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Neuraminic Acid and Related Compounds. I. Syntheses of Biologically Active 4',7',8',9'-Tetra-O-acetyl-sialyl- and Sialyl-(α2—6)-D-glucosamine-4-phosphate Analogues of Lipid A

CHIKAKO SHIMIZU, KIYOSHI IKEDA, and KAZUO ACHIWA*

School of Pharmaceutical Sciences, University of Shizuoka, Oshika 2-2-1, Shizuoka 422, Japan

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The syntheses of novel tetraacetylsialyl- and sialyl- $(\alpha 2-6)$ -D-glucosamine-4-phosphate analogues of lipid A containing sialic acid in place of 3-deoxy-D-manno-2-octulosonic acid (KDO) are described. Preliminary examination of the biological activity revealed that two synthetic disaccharides possessed weak mitogenic activities.

Keywords—sialic acid; sialyl-glucosamine-4-phosphate; lipid A analog; mitogenic activity

Lipopolysaccharide (LPS) of gram-negative bacteria possesses a variety of biological activities and is composed of three structural regions; an inner part, a polysaccharide region and a lipid part called lipid A.¹⁾ 3-Deoxy-D-manno-2-octulosonic acid (KDO) as a ketosidic component of the polysaccharide part is attached to lipid A, a biologically active fragment of LPS. Recently, two research groups reported the structure of the KDO region of LPS from Salmonella minnesota²⁾ and Escherichia coli,³⁾ in which the KDO group was attached to lipid A with a $(\alpha 2-6)$ linkage.

It was also found by us and the other groups that the nonreducing sugar moiety of lipid A is more important than the other parts (lipid X and lipid Y) for expressing the biological activities of LPS.⁴⁾ Furthermore, tetraacetyl-KDO-⁵⁾ and KDO⁶⁾ (α 2—6)-D-glucosamine-4-phosphate analogues (1 and 2 in Chart 1) possess mitogenic activity comparable to that of lipid A.

Chart 1

We report herein the syntheses of two new compounds, tetraacetylsialyl- and sialyl-($\alpha 2$ —6)-D-glucosamine-4-phosphate analogues (3 and 4 in Chart 1) of lipid A containing sialic acid, whose structure is similar to KDO and whose function has recently attracted interest.⁷⁾

First, we describe a synthetic sequence for the tetraacetyl-sialyl glucosamine-4-phosphate derivative (3). (Chart 2) Esterification of N-acetylneuraminic acid (5) with phenyldiazomethane in methanol at room temperature afforded the benzyl ester (6) in 76.9% yield. The hydroxyl groups of 6 were acetylated with acetic acid, pyridine, and 4-dimethylaminopyridine (DMAP) at room temperature for 20 h to give 7 in 73.8% yield. Treatment of 7 with acetyl chloride saturated with dry hydrogen chloride at room temperature for 24 h gave the 2-chloro compound (8) in nearly quantitative yield. The unstable chloride (8) was used for the subsequent glycosylation without further purification. In our previous investigations, 4e,f) 2-deoxy-4-O-phosphono-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyll-p-glucose showed some distinct biological activities, such as antitumor activity. Therefore, we used benzyl 2-deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O- $[(R)-3-tetradecanoyloxytetradecanoyl]-\beta-D-glucopyranoside (9) carrying the tetradecanoyloxyt$ oxytetradecanoyl group at both N-2 and O-3 of the glucosamine residue, as the reducing part. The 6-OH group of 9 should be much more reactive than the 4-OH group and is therefore suitable as a substrate for the reaction with the halide (8). Glycosylation of the glycosyl acceptor (9), easily prepared from benzyl 2-amino-2-deoxy-4,6-O-isopropylidene-β-Dglucopyranoside in 2 steps, 4c) with the glycosyl donor (8) in the presence of Hg(CN)₂, HgBr₂, and Molecular Sieves 4A in CH₂Cl₂ at room temperature for 4d gave the desired (2—6)linked disaccharide containing sialic acid (10) in 34.0% yield as a mixture of anomers. Separation by preparative thin layer chromatography (TLC (CHCl₃-MeOH = 20:1)) afforded 10α and 10β in yields of 23.4 and 10.6%, respectively. The by-product of the reaction was mainly the 2,3-dehydrosialic acid derivative. The presence of an α -glycosidic linkage in 10α was indicated by the proton nuclear magnetic resonance (1H-NMR) spectrum, which showed signals for the equatorial proton H-3_{eq} of sialic acid at 2.69 ppm (ranges for α -linked sialyl derivatives: H-3_{eq} at 2.6—2.8 ppm).⁸⁾ In the case of 10β , the H-3_{eq} signals overlapped with the methylene proton signals (2.22—2.67 ppm) of fatty acids in the ¹H-NMR spectrum.⁹ Therefore, the confirmation of the β linkage of 10β was based on the ¹H-NMR spectral data (the H-3_{eq} signals of 10β were not observed downfield from 2.67 ppm). In addition, signals at 99.7 ppm (singlet) of 10α and 98.5 ppm (singlet) of 10β in the carbon-13 nuclear magnetic resonance (13 C-NMR) spectrum were assigned to the α - and β -ketosidic carbon atom, respectively. 10) The structure of 10α was confirmed by the ¹H-NMR spectrum; the signals at 1.25 ppm were assigned to the methylene protons of fatty acids, and the signals at 1.84— 2.22 ppm to the acetyl group protons. The 4-hydroxyl group of 10α was phosphorylated with diphenylphosphorochloridate, pyridine, and DMAP in CH₂Cl₂ to give the phosphate (11) in 57% yield. The infrared (IR) spectrum of 11 showed a characteristic band of >P-O-Ph at 955 cm⁻¹. The protective benzyl and phenyl groups were cleaved stepwise by hydrogenation in the presence of palladium-carbon and then of platinum oxide as catalysts in methanol solution to yield the tetraacetylsialyl glucosamine-4-phosphate derivative (3) in 23% yield as the free acid form. Compound 3 gave a positive test with the specific spray-reagent for phosphate.11)

Next, for the synthesis of the sially glucosamine-4-phosphate derivative (4), we used the monochloroacetyl group in place of the acetyl group as a protecting group for the hydroxyl groups of sialic acid. The monochloroacetyl group can be removed selectively in the presence of other ester functions. The synthesis of 4 was carried out by a method similar to that used for 3, as shown in Chart 2.

The hydroxyl groups of the benzyl ester (6) were acylated with monochloroacetic anhydride, pyridine, and DMAP to give the monochloroacetyl compound (12) in 77.7% yield.

 $R: CH_3CO^-$ or $CICH_2CO^-, \ R': \ CH_3(CH_2)_{10}CHCH_2CO^ CH_3(CH_2)_{12}CO$ O

Chart 2

The anomeric monochloroacetyl group of 12 was quantitatively converted into the 2-chloro derivative (13) as described for the preparation of 8. Condensation of the two components, 9 and 13 in the presence of Hg(CN)₂, HgBr₂, and Molecular Sieves 4A in CH₂Cl₂ at room temperature for 2 d gave the expected ($\alpha 2$ —6) sially disaccharide (14) in poor yield (4.3%), as a single anomer. The ¹H-NMR spectrum of 14 showed signals of H-3_{eq} at 2.65 ppm, suggestive of the α-anomeric configuration. The low yield can be accounted for by the lack of reactivity of 13, probably due to the steric hindrance of monochloroacetyl groups. The stereoselective formation of the thermodynamically more stable α -anomer (14 α) in glycosylation may be attributable to the low reactivity of 13. Phosphorylation of the 4-hydroxyl group of 14 was effected with diphenylphosphorochloridate, pyridine, and DMAP to give the 4-phosphate (15). The monochloroacetyl groups of 15 were selectively removed by diisopropylethylamine and thiourea to afford 16 in 43% yield. The ¹H-NMR spectrum of 16 showed that 16 has no chloroacetyl groups. Finally, deprotection of 16 as described for 11 gave the $(\alpha 2-6)$ sialyl glucosamine-4-phosphate derivative (4) in 56% yield. The structure of 4 was assigned on the basis of the positive fast atom bombardment mass spectrometry (FAB-HS), which showed an $(M+K)^+$ ion at m/z 1462. The structures of all compounds were characterized by ¹H-NMR and ¹³C-NMR spectroscopies, as well as IR spectroscopy and elemental analyses.

Preliminary examination of the biological activity revealed that two synthetic disaccharides, 3 and 4, possessed weak mitogenic activities.

Experimental

All melting points were determined with a micro-melting point apparatus (Yanagimoto) and are uncorrected. Optical rotations were measured on a JASCO DIP-140 digital polarimeter. IR spectra were measured on a JASCO A-202 infrared spectrophotometer. 1 H-NMR spectra were recorded on a JEOL JNM-FX90Q (90 MHz) FT-NMR spectrometer using tetramethylsilane (TMS) as an internal standard. 13 C-NMR spectra were recorded on a JEOL JNM-FX90Q (22.5 MHz) FT-NMR spectrometer using tetramethylsilane (in CDCl₃) as an internal standard. Chemical shifts were recorded in values (δ) downfield from TMS, and the abbreviations of signal patterns are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. Thin layer chromatography (TLC) was performed on silica gel (Kiesel $60F_{254}$ on aluminium sheet, Merck). All compounds were located by spraying with the phosphate-specific spray-reagent. 11 Preparative TLC was performed on preparative layer chromatography plates (Kieselgel $60F_{254}$, Merck). Column chromatography was performed on silica gel (Kieselel 60, 70—230 mesh, Merck). Evaporations were carried out under reduced pressure at 35 °C.

5-Acetamido-3,5-dideoxy-D-glycero-β-D-galacto-2-nonulosonic Acid Benzyl Ester (6)—Phenyldiazomethane (about 9 mmol) was added dropwise to a suspension of N-acetylneuraminic acid (5) (1.50 g, 4.85 mmol) and methanol (80 ml) with stirring. The mixture was stirred at room temperature for 3 h, then the excess of phenyldiazomethane was decomposed with acetic acid until the red color disappeared. The resulting solution was evaporated to dryness. The residue was purified on a column of silica gel (chloroform: methanol = 3:1) to give 6 (1.24 g, 76.9%), mp 163—165 °C. [α] $_{D}^{20}$ - 21.9° (c = 1.0, methanol). IR (KBr): 3360, 1755, 1661, 687 cm $^{-1}$. ¹H-NMR (CD₃OD) δ: 2.01 (3H, s, -COCH₃), 2.25 (1H, dd, J = 4.5, 12.8 Hz, 3-H_{eq}), 5.22 (2H, s, -COOCH₂Ph), 7.31—7.36 (5H, m, -CH₂Ph). Anal. Calcd for C₁₈H₂₅NO₉: C, 54.13; H, 6.13; N, 3.51. Found: C, 54.23; H, 6.23; N, 3.73.

5-Acetamido-2,4,7,8,9-penta-O-acetyl-3,5-dideoxy-D-glycero- β -D-galacto-2-nonulosonic Acid Benzyl Ester (7) — Acetic anhydride was added to a solution of 6 (1.24 g, 4 mmol), pyridine (3.16 g, 40 mmol), and 4-dimethylamino-pyridine (1.22 g, 10 mmol) in dichloromethane (20 ml) cooled at 0 °C with stirring. The mixture was warmed at room temperature and stirred for 15 h. The resulting mixture was washed with 1 n HCl, saturated aqueous NaCl, saturated aqueous NaHCO₃, and saturated aqueous NaCl. The organic phase was dried (MgSO₄) and concentrated to dryness. The residue was purified on a column of silica gel (chloroform: ether: acetone = 1:2:1) to afford 7 (1.80 g, 73.8%), mp 113—115 °C. [α]_D²⁰ – 29.1° (c = 1.0, CHCl₃). IR (KBr): 3400, 1756, 1671, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.83—2.28 (19H, m, –COCH₃ × 6 and 3-H_{ax}), 2.56 (1H, dd, J = 5.1, 13.2 Hz, 3-H_{eq}), 5.20 (2H, s, –COOCH₂Ph), 7.35 (5H, s, –CH₂Ph). Anal. Calcd for C₂₈H₃₅NO₁₄: C, 55.17; H, 5.79; N, 2.30. Found: C, 55.13; H, 6.16; N, 2.18.

5-Acetamido-4,7,8,9-tetra-O-acetyl-2-chloro-2,3,5-tri-deoxy-D-glycero- β -D-galacto-2-nonulosonic Acid Benzyl Ester (8)—Compound 7 (200 mg, 0.33 mmol) was dissolved in acetyl chloride (7 ml) and cooled in -5 °C—-10 °C. Dry hydrogen chloride gas was passed through the solution for 10 min. The mixture was kept at room temperature for 24 h and concentrated to a syrup. After further coevaporations of the syrup from toluene (3 × 5 ml)

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and then ether $(5 \times 5 \text{ ml})$, the residue was dried under reduced pressure to give a white powder (8) (quantitative). TLC (ethyl acetate, Rf = 0.62) showed the reaction to be complete.

Benzyl 2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(benzyl-5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α,β-D-galacto-2-nonulopyranosyl)-β-D-glucopyranoside (10α and 10β)—Benzyl 2-deoxy-2-[(R)-3-tetradecanoloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]- β -D-glucopyranoside (9) (366 mg, 3.28 × 10⁻⁴ mol), pulverized Molecular Sieves 4A (230 mg), $Hg(CN)_2$ (58 mg, 2.30×10^{-4} mol), and $HgBr_2$ (28 mg, 7.65×10^{-5} mol) were dried by the use of a high vacuum-pump for 2 h. The mixture was dissolved in dichloromethane (5 ml) and stirred at room temperature for 1 h under argon. The chloride (8) (192 mg, 3.28×10^{-4} mol) in CH₂Cl₂ (5 ml) was added to the mixture over a period of 1 h, and then the suspension was stirred at room temperature for 4 d. The resulting mixture was filtered and the filtrate was washed with 10% aqueous KI solution and saturated aqueous NaCl. The organic phase was concentrated to dryness and the residue was purified on a column of silica gel with ether to yield 10α and 10β . The anomeric mixture (10α and 10β) was further purified by preparative TLC to afford (α -linked) 10 α (108 mg, 23.4%) and (β -linked) 10 β (51 mg, 10.6%). 10 α : mp 73—76 °C. [α]_D²⁰ -6.91° (c = 0.9, CHCl₃). IR (KBr): 3352, 1740, 1658, 697 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (12H, t, $J = 5.9 \,\mathrm{Hz}$, $-\mathrm{CH} - (\mathrm{CH}_2)_{10} \,\mathrm{CH}_3 \times 2$ and $-\mathrm{CO}(\mathrm{CH}_2)_{12} \,\mathrm{CH}_3 \times 2$), 1.25 (80H, s, $-\mathrm{CH} - \mathrm{CH}_2 (\mathrm{CH}_2)_9 \,\mathrm{CH}_3 \times 2$ and $-\text{COCH}_2(\text{CH}_2)_{11}\text{CH}_3 \times 2$), 1.84—2.22 (16H, m, $\text{CH}_3\text{CO} \times 5$ and H-3_{ax}), 2.22—2.64 (8H, m, $\text{COCH}_2 \times 4$), 2.69 (1H, dd, J=4.4, 14.4 Hz, $3-H_{eq}$), 5.21 (2H, s, $-COOCH_2Ph$), 5.85 (1H, d, J=9.0 Hz, -NH-), 7.29, 7.36 (10H, s, $-CH_2Ph \times 2$). ¹³C-NMR (CDCl₃) δ : 99.7 (s, ketosidic carbon atom), 100.3 (d, glycosidic carbon atom). *Anal.* Calcd for $C_{95}H_{154}N_2O_{23}$: C, 67.42; H, 9.17; N, 1.65. Found: C, 67.75; H, 8.93; N, 1.63. **10** β : Amorphous. $[\alpha]_D^{20} - 13.1^{\circ}$ (c =0.94, CHCl₃). IR (KBr): 3400, 1754, 1668, $702 \,\mathrm{cm}^{-1}$. ¹H-NMR (CDCl₃) δ : 0.88 (12H, t, $J=5.9 \,\mathrm{Hz}$, -CH- $(CH_2)_{10}CH_3 \times 2$ and $-CO(CH_2)_{12}CH_3 \times 2$), 1.25 (80H, s, $-CH-CH_2(CH_2)_9CH_3 \times 2$ and $-COCH_2(CH_2)_{11}CH_3 \times 2$), 1.89—2.18 (16H, m, $C\underline{H}_3CO \times 5$ and $H-3_{ax}$), 2.22—2.67 (9H, m, $COC\underline{H}_2 \times 4$ and $H-3_{eq}$), 5.23 (2H, s, $-COOC\underline{H}_2Ph$), 5.85 (1H, d, $J = 9.3 \,\text{Hz}$, -NH-), 7.32, 7.35 (10H, s, $-\text{CH}_2\text{Ph} \times 2$). $^{13}\text{C-NMR}$ (CDCl₃) δ : 98.5 (s, ketosidic carbon atom), 100.2 (s, glycosidic carbon atom). Anal. Calcd for C₉₅H₁₅₄N₂O₂₃·2H₂O: C, 66.02; H, 9.21; N, 1.62. Found: C, 66.20; H, 9.07; N, 1.52.

Benzyl 2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(benzyl-5-acetamido-4,7;8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosyl)-4-O-diphenylphosphono-β-D-glucopyranoside (11)—Diphenylphosphorochloridate (36 mg, 1.33×10^{-4} mol) was added to a solution of 10α (45 mg, 2.66×10^{-5} mol), 4-dimethylaminopyridine (16 mg, 1.33×10^{-4} mol), and pyridine (11 mg, 1.33×10^{-4} mol) in CH₂Cl₂ (5 ml) at 0 °C under argon. The reaction mixture was warmed at room temperature and stirred for 3 h. The solution was washed with saturated aqueous NaHCO₃ and saturated aqueous NaCl. The organic phase was dried with MgSO₄ and evaporated. Purification of the residue was achieved with preparative TLC (chloroform: methanol = 20:1) to give 11 (29 mg, 57%), mp 49—52 °C. [α]_D²¹ -8.89° (c=0.36, CHCl₃). IR (KBr): 3352, 1740, 1658, 955, 697 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (12H, t, J=5.6 Hz, -CH-(CH₂)₁₀CH₃×2 and -CO(CH₂)₁₂CH₃×2), 1.25 (80H, s, -CH-CH₂(CH₂)₉CH₃×2 and -COCH₂(CH₂)₁₁CH₃×2), 1.84—2.22 (16H, m, CH₃CO×5 and H-3_{ax}), 2.69 (1H, dd, J=4.6, 12.9 Hz, 3-H_{eq}), 5.21 (2H, s, -COOCH₂Ph), 5.81 (1H, d, J=8.8 Hz, -NH-), 7.29, 7.36 (20H, s, -CH₂Ph×2 and (PhO)₂PO-). Anal. Calcd for C₁₀₇H₁₆₃N₂O₂₆P: C, 66.78; H, 8.54; N, 1.46. Found: C, 66.39; H, 9.01; N, 1.33.

2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosyl)-4-O-phosphono-D-glucopyranose (3)—The phosphate (11) (16 mg, 8.31×10^{-6} mol) in methanol (0.3 ml) was hydrogenated in the presence of 10% Pd-on-carbon (8 mg) at 40 °C for 4 h. The catalyst was filtered off and Adams platinum catalyst (10 mg) was added to the filtrate. Hydrogenolysis was continued at 35 °C for 15 h and TLC (chloroform: methanol = 5:1) showed the reaction to be complete. The catalyst was filtered off and the filtrate was evaporated to dryness. The residue was purified by preparative TLC (chloroform: methanol = 10:3) to give 3 (3 mg, 23%), mp 41—44 °C. [α]²¹₄₅₀—55.6° (c = 0.018, methanol). IR (KBr): 3352, 1742, 1650, 1262 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (12H, t, J = 6.1 Hz, J = 6.1 Hz,

5-Acetamido-2,4,7,8,9-penta-*O*-monochloroacetyl-3,5-dideoxy-D-*glycero*-α-D-*galacto*-2-nonulosonic Acid Benzyl Ester (12) — Monochloroacetic anhydride (4.44 g, 2.60×10^{-2} mol) in CH₂Cl₂ (45 ml) was added to a suspension of pyridine (2.06 g, 2.60×10^{-2} mol), 4-dimethylaminopyridine (790 mg, 6.50×10^{-3} mol), and the benzyl ester (6) (1.58 g, 2.60×10^{-3} mol) in CH₂Cl₂ (45 ml) at 0 °C. The resulting mixture was stirred at 0 °C for 3 h, and then at room temperature for 15 h. The solution was worked up in the same manner as described for 7. The residue was chromatographed on silica gel with CH₂Cl₂: ether (2:1) to afford 12 (1.58 g, 77.7%), mp 58—59 °C. [α]_D²¹ – 20.1° (c = 1.6, CHCl₃). IR (KBr): 3276, 1769, 1689, 698 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.90 (3H, s, CH₃CO-), 2.65 (1H, dd, J = 5.0, 13.2 Hz, 3-H_{eq}), 3.99—4.25 (10H, m, ClCH₂CO-×5), 5.23 (2H, s, PhCH₂OCO-), 6.14 (1H, d, J = 9.3 Hz, -NH₋), 7.36 (5H, s, -CH₂Ph). *Anal.* Calcd for C₂₈H₃₀Cl₅NO₁₄: C, 43.02; H, 3.87; N, 1.79. Found: C, 43.38; H, 3.80; N, 1.90.

5-Acetamido-4,7,8,9-tetra-O-monochloroacetyl-2-chloro-2,3,5-trideoxy-D-glycero- α -D-galacto-2-nonulosonic Acid Benzyl Ester (13)—The monochloroacetyl compound (12) (250 mg, 3.20×10^{-4} mol) was dissolved in acetyl

chloride (6 ml) saturated with dry hydrogen chloride gas at -5 °C — -10 °C. The solution was stirred at room temperature for 24 h and concentrated to dryness. Traces of acetyl chloride, monochloroacetic acid, and hydrogen chloride were removed by coevaporation from toluene (5 × 5 ml) and then ether (5 × 5 ml) to yield 13 (quantitative). TLC (CH₂Cl₂: ehter = 5:1) showed the reaction to be complete.

Benzyl 2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-tetradecanoyloxytetradecanoyl]-6-O-(benzyl-5-acetamido-3,5-di-deoxy-D-glycero-α-D-galacto-2-nonulopyranosyl)-β-D-glucopyranoside (14) — The glucosamine derivative (9) (366 mg, 3.23×10^{-4} mol), pulverized Molecular Sieves 4A (400 mg), Hg(CN)₂ (58 mg, 2.30×10^{-4} mol), and HgBr₂ (28 mg, 7.65×10^{-5} mol), dried with a high vacuum-pump for 2 h, were dissolved in CH₂Cl₂ (7 ml) and the mixture was stirred under argon for 1 h. The chloride (13) (231 mg, 3.20×10^{-4} mol) in CH₂Cl₂ (7 ml) was then added dropwise, and the whole was stirred at room temperature for 2 d. The resulting mixture was worked up as described for 10. The crude product (14) was purified by a column of silica gel (CH₂Cl₂: ether = 5:1) to afford 14 (25 mg, 4.3%), mp 32-34 °C. [α]_D²¹ – 13.3° (c=0.18, CHCl₃). IR (KBr): 3296, 1746, 1663, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (12H, t, J=5.9 Hz, -CH-(CH₂)₁₀CH₃ × 2 and -CO(CH₂)₁₂CH₃ × 2), 1.25 (80H, s, -CH-CH₂(CH₂)₉CH₃ × 2 and -COCH₂(CH₂)₁₁CH₃ × 2), 1.89 (3H, s, CH₃CO-), 2.65 (1H, dd, J=5.4, 14.4 Hz, $3-H_{eq}$), 3.98-4.26 (8H, m, -COCH₂Cl×4), 5.22 (2H, s, -COOCH₂Ph), 7.42, 8.19 (10H, m, PhCH₂-×2). Anal. Calcd for C₉₅H₁₅₀Cl₄N₂O₂₃: C, 62.35; H, 8.26; N, 1.63. Found: C, 62.42; H, 8.26; N, 1.42.

Benzyl 2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanol]-6-O-(benzyl-5-acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosyl)-4-O-diphenylphosphono-β-D-glucopyranoside (15)—Diphenylphosphorochloridate (22 mg, 8.32×10^{-6} mol) was added dropwise to a solution of the disaccharide (14), pyridine (7 mg, 8.32×10^{-5} mol), and 4-dimethylaminopyridine (10 mg, 8.32×10^{-5} mol) in CH₂Cl₂ under argon at 0 °C. The mixture was stirred at room temperature for 3 h and worked up as described for 11. The crude product (11), thiourea (13 mg, 1.66×10^{-4} mol), and diisopropylethylamine (22 mg, 1.66×10^{-4} mol) were dissolved in tetrahydrofuran (THF) (3 ml), and then refluxed for 3 h. The solvent was evaporated off and the residue was purified by preparative TLC (CHCl₃: methanol = 5:1) to give 15 (6 mg, 43%), mp 64—67 °C. [α]_D²¹ – 7.7° (c = 0.26, CHCl₃). IR (KBr): 3400, 1742, 1675, 972, 695 cm⁻¹. ¹H-NMR (CDCl₃): 0.88 (12H, t, J=6.1 Hz, -CH-(CH₂)₁₀CH₃×2 and -CO(CH₂)₁₂CH₃×2), 1.26 (80H, s, -CH-CH₂(CH₂)₉CH₃×2 and -COCH₂(CH₂)₁₁CH₃×2), 1.97 (3H, s, CH₃CO-), 7.26—7.45 (20H, m, PhCH₂-×2 and (PhO)₂PO-). Anal. Calcd for C₉₉H₁₅₅N₂O₂₂P: C, 67.70; H, 8.90; N, 1.59. Found: C, 67.98; H, 8.40; N, 1.55.

2-Deoxy-2-[(R)-3-tetradecanoloxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-(5-acetamide-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosyl)-4-O-phosphono-D-glucopyranose (4)—The phosphate (12) (11 mg, 6.26×10^{-6} mol) in methanol (0.1 ml) was hydrogenated in the presence of Pd-on-carbon (5 mg) at 40 °C for 3 h. The catalyst was filtered off and Adams platinum catalyst (10 mg) was added to the filtrate. Hydrogenolysis was run at 35 °C for 18 h and TLC (chloroform: methanol=2:1) showed the reaction to be complete. The catalyst was filtered off and the filtrate was concentrated to dryness. The residue was purified by preparative TLC (chloroform: methanol=2:1) to give 4 (5 mg, 56%), mp 183—187 °C. [α] $_D^{22}$ -25° (c=0.04, methanol). IR (KBr): 3448, 1743, 1645, 1262 cm $^{-1}$. 1 H-NMR (CDCl $_3$) δ : 0.88 (12H, t, J=6.1 Hz, -CH-(CH $_2$) $_{10}$ CH $_3$ × 2 and -CO(CH $_2$) $_{12}$ CH $_3$ × 2), 1.26 (80 H, s, -CH-CH $_2$ (CH $_2$) $_9$ CH $_3$ × 2 and -COCH $_2$ (CH $_2$) $_{11}$ CH $_3$ × 2), 1.93 (3H, s, CH $_3$ CO-). Positive FAB-MS (M+K) $^+$ at m/z 1462.

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