

Synthesis of a neoglycoprotein containing the Lewis X analogous trisaccharide β -D-GalpNAc- $(1\rightarrow 4)[\alpha$ -L-Fucp- $(1\rightarrow 3)]$ - β -D-GlcpNAc

Johannes Bartek, Renate Müller, Paul Kosma*

Institute of Chemistry, University of Agricultural Sciences Vienna, Muthgasse 18, A-1190 Vienna, Austria

Received 10 December 1997; accepted 10 March 1998

Abstract

The trisaccharide allyl glycoside **36** and related disaccharide part structures have been prepared using the 2-trichloroacetamido-2-deoxy- α -D-galactopyranosyl trichloroacetimidate derivative **9** as glycosyl donor under promotion with TMSOTf or Sn(OTf)₂, respectively, to produce the β -(1 \rightarrow 4) linkage to suitably protected glucosamine derivatives in fair yields. Fucosylation was effected employing the ethyl 1-thio glycosyl donor **20** in the presence of IDCP. Deprotection of the intermediates afforded the disaccharide allyl glycosides β -D-GalpNAc-(1 \rightarrow 4)- β -D-GlcpNAc **13**, β -D-GalpNClAc-(1 \rightarrow 4)- β -D-GlcpNAc **14**, α -L-Fucp-(1 \rightarrow 3)- β -D-GlcpNAc **24**, α -L-Fucp-(1 \rightarrow 4)- β -D-GlcpNAc **31** and the branched trisaccharide allyl glycoside β -D-GalpNAc-(1 \rightarrow 4)[α -L-Fucp-(1 \rightarrow 3)]- β -D-GlcpNAc **36**. The trisaccharide which corresponds to a structural motif occurring in N-glycoprotein glycans from human urokinase, human recombinant protein C, phospholipase A₂ as well as O-glycans, was converted into a neoglycoprotein following introduction of a cysteaminederived spacer group and subsequent activation with thiophosgene. © 1998 Elsevier Science Ltd. All rights reserved

Keywords: Lewis X; N-Acetyl-D-galactosamine; Oligosaccharide synthesis

1. Introduction

The terminal trisaccharide determinant β -D-GalpNAc- $(1\rightarrow 4)[\alpha$ -L-Fucp- $(1\rightarrow 3)]$ - β -D-GlcpNAc has been found in N- and O-glycans from various organisms. Thus, it has been isolated from allergenic H-antigens of sea squirt [1] and described as a structural element occurring in phospholipase A₂ and hyaluronidase of the honey bee venom [2,3] and in a serine protease from the snake venom of

Bothrops moojeni toxins [4]. The presence of this trisaccharide has furthermore been demonstrated for egg jelly coats from Axolotl maculatum [5] and N-glycans of Schistosomas mansoni [6]. Moreover, it constitutes an important determinant in the N-glycans of bovine pro-opiomelanocortin [7], human urokinase [8] and recombinant human protein C expressed in kidney 293 cells [9]. It has been postulated that the trisaccharide unit effectively inhibits selectin-mediated cell adhesion and thus seems to be a more potent ligand for E-selectin than sialyl Lewis X [10].

^{*} Corresponding author.

Previously, the trisaccharide has been prepared as the methyl glycoside by a chemo-enzymatic approach employing human milk fucosyltransferase [11]. For further biological studies, we have set out to prepare neoglycoproteins containing this trisaccharide determinant. Here, we report on the synthesis of the corresponding allyl glycoside which may be used for the introduction of a spacer function and the design of glycopolymers. A 2-deoxy-2-trichloro-acetamido-galactopyranosyl donor was elaborated to provide access for additional chemical transformations at the amino group of the terminal galactosamine unit.

2. Results and discussion

For the synthesis of the 2'-amino-2'-deoxy-lactosamine derivatives **13** and **14**, known [12] allyl 2-acetamido-2-deoxy-4,6-*O*-*p*-methoxybenzylidene- β -D-glucopyranoside **1** was subjected to reductive ring-opening [13] in *N*,*N*-dimethylformamide in the presence of sodium cyanoborohydride–trifluoroacetic acid which proceeded with low selectivity to give the 4-*O*- and 6-*O*-*p*-methoxybenzyl ether derivatives **2** and **3** in a 1:1.1 ratio and 77% yield (Scheme 1). The isomers were separated by chromatography and elaborated into suitable glycosyl acceptors. Thus, compound **2** was converted into the 3,6-di-*O*-acetyl derivative **5** *via* acetylation $(\rightarrow 4)$ and subsequent oxidative removal of the 4-O*p*-methoxybenzyl group, which was accomplished in 72% yield by treatment with ceric ammonium nitrate and sonication. As the glycosyl donor for the galactosamine unit, the 2-deoxy-2-trichloroacetamido trichloroacetimidate derivative 9 was elaborated, since related glucosyl donors have been used for efficient synthesis of chitobiosyl units [14]. Starting from previously described [15] N-p-methoxybenzylidene derivative 6, amine 7 was obtained by acidic hydrolysis with 1 M HCl in 86% yield, which was then acylated with trichloroacetyl chloride-triethylamine in dichloromethane to give compound 8 in 99% yield. The anomeric acetate was selectively removed with hydrazine acetate [16] and the resulting reducing galactosamine derivative was converted into the α -trichloroacetimidate 9 by reaction with trichloroacetonitrile and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) [17] in dichloromethane in 78% yield. Trimethylsilyl trifluoromethanesulfonate-promoted coupling of the trichloroacetimidate donor 9 with acceptor 5 afforded the β -(1 \rightarrow 4)-linked crystalline disaccharide derivative 10 in 37% yield. The structural assignment was evident from the value of the coupling constant $J_{1'2'}$ (8.3 Hz) of the galactosamine unit and the chemical shift of H-4 (δ 3.78) of the glucosamine residue. Promotion of the glycosidation reaction by either stannous trifluoromethanesulfonate or zinc trifluoromethanesulfonate [18] resulted in even



lower yields (<20%) of the disaccharide product. Reduction [19] of the *N*-trichloroacetamido group of **10** could be achieved in the presence of sodium borohydride in ethanol to give after *N*-acetylation the disaccharide derivative **11** in 68% yield. Moreover, partial reduction of the trichloroacetamide was carried out in 78% yield using zinc dust in acetic acid at 110 °C to furnish the *N*-chloroacetamido derivative **12**.

Deprotection of both protected disaccharide derivatives 11 and 12 with sodium methoxide in methanol proceeded in near quantitative yield to give the *N*-acetyl and *N*-chloroacetyl compounds 13 and 14, respectively. Due to the downfield-shift of the chloromethylene protons, the remaining methyl group of the reducing glucosamine unit and the chloroacetamido group in compound 14 are amenable for selective NOE experiments in conformational studies.

Since the anomeric allyl group was chosen for the introduction of the spacer moiety, the use of benzyl-protected fucopyranosyl donors was not feasible. Instead, the previously described [20] tri-*O*-benzoyl-L-fucopyranosyl bromide **15** was employed (Scheme 2). Glycosylation of either *N*acetyl or *N*-phthaloyl protected 4,6-*O*-*p*-methoxybenzylidene derivative **1** or **17** [21] was promoted by mercuric cyanide–mercuric bromide to give the β -(1 \rightarrow 3)-linked disaccharide 16 in 75% yield and a 1:1 anomeric mixture of the N-phthalimido derivatives 18 and 19 in 38% yield, respectively, in accordance with the results of Nifant'ev et al. [20]. By using the ethyl 1-thio glycoside 20 [22] under promotion of iodonium di-sym-collidinium perchlorate (IDCP) [23], the α -(1 \rightarrow 3)-linked disaccharide derivative 21 was formed with greatly enhanced selectivity (α/β ratio 12:1) and in fair yield (49%). Oxidative removal of the 4,6-O-pmethoxybenzylidene group with ceric ammonium nitrate furnished the 4,6-diol derivative 22, whereas treatment of 21 with 2,3-dichloro-5,6-dicyano-1,4benzoquinone (DDQ) lead to simultaneous cleavage of the 2'-O-p-methoxybenzyl ether and the 3',4'-O-isopropylidene group, respectively, to give, after O-acetylation, the acetylated disaccharide derivative 23 in 92% overall yield. Deprotection in the presence of sodium methoxide afforded the disaccharide α -L-Fucp-(1 \rightarrow 3)- β -D-GlcpNAc allyl glycoside 24. (Scheme 2).

Since the reductive ring opening of the 4,6-O-pmethoxybenzylidene group of **21** in the presence of sodium cyanoborohydride–trifluoroacetic acid met with difficulties due to the acid lability of the fucosidic bond, the disaccharide acceptor **25** was prepared *via* regioselective glycosylation of the 3,4-diol compound **3** (Scheme 3). Iodonium



Scheme 2.



Scheme 3.

di-sym-collidinium perchlorate-promoted coupling of 3 with 1.1 equivalents of the ethyl 1-thioglycoside donor 20 in 1:5 N,N-dimethylformamidedichloromethane afforded a mixture of three disaccharides which was separated by column chromatography to give 32% of the α -(1 \rightarrow 3)-linked disaccharide 25, a minor portion (4%) of the corresponding β -isomer 26 and the α -(1 \rightarrow 4)-linked disaccharide derivative 28 in 11% yield. Whereas the anomeric configurations were deduced from the values of the proton coupling constants $J_{1',2'}$ (3.5 Hz for 25 and 28, 8.3 Hz for 26), the attachment site was assigned after subsequent O-acetylation to give 27 and 29, respectively. Upon acetylation, H-4 of compound 27 experienced a downfield shift (to 4.88 ppm), whereas H-3 of 29 was shifted to 5.08 ppm. Compound 28, corresponding to a synthetic part structure of the Lewis A bloodgroup antigenic determinant [24] was deprotected via hydrolysis of the blocking groups in aqueous 90% trifluoroacetic acid followed by reacetylation to give 30 in 83% yield. Zemplén de-O-acetylation of 30 gave the disaccharide α -L-Fucp-(1 \rightarrow 4)- β -D-GlcpNAc allyl glycoside **31**.

Various attempts to introduce the β -(1 \rightarrow 4)linked galactosamine residues by coupling of the disaccharide acceptor **25** with 2-acetamido-3,4,6tri-*O*-acetyl-2-deoxy- α -D-galactopyranosyl donors (anomeric bromide or trichloroacetimidate) in the presence of silver trifluoromethanesulfonate or borontrifluoride etherate, respectively, under standard and "inverse conditions" [25] did not lead to significant product formation. Employing the Ntrichloroacetamido donor 9 under promotion of trimethylsilyl trifluoromethanesulfonate, silver trifluoromethanesulfonate, zinc trifluoromethanesulfonate, stannic chloride, or stannous chloride resulted in the formation of by-products or hydrolysis of the protecting groups of the acceptor disaccharide. Finally, stannous trifluoromethanesulfonate-mediated glycosylation [26] under carefully controlled conditions proved to be effective. Thus, using 1.5 equivalents of 9, 1.3 equivalents of promoter and 1.1 equivalents of N, N, N', N'-tetramethyl urea in dichloromethane-toluene for 1.5 h at room temperature furnished the trichloromethyloxazoline derivative **32** (36%), the β -(1 \rightarrow 4)linked trisaccharide derivative 33 (41%) and unreacted starting material 25 (26%), which were separated by column chromatography. The β anomeric configuration of 33 was evident from the value $J_{1''2''}$ (9.0 Hz) indicating a *trans*-diaxial arrangement of the respective protons. Deblocking was achieved by first removing the *p*-methoxybenzyl ether groups of 33 by treatment with 2,3dichloro-5,6-dicyano-1,4-benzoquinone followed

by O-acetylation-to stabilize the fucosidic linkage—which afforded 34 in 96% yield. Hydrolysis of the isopropylidene acetal group was accomplished in the presence of aqueous 90% trifluoroacetic acid at -20 °C, followed by reduction of the trichloroacetamido group with sodium cyanoborohydride at 60 °C. Subsequent O-acetylation furnished the per-O-acetyl trisaccharide compound 35 in 60% yield (3 steps). Removal of the O-acetyl groups by treatment with sodium methoxide afforded the target trisaccharide derivative 36 in near quantitative yield. Due to the branched substitution pattern, the ¹³C NMR signals of C-4 and C-3 of the glucosamine unit of 36 displayed no significant glycosylation shifts in contrast to the respective signals observed for the disaccharide part structures (Table 1).

Finally, the allyl glycoside was transformed in 96% yield into the 3-(2-aminoethylthio)propyl spacer [27] derivative **37** by radical addition of cysteamine hydrochloride under UV irradiation [28]. The spacer compound **37** was isolated as the acetate, then activated with thiophosgene and coupled to the ε -amino groups of lysine residues of bovine serum albumin (BSA). The ligand/

Table 1

¹³ C NMR data	^a for disaccharides	13, 14, 24, 31 an	d trisaccharide 36
--------------------------	--------------------------------	-------------------	--------------------

protein ratio of the BSA conjugate **38** based on protein [29] and aminosugar analysis [30] was 6:1 mol/mol.

3. Experimental

General Methods.--Melting points were determined with a Kofler hot stage and are uncorrected. Optical rotations were measured with a Perkin-Elmer 243 B polarimeter; $[\alpha]_{D}^{20}$ values are given in units of 10⁻¹ degcm² g⁻¹. ¹H NMR spectra were recorded at 297 K with a Bruker AC 300F instrument operating at 300 MHz for ¹H using CDCl₃ as solvent and tetramethylsilane as internal standard, unless stated otherwise. 500 MHz ¹H spectra were recorded on a Bruker AMX instrument. Coupling constants are given in Hz (first order values). ¹³C NMR spectra were measured at 75.47 MHz and referenced to 1,4-dioxane (δ 67.40). Homo- and heteronuclear 2D NMR spectroscopy was performed with Bruker standard software. TLC was performed on Merck precoated plates $(5 \times 10 \text{ cm})$, layer thickness 0.25 mm, Silica Gel 60F₂₅₄); spots were detected by spraying with anisaldehyde-H₂SO₄.

 $\delta(nnm)$

Residue	Carbon	- (FF)					
		13	14	24	31	36	
α-L-Fucp	1			100.83	100.44	99.27	
	2			68.86	68.88	68.59	
	3			70.43	70.22	70.05	
	4			72.73	72.71	72.86	
	5			67.79	67.82	67.76	
	6			16.06	16.02	16.21	
β-d-GalpNR	1	102.58	102.01			101.57	
	2	53.39	53.98			53.21	
	3	71.52	71.26			71.60	
	4	68.45	68.51			68.19	
	5	76.18	76.22			75.67	
	6	61.80	61.82			62.26	
β-D-GlcpNAc	1	100.82	100.85	100.75	100.78	100.71	
	2	55.66	55.74	56.13	56.82	56.39	
	3	73.44	73.41	81.31	73.47	75.54	
	4	79.77	79.59	69.50	78.25	74.26	
	5	75.39	75.38	76.77	76.08	76.31	
	6	61.00	61.18	61.67	60.82	60.89	
Aglycon	1	71.34	71.34	71.37	71.24	71.40	
	2	134.14	134.18	134.23	134.18	134.18	
	3	119.04	119.03	119.06	119.00	119.07	
NHAc		23.03	23.00	23.07	22.94	23.02	
		22.98				23.02	
СО		175.63	174.45	175.55	175.20	175.68	
		175.45	171.31			175.14	
CH ₂ Cl			43.14				

^a Spectra (75.47 MHz) were recorded at 297 K and referenced to 1,4-dioxane (δ 67.40).

For column chromatography, silica gel (0.040-0.063 mm) was used. Concentration of solutions was performed at reduced pressure at temperatures $< 40 \,^{\circ}$ C. Elemental analyses were provided by Dr. J. Theiner, Mikroanalytisches Laboratorium, Institut für Physikalische Chemie, University of Vienna. Electrospray-ionisation mass spectra (ES-MS) were obtained on a Finnigan TSQ7000 instrument in the positive ion mode using 1:1 MeOH–water in 0.1 mM NH₄OAc containing 1% HOAc as the matrix.

Allyl 2-acetamido-2-deoxy-4-O-p-methoxybenzyl- β -D-glucopyranoside (2) and allyl 2-acetamido-2deoxy-6-O-p-methoxybenzyl-β-D-glucopyranoside (3).—A suspension of 1 (1.58 g, 4.15 mmol) and 4 A molecular sieves (0.5 g) in DMF (20 mL) was stirred for 30 min under N₂ at room temperature. NaCNBH₃ (5.2 g, 83 mmol) was added followed by addition of CF₃CO₂H dropwise (3.23 mL, 42 mmol) in DMF (5 mL) for 30 min. The suspension was stirred for 24 h, then solid NaHCO₃ (8 g) was slowly added. The suspension was diluted with $CHCl_3$ (50 mL), filtered, and the filtrate was washed with ice-cold water. The organic layer was dried (Na_2SO_4) , concentrated, and the residue was purified on a column of silica gel $(8:1 \rightarrow 4:1 \text{ EtOAc} \rightarrow 4:$ MeOH). Pooling of the fractions containing the faster moving component and evaporation afforded 2. Yield: 568 mg (36%); colourless crystals, mp 227–229 °C (CH₂Cl₂–hexane); $[\alpha]^{20}_{D}$ –31° (*c* 1.2, MeOH). ¹H NMR (CD₃OD): δ 7.30–7.27 (m, 2 H, arom. H), 6.90–6.86 (m, 2 H, arom. H), 5.89 (m, 1 H, CH=), 5.26 (dq, 1 H,=CH_{2trans}), 5.13 (dq, 1 H, = CH_{2cis}), 4.85 and 4.57 (AB, 2 H, J_{A,B} 10.5 Hz, OCH₂Ar), 4.40 (d, 1 H, *J*_{1,2} 8.5 Hz, H-1), 4.33 (ddt, 1H, OCH₂), 4.05 (ddt, 1 H, OCH₂), 3.81 (dd, 1 H, J_{6a,5} 2.0, J_{6a,6b} 12.0 Hz, H-6a), 3.78 (s, 3 H, OCH₃), 3.72 (dd, 1 H, J_{4,3} 8.0, J_{4,5} 10.0 Hz, H-4), 3.64 (dd, 1 H, J_{6b.5} 5.0 Hz, H-6b), 3.61 (dd, 1 H, J_{2,1} 8.5, J_{2,3} 10.0 Hz, H-2), 3.38 (dd, 1 H, H-3), 3.26 (ddd, 1 H, H-5), 1.98 (s, 3 H, Ac). Anal. Calcd for C₁₉H₂₇NO₇: C, 59.83; H, 7.13; N, 3.67. Found: C, 59.47; H, 7.09; N, 3.60.

Further elution furnished **3** (652 mg, 41%) as colourless crystals, mp 130–132 °C (CH₂Cl₂–hexane); $[\alpha]^{20}{}_{\rm D}$ +2° (*c* 1.1, MeOH). ¹H NMR (CD₃OD): δ 7.31–7.26 (m, 2 H, arom. H), 6.91–6.86 (m, 2 H, arom. H), 5.89 (m, 1 H, CH=), 5.29 (dq, 1 H, =CH_{2trans}), 5.13 (dq, 1 H, =CH_{2cis}), 4.52 (s, 2 H, OCH₂Ar), 4.38 (d, 1 H, J_{1,2} 8.5 Hz, H-1), 4.30 (ddt, 1 H, OCH₂), 4.05 (ddt, 1 H, OCH₂), 3.82 (dd, 1 H, J_{6a,5} 2.0, J_{6a,6b} 11.0 Hz, H-6a), 3.78 (s, 3 H,

OCH₃), 3.67 (dd, 1 H, $J_{2,3}$ 10.0 Hz, H-2), 3.64 (dd, 1 H, $J_{6b,5}$ 5.8 Hz, H-6b), 3.43 (dd, 1 H, $J_{3,4}$ 8.5 Hz, H-3), 3.39 (ddd, 1 H, $J_{5,4}$ 9.5 Hz, H-5), 3.34–3.28 (unresolved signal, 1 H, H-4), 1.96 (s, 3 H, Ac). Anal. Calcd for C₁₉H₂₇NO₇: C, 59.83; H, 7.13; N, 3.67. Found: C, 59.85; H, 7.27. N, 3.61.

Allyl 2-acetamido-3,6-di-O-acetyl-2-deoxy-4-O-pmethoxybenzyl- β -D-glucopyranoside (4).—A solution of 2 (200 mg, 0.52 mmol) and Ac₂O (0.25 mL, 2.65 mmol) in dry pyridine (5 mL) was stirred for 24 h at room temperature. The reaction was quenched by adding MeOH (2.5 mL) at 0 °C. The solution was coevaporated with toluene (10 mL) and taken to dryness. Crystallization of the residue from EtOAc-hexane gave 4 as colourless crystals (230 mg, 95%), mp 151–153 °C (EtOAc–hexane); $[\alpha]^{20}_{D} - 21^{\circ}$ (c 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 7.19–7.14 (m, 2 H, arom. H), 6.89–6.84 (m, 2 H, arom. H), 5.84 (m, 1 H, CH =), 5.55 (d, 1 H, $J_{\rm NH,2}$ 9.5 Hz, NH), 5.25 (dq, 1 H, = CH_{2trans}), 5.17 (dq, 1 H, = CH_{2cis}), 5.09 (dd, 1 H, $J_{3,2}$ 10.5, $J_{3,4}$ 8.5 Hz, H-3), 4.56 and 4.47 (AB, 2 H, J_{A,B} 10.5 Hz, OCH₂Ar), 4.42 (d, 1 H, *J*_{1,2} 8.0 Hz, H-1), 4.35 (dd, 1 H, J_{6a,5} 2.5, J_{6a,6b} 12.0 Hz, H-6a), 4.31 (ddt, 1 H, OCH₂), 4.18 (dd, 1 H, J_{6b.5} 4.5 Hz, H-6b), 4.08 (ddd, 1 H, H-2), 4.03 (ddt, 1 H, OCH₂), 3.80 (s, 3 H, OCH₃), 3.65 (t, 1 H, H-4), 3.53 (ddd, 1 H, J_{5,4} 9.5 Hz, H-5), 2.06, 2.05, 1.95 (3 s, each 3 H, 3 Ac). Anal. Calcd for C₂₃H₃₁NO₉: C, 59.35; H, 6.71; N, 3.01. Found: C, 59.39; H, 6.86; N, 3.15.

Allyl 2-acetamido-3,6-di-O-acetyl-2-deoxy-β-Dglucopyranoside (5).—A solution of 4 (300 mg, 0.64 mmol) in 18:1 CH₂Cl₂-H₂O (7 mL) was treated with ceric ammonium nitrate (710 mg, 1.3 mmol) at 4 °C with ultrasonication for 70 min. NaHCO₃ was added until the solution had pH 5. The solution was dried (Na_2SO_4) and filtered. Solids were washed thoroughly with EtOAc, the organic phases were combined and concentrated. Purification of the residue on silica gel (5:5:1 CH₂Cl₂-hexane–EtOH) furnished 5 (159 mg, 72%) as colourless crystals, mp 146-148 °C (EtOAchexane); $[\alpha]_{D}^{20} - 69^{\circ}$ (c 0.5, CHCl₃). ¹H NMR $(CDCl_3)$: δ 5.87 (m, 1 H, CH =), 5.70 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 5.27 (dq, 1 H, = CH_{2trans}), 5.19 (dq, 1 H, = CH_{2cis}), 5.10 (dd, 1 H, $J_{3,2}$ 10.5, $J_{3,4}$ 8.5 Hz, H-3), 4.57 (d, 1 H, J_{1,2} 8.5 Hz, H-1), 4.51 (dd, 1 H, J_{6a.5} 4.0, J_{6a.6b} 12.0 Hz, H-6a), 4.38 (ddt, 1 H, OCH₂), 4.31 (dd, 1 H, J_{6b.5} 2.5 Hz, H-6b), 4.09 (ddt, 1 H, OCH₂), 3.92 (ddd, 1 H, H-2), 3.62-3.54 (m, 1 H, H-4), 3.52 (ddd, 1 H, J_{5.4} 10.0 Hz, H-5), 3.14 (d, 1 H, J_{OH,4} 5.0 Hz, OH), 2.13, 2.11, and 1.96 (3 s, each 3 H, 3 Ac). Anal. Calcd for $C_{15}H_{23}NO_8$: C, 52.17; H, 6.71; N, 4.06. Found: C, 52.15; H, 6.61; N, 3.98.

1,3,4,6-Tetra-O-acetyl-2-amino-2-deoxy-β-D-galactopyranose hydrochloride (7).—A solution of 6 (3.5 g, 7.5 mmol) in acetone (30 mL) was heated to reflux temperature. After addition of 1 M HCl (11.3 mmol), the solution was cooled to $0 \,^{\circ}$ C. The crystalline mass formed was filtered, washed with acetone, and dried to give 7 (2.9 g, 86%) as colourless needles, mp 210 °C (dec.); $[\alpha]_{D}^{20} + 30^{\circ}$ (c 0.3, H₂O). ¹H NMR (D₂O): δ 5.97 (d, 1 H, J_{1.2} 9.0 Hz, H-1), 5.54 (dd, 1 H, J_{4,3} 3.5, J_{4,5} 1.0 Hz, H-4), 5.34 (dd, 1 H, J_{3,2} 11.0 Hz, H-3), 4.42 (t, 1 H, $J_{5.6a} = J_{5.6b} = 5.5 \text{ Hz}, \text{ H-5}$, 4.26–4.24 (m, 2 H, H-6a,6b), 3.87 (dd, 1 H, H-2), 2.25, 2.22, 2.11, and 2.09 (4 s, each 3 H, 4 Ac). Anal. Calcd for C₁₄H₂₂C1NO₉: C, 43.81; H, 5.77; N, 3.65. Found: C, 43.65; H, 5.51; N, 3.51.

1,3,4,6-Tetra-O-acetyl-2-deoxy-2-trichloroacetamido- β -D-galactopyranose (8).—A suspension of 7 (200 mg, 0.52 mmol) in CH₂Cl₂ (5 mL) was cooled to 0 °C. Et₃N (0.2 mL, 1.5 mmol) and trichloroacetyl chloride (0.09 mL, 0.81 mmol) was added. After 50 min the reaction mixture was diluted with CH_2Cl_2 (50 mL) and washed with satd aq NaHCO₃. The organic layer was dried (Na_2SO_4), concentrated, and the residue was purified on a column of silica gel (2:1 toluene–EtOAc) to give 8 (255 mg, 99%) as colourless syrup; $[\alpha]^{20}_{D} + 4^{\circ} (c$ 0.8, CHCl₃). ¹H NMR (CDCl₃): δ 6.84 (d, 1 H, J_{NH.2} 9.5 Hz, NH), 5.84 (d, 1 H, J_{1.2} 9.0 Hz, H-1), 5.42 (dd, 1 H, *J*_{4,3} 3.3, *J*_{4,5} 1.1 Hz, H-4), 5.25 (dd, 1 H, J_{3,2} 11.3 Hz, H-3), 4.45 (ddd, 1 H, H-2), 4.20 (dd, 1 H, J_{6a.5} 6.5, J_{6a,6b} 11.5 Hz, H-6a), 4.14 (dd, 1 H, J_{6b,5} 6.5 Hz, H-6b), 4.07 (dt, 1 H, H-5), 2.20, 2.14, 2.07, and 2.03 (4 s, each 3 H, 4 Ac). Anal. Calcd for $C_{16}H_{20}Cl_3NO_{10}$: C, 39.00; H, 4.11; N, 2.85. Found: C, 39.67; H, 3.95; N, 2.72.

3,4,6-Tri-O-acetyl-2-deoxy-2-trichloroacetamido- α -D-galactopyranosyl trichloroacetimidate (9).—A solution of 8 (1.7 g, 3.45 mmol) in DMF (15 mL) was stirred with hydrazine acetate (479 mg, 5.2 mmol) for 1 h at room temperature. The solution was diluted with CH_2Cl_2 (50 mL), washed with aq NaHCO₃, dried (Na₂SO₄), and evaporated to dryness. A solution of the residue in CH₂Cl₂ stirred with DBU $(25 \,\mathrm{mL})$ was $(0.13 \,\mathrm{mL},$ 34.5 mmol) and trichloroacetonitrile (3.5 mL, 34.5 mmol) for 75 min. After concentration, the residue was purified on silica gel (2:1 hexane-EtOAc containing 0.1% Et₃N). Yield for 9: 1.6g (78%), colourless crystals, mp 91–92 °C (hexane– EtOAc); $[\alpha]^{20}{}_{D}$ +81° (*c* 0.7, CHCl₃). ¹H NMR (CDCl₃): δ 8.83 (s, 1 H, = NH), 6.83 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 6.52 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 5.54 (dd, 1 H, $J_{4,3}$ 3.0, $J_{4,5}$ 1.0 Hz, H-4), 5.42 (dd, 1 H, $J_{3,2}$ 11.0 Hz, H-3), 4.72 (ddd, 1 H, H-2), 4.41 (dt, 1 H, $J_{5,6a} = J_{5,6b} = 6.5$ Hz, H-5), 4.20 (dd, 1 H, $J_{6a,6b}$ 11.5 Hz, H-6a), 4.09 (dd, 1 H, H-6b), 2.22, 2.05, and 2.03 (3 s, each 3 H, 3 Ac). Anal. Calcd for C₁₆H₁₈Cl₆N₂O₉: C, 32.30; H, 3.05; N, 4.70. Found: C, 32.60; H, 3.06; N, 4.47.

Allyl (3,4,6-tri-O-acetyl-2-deoxy-2-trichloroacetamido- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranoside (10).— A solution of TMSOTf (0.311 mmol) in toluene (1 mL) was added at 0 °C to a solution of 5 (72 mg, 0.208 mmol) and 9 (185 mg, 0.311 mmol) in dry CH_2Cl_2 (5 mL) under N₂. The solution was stirred for 30 min at 0 °C, then diluted with CH2Cl2 (50 mL) and washed with satd aq NaHCO₃. The organic layer was dried (Na₂SO₄), concentrated, and the residue was applied on silica gel. Elution with EtOAc gave 10 (60 mg, 37%) as the major reaction product and 32 (not isolated) as side-product. Colourless crystals, mp 211–213 °C (hexane– EtOAc); $[\alpha]_{D}^{20} - 38^{\circ}$ (*c* 0.2, CHCl₃). ¹H NMR (CDCl₃): δ 7.20 (d, 1 H, J_{NH',2'} 9.5 Hz, NH'), 5.85 (m, 1 H, CH=), 5.72 (d, 1 H, $J_{NH,2}$ 9.5 Hz, NH), 5.37 (dd, 1 H, *J*_{4',3'} 3.4, *J*_{4',5'} 0.9 Hz, H-4'), 5.26 (dq, 1 H, = CH_{2trans}), 5.22 (dq, 1 H, = CH_{2cis}), 5.20 (dd, 1 H, $J_{3',2'}$ 11.3 Hz, H-3'), 5.12 (dd, 1 H, $J_{3,2}$ 9.5, *J*_{3,4} 8.0 Hz, H-3), 4.64 (d, 1 H, *J*_{1',2'} 8.3 Hz, H-1'), 4.51 (d, 1 H, J_{1,2} 7.4 Hz, H-1), 4.45 (dd, 1 H, J_{6a,5} 4.7, J_{6a,6b} 12.1 Hz, H-6a), 4.35–4.28 (m, 2 H, H-6b, OCH₂), 4.18–4.00 (m, 5 H, H-2,2',6a',6b', and OCH₂), 3.91 (dt, 1 H, $J_{5',6a'} = J_{5',6b'} = 6.6$ Hz, H-5'), 3.78 (t, 1 H, J_{4,5} 8.0 Hz, H-4), 3.67–3.61 (m, 1 H, H-5), 2.17, 2.13, 2.08, 2.07, 1.99, and 1.987 (6 s, each 3 H, 6 Ac). Anal. Calcd for C₂₉H₃₉Cl₃ N₂O₁₆: C, 44.77; H, 5.05; N, 3.60. Found: C, 45.17; H, 4.92; N, 3.33.

Allyl (2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranoside (11).—A solution of 10 (38 mg, 0.05 mmol) and NaBH₄ (67 mg, 1.77 mmol) in EtOH (8 mL) was stirred for 48 h at 60 °C. The solution was diluted with EtOH (5 mL), cooled to 0 °C and neutralized with HOAc (0.1 mL). The solution was coevaporated five times with MeOH (10 mL portions) and taken to dryness. The residue was dissolved in pyridine (8 mL) and stirred with Ac₂O (0.2 mL) for 60 h.

The solution was concentrated and the residue was purified on silica gel (10:1 EtOAc-MeOH). The crude product obtained (28 mg) was further subjected to LC on silica gel (10 µm, 10:1 EtOAc-MeOH) which gave 11 as colourless crystals. Yield: 22.5 mg (68%), mp 223–225 °C (EtOAc); $[\alpha]^{20}_{D}$ -23° (c 0.5, MeOH). ¹H NMR (CDCl₃): δ 5.86 (m, 1 H, CH =), 5.78 (d, 1 H, $J_{NH',2'}$ 8.8 Hz, NH'), 5.77 (d, 1 H, J_{NH,2} 9.8 Hz, NH), 5.32 (dd, 1 H, J_{4'.3'} 3.5, $J_{4',5'} < 1.0 \text{ Hz}, \text{H-4'}$, 5.26 (dq, 1 H, = CH_{2trans}), 5.18 $(dq, 1 H, = CH_{2cis}), 5.15 (dd, 1 H, J_{3',2'} 11.5 Hz, H-$ 3'), 5.12 (t, 1 H, $J_{3,4} = J_{3,2} = 8.5$ Hz, H-3), 4.60 (d, 1 H, *J*_{1',2'} 8.3 Hz, H-1'), 4.51 (d, 1 H, *J*_{1,2} 7.3 Hz, H-1), 4.41 (dd, 1 H, $J_{6a,5}$ 3.0, $J_{6a,6b}$ 12.0 Hz, H-6a), 4.35–4.28 (m, 1 H, OCH₂), 4.31 (dd, 1 H, J_{6b.5} 5.0 Hz, H-6b), 4.14–3.94 (m, 4 H, H-2,2',6b', and OCH₂), 4.09 (dd, 1 H, *J*_{6a',5'} 6.5, *J*_{6a',6b'} 11.0 Hz, H-6a'), 3.87 (td, 1 H, J_{5',6b'} 6.5 Hz, H-5'), 3.76 (t, 1 H, J_{4.5} 8.5 Hz, H-4), 3.69–3.64 (m, 1 H, H-5), 2.14, 2.13, 2.08, 2.05, 1.99, 1.96, and 1.958 (7 s, each 3 H, 7 Ac). Anal. Calcd for C₂₉H₄₂N₂O₁₆: C, 51.62; H, 6.27; N, 4.15. Found: C, 51.51; H, 5.96; N, 4.00.

Allyl (3,4,6-tri-O-acetyl-2-chloroacetamido-2 $deoxy-\beta$ -D-galactopyranosyl)- $(1\rightarrow 4)$ -2-acetamido-3.6-di-O-acetyl-2-deoxy- β -D-glucopyranoside (12). A solution of 10 (7 mg, 9 μ mol) in HOAc (2 mL) was treated with zinc-dust (35 mg, 0.535 mmol) for 4h at 110 °C. The solution was diluted with CH_2Cl_2 (50 mL), filtered, and the filtrate was concentrated. Purification of the residue on silica gel (EtOAc) gave 12 as an amorphous solid. Yield: 5 mg (78%). $[\alpha]^{20}_{D} - 23^{\circ} (c \ 0.4, \text{ MeOH})$. ¹H NMR (CDCl₃): δ 6.70 (d, 1 H, J_{NH',2'} 8.5 Hz, NH'), 5.86 (m, 1 H, CH=), 5.72 (d, 1 H, $J_{\rm NH,2}$ 9.0 Hz, NH), 5.35 (dd, 1 H, *J*_{4',3'} 3.5, *J*_{4',5'} 1.0 Hz, H-4'), 5.28 (dd, 1 H, $J_{3',2'}$ 11.5 Hz, H-3'), 5.26 (dq, 1 H, $= CH_{2trans}$), 5.19 (dq, 1 H, $= CH_{2cis}$), 5.12 (dd, 1 H, $J_{3,4}$ 8.0, $J_{3,2}$ 9.5 Hz, H-3), 4.75 (d, 1 H, $J_{1',2'}$ 7.2 Hz, H-1'), 4.51 (d, 1 H, J_{1.2} 7.2 Hz, H-1), 4.42 (dd, 1 H, J_{6a.5} 3.0, J_{6a.6b} 12.0 Hz, H-6a), 4.31 (ddt, 1 H, OCH₂), 4.18 (dd, 1 H, J_{6b,5} 5.6 Hz, H-6b), 4.14–4.00 (m, 4 H, H-2,6a',6b', and OCH₂), 4.03 (s, 2 H, ClCH₂), 3.96 (dd, 1 H, H-2'), 3.90 (dt, 1 H, $J_{5',6a'} = J_{5',6b'} = 7.0 \text{ Hz}, \text{ H-5'}, 3.78 \text{ (t, 1 H, } J_{4,5})$ 8.0 Hz, H-4), 3.63 (m, 1 H, H-5), 2.14, 2.12, 2.08, 2.06, 1.99, and 1.98 (6 s, each 3 H, 6 Ac). Anal. Calcd for C₂₉H₄₁ClN₂O₁₆: C, 49.12; H, 5.83; N, 3.95. Found: C, 48.84; H, 5.77; N, 3.89.

Allyl (2-acetamido-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (13).—A solution of 11 (16 mg, 0.024 mmol) in MeOH (4 mL) was stirred with

0.1 M methanolic NaOMe (1 mL) for 24 h at room temperature. The solution was made neutral by addition of Dowex 50 (H^+) resin and filtered. The filtrate was evaporated and the residue was dissolved in water (2 mL) and lyophilized giving 13 as an amorphous solid. Yield: 11 mg (quant.); $[\alpha]^{20}_{D}$ -3° (c 0.6, H₂O). ¹H NMR (500 MHz, D₂O): δ 5.84 (m, 1 H, CH=), 5.24 (dq, 1 H, =CH_{2trans}), 5.20 (dq, 1 H, = CH_{2cis}), 4.50 (d, 1 H, $J_{1,2}$ 8.2 Hz, H-1), 4.46 (d, 1 H, $J_{1^\prime,2^\prime}$ 8.4 Hz, H-1^\prime), 4.27 (ddt, 1 H, OCH₂), 4.09 (ddt, 1 H, OCH₂), 3.88 (dd, 1 H, $J_{4',3'}$ 3.3, $J_{4',5'}$ < 1.0 Hz, H-4'), 3.87 (dd, 1 H, $J_{2',3'}$ 10.9 Hz, H-2'), 3.79 (dd, 1 H, $J_{6a,5}$ 2.1, $J_{6a,6b}$ 12.1 Hz, H-6a), 3.74 (dd, 1 H, J_{2.3} 11.8 Hz, H-2), 3.72-3.65 (m, 3 H, H-5', 6a', 6b'), 3.70 (dd, 1 H, H-3'), 3.64 (dd, 1 H, J_{3,4} 8.2 Hz, H-3), 3.60 (dd, 1 H, J_{6b,5} 5.6 Hz, H-6b), 3.56 (dd, 1 H, J_{4,5} 9.6 Hz, H-4), 3.45 (ddd, 1 H, H-5), 2.01 and 1.97 (2 s, each 3 H, 2 Ac). ES-MS ($C_{19}H_{32}N_2O_{11}$): m/z 465.1 (M+H)⁺.

Allyl (2-chloroacetamido-2-deoxy-β-D-galactopyranosyl)- $(1 \rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranoside (14).—A solution of 12 (3.5 mg, 4.9 µmol) in MeOH (2 mL) was stirred with 0.1 M methanolic NaOMe (0.3 mL) for 24 h at room temperature. Work-up as described for 13 gave 14 as amorphous powder. Yield 2.5 mg (\sim quant.); $[\alpha]^{20}_{D} - 4^{\circ} (c \ 0.2, H_2O)$. ¹H NMR (D₂O): δ 5.90 (m, 1 H, CH =), 5.33-5.22 (m, 2 H, = CH₂), 4.59 (d, 1 H, *J*_{1',2'} 8.3 Hz, H-1'), 4.55 (d, 1 H, *J*_{1.2} 8.1 Hz, H-1), 4.32 (ddt, 1 H, OCH₂), 4.21 and 4.18 (AB, 2 H, J_{AB} 14.3 Hz, ClCH₂), 4.14 (ddt, 1 H, OCH₂), 3.98 (dd, 1 H, $J_{2',3'}$ 10.9 Hz, H-2'), 3.95 (dd, 1 H, $J_{4',3'}$ 2.8, $J_{4',5'} < 1.0$ Hz, H-4'), 3.86 (dd, 1 H, $J_{6a,5}$ 2.6, J_{6a,6b} 11.9 Hz, H-6a), 3.82 (dd, 1 H, H-3'), 3.79 (dd, 1 H, $J_{6a',5'}$ 3.3, $J_{6a',6b'}$ 10.9 Hz, H-6a'), 3.77-3.61 (m, 6 H, H-2,3,4,6b,5',6b'), 3.50 (ddd, 1 H, H-5), 2.02 (s, 3 H, Ac). ES-MS ($C_{19}H_{31}ClN_2O_{11}$): m/z 499.1 $(M + H)^+$.

Allyl (2,3,4-tri-O-benzoyl-β-L-fucopyranosyl)-(1→3)-2-acetamido-2-deoxy-4,6-O-p-methoxybenzylidene-β-D-glucopyranoside (16).—Compound 15 (170 mg, 0.317 mmol) and 1 (105 mg, 0.264 mmol) were dissolved in dry nitromethane (10 mL) under N₂. After addition of 4 Å molecular sieves and stirring for 0.5 h, Hg(CN)₂ (72 mg, 0.285 mmol) and HgBr₂ (34 mg, 0.095 mmol) were added, and stirring was continued for 14 h at ambient temperature. The mixture was diluted with CH₂Cl₂ (50 mL) and filtered through Celite[®]. The filtrate was washed with aq 10% KI and satd aq NaHCO₃ (2×), dried over MgSO₄, and concentrated. Purification of the residue by column

chromatography (2:1 toluene-EtOAc) yielded 16 (175 mg, 75%) as crystals, mp 228–234 °C (EtOAc– MeOH); $[\alpha]_{D}^{20} - 110^{\circ}$ (c 0.8, CHCl₃). ¹H NMR (CDCl₃): 8 8.09-6.96 (m, 19 H, arom. H), 5.92 (d, 1 H, $J_{\rm NH,2}$ 7.0 Hz, NH), 5.88 (m, 1 H, CH=), 5.70 $(dd, 1 H, J_{3',4'} 3.5, J_{4',5'} 1.0 Hz, H-4'), 5.67 (dd, 1 H,$ $J_{2',3'}$ 10.5, $J_{2',1'}$ 8.0 Hz, H-2'), 5.47 (dd, 1 H, H-3'), 5.28 (dq, 1 H, = CH_{2trans}), 5.21 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 5.19 (dq, 1 H, =CH_{2cis}), 5.07 (s, 1 H, OCHO), 5.01 (d, 1 H, H-1'), 4.64 (dd, 1 H, J_{3.2} 10.0, J_{3.4} 9.0 Hz, H-3), 4.32 (ddt, 1 H, OCH₂), 4.24 $(dd, 1 H, J_{6a,6b} 10.0, J_{6b,5} 4.5 Hz, H-6b), 4.10 (ddt,$ 1 H, OCH₂), 4.07 (dq, 1 H, *J*_{5'.6'} 6.5 Hz, H-5'), 3.87 (s, 3 H, OMe), 3.60 (t, 1 H, J_{6a,5} 10.0 Hz, H-6a), 3.49 (ddd, 1 H, J_{5.4} 9.0 Hz, H-5), 3.37 (t, 1 H, H-4), 3.15 (ddd, 1 H, H-2), 1.99 (s, 3 H, Ac), 1.32 (d, 3 H, H-6'). Anal. Calcd for C₄₆H₄₇NO₁₄: C, 65.94; H, 5.65; N, 1.67. Found: C, 65.61; H, 5.72; N, 1.67.

Allyl $(2,3,4-tri-O-benzoyl-\beta-L-fucopyranosyl)$ - $(1 \rightarrow 3)$ -2-deoxy-4,6-O-p-methoxybenzylidene-2-phthal*imido*- β -D-glucopyranoside (18) and allyl(2,3,4-tri-O-benzoyl- α -L-fucopyranosyl)- $(1 \rightarrow 3)$ -2-deoxy-4,6-O-p-methoxybenzylidene-2-phthalimido-β-D-gluco*pyranoside* (19).—Donor 15 (5.06 g, 9.38 mmol) and compound 17 (3.95g, 8.44 mmol) were dissolved in dry nitromethane (60 mL) under N₂. After addition of 4 Å molecular sieves and stirring for 0.5 h, Hg(CN)₂ (2.13 g, 8.44 mmol) and HgBr₂ (1.01 g, 2.81 mmol) were added, and stirring at ambient temperature was continued for 24 h. The mixture was diluted with CH₂Cl₂ and filtered through Celite[®]. The filtrate was washed with aq 10% KI and satd aq NaHCO₃ (2×), dried over MgSO₄, and concentrated. Purification of the residue by column chromatography (12:1 toluene-EtOAc) yielded 18 (1.46 g, 19%) as crystals, mp 131–135 °C (CH₂Cl₂–MeOH); $[\alpha]^{20}_{D}$ – 38° (c 0.57, CHCl₃). ¹H NMR (CDCl₃): δ 7.82–6.93 (m, 23 H, arom. H), 5.72 (m, 1 H, CH =), 5.49 (dd, 1 H, $J_{2',3'}$ 10.3, $J_{2',1'}$ 7.8 Hz, H-2'), 5.42 (dd, 1 H, $J_{4',3'}$ 3.5, $J_{4',5'} < 1.0 \,\text{Hz}, \text{H-4'}$, 5.42 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 5.32 (dd, 1 H, H-3'), 5.20 (s, 1 H, OCHO), 5.15 (dq, $1 \text{ H}, = \text{CH}_{2\text{trans}}$, 5.05 (dq, $1 \text{ H}, = \text{CH}_{2\text{cis}}$), 4.89 (d, 1 H, H-1'), 4.71 (dd, 1 H, J_{3,2} 10.0, J_{3,4} 9.0 Hz, H-3), 4.33 (dd, 1 H, H-2), 4.31 (dd, 1 H, $J_{6a,6b}$ 10.0, $J_{6b,5}$ 4.5 Hz, H-6b), 4.28 (ddt, 1 H, OCH₂), 4.04 (ddt, 1 H, OCH₂), 3.88 (s, 3 H, OMe), 3.82 (dq, 1 H, $J_{5',6'}$ 6.5 Hz, H-5'), 3.72 (t, 1 H, $J_{6a,5} \sim 10$ Hz, H-6a), 3.65–3.55 (m, 2 H, H-4,5), 0.55 (d, 3 H, H-6'). Anal. Calcd for C₅₂H₄₇NO₁₅: C, 67.45; H, 5.12; N, 1.51. Found: C, 67.03; H, 5.28; N, 1.48.

Further elution gave **19** (1.49 g, 19%) as crystals, mp 111–115 °C (CH₂Cl₂–MeOH); $[\alpha]^{20}_{D}$ –158° (c 0.75, CHCl₃). ¹H NMR (CDCl₃): δ 7.89–6.88 (m, 23 H, arom. H), 5.85 (dd, 1 H, $J_{3',2'}$ 11.0, $J_{3',4'}$ 3.3 Hz, H-3'), 5.58 (s, 1 H, OCHO), 5.53 (m, 1 H, CH =), 5.49 (dd, 1 H, $J_{4',5'}$ 1.0 Hz, H-4'), 5.38 (dd, 1 H, *J*_{2',1'} 3.8 Hz, H-2'), 5.16 (d, 1 H, H-1'), 5.09 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 5.01 (dq, 1 H, = CH_{2trans}), 4.96 (dd, 1 H, J_{3,2} 10.5, J_{3,4} 9.0 Hz, H-3), 4.92 (dq, 1 H, = CH_{2cis}), 4.56 (dq, 1 H, $J_{5',6'}$ 6.5 Hz, H-5'), 4.40 (dd, 1 H, J_{6a.6b} 10.0, J_{6b.5} 4.5 Hz, H-6b), 4.33 (dd, 1 H, H-2), 4.20 (ddt, 1 H, OCH₂), 3.91 (ddt, 1 H, OCH₂), 3.87 (t, 1 H, $J_{6a,5} \sim 10$ Hz, H-6a), 3.82 (m, 1 H, H-4), 3.78 (s, 3 H, OMe), 3.71 (dt, 1 H, $J_{5,4} \sim 9.5$ Hz, H-5), 0.55 (d, 3 H, H-6'). Anal. Calcd for C₅₂H₄₇NO₁₅: C, 67.45; H, 5.12; N, 1.51. Found: C, 67.44; H, 5.33; N, 1.52.

Allyl (3,4-O-isopropylidene-2-O-p-methoxy $benzyl-\alpha-L-fucopyranosyl)-(1\rightarrow 3)-2-acetamido-2$ *deoxy-4,6-O-p-methoxybenzylidene-β-D-glucopyra*noside (21).—Compound 1 (1.423 g, 3.75 mmol) and 20 (1.656 g, 4.5 mmol) were dissolved in 5:1 CH_2Cl_2 -DMF (96 mL) under N₂. After addition of 4 Å molecular sieves, and stirring for 0.5 h, IDCP (1.757 g) was added in 5 portions over 15h, and stirring at room temperature was continued for further 25h. Then the mixture was diluted with CH₂Cl₂, and filtered through Celite[®]. The filtrate was washed with aq 5% $Na_2S_2O_3$, dried over MgSO₄, and concentrated. Purification of the residue by column chromatography (1:1 toluene-EtOAc) and crystallization from EtOAchexane yielded **21** (1.262 g, 49%), mp 184–186 °C; $[\alpha]^{20}_{D} - 78^{\circ}$ (c 1.1, CHCl₃). ¹H NMR (CDCl₃): δ 7.43–7.38 (m, 2 H, arom. H), 7.30–7.25 (m, 2 H, arom. H), 6.91–6.85 (m, 4 H, arom. H), 5.84 (m, 1 H, CH=), 5.75 (d, 1 H, $J_{\rm NH,2}$ 7.0 Hz, NH), 5.46 (s, 1 H, OCHO), 5.25 (dq, 1 H, = CH_{2trans}), 5.17 (dq, 1 H, =CH_{2cis}), 5.07 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 4.90 (d, 1 H, J_{1',2'} 3.5 Hz, H-1'), 4.84 (d, 1 H, J_{gem} 11.5 Hz, OCH₂Ar), 4.35 (dq, 1 H, $J_{5',6'}$ 6.5, $J_{5',4'}$ 2.5 Hz, H-5'), 4.35 (d, 1 H, OCH₂Ar), 4.23 (dd, 1 H, $J_{3',2'}$ 8.0, $J_{3',4'}$ 5.5 Hz, H-3'), 4.40–4.22 (m, 3 H, H-3,6a, OCH₂), 4.06 (ddt, 1 H, OCH₂), 3.93 (dd, 1 H, H-4'), 3.80 (2 s, each 3 H, 2 OMe), 3.74 (br t, 1 H, $J_{6a,6b} = J_{6b,5}$ ~10 Hz, H-6b), 3.53 (dd, 1 H, H-2'), 3.45–3.56 (m, 2 H, H-4, 5), 3.23 (ddd, 1 H, $J_{2,3}$ 10.0 Hz, H-2), 1.73 (s, 3 H, Ac), 1.43, and 1.31 (2 s, each 3 H, Me_2C), 0.69 (d, 3 H, H-6'). Anal. Calcd for C₃₆H₄₇NO₁₂: C, 63.05; H, 6.91; N, 2.04. Found: C, 63.07; H, 6.92; N, 2.02.

Allyl (3,4-O-isopropylidene-2-O-p-methoxybenzyl- α -L-fucopyranosyl)- $(1 \rightarrow 3)$ -2-acetamido-2-deoxy- β -D-glucopyranoside (22).—To a solution of 21 (590 mg, 0.86 mmol) in 9:1 CH₃CN-water (150 mL) was added ceric ammonium nitrate (456 mg, 0.833 mmol), and the mixture was stirred for 1 h at room temperature. Then, NaHCO₃ (0.5 g) was added, and the mixture was concentrated to dryness. The residue was purified by column chromatography (9:1 EtOAc–MeOH) to give 22 (267 mg, 55%), which crystallized from Et₂O-CH₂Cl₂, mp 175–177 °C; $[\alpha]_{\rm D}^{20}$ –45° (c 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 7.31–7.25 (m, 2 H, arom. H), 6.92–6.86 (m, 2 H, arom. H), 5.87 (m, 1 H, CH=), 5.56 (d, 1 H, $J_{\rm NH.2}$ 7.5 Hz, NH), 5.27 (dq, 1 H, = CH_{2trans}), 5.19 (dq, 1 H, = CH_{2cis}), 4.94 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 4.85 (d, 1 H, *J*_{1',2'} 3.5 Hz, H-1'), 4.72 (d, 1 H, J 11.7 Hz, OCH₂Ar), 4.60 (d, 1 H, OCH₂Ar), 4.35 (dq, 1 H, *J*_{5',6'} 6.5, *J*_{5',4'} 2.0 Hz, H-5'), 4.32 (m, 1 H, H-3'), 4.31 (ddt, 1 H, OCH₂), 4.10 (ddt, 1 H, OCH₂), 4.06 (dd, 1 H, J_{4',3'} 6.5 Hz, H-4'), 3.96 (dd, 1 H, J_{3,2} 10.5, J_{3,4} 8.0 Hz, H-3), 3.90 (dd, 1 H, J_{6a,6b} \sim 12.0 Hz, H-6b), 3.81 (s, 3 H, OMe), 3.80 (dd, 1 H, H-6a), 3.58 (dd, 1 H, J_{2',3'} 3.5 Hz, H-2'), 3.48 (m, 1 H, *J*_{4,5} 9.0 Hz, H-4), 3.40 (ddd, 1 H, *J*_{5,6a} 5.0, J_{5,6b} 3.0 Hz, H-5), 3.26 (ddd, 1 H, H-2), 2.20 (br m, 2 H, OH), 1.77 (s, 3 H, Ac), 1.43 and 1.34 (2 s, each 3 H, Me₂C), 1.31 (d, 3 H, H-6'). Anal. Calcd for C₂₈H₄₁NO₁₁: C, 59.25; H, 7.28; N, 2.47. Found: C, 59.26; H, 7.12; N, 2.40.

Allyl (2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-glucopy-(23).—A solution of 21 (15 mg, ranoside 0.021 mmol) in 18:1 CH₂Cl₂-water (9 mL) was vigorously stirred with DDQ (21 mg, 0.093 mmol) at room temperature for 24 h. The solution was taken to dryness, the residue was dissolved in pyridine (6 mL) and stirred with Ac_2O (1.8 mL) for 20 h at room temperature. MeOH (1mL) was added and the solution was concentrated. Purification of the residue on silica gel (EtOAc) gave 23 as colourless syrup. Yield: 12 mg (92%); $[\alpha]_{D}^{20} -57^{\circ}$ (c 0.7, CHCl₃). ¹H NMR (CDCl₃): δ 5.87 (m, 1 H, CH =), 5.70 (d, 1 H, J_{NH,2} 7.1 Hz, NH), 5.31 (dd, 1 H, J_{3',4'} 3.2, $J_{3',2'}$ 11.0 Hz, H-3'), 5.27 (dq, 1 H, = CH_{2trans}), 5.26 (d, 1 H, H-1'), 5.24 (dd, 1 H, J_{5',4'} 1.5 Hz, H-4'), 5.21 (dq, 1 H, = CH_{2cis}), 5.09 (dd, 1 H, $J_{1',2'}$ 3.4 Hz, H-2'), 5.03 (d, 1 H, J_{1,2} 8.0 Hz, H-1), 4.96 (t, 1 H, $J_{3,4} \sim J_{4,5} = 9.3$ Hz, H-4), 4.45 (t, 1 H, $J_{3,2}$ 9.3 Hz, H-3), 4.32 (m, 1 H, OCH₂), 4.19 (dd, 1 H, $J_{6a,6b}$ 12.5, $J_{6a,5}$ 5.5 Hz, H-6a), 4.16 (m, 1 H, H-5'), 4.08 (dd, 1 H, $J_{6b,5}$ 2.9 Hz, H-6b), 4.08 (m, 1 H,

OCH₂), 3.61 (ddd, 1 H, H-5), 3.25 (m, 1 H, H-2), 2.15, 2.11, 2.09, 2.00, 1.99, and 1.98 (6 s, each 3 H, 6 Ac), 1.07 (d, 3 H, H-6'). Anal. Calcd for $C_{27}H_{39}NO_{15}$: C, 52.51; H, 6.36; N, 2.27. Found: C, 52.44; H, 6.38; N, 2.10.

Allyl $(\alpha$ -L-fucopyranosyl)- $(1 \rightarrow 3)$ -2-acetamido-2*deoxy*-β-D-glucopyranoside (24).—A solution of 23 (7.2 mg) in dry MeOH (5 mL) was stirred with 0.1 M methanolic NaOMe (0.5 mL) for 2 h at room temperature. The pH of the solution was adjusted to 7.0 by addition of Dowex 50 (H^+) resin, the resin was filtered off and the filtrate was taken to dryness. Purification of the residue on a Bio-Gel P-2 column (1.6×80 cm, aq 5% EtOH) gave 24 as amorphous solid. Yield: 4.1 mg (87%); $[\alpha]^{20}_{D}$ -105° (c 0.4, H₂O). ¹H NMR (D₂O): δ 5.92 (m, 1 H, CH=), 5.32 (dq, 1 H, =CH_{2trans}), 5.28 (dq, 1 H, = CH_{2cis}), 5.00 (d, 1 H, $J_{1',2'}$ 4.1 Hz, H-1'), 4.59 (d, 1 H, J_{1,2} 8.6 Hz, H-1), 4.37 (m, 1 H, OCH₂), 4.35 (m, 1 H, H-5'), 4.18 (m, 1 H, OCH₂), 3.96 (dd, 1 H, J_{6a,5} 2.2, J_{6a,6b} 12.4 Hz, H-6a), 3.87 (dd, 1 H, H-2), 3.85 (dd, 1 H, $J_{3',2'}$ 10.1, $J_{3',4'}$ 3.2 Hz, H-3'), 3.8 (dd, 1 H, $J_{4',5'}$ 1.0 Hz, H-4'), 3.77 (dd, 1 H, $J_{6b,5}$ 5.4 Hz, H-6b), 3.71 (dd, 1 H, H-2'), 3.66 (dd, 1 H, J_{3.2} 10.1, J_{3.4} 9.8 Hz, H-3), 3.54 (t, 1 H, J_{4.5} 9.8 Hz, H-4), 3.50 (m, 1 H, H-5), 2.04 (s, 3 H, Ac), 1.15 (d, 3 H, *J*_{6',5'} 6.6 Hz, H-6').

Allyl (3,4-O-isopropylidene-2-O-p-methoxybenzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-2-deoxy-6-O-p-methoxybenzyl- β -D-glucopyranoside (25), allyl $(3,4-O-isopropylidene-2-O-p-methoxybenzyl-\beta-L$ fucopvranosvl)- $(1 \rightarrow 3$)-2-acetamido-2-deoxy-6-O-pmethoxybenzyl- β -D-glucopyranoside (26) and allyl $(3,4-O-isopropylidene-2-O-p-methoxybenzyl-\alpha-L$ fucopyranosyl)- $(1 \rightarrow 4)$ -2-acetamido-2-deoxy-6-O-p*methoxybenzyl*-β-D-glucopyranoside (28).—A suspension of **3** (600 mg, 1.57 mmol), **20** (636 mg, 1.73 mmol) and 4 A molecular sieves (1g) in 1:5 dry DMF-CH₂Cl₂ (14 mL) was stirred at room temperature for 75 min under N₂. Iodonium disym-collidinium perchlorate (IDCP, 785 mg, 1.57 mmol) was added in 7 portions over a period of 7h. Stirring was continued for one additional hour, the suspension was diluted with CH₂Cl₂ (50 mL) and filtered through Celite^{\mathbb{R}}. The filtrate was washed twice with aq 5% Na₂S₂O₃, dried (Na₂SO₄), and concentrated. Purification of the residue on silica gel (1:3 toluene-EtOAc) gave 25 as colourless crystals. Yield: 260 mg (32%), mp 162-164 °C (EtOAc-hexane); $[\alpha]^{20}_{D}$ -59° (c 0.5, CHCl₃). ¹H NMR (CDCl₃): δ 7.29–7.25 (m, 4 H, arom. H), 6.89–6.85 (m, 4 H, arom. H), 5.88 (m, 1

H, CH=), 5.51 (d, 1 H, $J_{NH,2}$ 7.7 Hz, NH), 5.25 (dq, 1 H, =CH_{2trans}), 5.18–5.14 (m, 1 H, =CH_{2cis}), 4.89 (d, 1 H, $J_{1,2}$ 8.2 Hz, H-1), 4.86 (d, 1 H, $J_{1',2'}$ 3.5 Hz, H-1'), 4.72 and 4.60 (AB, 2 H, J_{AB} 12.0 Hz, OCH₂Ar), 4.59–4.51 (m, 2 H, OCH₂Ar), 4.36 (dq, 1 H, $J_{5',4'}$ 2.5, $J_{5',6'}$ 6.7 Hz, H-5'), 4.31 (dd, 1 H, $J_{3',2'}$ 6.7, $J_{3',4'}$ 6.0 Hz, H-3'), 4.39–4.29 (m, 1 H, OCH₂), 4.11–4.04 (m, 1 H, OCH₂), 4.05 (dd, 1 H, H-4'), 3.96 (br dd, 1 H, H-3), 3.83–3.80 (m, 8 H, 2 OMe, H-6a,6b), 3.68 (ddd, 1 H $J_{5,6a}$ 5.0, $J_{5,4}$ 10.4 Hz, H-5), 3.75 (dd, 1 H, H-2'), 3.48–3.43 (m, 2 H, H-4, OH), 3.26 (m, 1 H, H-2), 1.76 (s, 3 H, Ac), 1.42 and 1.33 (2 s, each 3 H, Me₂C), 1.31 (d, 3 H, H-6'). Anal. Calcd for C₃₆H₄₉NO₁₂: C, 62.87; H, 7.18; N, 2.04. Found: C, 62.94; H, 7.12; N, 1.99.

The column was then eluted with 5:1 EtOAc-MeOH and the eluate was concentrated. The residue was purified on silica gel (10:10:1 CH₂Cl₂hexane-EtOH), which afforded 26 as colourless crystals. Yield: 33 mg (4%); mp 171–173 °C (CH₂Cl₂-hexane); $[\alpha]^{20}_{D} - 8^{\circ}$ (*c* 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 7.26 (m, 4 H, arom. H), 6.84 (m, 4 H, arom. H), 5.91 (m, 1 H, CH =), 5.78 (d, 1 H, $J_{\rm NH,2}$ 6.8 Hz, NH), 5.27 (dq, 1 H, = CH_{2trans}), 5.17 $(dq, 1 H, = CH_{2cis}), 5.04 (d, 1 H, J_{1,2} 8.1 Hz, H-1),$ 4.85 and 4.66 (AB, 2 H, J_{AB} 11.5 Hz, OCH₂Ar), 4.52 (s, 2 H, OCH₂Ar), 4.41 (d, 1 H, $J_{1',2'}$ 8.3 Hz, H-1'), 4.34 (ddt, 1 H, OCH₂), 4.23 (br dd, 1 H, H-3), 4.18 (dd, 1 H, $J_{3',2'}$ 7.0, $J_{3',4'}$ 5.5 Hz, H-3'), 4.09 (ddt, 1 H, OCH₂), 3.97 (dd, 1 H, J_{4',5'} 2.1 Hz, H-4'), 3.86–3.78 (m, 1 H, H-6a), 3.82 (dq, 1 H, J_{5'.6'} 6.6 Hz, H-5'), 3.79 and 3.76 (2 s, each 3 H, 2 OMe), 3.73 (dd, 1 H, *J*_{6b,5} 2.0, *J*_{6b,6a} 11.0 Hz, H-6b), 3.62– 3.56 (m, 1 H, H-5), 3.52-3.47 (m, 2 H, H-4, OH), 3.43 (dd, 1 H, H-2'), 3.04 (br td, 1 H, H-2), 1.97 (s, 3 H, Ac), 1.44 and 1.34 (2 s, each 3 H, Me₂C), 1.36 (d, 3 H, H-6'). Anal. Calcd for $C_{36}H_{49}NO_{12}$: C, 62.87; H, 7.18; N, 2.04. Found: C, 62.89; H, 7.19; N, 2.00.

Further elution gave **28** as crystals. Yield: 90 mg (11%); mp 160 °C (EtOAc–hexane); $[\alpha]^{20}{}_{\rm D}$ –72° (*c* 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 7.22–7.19 (m, 4 H, arom. H), 6.86–6.81 (m, 4 H, arom. H), 5.90 (m, 1 H, CH=), 5.73 (d, 1 H, $J_{\rm NH,2}$ 6.7 Hz, NH), 5.28 (dq, 1 H, =CH_{2trans}), 5.19 (dq, 1 H, =CH_{2cis}), 4.88 (d, 1 H, $J_{1',2'}$ 3.5 Hz, H-1'), 4.87 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 4.61 and 4.55 (AB, 2 H, $J_{\rm AB}$ 11.0 Hz, OCH₂Ar), 4.43 and 4.38 (AB, 2 H, $J_{\rm AB}$ 11.0 Hz, OCH₂Ar), 4.39 (dq, 1 H, $J_{5',4'}$ 2.5, $J_{5',6'}$ 6.7 Hz, H-5'), 4.40–4.31 (m, 1 H, OCH₂), 4.26 (t, 1 H, $J_{3',4'} = J_{3',2'} = 5.5$ Hz, H-3'), 4.11–4.01 (m, 2 H, H-3, OCH₂), 4.02 (dd, 1 H, H-4'), 3.79 and 3.77 (2 s,

each 3 H, 2 OMe), 3.74 (dd, 1 H, $J_{6a,5}$ 1.9, $J_{6a,6b}$ 10.5 Hz, H-6a), 3.69 (dd, 1 H, $J_{6b,5}$ 3.9 Hz, H-6b), 3.57–3.47 (m, 4 H, H-4,5,2', and OH), 3.31 (ddd, 1 H, $J_{2,3}$ 10.0 Hz, H-2), 2.01 (s, 3 H, Ac), 1.42 and 1.34 (2 s, each 3 H, Me₂C), 1.29 (d, 3 H, H-6'). Anal. Calcd for C₃₆H₄₉NO₁₂: C, 62.87; H, 7.18; N, 2.04. Found: C, 63.08; H, 7.34; N, 1.99. Further elution afforded unreacted **3** (150 mg, 25%).

Acetvlation of 25.—A solution of 25 (5 mg) in pyridine (1 mL) was treated with $Ac_2O(0.1 \text{ mL})$ for 64 h at room temperature. MeOH (0.2 mL) was added and the solution was taken to dryness. Purification of the residue on silica gel (1:2 toluene-EtOAc) furnished 27 as syrup. Yield: 5 mg (95%). ¹H NMR (CDCl₃): δ 7.27–7.23 (m, 4 H, arom. H), 6.91-6.85 (m, 4 H, arom. H), 5.90-5.78 (m, 1 H, CH =), 5.81 (d, 1 H, $J_{NH,2}$ 7.5 Hz, NH), 5.24 (dq, 1 H, = CH_{2trans}), 5.15 (dq, 1 H, = CH_{2cis}), 4.88 (t, 1 H, $J_{4,3} = J_{4,5} = 9.0$ Hz, H-4), 4.78 and 4.58 (AB, 2 H, J_{AB} 11.5 Hz, OCH₂Ar), 4.76 (d, 1 H, $J_{1',2'}$ 3.0 Hz, H-1), 4.64 (d, 1 H, J_{1,2} 8.3 Hz, H-1), 4.44 and 4.47 (AB, 2 H, J_{AB} 11.0 Hz, OCH₂Ar), 4.32 (ddt, 1 H, OCH₂), 4.20 (dd, 1 H, $J_{3',2'}$ 8.0, $J_{3',4'}$ 5.5 Hz, H-3'), 4.15 (dq, 1 H, J_{5',6'} 6.7 Hz, H-5'), 4.07 (ddt, 1 H, OCH₂), 3.97 (dd, 1 H, H-4'), 3.81 and 3.80 (2 s, each 3 H, 2 OMe), 3.82–3.76 (m, 1 H, H-3), 3.56–3.51 (m, 4 H, H-2,5,6a,6b), 3.47 (dd, 1 H, H-2'), 1.98 (s, 6 H, 2 Ac), 1.44 and 1.33 (2 s, each 3 H, Me₂C), 1.23 (d, 3 H, H-6').

Acetylation of 28.—Compound 28 (5 mg) was treated as described for 27 to give 5 mg (95%) of 29as a syrup. ¹H NMR (CDCl₃): δ 7.25–7.18 (m, 4 H, arom. H), 6.87-6.82 (m, 4 H arom. H), 5.86 (m, 1 H, CH=), 5.45 (d, 1 H, $J_{\rm NH,2}$ 9.0 Hz, NH), 5.26 $(dq, 1 H, = CH_{2trans}), 5.17 (dq, 1 H, = CH_{2cis}),$ 5.03 (dd, 1 H, J_{3,2} 10.5, J_{3,4} 8.5 Hz, H-3), 4.81 (d, 1 H, $J_{1',2'}$ 3.5 Hz, H-1'), 4.67 and 4.53 (AB, 2 H, J_{AB} 11.5 Hz, OCH₂Ar), 4.47 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.41 and 4.35 (AB, 2 H, J_{AB} 11.7 Hz, OCH₂Ar), 4.32 (ddt, 1 H, OCH₂), 4.15 (dd, 1 H, J_{3',2'} 8.0, J_{3',4'} 5.5 Hz, H-3'), 4.11–4.03 (m, 2 H, H-5', OCH₂), 4.01-3.90 (m, 3 H, H-2,6a,4'), 3.79 and 3.78 (2 s, each 3 H, 2 OMe), 3.67 (dd, 1 H, $J_{6b,5}$ 5.5, $J_{6b,6a}$ 9.5 Hz, H-6b), 3.63 (dd, 1 H, J_{4,5} 9.5 Hz, H-4), 3.52 (ddd, 1 H, *J*_{5,6a} 2.0 Hz, H-5), 3.42 (dd, 1 H, H-2'), 2.06 and 1.92 (2 s, each 3 H, 2 Ac), 1.42 and 1.33 (2 s, each 3 H, Me₂C), 1.23 (d, 3 H, H-6').

Allyl (2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 4)-2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranoside (**30**).—A solution of **28** (28 mg, 0.041 mmol) in CH₂Cl₂ (7 mL) was cooled to -20 °C and treated with aq 90% CF₃CO₂H (0.175 mL) for 2.5 h. After concentration, the residue was dissolved in pyridine (6 mL) and treated with Ac_2O (3 mL) for 96 h. MeOH (5 mL) was added at 0 °C and the solution was concentrated. Purification of the residue on silica gel (EtOAc) furnished **30** as a syrup. Yield: 21 mg (83%); $[\alpha]^{20}_{D}$ -93° (c 0.8, CHCl₃). ¹H NMR (CDCl₃): δ 5.86 (m, 1 H, CH =), 5.46 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 5.31– 5.10 (m, 6 H, H-1', 2', 3', 4', and = CH₂), 5.08 (dd, 1 H, $J_{3,2}$ 10.5, $J_{3,4}$ 8.0 Hz, H-3), 4.66 (dd, 1 H, $J_{6a,5}$ 2.0, J_{6a.6b} 11.5 Hz, H-6a), 4.55 (d, 1 H, J_{1.2} 8.5 Hz, H-1), 4.32 (ddt, 1 H, OCH₂), 4.13 (dd, 1 H, J_{6b,5} 5.5 Hz, H-6b), 4.10 (dd, 1 H, *J*_{5',4'} 1.5, *J*_{5',6'} 6.5 Hz, H-5'), 4.08 (ddt, 1 H, OCH₂), 3.92 (ddd, 1 H, H-2), 3.71–3.63 (m, 1 H, H-5), 3.60 (dd, 1 H, J_{4.5} 9.5 Hz, H-4), 2.15, 2.11, 2.09, 2.07, 1.98, and 1.94 (6 s, each 3 H, 6 Ac), 1.06 (d, 3 H, H-6'). Anal. Calcd for C₂₇H₃₉NO₁₅: C, 52.51; H, 6.36; N, 2.27. Found: C, 52.64; H, 6.28; N, 2.18.

Allyl (α -L-fucopyranosyl)-(1 \rightarrow 4)-2-acetamido-2deoxy- β -D-glucopyranoside (31).—A solution of 30 (9.8 mg) in dry MeOH (6 mL) was stirred with 0.1 M methanolic NaOMe (0.8 mL) for 24 h at room temperature. The pH of the solution was adjusted to 7.0 by addition of Dowex 50 (H^+) resin, the resin was filtered off, and the filtrate was taken to dryness to give 31 as an amorphous solid. Yield: 6.5 mg (~ quant.); $[\alpha]^{20}_{D} - 99^{\circ}$ (c 0.7, H₂O). ¹H NMR (D₂O): δ 5.91 (m, 1 H, CH =), 5.34–5.23 (m, 2 H, = CH₂), 4.95 (d, 1 H, $J_{1',2'}$ 3.5 Hz, H-1'), 4.56 (d, 1 H, J_{1.2} 8.0 Hz, H-1), 4.33 (ddt, 1 H, OCH₂), 4.34 (dq, 1 H, *J*_{5',4'} 1.5, *J*_{5',6'} 6.5 Hz, H-5'), 4.15 (ddt, 1 H, OCH₂), 3.98 (dd, 1 H, J_{6a.5} 1.8, $J_{6a,6b}$ 12.5 Hz, H-6a), 3.87–3.80 (m, 2 H, H-6b,2'), 3.80 (dd, 1 H, *J*_{4',3'} 3.5 Hz, H-4'), 3.72 (dd, 1 H, *J*_{2,3} 10.0 Hz, H-2), 3.77-3.50 (m, 4 H, H-3,4,5,3'), 2.03 (s, 3 H, Ac), 1.15 (d, 3 H, H-6'). ES-MS $(C_{17}H_{29}NO_{10}): m/z 407.9 (M+H)^+.$

Allyl (3,4-O-isopropylidene-2-O-p-methoxy $benzyl-<math>\alpha$ -L-fucopyranosyl)- $(1\rightarrow 3)$ -[(3,4,6-tri-O-acetyl- $2-deoxy-2-trichloroacetamido-\beta-D-galactopyrano$ $syl-<math>(1\rightarrow 4)$]-2-acetamido-2-deoxy-6-O-p-methoxybenzyl- β -D-glucopyranoside (33).—A suspension of 25 (183 mg, 0.266 mmol), 9 (240 mg, 0.403 mmol), N,N,N',N'-tetramethyl urea (0.035 ml, 0.292 mmol) and 4 Å molecular sieves (500 mg) in CH₂Cl₂ (8 mL) containing toluene (0.4 mL) was stirred for 45 min at room temperature. Then Sn(OTf)₂ (146 mg, 0.348 mmol) was added in two portions during 30 min, and the suspension was stirred for one additional hour. Et₃N (0.075 mL) and CH₂Cl₂ (50 mL) were added, solids were removed by filtration, and the filtrate was washed with satd aq NaHCO₃, dried (Na₂SO₄), and concentrated. Purification of the residue on silica gel (1:2 toluene– EtOAc) gave 2-trichloromethyl-4,5-dihydro-(3,4, 6-tri-*O*-acetyl-1,2-dideoxy- α -D-galactopyranoso)-[2,1-d]-1,3-oxazole (**32**) as a syrup. Yield: 65 mg (36%); [α]²⁰_D+108° (*c* 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 6.33 (d, 1 H, $J_{1,2}$ 7.0 Hz, H-1), 5.48 (dd, 1 H, $J_{4,3}$ 3.7, $J_{4,5}$ 2.5 Hz, H-4), 4.99 (dd, 1 H, $J_{3,2}$ 7.5 Hz, H-3), 4.31 (td, 1 H, $J_{5,6a}$ = $J_{5,6b}$ =6.5 Hz, H-5), 4.29 (dd, 1 H, H-2), 4.22 (dd, 1 H, $J_{6a,6b}$ 11.5 Hz, H-6a), 4.15 (dd, 1 H, H-6b), 2.15, 2.09, and 2.08 (3 s, each 3 H, 3 Ac).

Further elution afforded 33 as a syrup. Yield: 90 mg (41%); $[\alpha]^{20}_{D}$ -67° (c 0.6, CHCl₃). ¹H NMR (CDCl₃): δ 7.31–7.25 (m, 4 H, arom. H), 6.94–6.91 (m, 2 H, arom. H), 6.88-6.85 (m, 2 H, arom. H), 6.46 (d, 1 H, J_{NH",2"} 9.0 Hz, NH"), 6.08 (d, 1 H, $J_{\rm NH,2}$ 8.0 Hz, NH), 5.83 (m, 1 H, CH=), 5.22 (dd, 1 H, $J_{4'',3''}$ 3.5, $J_{4'',5''}$ 1.0 Hz, H-4"), 5.23 (dq, 1 H, = CH_{2trans}), 5.13 (dq, 1 H, = CH_{2cis}), 5.06 (d, 1 H, $J_{1',2'}$ 3.4 Hz, H-1'), 4.98 (dd, 1 H, $J_{3'',2''}$ 11.3 Hz, H-3"), 4.78–4.73 (m, 2 H, H-1, OCH₂Ar), 4.63 (d, 1 H, J_{AB} 12.0 Hz, OCH₂Ar), 4.63 and 4.36 (AB, 2 H, J_{AB} 12.0 Hz, OCH₂Ar), 4.59 (d, 1 H, J_{1".2"} 9.0 Hz, H-1"), 4.53 (dq, 1 H, $J_{5',4'}$ 2.5, $J_{5',6'}$ 6.7 Hz, H-5'), 4.25 (dd, 1 H, *J*_{3',2'} 7.5, *J*_{3',4'} 6.0 Hz, H-3'), 4.20 (dd, 1 H, J_{6a",5"} 8.0, J_{6a",6b"} 11.0 Hz, H-6a"), 4.28–4.17 (m, 1 H, OCH₂), 4.10–3.96 (m, 6 H, H-3,4,4',2",6b", and OCH₂), 3.81 and 3.80 (2 s, each 3 H, 2 OMe), 3.74–3.60 (m, 4 H, H-2,6a,6b,5"), 3.57 (dd, 1 H, H-2'), 3.54–3.49 (m, 1 H, H-5), 2.15, 2.06, 1.99, and 1.90 (4 s, each 3 H, 4 Ac), 1.43 and 1.35 (2 s, each 3 H, Me₂C), 1.35 (d, 3 H, H-6'). Anal. Calcd for C₅₀H₆₅Cl₃N₂O₂₀: C, 53.59; H, 5.85; N, 2.50. Found: C, 53.57; H, 6.02; N, 2.33. Further elution of the column gave unreacted starting material **25** (47 mg, 26%).

Allyl (2-O-acetyl-3,4-O-isopropylidene- α -L-fucopyranosyl)- $(1\rightarrow 3)$ -[(3,4,6-tri-O-acetyl-2-deoxy-2trichloroacetamido- β -D-galactopyranosyl)- $(1\rightarrow 4)$]-2-acetamido-6-O-acetyl-2-deoxy- β -D-glucopyranoside (34).—A solution of 33 (40 mg, 0.036 mmol) and DDQ (41 mg, 0.18 mmol) in 18:1 CH₂Cl₂-H₂O (10 mL) was vigorously stirred for 7 h at room temperature. The reaction was quenched by addition of aq 20% Na₂S₂O₃ (2.5 mL), the solution was extracted with EtOAc, the organic layer was dried (Na₂SO₄), and concentrated. The residue was purified by flash-chromatogrphy on silica gel (24:1 EtOAc-MeOH). The product obtained (28.5 mg) was dissolved in pyridine (6 mL) and treated with Ac₂O (0.7 mL) for 24 h at 0 °C. MeOH (2 mL) was added and the solution was taken to dryness. Chromatography of the residue on silica gel (1:5 toluene-EtOAc) gave 34 as a syrup. Yield: 30 mg $(96\%); \ [\alpha]_{D}^{20} -91^{\circ} \ (c \ 0.2, \ \text{CHCl}_3).$ ¹H NMR (CDCl₃): δ 7.08 (d, 1 H, J_{NH",2"} 9.0 Hz, NH"), 5.90 (d, 1 H, $J_{\rm NH,2}$ 9.0 Hz, NH), 5.83 (m, 1 H, CH=), 5.38 (dd, $J_{4'',3''}$ 3.5, $J_{4'',5''}$ 0.8 Hz, H-4''), 5.29 (d, 1 H, $J_{1',2'}$ 3.5 Hz, H-1'), 5.29–5.22 (m, 1 H, = CH_{2trans}), 5.19–5.12 (m, 1 H, = CH_{2cis}), 5.14 (dd, 1 H, $J_{3'',2''}$ 11.5 Hz, H-3"), 4.84 (dd, 1 H, $J_{2',3'}$ 8.0 Hz, H-2'), 4.58–4.56 (m, 1 H, H-5'), 4.56 (d, 1 H, H-1), 4.47 (d, 1 H, *J*_{1",2"} 8.0 Hz, H-1"), 4.43 (dd, 1 H, J_{6a,5} 5.5, J_{6a,6b} 12.0 Hz, H-6a), 4.33 (dd, 1 H, J_{6b,5} 4.0 Hz, H-6b), 4.30 (dd, 1 H, $J_{3',4'}$ 5.0 Hz, H-3'), 4.29–4.21 (m, 3 H, H-2",6a", and OCH₂), 4.15 (dd, 1 H, *J*_{6b",6a"} 11.0 Hz, H-6b"), 4.19–4.14 (m, 1 H, H-4'), 4.04–3.96 (m, 3 H, H-2,3, and OCH₂), 3.90 (td, 1 H, *J*_{5",6a"} 6.5 Hz, H-5"), 3.83–3.79 (m, 1 H, H-4), 3.72–3.68 (m, 1 H, H-5), 2.19, 2.13, 2.128, 2.08, 2.02, and 2.01 (6 s, each 3 H, 6 Ac), 1.52 and 1.36 $(2 \text{ s, each 3 H, Me}_2\text{C}), 1.42 \text{ (d, 3 H, } J_{6',5'} \text{ 6.7 Hz, H-}$ 6'). Anal. Calcd for C₃₈H₅₃Cl₃N₂O₂₀: C, 47.33; H, 5.54; N, 2.90. Found: C, 46.86; H, 5.21; N, 2.75.

Allyl $(2,3,4-tri-O-acetyl-\alpha-L-fucopyranosyl)$ - $(1 \rightarrow 3)$ -[(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl)- $(1 \rightarrow 4)$]-2-acetamido-6-O-acetyl-2-deoxy- β -D-glucopyranoside (35).—A solution of **34** (25 mg, 0.026 mmol) in CH_2Cl_2 (10 mL) was treated with aq 90% CF₃CO₂H (0.2 mL) at -20 °C for 2h. The solution was evaporated to dryness. The residue was dissolved in EtOH (8 mL) and stirred with NaBH₄ (51 mg, 1.35 mmol) for 24 h at 60 °C. After addition of EtOH (10 mL) the solution was cooled to 0 °C and HOAc (0.076 mL) was added. The solution was coevaporated 5 times with MeOH (10 mL). The residue was dissolved in pyridine (8 mL) and stirred with Ac₂O (4 mL) for 96 h at 0 °C. MeOH (5 mL) was added and the solution was concentrated. Purification of the residue on silica gel (24:1 EtOAc-MeOH) furnished 14 mg (60%) of **35** as syrup; $[\alpha]^{20}_{D} - 76^{\circ}$ (*c* 0.3, MeOH). ¹H NMR (CDCl₃): δ 5.95 (d, 1 H, $J_{\text{NH},2}$ 9.0 Hz, NH), 5.88 (d, 1 H, *J*_{NH",2"} 9.0 Hz, NH"), 5.85 (m, 1 H, CH =), 5.41 (d, 1 H, $J_{1',2'}$ 4.0 Hz, H-1'), 5.37– 5.36 (m, 2 H, H-4',4"), 5.30–5.22 (m, 1 H, = CH_{2trans}), 5.20–5.36 (m, 1 H, = CH_{2cis}), 5.20 (dd, 1 H, J_{3',2'} 11.0, J_{3',4'} 3.5 Hz, H-3'), 5.06 (dd, 1 H, H-2'), 4.98 (dd, 1 H, $J_{3'',2''}$ 11.0, $J_{3'',4''}$ 3.5 Hz, H-3''), 4.69 (m, 1 H, H-5'), 4.60 (d, 1 H, J_{1,2} 6.0 Hz, H-1), 4.45 (dd, 1 H, J_{6a.5} 3.5, J_{6a.6b} 11.5 Hz, H-6a), 4.41 (dd, 1 H, J_{6a",5"} 6.5, J_{6a",6b"} 11.5 Hz, H-6a"), 4.32 (dd, 1 H, $J_{6b,5}$ 5.5 Hz, H-6b), 4.31 (d, 1 H, $J_{1'',2''}$ 8.5 Hz, H-1''), 4.28 (dd, 1 H, $J_{6b'',5''}$ 6.5 Hz, H-6b''), 4.20 (ddd, 1 H, H-2''), 4.31–4.23 (m, 1 H, OCH₂), 4.06 (t, 1 H, $J_{3,2}=J_{3,4}=7.0$ Hz, H-3), 4.05–3.99 (m, 1 H, OCH₂), 3.91 (m, 1 H, H-2), 3.84 (m, 2 H, $J_{4,5}$ 7.0 Hz, H-4,5), 3.67 (m, 1 H, H-5), 2.19, 2.15, 2.149, 2.094, 2.087, 2.08, 2.014, 2.01, and 1.97 (9 s, each 3 H, 9 Ac), 1.24 (d, 3 H, $J_{6',5'}$ 6.6 Hz, H-6'). Anal. Calcd for C₃₉H₅₆N₂O₂₂: C, 51.76; H, 6.24; N, 3.09. Found: C, 52.25; H, 6.08; N, 3.14.

Allyl $(\alpha$ -L-fucopyranosyl)- $(1 \rightarrow 3)$ -[(2-acetamido-2-deoxy- β -D-galactopyranosyl)- $(1 \rightarrow 4)$]-2-acetamido-2-deoxy- β -D-glucopyranoside (36).—A solution of 35 (9.0 mg, 0.010 mmol) in MeOH (2 mL) was stirred with 0.1 M methanolic NaOMe (0.8 mL) for 24 h at ambient temperature. Dowex (H^+) resin was added to adjust the pH of the solution to 7.0. The resin was filtered off, and the filtrate was concentrated to give 6.0 mg (99%) of 36 as amorphous solid; $[\alpha]_{D}^{20} - 69^{\circ}$ (c 0.6, H₂O). ¹H NMR $(500 \text{ MHz}, \text{ D}_2\text{O}): \delta 5.84 \text{ (m, 1 H, CH} =), 5.24 \text{ (dq,})$ $1 \text{ H}_{1} = \text{CH}_{2\text{trans}}$, 5.20 (dq, $1 \text{ H}_{2\text{cis}}$), 5.05 (d, 1 H, $J_{1',2'}$ 4.0 Hz, H-1'), 4.79 (m, 1 H, H-5'), 4.49 (d, 1 H, *J*_{1,2} 8.4 Hz, H-1), 4.40 (d, 1 H, *J*_{1",2"} 8.4 Hz, H-1"), 4.26 (ddt, 1 H, OCH₂), 4.08 (ddt, 1 H, OCH₂), 3.91 (dd, 1 H, J_{2",3"} 10.8 Hz, H-2"), 3.88 (dd, 1 H, $J_{3',4'}$ 3.3 Hz, H-3'), 3.87 (dd, 1 H, $J_{2,3}$ 12.2 Hz, H-2), 3.88–3.85 (m, 2 H, H-6a,4"), 3.82 (t, 1 H, $J_{4,3} = J_{4,5} = 9.0$ Hz, H-4), 3.78 (t, 1 H, H-3), 3.78 (m, 1 H, H-4'), 3.71 (dd, 1 H, $J_{6a'',5''}$ 7.8, $J_{6a'',6b''}$ 11.6 Hz, H-6a''), 3.68 (dd, 1 H, $J_{6b,5}$ 5.2, J_{6a,6b} 12.3 Hz, H-6b), 3.68 (dd, 1 H, J_{6b",5"} 4.4 Hz, H-6b"), 3.66 (dd, 1 H, *J*_{3",4"} 3.3 Hz, H-3"), 3.64 (dd, 1 H, $J_{2',3'}$ 10.1 Hz, H-2'), 3.52 (dd, 1 H, $J_{5'',4''}$ <1.0 Hz, H-5"), 3.45 (ddd, 1 H, *J*_{5.6a} 2.3 Hz, H-5), 1.99 and 1.96 (2 s, each 3 H, 2 Ac), 1.20 (d, 3 H, $J_{6',5'}$ 6.6 Hz, H-6'). ES-MS (C₂₅H₄₂N₂O₁₅): m/z $611.3 (M + H)^+$.

3-(2-Aminoethylthio)propyl (α-L-fucopyranosyl)-(1→3)-[(2-acetamido-2-deoxy-β-D-galactopyanosyl)-(1→4)]-2-acetamido-2-deoxy-β-D-glucopyranoside (**37**).—A solution of **36** (5.0 mg, 8.2 µmol) and cysteamine hydrochloride (7.0 mg, 0.62 mmol) in water (0.2 mL) was irradiated at 254 nm for 6 h at room temperature. The solution was passed through a column of Dowex AGWX8 resin (NH₄⁺ form) using 0.1 M aq NH₃ as eluent. Carbohydrate-containing fractions were pooled, lyophilized and further purified on Sephadex G-10 (0.05 M aq NH₄Ac) to give, after lyophilization, **37** as amorphous powder. Yield: 5.9 mg (96%); [*a*]²⁰_D -46° (*c* 0.5, H₂O). ¹H NMR (D₂O): δ 5.10 (d, 1 H,

J_{1',2'} 3.9 Hz, H-1'), 4.83 (br d, 1 H, H-5'), 4.48 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.45 (d, 1 H, $J_{1'',2''}$ 8.3 Hz, H-1"), 3.99–3.82 (m, 8 H, H-2,6a,3',4',2",3",4", and OCH₂), 3.80-3.62 (m, 7 H, H-3,4,6b,2',6a",6b", and OCH₂), 3.56 (dd, 1 H, J_{5",6a}" 4.5, J_{5",6b}" 7.6 Hz, H-5"), 3.53-3.48 (m, 1 H, H-5), 3.19 (t, 2 H, CH₂N), 2.82 (t, 2 H, SCH₂), 2.60 (t, 2 H, CH₂S), 2.04 and 2.03 (2 s, each 3 H, 2 Ac), 1.84 (br t, 2 H, CH₂), 1.25 (d, 3 H, $J_{6',5'}$ 6.6 Hz, H-6'). ¹³C NMR (D_2O) : δ 175.73 (C=O), 175.01 (C=O), 101.94 (C-1"), 101.63 (C-1), 99.32 (C-1'), 76.34 (C-5), 75.73 (C-5"), 75.55 (C-3), 74.26 (C-4), 72.89 (C-4'), 71.62 (C-3"), 70.08 (OCH₂), 69.57 (C-3'), 68.61 (C-2'), 68.25 (C-4"), 67.82 (C-5'), 62.34 (C-6"), 60.92 (C-6), 56.55 (C-2), 53.27 (C-2"), 39.55 (CH₂N), 30.40 and 29.41 (2 CH₂S), 27.97 (CH₂), 23.18 and 23.08 (2 CH₃), 16.25 (C-6'). ES-MS (C₂₇H₄₉N₃O₁₅S): m/z $688.3 (M+H)^+$.

Synthesis of BSA-conjugate 38.—A solution of thiophosgene (1 µL, 13 µmol) in CHCl₃ (1 mL) was added to a solution of compound 37 (3.7 mg, $5 \mu mol)$ in 0.1 M aq NaHCO₃ (1.5 mL) and the mixture was vigorously stirred for 3h at room temperature. The organic phase was separated and the aqueous phase was washed three times with $CHCl_3$ (2 mL portions) and finally purged with N_2 until a clear solution was obtained. The solution was transferred to a solution of BSA (SIGMA[®], 4.0 mg in 0.1 M aq NaHCO₃-0.3 M NaCl (1 mL)and stirred for 48 h at ambient temperature, then passed through a Sephadex G-25 column $(1.6 \times 50 \text{ cm}, 0.01 \text{ M} \text{ aq}, \text{NaHCO}_3)$. Ninhydrinpositive fractions were combined and dialysed twice against water (2 L) for 24 h. Lyophilization gave the BSA-conjugate 38 as a fluffy white powder. Yield: 4.6 mg.

Acknowledgements

The authors gratefully acknowledge financial support by the Austrian Science Foundation (FWF-grant P 9038 MOB) and the Jubiläumsfonds der Österreichischen Nationalbank (Project N5427). Furthermore, the authors thank Frank Unger for helpful discussions, Andreas Hofinger for recording NMR spectra and Thomas Dalik for technical assistance. Finally, accessibility and use of NMR and mass spectrometry facilities at the Department of Organic Chemistry, Eidgenössische Technische Hochschule Zürich (Group of Professor Diederich) is gratefully acknowledged.

References

- M. Otha, F. Matsuura, Y. Kobayashi, S. Shigeta, K. Ono, and S. Oka, *Arch. Biochem. Biophys.*, 290 (1991) 474–483.
- [2] V. Kubelka, F. Altmann, E. Staudacher, V. Tretter, L. März, K. Hård, J.P. Kamerling, and J.F.G. Vliegenthart, *Eur. J. Biochem.*, 213 (1993) 1193– 1204.
- [3] V. Kubelka, F. Altmann, and L. März, *Glyco-conjugate J.*, 12 (1995) 77–83.
- [4] G. Lochnit and R. Geyer, Eur. J. Biochem., 228 (1995) 805–816.
- [5] G. Strecker, J.M. Wieruszeski, M.D. Fontaine, and Y. Plancke, *Glycobiology*, 4 (1994) 605–609.
- [6] J. Srivatsan, D.F. Smith, and R.D. Cummings, *Glycobiology*, 2 (1992) 445–452.
- [7] R.A. Siciliano, H.R. Morris, R.A. McDowell, P. Azadi, M.E. Rogers, H.P.J. Bennett, and A. Dell, *Glycobiology*, 3 (1993) 225–239.
- [8] A.A. Bergwerff, J.E. Thomas-Oates, J. van Oostrum, J.P. Kamerling, and J.F.G. Vliegenthart, *FEBS Lett.*, 314 (1992) 389–394.
- [9] S.B. Yan, Y.B. Chao, and H. van Halbeek, *Glyco*biology, 3 (1993) 597–608.
- [10] B.W. Grinnell, R.B. Hermann, and S.B. Yan, *Gly-cobiology*, 4 (1994) 221–225.
- [11] A.A. Bergwerff, J.A. van Kuik, W.E.C.M. Schiphorst, C.A.M. Koeleman, D.H. Van den Eijnden, J.P. Kamerling, and J.F.G. Vliegenthart, *FEBS Lett.*, 334 (1993) 133–138.
- [12] V. Poszgay, J.R. Brisson, and H.J. Jennings, Carbohydr. Res., 205 (1990) 133–146.
- [13] R. Johansson and B. Samuelsson, J. Chem. Soc. Perkin Trans I, (1984) 2371–2374.
- [14] G. Blatter, J.M. Beau, and J.C. Jacquinet, *Carbohydr. Res.*, 260 (1994) 189–202; C. Coutant and J.C. Jacquinet, *J. Chem. Soc. Perkin Trans. I*, (1995) 1573–1581.
- [15] N.K. Kochetkov, V.A. Derevitskaya, and O.S. Novikova, *Bull. Acad. Sci. USSR*, 29 (1980) 302– 305.
- [16] G. Excoffier, D. Gagnaire, and J.P. Utille, *Carbo-hydr. Res.*, 39 (1975) 368–373.
- [17] R.R. Schmidt, J. Michel, and M. Roos, *Liebigs Ann. Chem.*, (1984) 1343–1357.
- [18] T. Mukayama and K. Matsubara, Chem. Lett., (1992) 1755–1758.
- [19] F. Weygand and E. Frauendorfer, *Chem. Ber.*, 103 (1970) 2437–2449.
- [20] N.E. Nifant'ev, V.Y. Amochaeva, A.S. Shashkov, and N.K. Kochetkov, *Carbohydr. Res.*, 242 (1993) 77–89.
- [21] Y. Hasegawa, T. Kawada, Y. Kikkawa, and T. Sakuno, *Mokuzai Gakkaishi*, 41, (1995) 83–91.

- [22] C. Steindl, P. Kosma, L. März, and A. Neszmélyi, *Carbohydr. Res.*, 246 (1993) 353–360.
- [23] P. Smid, G.A. de Ruiter, G.A. van der Marel, F.M. Rombouts, and J.H. van Boom, J. Carbohydr. Chem., 10 (1991) 833–849.
- [24] S.S. Rana and K.L. Matta, Carbohydr. Res., 117 (1983) 101–111.
- [25] R.R. Schmidt and A. Toepfer, *Tetrahedron Lett.*, 32 (1991) 3353–3356.
- [26] A. Lubineau and A. Malleron, *Tetrahedron Lett.*, 26 (1985) 1713–1716.
- [27] R.T. Lee and Y.C. Lee, *Carbohydr. Res.*, 37 (1974) 193–201.
- [28] R. Roy and C.A. Laferrière, J. Chem. Soc. Chem. Commun., (1990) 1709–1711.
- [29] M.M. Bradford, Anal. Biochem., 72 (1976) 248-254.
- [30] F. Altmann, Anal. Biochem., 204 (1992) 215-219.