Tetrahedron Letters 52 (2011) 2443-2445

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet



Total synthesis of (–)-cleistenolide

Palakuri Ramesh, H. M. Meshram*

Organic Chemistry Division-I, Indian Institute of Chemical Technology, Hyderabad 500 007, India

ARTICLE INFO

ABSTRACT

Article history: Received 16 October 2010 Revised 24 January 2011 Accepted 27 January 2011 Available online 1 February 2011 The total synthesis of (–)-cleistenolide, a novel natural product recently isolated from the annonaceae species cleistochlamys kirkii oliver, is described. The synthesis proceeds starting from easily accessible p-manitol using a selective benzoylation, a selective acetonide deprotection, silylprotection and ring-closing metathesis reaction.

© 2011 Elsevier Ltd. All rights reserved.

Naturally isolated 6-substituted- α , β -unsaturated- δ -lactones gained great attention of researchers due to their cytotoxic and anti-tumor properties.¹ In addition they inhibit HIV protease,² induce apoptosis,^{3,4} and have proven to be anti-leukemic,⁵ along with having many other relevant pharmacological properties.⁶ Synargentolide A,⁷ spicegerolide,⁸ hyptolide,⁹ synrotolide,¹⁰ and anamarine¹¹ isolated from syncolostemon and hyptis species are examples of α , β -unsaturated- δ -lactones. Due to their pharmacological properties, these molecules became the interesting synthetic goals.

Cleistochlamys kirkii oliver is an annonaceae species occurring in Tanzania and Mozambique.¹² Extracts of this plant are used in traditional medicine as a remedy for the treatment of wound infections, rheumatism, and tuberculosis.¹³ Recently, Nkunya coworkers¹⁴ investigated the chemical composition of organic extracts from fruits, leaves, stem and root barks of C. kirkii oliver and discovered two novel plant constituents, which were named cleistenolide (**1**) and cleistodienol (**2**) (Fig. 1).¹⁴ cleistenolide is known to possess antibacterial activity against *staphylococcus aureus* and *bacillus anthracis*, as well as antifungal activity against *Candida albicans*.

Recently, the first total synthesis of cleistenolide was published by Schmidt et al.¹⁵ in overall 18% yield, by applying a ring-closing metathesis (RCM) protocol to prepare the key building block, an α , β -unsaturated lactone. Very recently synthesis of cleistenolide was reported by Cai et al. from the α , β -unsaturated acid through an intramolecular Yamaguchi esterification.¹⁶

Herein, we report the total synthesis of cleistenolide (1), starting from an enantiomerically pure p-manitol. Our synthesis confirms the absolute configuration assigned (–)-cleistenolide. As depicted in the retro synthetic analysis of the (–)-cleistenolide the crucial dihydropyran-2-one was prepared from ring-closing metathesis (RCM). Compound **4** would be prepared from benzoylated diacetonide **5**. Precursor **5** would be constructed from natural chiral template *D*-manitol **6** through the regioselective deprotection of acetonide, and monobenzoylation (Scheme 1).

The present synthesis commences (Scheme 2) from D-(+)-mannitol. The diol **7** is readily accessible from tri-*O*-isopropylidene-D-(+)-manitol,¹⁸ with the terminal acetonides acting as



Figure 1. Chemical structures of cleistenolide (1) and cleistodienol (2).



Scheme 1. Retrosynthesis of (-)-cleistenolide.



^{*} Corresponding author. Tel.: +91 40 27160123 2642; fax: +91 40 27160512. *E-mail address:* hmmeshram@yahoo.com (H.M. Meshram).

^{0040-4039/\$ -} see front matter \odot 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2011.01.124



Scheme 2. Reagents and conditions: (a) Pyridine, benzoyl chloride, DMAP, CH₂Cl₂, -78-20 °C, 4 h, 83%; (b) TBDPS-Cl, imidazole, CH₂Cl₂, 0 °C to rt, 86%; (c) CuCl₂·2H₂O, CH₃CN, 0 °C, 45 min, 99% (with recovered starting material); (d) PPh₃-imidazole-iodine, toluene, 110 °C, 4 h, 84%; (e) PPTS (cat)/MeOH, rt, 86% (or) CuCl₂·2H₂O, CH₃CN, rt, 20 h, 84%; (f) TBS-OTf (1 equiv), 2,6-lutidine, CH₂Cl₂, -78 °C, 85%; (g) acryloyl chloride, DIPEA, CH₂Cl₂, 0 °C to rt, 81%; (h) 5 mol % Grubb's 2nd generation catalyst, toluene, 110 °C, 68%; (i) TBAF, THF then Ac₂O, 62%.

surrogates for the generation of olefin. The formation of the terminal diol **9** (Scheme 2) from the diacetonide **8** was achieved by the slight modification of a literature method.^{18,19}

Monobenzoylation of diol **7** by using benzoyl chloride and DMAP in pyridine gave **5** in good yield.¹⁹ The hydroxy group of compound **5** was protected with TBDPS–Cl to give silyl ether **8** with 86% yields. The diacetonide **8** underwent selective hydrolysis of the terminal acetonide using an equivalent amount of CuCl₂·2H₂O at 0 °C to generate the diol **9** in quantitative yield.²⁰ Pb(OAc)₄-cleavage of **9** followed by Wittig reaction provided the terminal olefin **4** in poor yield along with an unidentified mixture of products. However, it was smoothly converted **4** by the reaction with Ph₃P-imidazole-iodine in toluene and gave 84% yield.²¹ Thus, acetonide cleavage of **4** by PPTS/MeOH (or) CuCl₂·2H₂O at room temperature produced the diol **10** (Scheme 2).¹⁹



2nd generation Grubb's catalyst

The key building block **10** is thus synthesized in a facile manner from D-(+)-mannitol in a combined 44% overall yield. The reaction of compound **10** on treatment with 1 equiv. of TBS–OTf, 2,6-luti-

dine, DCM, -78 °C selectively gave allylic silylether 11, in 85% yield. Esterification of 11 with acryloyl chloride led to the formation of 3 in 81% yield. To avoid isomerization of the double bond by prolonged exposure to the reaction conditions, the reaction was arrested before complete disappearance of the starting material. Ring-closing metathesis of 3 proceeded well with 5 mol % of Grubbs catalyst. In dilute reaction conditions, the six-membered ring δ-lactone **12** was isolated in 68% yield (Scheme 2).^{17,20b} Completion of the synthesis required desilylation of 12 and acetylation of the two secondary alcohols. This was most conveniently achieved as a one-flask reaction in THF by the addition of TBAF and subsequent addition of acetic anhydride. Notably, the desilvlation/double acetylation proceeds in the absence of base and without any migration of the benzoyl group. Thus, analytically pure (-)-cleistenolide (1) was obtained as a single regio and stereoisomer in a yield of 62%. The physical and spectral data²² of synthesised sample (–)-cleistenolide **1** were identical to those reported in the literature.^{14,16} Our results, as well as Schmidt's report,¹⁵ support the absolute configuration assigned to natural product as (-)-cleistenolide.

In summary, we have achieved the total syntheses of (–)cleistenolide from a common precursor simply by changing the sequence of carbinol protection and thus allowing the formation of δ -lactone. Key features of present route to (–)-cleistenolide are the efficient utilization of the C_2 -symmetry of the starting material (D-(+)-mannitol), a selective benzoylation, a selective acetonide deprotection, selective silyl protection and a ring-closing metathesis reaction.

Acknowledgments

P.R. thanks CSIR for the award of a fellowship and to Dr. I. S. Yaday, Director IICT, for his support and encouragement.

References and notes

- 1. Hoffmann, H. M. R.; Rabe, J. Angew. Chem., Int. Ed. Engl. 1985, 24, 94.
- (a) Romines, K. R.; Chrusciel, R. A. Curr. Med. Chem. 1995, 2, 825; (b) Aristoff, P. 2. A. Drugs Future 1998, 23, 995; (c) Hagen, S. E.; Vara-Prasad, J. V. N.; Tait, B. D. Adv. Med. Chem. 2000, 5, 159; (d) Hagen, S. E.; Domagala, J. M.; Gajda, C.; Lovdahl, M.; Tait, B. D.; Wise, E.; Holler, T.; Hupe, D.; Nouhan, C.; Urumov, A.; Zeikus, G.; Zeikus, E.; Lunney, E. A.; Pavlovsky, A.; Gracheck, S. J.; Saunders, J. M.; Vander, R. S.; Brodfuehrer, J. J. Med. Chem. 2001, 44, 2319; (e) Agrawal, V. K.; Singh, J.; Mishra, K. C.; Khadikar, P. V.; Jaliwala, Y. A. ARKIVOC 2006, 162.
- (a) Inayat, H. S. H.; Annuar, B. O.; Din, L. B.; Taniguchi, N. Toxicol. Lett. 2002, 131, 153; (b) Inayat, H. S. H.; Annuar, B. O.; Din, L. B.; Ali, A. M.; Ross, D. Toxicol. In Vitro 2003, 17, 433; (c) Chan, K. M.; Rajab, N. F.; Ishak, M. H. A.; Ali, A. M.; Yusoff, K.; Din, L. B.; Inayat, H. S. H. Chem. Biol. Interact. 2006, 159, 129.
- 4. For further literature related to this important biological property, see, for example: (a) Blatt, N. B.; Glick, G. D. Bioorg. Med. Chem. 2001, 9, 1371; (b) Huang, Z. W. Chem. Biol. 2002, 9, 1059.
- Kikuchi, H.; Sasaki, K.; Sekiya, J.; Maeda, Y.; Amagai, A.; Kubohara, Y.; Ohsima, Y. Bioorg. Med. Chem. **2004**, 12, 3203.
- See, for example: (a) Stampwala, S. S.; Bunge, R. H.; Hurley, T. R.; Willmer, N. E.; Brankiewicz, A. J.; Steinman, C. E.; Smitka, T. A.; French, J. C. J. Antibiot. **1983**, 36, 1601; (b) Nagashima, H.; Nakamura, K.; Goto, T. Biochem. Biophys. Res. Commun. 2001, 287, 829; (c) Raoelison, G. E.; Terreaux, C.; Queiroz, E. F.; Zsila, F.; Simonyi, M.; Antus, S.; Randriantsoa, A.; Hostettmann, K. *Helv. Chim. Acta* 2001, 84, 3470; (d) Lewy, D. S.; Gauss, C. M.; Soenen, D. R.; Boger, D. L. Curr. Med. Chem. 2002, 9, 2005; (e) Larsen, A. K.; Escargueil, A. E.; Skladanowski, A. Pharmacol. Ther. 2003, 99, 167; (f) Richetti, A.; Cavallaro, A.; Ainis, T.; Fimiani, V. Immunopharmacol. Immunotoxicol. 2003, 25, 441; (g) Koizumi, F.; Ishiguro, H.; Ando, K.; Kondo, H.; Yoshida, M.; Matsuda, Y.; Nakanishi, S. J. Antibiot. 2003, 56, 603
- Collett, L. A.; Davies, C. M. T.; Rivett, D. E. A. Phytochemistry 1998, 48, 651. 7
- Pereda, M. R.; Fragoso, S. M.; Cerda, G. R. C. M. Tetrahedron 2001, 57, 47. 8
- Achmad, S. A.; Hoyer, T.; Kjaer, A.; Makmur, L.; Norrestam, R. Acta Chem. Scand. 9. 1987. 41B. 599.
- Coleman, M. T. D.; English, R. B.; Rivett, D. E. A. Phytochemistry 1987, 26, 1497. 10 Alemany, A.; Marquez, C.; Pascual, C.; Valverde, S.; Martinez, R. M.; Fayos, J.; 11. Perales, A. Tetrahedron Lett. 1979, 20, 3583.
- Nkunya, M. H. H. Pure Appl. Chem. 2005, 77, 1943. 12
- Verzar, R.; Petri, G. J. Ethnopharm. 1987, 19, 67. 13.
- Samwel, S.; Mdachi, S. J. M.; Nkunya, M. H. H.; Irungu, B. N.; Moshi, M. J.; 14
- Moulton, B.; Luisi, B. S. Nat. Prod. Commun. 2007, 2, 737.
- Schmidt, B.; Kunz, O.; Biernat, A. J. Org. Chem. 2010, 75, 2389. 15.
- Chao, C.; Jun, L.; Yuguo, D.; Robert, J. L. J. Org. Chem. 2010, 75, 5754.
 Feurstner, A.; Thiel, O. R.; Ackermann, L.; Schanz, H. J.; Nolan, S. P. J. Org. Chem. 16
- 17. **2000**, 65, 220; (b) Samojlowicz, C.; Bieniek, M.; Zarecki, A.; Kadyrov, R.; Grela, K. *Chem. Commun.* **2008**, 6282; (c) Rost, D.; Porta, M.; Gessler, S.; Blechert, S. Tetrahedron Lett. 2008, 49, 5968.
- 18 Wiggins, L. F. J. Chem. Soc. 1946, 13.
- Chandrasekhar, M.; Kusum, L. C.; Singh, V. K. J. Org. Chem. 2003, 68, 4039. 19
- (a) Saravanan, P.; Chandrasekhar, M.; Anand, R. V.; Singh, V. K. Tetrahedron Lett. 20. 1998, 39, 3091; (b) Yadav, V. K.; Agrawal, D. Chem. Commun. 2007, 5232. 21
- Garegg, P. J.; Samuelsson, B. Synthesis 1979, 469.
- Spectral data for selected compounds: (R)-2-(tert-butyldiphenylsilyloxy)-2-22. ((4S,5R)-2,2-dimethyl-5-vinyl-1,3-dioxolan-4-yl)ethylbenzoate (4): $[\alpha]_D^{DS}$ -2.46 (*c* 1.4, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 1.05 (s, 9H), 1.36 (s, 3H),

1.39 (s, 3H), 3.86 (dd, 1H, J = 4.53, 3.39 Hz), 4.15 (dd, 1H, J = 6.79, 3.23 Hz), 4.22 (dd, 1H, J = 6.04, 5.66 Hz), 4.35 (dd, 1H, J = 7.93, 3.77 Hz), 4.45 (t, 1H, J = 7.36 Hz), 5.10 (d, 1H, J = 10.38 Hz), 5.30 (d, 1H, J = 17.18 Hz), 5.65 (ddd, 1H, J = 16.9, 10.57, 6.79 Hz), 7.30–7.50 (m, 8H), 7.62–7.78 (m, 7H); ¹³C NMR (75 MHz, CDCl₃); δ 19.2, 26.4, 26.6, 66.2, 69.8, 78.3, 79.5, 111.5, 119.2, 128.6, 129.4, 129.6, 130.0, 130.2, 134.2, 134.6, 165.9; IR (KBr): 3068, 2929, 2857, 1592, 1428, 1260, 1108 cm⁻¹; ESI-MS: *m*/*z* 553 [M⁺+Na]; ESI-HRMS: *m*/*z* calcd for C32H38O5SiNa: 553.2386, found: 553.2382.

 $\begin{array}{l} (2R_{3}S_{4}R_{1})-2-(tert-Butyldiphenylsilyloxy)-3,4-dihydroxyhex-5-enyl \ benzoate \ (10): \\ [\alpha]_{D}^{25}-4.24 \ (c \ 0.96, \ CHCl_{3}); \ ^{1}H \ NMR \ (300 \ MHz, \ CDCl_{3}): \ \delta \ 1.05 \ (s, \ 9H), \ 2.22 \ (s, \ 1H), \ 2.65 \ (s, \ 1H), \ 3.58 \ (dd, \ 1H, \ J=8.80, \ 5.62, \ 2.80 \ Hz), \ 4.12 \ (dd, \ 1H, \ J=9.44, \ 1H, \ J=9.44, \ 1H, \ J=9.44, \ J=9.44,$ 3.68 Hz), 4.32 (dd, 1H, J = 9.06, 3.02 Hz), 4.42 (dd, 2H, J = 7.36, 4.53 Hz), 5.18 (d, 1H, J = 10.57 Hz), 5.35 (d, 1H, J = 17.18 Hz), 5.80 (ddd, 1H, J = 16.9, 11.57, 6.42 Hz), 7.20–7.40 (m, 8H), 7.50 (t, 1H, *J* = 7.55 Hz), 7.70 (d, 4H, *J* = 6.61 Hz), 7.83 (d, 2H, *J* = 7.17 Hz); ¹³C NMR (75 MHz, CDCl₃): δ 19.5, 26.7, 66.2, 71.4, 71.6, 78.8, 115.6, 128.4, 129.5, 130.0, 130.8, 133.4, 134.0, 138.6, 165.8; IR (KBr): 3068, 2929, 2857, 1592, 1428, 1260, 1108 cm⁻¹; ESI-MS: *m/z* 513 [M⁺+Na]; ESI-HRMS: m/z calcd for C₂₉H₃₄O₅SiNa: 513.2073, found: 513.2069.

(2R,3S,4R)-4-(tert-Butyldimethylsilyloxy)-2-(tert-butyldiphenylsilyloxy)-3-hydro-xyhex-5-enyl benzoate (**11**): $[x]_{25}^{25}$ +26.4 (*c* 2.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 0.18 (s, 6H), 1.03 (s, 9H), 1.20 (s, 9H), 2.75 (d, 1H, *J* = 4.91 Hz), 3.68 (dd, 1H, J = 4.91, 4.70 Hz), 4.20 (dd, 1H, J = 6.40, 3.20 Hz), 4.45 (dd, 2H, J = 11.52, 5.09 Hz), 4.60 (dd, 1H, J = 9.25, 2.62 Hz), 5.20 (d, 1H, J = 10.57 Hz), 5.35 (d, 1H, J = 17.34 Hz), 5.80 (ddd, 1H, J = 17.56, 9.63, 7.36 Hz), 7.40–7.60 (m, 9H), 7.70 (t, 1H, J = 7.55 Hz), 7.82–7.95 (m, 5H); ¹³C NMR (75 MHz, CDCl₃): δ –5.3, –4.9, 18.6, 19.4, 25.9, 26.6, 66.2, 71.5, 71.9, 79.8, 115.7, 128.6, 129.4, 129.9, 130.08, 130.1, 134.0, 137.4, 165.9; IR (KBr): 3442, 2935, 2930, 1728, 1518, 1418, 1398, 1246, 1156, 836 cm⁻¹; ESI-MS: *m/z* 627 [M⁺+Na]; ESI-HRMS: *m/z* calcd for C35H48O5Si2Na: 627.2432, found: 627.2435.

(2R,3S,4R)-3-(Acryloyloxy)-4-(tert-butyldimethylsilyloxy)-2-(tertbutyldiphenylsil-4.62 (ddd, 2H, J = 11.33, 6.79, 4.53 Hz), 4.81 (dd, 1H, J = 8.30, 3.77 Hz), 5.43 (d, 1H, J = 9.80 Hz), 5.48 (dd, 1H, J = 6.04, 3.02 Hz), 5.58 (dd, 1H, J = 17.34, 3.77 Hz), 5.90 (ddd, 1H, J = 17.34, 9.82, 6.70 Hz), 6.20 (dd, 1H, J = 9.06, 3.02 Hz), 6.45 (dd, (t, 2H, J = 7.55 Hz), 7.70 (t, 2H, J = 7.55 Hz), 7.85 (m, 3H); ¹³C NMR (75 MHz, $CDCl_3$): $\delta = 4.7, -5.2, 18.2, 19.5, 25.7, 26.8, 65.7, 69.0, 72.9, 73.3, 117.6, 127.8,$ 128.3, 129.6, 129.7, 129.8, 130.0, 131.9, 133.2, 134.1, 135.3, 164.9, 166.5; IR (KBr): 2949, 2895, 2854, 1727, 1535, 1423, 1240, 1198, 1154 cm⁻¹; ESI-MS: m/ z 681 [M⁺+Na]; ESI-HRMS: *m*/z calcd for C₃₈H₅₀O₆Si₂Na: 681.3043, found: 681.3047.

(R)-2-((2S,3R)-3-(tert-Butyldimethylsilyloxy)-6-oxo-3,6-dihydro-2H-pyran-2-yl)-2-(tert-butyldiphenylsilyloxy)ethyl benzoate (**12**): $[\alpha]_D^{25}$ –156 (c 0.5, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 0.2 (s, 6H), 1.10 (s, 9H), 1.23 (s, 9H), 4.26 (dd, 1H, J = 9.77, 2.32 Hz), 4.29 (m, 1H), 4.47 (dd, 1H, J = 5.86, 2.22 Hz), 4.54 (dd, 1H, *I* = 12.56, 4.57 Hz), 4.86 (dd, 1H, *I* = 12.34, 2.32 Hz), 6.10 (d, 1H, *I* = 10.57 Hz), 6.93 (dd, 1H, J = 9.77, 5.80 Hz), 7.38–7.53 (m, 8H), 7.62 (t, 2H, J = 7.48 Hz), 7.73 (t, 1H, J = 7.48 Hz), 7.83–7.88 (m, 4H); ¹³C NMR (75 MHz, CDCl₃): δ –5.2, –4.6, 18.0, 19.5, 25.6, 60.0, 66.0, 67.8, 80.0, 122.5, 128.4, 129.4, 129.6, 129.8, 133.3, 144.4, 162.8, 166.9; IR (KBr): 2939, 2855, 1717, 1505, 1423, 1248, 1148, 1054 cm⁻¹; ESI-MS: *m*/*z* 631 [M⁺+H]]; ESI-HRMS: *m*/*z* calcd for C₃₆H₄₇O₆Si₂: 631.2906, found: 631.2909.

J = 6.02, 2.50 Hz), 5.51 (ddd, 1H, J = 9.50, 4.42, 2.32 Hz), 6.29 (d, 1H, J = 9.60 Hz), 7.00 (dd, 1H, *J* = 9.60, 6.10 Hz), 7.45 (t, 2H, *J* = 7.65 Hz), 7.57 (t, 1H, *J* = 7.42 Hz), 8.01 (d, 2H, J = 7.65 Hz); ¹³C NMR (75 MHz, CDCl₃): δ 20.5, 20.7, 59.7, 62.0, 67.7, 75.4, 125.3, 128.5, 129.6, 129.7, 129.7, 133.2, 139.7, 161.0, 166.0, 169.5, 169.9; IR (KBr): 2954, 1723, 1456, 1367, 1218, 1107, 1070; ESI-MS: *m*/*z* 385 [M⁺+Na]; ESI-HRMS: *m*/*z* calcd for C₁₈H₁₈O₈Na: 385.0894, found: 385.0897.