive antibacterial activity (Table 1).⁷

Although compounds **1a**,**b** and **2a**,**b** were active in vitro, they had apparently no in vivo antibacterial activity and

Keywords: DNA gyrase inhibitor; 5-Vinylpyrazole analogues; Multidrug resistant strains.

^{*} Corresponding author. Tel.: +81-6-6337-5906; fax: +81-6-6338-7656; e-mail: akihiko-tanitame@dainippon-pharm.co.jp

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2004.03.045

Table 1. Inhibitory activity against *E. coli* DNA gyrase and topoisomerase IV (IC_{50}) and minimum inhibitory concentration (MIC) of compounds 1a,b and 2a,b

No.		IC ₅₀ (µg/mL)			MIC (µg/mL)			
	Gyrase ^a	Topo IV ^b	Topo II ^c	S. aureus		E. faecalis		
				FDA 209P ^d	KMP9 ^e	ATCC 29212 ^d	KU 1777 ^f	
1a	4	>128	>400	4	4	8	4	
1b	1	>128	>400	4	4	4	4	
2a	14	8.0	>400	1	1	2	2	
2b	14	8.0	>400	1	1	2	2	

(CAM: clarithromycin, ABPC: ampicillin, SPFX: sparfloxacin, VCM: vancomycin).

^a DNA gyrase supercoiling activity.

^b Topoisomerase IV decatenation activity.

^cHuman topoisomerase II relaxation activity.

^d Susceptible strain.

^eSPFX-, CAM-, ABPC-resistant strain.

^fSPFX-, VCM-resistant strain.



Figure 2. Schematic representation of the indazole analogue 1a bound to the 43 kDa fragment of DNA gyrase B from *E. coli*.

showed weak antibacterial activity in the presence of 10% horse blood, indicating that these compounds, due to their lipophilicity, bind to serum proteins.

From the findings above and in order to improve the bioavailability and antibacterial activity of **1a**,**b**, we generated the novel 5-vinylpyrazoles **3** from the 3-[(3-methoxycarbonyl)cyclohexylaminomethyl]indazole derivatives **1a**,**b** and the cited 5-vinylpyrazole analogues **2a**,**b** (Fig. 1). The idea behind our strategy was to decrease the lipophilicity of compounds **1a**,**b** while keeping the van der Waals interaction with the lipophilic area around Ile94 of the DNA gyrase B.

The novel 5-vinylpyrazole analogues were prepared as shown in Scheme 1. Condensation of the aldehydes 4a,b with acetone in the presence of aq NaOH in EtOH,⁸ followed by condensation reaction with $(COOEt)_2$ and NaOEt gave the 1,3-diketones 5a,b. Cyclization of 5a,b with NH₂NH₂ in EtOH, and reduction with LiAlH₄ afforded the 3-hydroxymethylpyrazoles 6a,b. Chlorination of **6a**,**b** with (COCl)₂, followed by reaction with excess 3-(methoxycarbonyl)cyclohexylamine gave the desired products 7a,b. Next, separation of the diastereomeric mixture of 7b using CHP-20P (reverse phase column) furnished the less polar diastereoisomer 7ba and the more polar diastereoisomer **7bb**, respectively.⁹ Moreover, we found out that the (E)-forms 7a, 7ba and **7bb** were easily isomerized to the (Z)-forms **8a**, **8ba** and **8bb**, respectively, by irradiation with visible light in highly diluted MeOH. The stereochemistry of each isomer of 7 and 8 was determined on the basis of NOE experiments. For example, in compound 8a, an enhancement was observed between the two protons of the vinyl moiety. On the other hand, in compound 7a, no enhancement was observed between the two protons of the vinyl moiety.

Next, compounds **7a**, **7ba**, **7bb**, **8a**, **8ba** and **8bb** were tested for inhibition of DNA gyrase and topoisomerase IV,^{10,11} and their MIC values against *Staphylococcus* (*S*.) *aureus* and *Enterococcus* (*E*.) *faecalis* including susceptible and multi-drug resistant strains were determined by a microdilution method (Table 2).



Scheme 1. Synthesis of the novel 5-vinylpyrazole analogues. Reagents and conditions: (a) acetone, aq NaOH, EtOH; (b) (COOEt)₂, NaOEt, Et₂O; (c) NH₂NH₂, EtOH; (d) LiAlH₄, THF (e) (COCl)₂, CH₂Cl₂; (f) 3-methoxycarbonyl-cyclohexylamine (mixture of two stereoisomers), CHCl₃–MeOH; (g) *hv*, MeOH.

MeOOC											
No.		R ³	IC ₅₀ (μg/mL) Gyrase	MIC (µg/mL)							
				S. aureus		E. faecalis					
				FDA 209P	KMP9	ATCC 29212	KU 1777				
7a	Ε	CI	16	4	8	16	16				
8a	Ζ	CI	16	4	8	16	16				
		CI									
7ba	E		1	8	8	32	32				
7bb	E	$\gamma > 1$	1	16	16	64	32				
8ba	Z	Ύ-N	1	4	8	8	8				
8bb	Z	Me	0.5	8	16	64	32				

Table 2. Inhibitory activity against E. coli DNA gyrase (IC₅₀) and minimum inhibitory concentration (MIC) of the novel 5-vinylpyrazole analogues

NI_NILI

Compounds **7a** and **8a** showed less potent DNA gyrase inhibitory activity and slightly less potent antibacterial activity against four Gram-positive strains than the parent compound **1b** (Table 2). These results suggest that the 2-[3,4-(dichlorophenyl)]vinyl moiety renders good permeability through bacterial cell membrane. Since **7a** and **8a** inhibited DNA gyrase with the same IC_{50} value, it was assumed that both compounds bind to DNA gyrase in a similar manner. In addition, both compounds **7a** and **8a** showed the same MIC value against four Gram-positive strains.

Compounds 7ba, 7bb, 8ba and 8bb exhibited the same or 2-fold more potent inhibitory activity against DNA gyrase compared with the parent compound 1b, although their antibacterial activity was less potent than that of 1b. Among these compounds, 8bb showed the most potent inhibitory activity against DNA gyrase. In addition, 8ba and 8bb showed slightly more potent antibacterial activity against four Gram-positive strains than 7ba and 7bb, respectively. Finally, the less polar diastereoisomers 7ba and 8ba exhibited slightly more potent antibacterial activity than the more polar diastereoisomers 7bb and 8bb, respectively (Table 2).

In this study, all the novel 5-vinylpyrazole analogues had moderate to strong inhibitory activity against DNA gyrase, but not against topoisomerase IV^{11} (IC₅₀s > 128 µg/mL) and human topoisomerase II¹² inhibitory activity (IC₅₀s > 400 µg/mL). In addition, all compounds exhibited antibacterial activity against both susceptible and multi-drug resistant strains.

Compound **8bb** exhibited 8-fold more potent inhibitory activity against DNA gyrase than **1a**. On the other hand, we previously reported that the 3-[(3-methoxycarbonyl)cyclohexylaminomethyl]indazole portion of **1a** has three H-bond interactions with the 43 kDa fragment of DNA gyrase B on ATP binding site (Fig. 2).⁴ Taken together, these results suggest that **8bb** interacts with the 43 kDa fragment of DNA gyrase B on ATP binding site as does **1a**. In summary, we have designed and synthesized novel 5-vinylpyrazole analogues by decreasing the lipophilicity of the parent compounds **1a**,**b** while keeping the van der Waals interaction with the lipophilic area around Ile94 of the DNA gyrase B. The selected compound 8bb (clogP = 4.630) was less lipophilic than 1a (clogP =6.445) or **1b** (clogP = 5.925),¹³ and showed more potent inhibitory activity against DNA gyrase than the parent compounds. Although 8bb showed weaker antibacterial activity against four Gram-positive strains than the parent compounds **1a**,**b**, it exhibited relatively good antibacterial activity against staphylococci and enterococci, including multi-drug resistant strains. These results provide a good strategy for the development of Gram-positive antibacterial agents targeting DNA gyrase.

Acknowledgements

We are grateful to Dr. K. Nagai, Dr. K. Chiba, Dr. S. Kato, Dr. H. Yoshida and Dr. H. Terauchi for their encouragement throughout this work.

References and notes

- 1. Witte, W. J. Antimicrob. Agents Chemother. 1999, 14, 1.
- 2. Marchese, A.; Schito, G. C.; Debbia, E. A. J. Chemother. 2000, 12, 12.
- Ferrero, L.; Cameron, B.; Manse, B.; Lagneaux, D.; Crouzet, J.; Famechon, A.; Blanche, F. Mol. Microbiol. 1994, 13, 641.
- Tanitame, A.; Oyamada, Y.; Ofuji, K.; Kyoya, Y.; Suzuki, K.; Ito, H.; Kawasaki, M.; Nagai, K.; Wachi, M.; Yamagishi, J. *Bioorg. Med. Chem. Lett.* 2004, 14, preceding paper in this issue. doi:10.1016/j.bmcl.2004.03.044.
- Wachi, M.; Iwai, N.; Kunihisa, A.; Nagai, K. Biochimie. 1999, 81, 909.
- Hiraga, S.; Niki, H.; Ogura, T.; Ichinose, C.; Mori, H.; Ezaki, B.; Jaffe, A. J. Bacteriol. 1989, 171, 1496.
- 7. J. Med. Chem., in press.

- Edwards, M. L.; Ritter, H. W.; Stemerich, D. M.; Stewart, K. T. J. Med. Chem. 1983, 26, 431.
- 9. Compounds 7a and 8a were an inseparable mixture of diastereomers.
- Sato, K.; Inoue, Y.; Fujii, T.; Aoyama, H.; Inoue, M.; Mitsuhashi, S. Antimicrob. Agents Chemother. 1986, 30, 777.
- 11. Peng, H.; Marians, K. J. J. Biol. Chem. 1993, 268, 24481.
- 12. Spitzner, J. R.; Chung, I. K.; Muller, M. T. Nucleic Acids Res. 1990, 18, 1.
- The clogPs of these compounds were calculated by cLogP for Windows Version 4.0 (Biobyte Corp., Claremont, CA, U.S.A).