

Note

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J. Org. Chem., **Just Accepted Manuscript** • DOI: 10.1021/acs.joc.9b02318 • Publication Date (Web): 09 Oct 2019

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Decarboxylative Alkylation of Heteroarenes using *N*-Hydroxybenzimidoyl Chloride Esters

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Supporting Information Placeholder

Imine versus amine: A general acid-free strategy to generate a portfolio of alkyl radicals, and it can be applied in Minisci reaction of heteroarenes under mild photocatalytic conditions.



- New radical precursor
- Portfolio of alkyl sources
- Versatile heteroarenes
- General functionalization of peptides

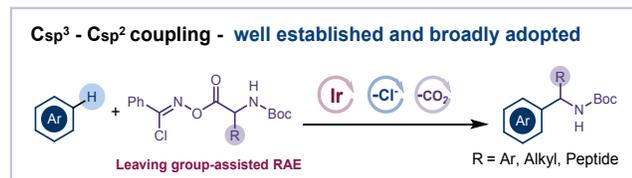
ABSTRACT: Functionalized *N*-heteroarenes are highly desired motifs in medicinal chemistry and pharmaceutical industry. Minisci-type reaction usually requires a protonated *N*-heteroarene for alkyl radical to attack. This work describes a leaving group assisted redox-active ester to enable direct coupling of amino acid with *N*-heteroarenes. The efficient and sustainable photoredox strategy provides a rapid access to alkylated heterocyclic manifold.

Introduction

Minisci-type additions of radical nucleophiles represent an attractive and versatile method for the rapid functionalization of heteroarenes.¹ Recent advances describe a variety of convenient radical precursors spawned under mild conditions.^{1c,1d,2} Particularly, photoredox catalysis³ has provided great opportunities for radical generation in Minisci-type additions with precursors that include alcohols,⁴ ethers,⁵ boronic acids,⁶ carboxylic acids,⁷ and redox-active esters (RAEs).⁸ A significant progress has been made with α -aminoalkylation of *N*-heteroarenes using α -amino acids and peptides. α -Aminoalkyl radicals could be readily generated *via* the activation of *N*-(acyloxy)-phthalimide⁹ in combination with photoredox and homolytically attacks protonated heteroarenes. Phipps developed enantioselective Minisci addition using chiral Brønsted acid catalyst.¹⁰

We recently developed a leaving group-assisted redox-active ester for photoinduced fluoroalkylations using *N*-hydroxybenzimidoyl chloride ester.¹¹ We envisioned that by simply installing a chlorine to oxime, this stabilized imine rivals with the electron-rich nitrogen of amino-acid, in favor of generating iminyl anion and α -amino carbon radical *via* decarboxylative fragmentation (Figure 1). Thus, α -amino alkylation of basic heteroarenes could furnish under similar photoredox strategy of fluoroalkylation. We herein reported a highly efficient Minisci addition using the newly developed *N*-hydroxybenzimidoyl chloride ester (NHBC) to activate amino acid.

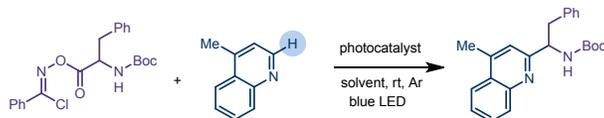
Figure 1 Functionalization of *N*-heteroarenes using Redox-active esters



Results and Discussion

Our experiments began with a brief survey of photocatalysts with NHBC ester and quinoline (Table 1). This study revealed that $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbby})]\text{PF}_6$ was able to promote the desired radical alkylation sequence (entry 1). Other iridium and ruthenium catalysts delivered poor conversions (entries 3-6). Phosphoric acid could not promote the radical addition (entry 7). The reaction efficiency was reduced in air (entry 8).

Table 1 Optimization of the reaction conditions^{a,b}

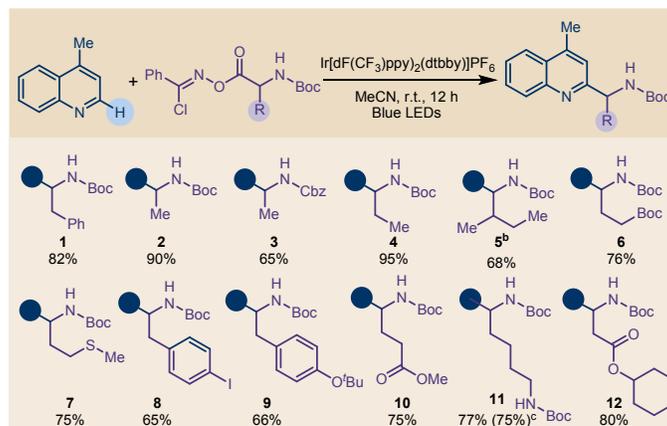


| Entry | Variations from conditions ^a | Yield |
|----------------|--|-------|
| 1 | none | 70% |
| 2 | 4CzIPN | 20% |
| 3 | $[\text{Ru}(\text{bpy})_3](\text{PF}_6)_2$ | 0% |
| 4 | $\text{Ir}(\text{ppy})_3$ | 45% |
| 5 | $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{bpy})]\text{PF}_6$ | 50% |
| 6 ^b | $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbby})]\text{PF}_6$ | 82% |
| 7 ^b | PA added | 80% |
| 8 | In air | 67% |

^a Heteroarene (0.40 mmol), NHBC esters (0.20 mmol), photocatalyst (2 mol%), in MeCN (2.0 mL), irradiated by 10 W blue LEDs for 12 h under Ar; ^b 60W blue LEDs used, PA: 1,1'-binaphthyl-2,2'-diyl hydrogenphosphate.

With the optimized reaction in hand, we studied the scope of the reaction with respect to the amino acid coupling partners. The reaction proceeds at room temperature without additional acid or oxidant, the reaction exhibits excellent functional group compatibility (Scheme 1). Various natural amino acid-derived redox-active esters are suitable substrates, including phenylalanine **1**, alanine **2** & **3**, isoleucine **5**, methionine **7**, tyrosine **9**, glutamic acid **10**, lysine **11**, aspartic acid **12**. Unnatural amino acids **4**, **6** and **8** are also tolerated under these conditions. Typical *N*-protecting groups in peptide chemistry, such as the tert-butyloxycarbonyl (-BOC) and benzyloxycarbonyl (-Cbz) groups, were suitable for their *N*-protection. The sulfide moiety in methionine, which is incompatible in the typical Minisci conditions, was well tolerated under this framework. This demonstrated the advantage of photoredox catalysis for selectively generating radicals while avoiding the use of stoichiometric oxidant.

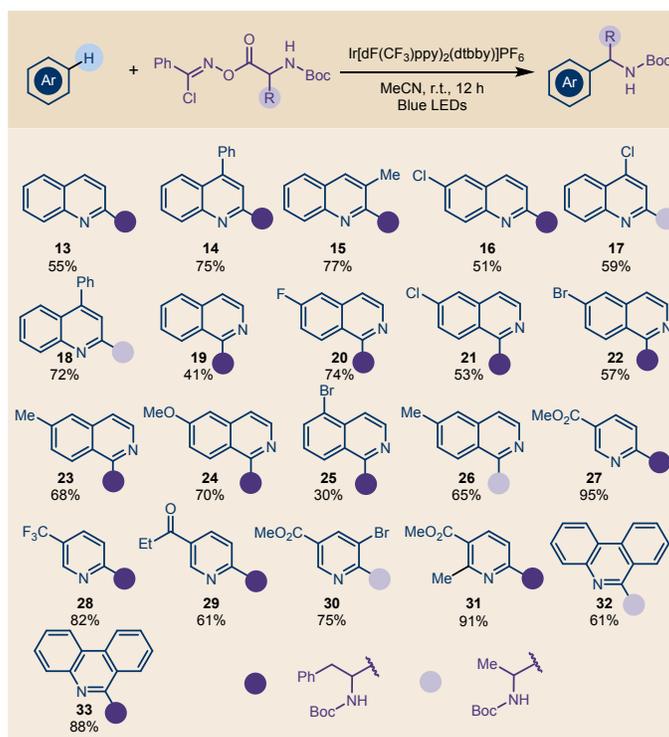
Scheme 1. Amino acid scope for the photocatalytic amination^{a,b,c}



^a Heteroarene (0.40 mmol), NHBC esters (0.20 mmol), photocatalyst (2 mol%), in MeCN (2.0 mL), irradiated by 60 W blue LEDs for 12 h under Ar. ^b 1:1 diastereomeric ratio. ^c Gram-scale; 2.5 mmol NHBC esters was added.

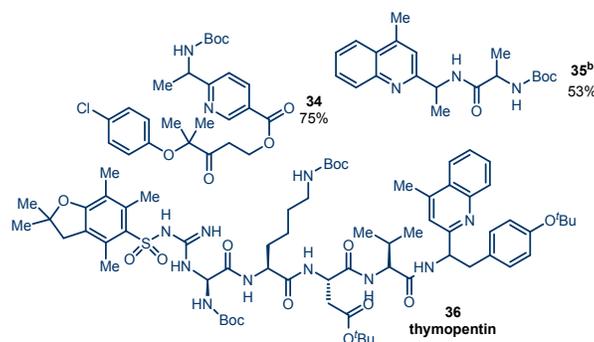
We then sought to demonstrate the generality of this approach with respect to *N*-heteroarenes (Scheme 2). A broad scope of *N*-heteroarenes was amenable despite their inherent differences in electrophilicity. Quinoline **13-18**, isoquinoline **19-26**, pyridine **27-31** and phenanthridine **32** and **33** can all be smoothly alkylated with good to excellent yields (50–90%). Reactions selectively took place on the most electrophilic position, which can be predicted by a radical nucleophilic attack mechanism.^{7,13} Electron-withdrawing (-F, -Cl, -Br) and electron-donating (-Me and -OMe) substituents at 4-, 5- and 6-positions can afford the corresponding alkylation products in good yields.

Scheme 2 Aromatic scope for the photocatalytic amination^a



^a Heteroarene (0.40 mmol), NHBC esters (0.20 mmol), Photocatalyst (2 mol%), in MeCN (2.0 mL), irradiated by 60 W blue LEDs for 12 h under Ar.

Scheme 3 Peptides and drugs for Minisci reaction^{a,b}

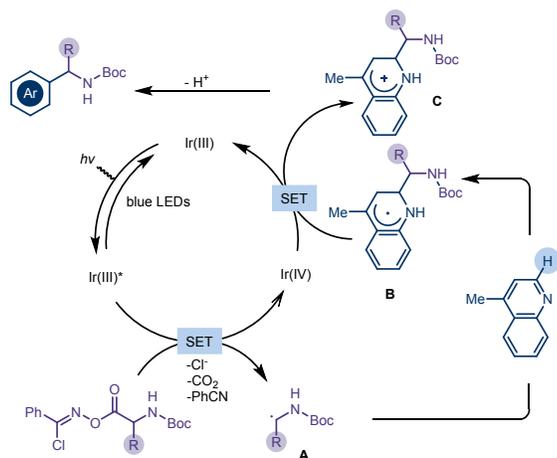


^a Heteroarene (0.40 mmol), NHBC esters (0.20 mmol), Photocatalyst (2 mol%), in MeCN (2.0 mL), irradiated by 60 W blue LEDs for 12 h under Ar. ^b 3:1 diastereomeric ratio.

The mild reaction conditions and the broad substrate scope prompt us to explore further elaboration of this reaction (Scheme 3). Bioactive etofibrate can be modified using this photocatalytic alkylation to form the pyridine derivative **34** in 75% yield. Direct installation of a *N*-heteroarene to peptides furnished various functionalized peptides at the C-terminal. Dipeptide-derived redox-active esters can be successfully used in the decarboxylative α -aminoalkylation reaction, generating desired peptide **35** in 53% yield. This protocol has been further applied to the functionalization of thymopentin, a pentapeptide, which could have potential biomedical applications (**36**, See Supporting Information).

Based on the previous reports⁸ and the quenching studies of photocatalysts (see Supporting Information), a possible mechanism for the Minisci reaction is illustrated in Scheme 4. The photoredox catalyst Ir(III) with suitable redox potential can interact with *N*-hydroxybenzimidoyl chloride ester to afford α -aminoalkyl radical **A** after decarboxylation. This radical **A** may not be further oxidized to form an iminium cation by photoredox catalyst, but able to attack a *N*-heteroarene to form a radical cation **B**, which is further oxidized by Ir(IV) and then deprotonation by a counteranion (Cl⁻) to afford the α -aminoalkylation product and regenerate the photoredox catalyst.

Scheme 4 Proposed mechanism for Minisci reaction



Conclusion

In summary, we have developed photoredox catalyzed α -aminoalkylation of *N*-heteroarenes using RAE esters of amino acids. This radical sequence exhibited wide substrate scope of natural and unnatural amino acids and heteroarenes. It could be further elaborated to access functionalized peptides at the C-terminal. Future investigation in chemical biology is underway.

EXPERIMENTAL SECTION

General Information. All reactions were performed in flame-dried glassware with magnetic stirring bar and sealed with a rubber septum. The solvents were distilled by standard methods. Reagents were obtained from commercial suppliers and used without further purification unless otherwise noted. Silica gel column chromatography was carried out using silica Gel 60 (230–400 mesh). Analytical thin layer chromatography (TLC) was done using silica Gel (silica gel 60 F254). TLC plates were analyzed by an exposure to ultraviolet (UV) light. NMR experiments were measured on a Bruker AVANCE III-400 or 500 spectrometer and carried out in deuterated chloroform (CDCl_3) ^1H NMR and ^{13}C NMR spectra were recorded at 400 MHz or 500 MHz and 100MHz or 125 MHz spectrometers, respectively. Chemical shifts are reported as δ values relative to internal TMS (δ 0.00 for ^1H NMR), chloroform (δ 7.26 for ^1H NMR), chloroform (δ 77.00 for ^{13}C NMR). The following abbreviations are used for the multiplicities: s: singlet, d: doublet, dd: doublet of doublet, t: triplet, q: quadruplet, m: multiplet, br: broad signal for proton spectra; Coupling constants (J) are reported in Hertz (Hz). Melting points were uncorrected. HRMS were recorded on a Bruker microTOF-Q111. A borosilicate glass tube was used as a reaction tube. The reaction mixture was irradiated with two Kessil LEDs (Saltwater Aquarium Light A360WE Series Tuna Blue; Rating: 19VDC 90W Max from 5 cm away with no filters). Unless otherwise noted, all reagents were weighed and handled in air, and all reactions were under argon.

General Procedure for the synthesis of NHBC esters (a) – (l)

1) Hydroxylamine (50% aq. soln.) (72 mmol, 1.2 equiv.) was added to corresponding aldehydes (60 mmol, 1 eq.) in ethanol (40 mL). The mixture was stirred at room temperature for 1 hour. After the reaction was completed by TLC

monitoring, the reaction mixture was evaporated in *vacuo*. Then, the reaction mixture was quenched with water and extracted with ethyl acetate (60 mL x 3). The organic layer was dried over Na₂SO₄, and evaporated in *vacuo*. The corresponding oximes were directly used in the next step without further purification.

2) To a colorless, homogeneous solution of the corresponding oximes (60 mmol) in *N,N*-dimethylformamide (40 mL) at room temperature was added *N*-chlorosuccinimide (72 mmol, 1.2 eq.) portion-wise over 1 h. During each addition, the reaction mixture would turn yellow and then gradually return to near colorlessness. Additionally, an exotherm was noted with each portion added to ensure that the reaction initiated after the addition of *N*-chlorosuccinimide. An ice bath was available, if required, to cool the exotherm. After the addition was complete, the mixture was stirred at room temperature and monitored by thin layer chromatography (TLC) until the starting material was not detected. The reaction mixture was diluted with 250 mL of water and extracted with ether (3 x 100 mL). The organic layers were combined, washed with water (2 x 100 mL), washed with a 10percent aqueous solution of lithium chloride (2 x 100 mL), and washed with brine (100 mL). The aqueous layers were back extracted with ether (100 mL), and the combined organic layers (400 mL) were dried over Na₂SO₄. Concentration under reduced pressure afforded hydroximoyl chlorides as a light yellow oil. The hydroximoyl chloride was directly used in the next step without further purification.

3) The corresponding alkyl acid (72 mmol, 1.2 equiv.), corresponding hydroximoyl chlorides (60 mmol, 1 equiv.), and 4-dimethylaminopyridine (3 mmol, 5 mol%) was mixed in a flask with a stirring bar. DCM (50 mL) was added. Then a solution of *N,N'*-dicyclohexylcatbodiimide (72 mmol, 1.2 equiv.) in DCM (20 mL) was added slowly at 0 °C. The reaction mixture was stirred at room temperature for 4h. Then the white precipitate was filtered off and the solution was concentrated under vacuum, then purified by column chromatography on silica gel.

***tert*-butyl (Z)-1-(((chloro(phenyl)methylene)amino)oxy)-1-oxo-3-phenylpropan-2-yl)carbamate (a)**

white solid (723.8mg, 90%); mp : 115-118 °C; Eluent: PE/EA (5:1); ¹H NMR (500 MHz, CDCl₃) δ 7.97 (d, *J* = 7.5 Hz, 2H), 7.53 (t, *J* = 7.4 Hz, 1H), 7.45 (t, *J* = 7.7 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 7.29 – 7.24 (m, 1H), 7.21 (d, *J* = 7.0 Hz, 2H), 5.03 (d, *J* = 8.6 Hz, 1H), 4.88 (q, *J* = 6.9 Hz, 1H), 3.22 (dd, *J* = 9.1, 6.2 Hz, 2H), 1.43 (s, 9H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 168.5 , 155.0 , 148.7 , 135.3 , 132.3 , 131.0 , 129.4 , 128.7 , 128.6 , 128.3 , 127.2 , 80.3 , 53.5 , 38.5 , 28.3 . HRMS (ESI) *m/z*: [M + H]⁺ Calcd for C₂₁H₂₄ClN₂O₄ 403.1419; found 403.1426.

***tert*-butyl (Z)-1-(((chloro(phenyl)methylene)amino)oxy)-1-oxopropan-2-yl)carbamate (b)**

white solid (599.8mg, 92%); mp : 115-117 °C; Eluent: PE/EA (5:1); ¹H NMR (500 MHz, CDCl₃) δ 7.97 (dt, *J* = 7.4Hz, 2H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.44 (t, *J* = 7.7 Hz, 2H), 5.11 (d, *J* = 8.0 Hz, 1H), 4.60 (q, *J* = 7.5 Hz, 1H), 1.53 (d, *J* = 7.3 Hz, 3H), 1.46 (s, 9H). ¹³C {¹H} (126 MHz, CDCl₃) δ 169.9 , 155.0 , 148.7 , 132.2 , 131.1 , 128.6 , 128.2 , 80.22 , 48.33 , 28.3 , 18.6 . HRMS(ESI) *m/z*: [M + H]⁺ Calcd for C₁₅H₂₀ClN₂O₄ 327.1106; found 327.1117.

benzyl (Z)-1-(((chloro(phenyl)methylene)amino)oxy)-1-oxopropan-2-yl)carbamate (c)

white solid (641.0mg, 89%); mp : 92-95 °C; Eluent: PE/EA (7:1); ¹H NMR (500 MHz, CDCl₃) δ 7.97 (d, *J* = 7.4 Hz, 2H), 7.54 (t, *J* = 7.4 Hz, 1H), 7.45 (t, *J* = 7.7 Hz, 2H), 7.40 – 7.29 (m, 5H), 5.39 (d, *J* = 8.0 Hz, 1H), 5.15 (s, 2H), 4.77-4.64 (m, *J* = 7.2 Hz, 1H), 1.58 (d, *J* = 7.3 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 169.5 , 155.5 , 149.03, 136.1 , 132.3 , 131.0 , 128.7 , 128.5 , 128.3 , 128.2 , 128.18, 67.17, 48.8 , 18.8 . HRMS(ESI) *m/z*: [M + H]⁺ Calcd for C₁₈H₁₈ClN₂O₄ 361.0950; found 361.0956.

***tert*-butyl (Z)-1-(((chloro(phenyl)methylene)amino)oxy)-1-oxobutan-2-yl)carbamate (d)**

white solid (578.2mg, 85%); mp : 92-95 °C; Eluent: PE/EA (7:1); ¹H NMR (400 MHz, CDCl₃) δ 7.98 (d, *J* = 7.6 Hz, 2H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.44 (t, *J* = 7.7 Hz, 2H), 5.11 (d, *J* = 8.6 Hz, 1H), 4.54 (q, *J* = 7.3 Hz, 1H), 1.98 (dt, *J* = 14.1, 6.9 Hz, 1H), 1.83 (dt, *J* = 14.2, 7.3 Hz, 1H), 1.47 (s, 9H), 1.05 (t, *J* = 7.4 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 169.3, 155.3 , 148.6 , 132.2 , 131.0, 128.6 , 128.2 , 80.18, 53.7 , 28.3 , 26.0, 9.7 . HRMS(ESI) *m/z*: [M + H]⁺ Calcd for C₁₆H₂₂ClN₂O₄ 341.1263; found 341.1267.

***tert*-butyl (Z)-1-(((chloro(phenyl)methylene)amino)oxy)-3-methyl-1-oxopentan-2-yl)carbamate (e)**

white solid (618.2mg, 84%); mp : 50-53 °C; Eluent: PE/EA (7:1); ¹H NMR (500 MHz, CDCl₃) δ 7.97 (d, *J* = 7.6 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.43 (t, *J* = 7.7 Hz, 2H), 5.13 (d, *J* = 9.1 Hz, 1H), 4.54 (dd, *J* = 9.1, 5.0 Hz, 1H), 2.05 – 1.85 (m, 1H), 1.56 (m, 1H), 1.45 (s, 9H), 1.37 – 1.18 (m, 1H), 1.02 (d, *J* = 6.9 Hz, 3H), 0.97 (t, *J* = 7.4 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 168.8 , 155.4 , 148.5 , 132.2 , 131.0 , 128.6 , 128.2 , 80.1 , 56.9 , 49.0 , 38.1 , 33.9 , 28.3 , 25.6 , 25.1 , 24.9 , 15.5 , 11.6 . HRMS(ESI) *m/z*: [M + H]⁺ Calcd for C₁₈H₂₆ClN₂O₄ 369.1576; found 369.1584.

di-tert-butyl (6-(((chloro(phenyl)methylene)amino)oxy)-6-oxohexane-1,5-diyl)(Z)-dicarbamate (f)

white solid (840.4mg, 87%); mp : 97-100 °C; Eluent: PE/EA (4:1); ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 7.4 Hz, 2H), 7.52 (t, J = 7.4 Hz, 1H), 7.43 (t, J = 7.6 Hz, 2H), 5.21 (d, J = 8.4 Hz, 1H), 4.74 – 4.36 (m, 2H), 3.13 (q, J = 6.4 Hz, 2H), 1.98 – 1.90 (m, 1H), 1.85 – 1.75 (m, 1H), 1.58 – 1.47 (m, 4H), 1.45 (s, 9H), 1.43 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 169.3 , 156.0 , 155.4 , 148.7 , 132.2 , 131.0 , 128.6 , 128.2 , 80.2 , 79.1 , 52.4 , 39.9 , 32.2 , 29.6 , 28.4 , 28.3 , 22.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₃₅ClN₃O₆ 484.2209; found 484.2216.

tert-butyl (Z)-(3-(4-(tert-butoxy)phenyl)-1-(((chloro(phenyl)methylene)amino)oxy)-1-oxopropan-2-yl)carbamate (g)

white solid (872.2mg, 92%); mp : 117- 120 °C; Eluent: PE/EA (4:1); ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 7.5 Hz, 2H), 7.53 (t, J = 7.0 Hz, 1H), 7.44 (t, J = 7.4 Hz, 2H), 7.10 (s, 2H), 6.93 (d, J = 8.0 Hz, 2H), 5.05 (d, J = 8.6 Hz, 1H), 4.84 (q, J = 7.6, 7.2 Hz, 1H), 3.16 (d, J = 6.3 Hz, 2H), 1.43 (s, 9H), 1.31 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 168.6 , 155.0 , 154.6 , 148.7 , 132.3 , 131.0 , 130.1 , 129.8 , 128.6 , 128.3 , 124.3 , 80.2 , 78.4 , 53.6 , 37.9 , 28.8 , 28.3 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₅H₃₂ClN₂O₅ 475.1994; found 475.2001.

(Z)-4-(2-((tert-butoxycarbonyl)amino)-3-(((chloro(phenyl)methylene)amino)oxy)-3-oxopropyl)phenylhypoidite (h)

white solid (971.5mg, 92%); mp : 125-128 °C; Eluent: PE/EA (5:1); ¹H NMR (400 MHz, CDCl₃) δ 7.97 (d, J = 7.4 Hz, 2H), 7.63 (d, J = 8.2 Hz, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.45 (t, J = 7.6 Hz, 2H), 6.96 (d, J = 8.2 Hz, 2H), 5.04 (d, J = 8.5 Hz, 1H), 4.86 (dd, J = 13.5, 6.3 Hz, 1H), 3.16 (dd, J = 8.8, 6.0 Hz, 2H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 168.3 , 154.9 , 149.0 , 137.8 , 135.1 , 132.4 , 131.4 , 130.9 , 128.7 , 128.3 , 92.8 , 53.3 , 38.1 , 28.3 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₁H₂₃ClN₂O₄ 529.0386; found 529.0382.

cyclohexyl (Z)-3-((tert-butoxycarbonyl)amino)-4-(((chloro(phenyl)methylene) amino)oxy)-4-oxobutanoate (i)

white solid (822.6mg, 92%); mp: 104-107 °C; Eluent: PE/EA (5:1); ¹H NMR (400 MHz, CDCl₃) δ 7.97 (d, J = 7.5 Hz, 2H), 7.53 (t, J = 7.3 Hz, 1H), 7.44 (t, J = 7.6 Hz, 2H), 5.61 (d, J = 9.2 Hz, 1H), 4.90 – 4.78 (m, 2H), 3.13 (dd, J = 17.1, 4.9 Hz, 1H), 2.91 (dd, J = 17.1, 4.6 Hz, 1H), 1.86 – 1.84 (m, 2H), 1.76 – 1.64 (m, 2H), 1.48 (s, 9H), 1.44 – 1.18 (m, 6H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 170.1 , 167.5 , 155.3 , 148.8 , 132.2 , 131.0 , 128.6 , 128.3 , 80.4 , 73.9 , 49.3 , 37.1 , 31.4 , 28.3 , 25.2 , 23.6 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₃₀ClN₂O₆ 453.1787; found 453.1794.

tert-butyl (Z)-(6-(tert-butoxy)-1-(((chloro(phenyl)methylene)amino)oxy)-1,5-dioxohexan-2-yl)carbamate (j)

white solid (783.2mg, 89%); mp : 115-118 °C; Eluent: PE/EA (5:1); ¹H NMR (500 MHz, CDCl₃) δ 7.97 (d, J = 7.4 Hz, 2H), 7.53 (t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.7 Hz, 2H), 5.19 (d, J = 8.7 Hz, 1H), 4.59 (q, J = 7.4, 6.7 Hz, 1H), 2.54 – 2.34 (m, 2H), 2.31 – 2.18 (m, 1H), 2.14 – 2.01 (m, 1H), 1.46 (s, 18H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 171.8 , 168.9 , 155.3 , 148.9 , 132.3 , 131.1 , 128.6 , 128.3 , 80.9 , 80.3 , 52.2 , 31.6 , 28.3 , 28.1 , 27.6 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₁H₃₀ClN₂O₆ 441.1787; found 441.1793.

tert-butyl (Z)-(1-(((chloro(phenyl)methylene)amino)oxy)-6-methoxy-1,5-dioxo hexan-2-yl)carbamate (k)

white solid (678.3mg, 85%); mp : 94-97 °C; Eluent: PE/EA (5:1); ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 7.4 Hz, 2H), 7.53 (t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.6 Hz, 2), 5.21 (d, J = 8.6 Hz, 1H), 4.62 (q, J = 7.7 Hz, 1H), 3.69 (s, 3H), 2.62 – 2.42 (m, 2H), 2.35 – 2.27 (m, 1H), 2.17 – 1.96 (m, 1H), 1.45 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 172.9 , 168.7 , 155.2 , 148.9 , 132.3 , 131.0 , 128.6 , 128.3 , 80.4 , 52.0 , 51.9 , 30.0 , 28.2 , 27.7 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₈H₂₄ClN₂O₆ 399.1317; found 399.1327.

tert-butyl (Z)-(1-(((chloro(phenyl)methylene)amino)oxy)-4-(methylthio)-1-oxobutan-2-yl)carbamate (l)

white solid (669.1mg, 82%); mp : 90-93 °C; Eluent: PE/EA (8:1); ¹H NMR (500 MHz, CDCl₃) δ 7.98 (d, J = 7.4 Hz, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.45 (t, J = 7.7 Hz, 2H), 5.19 (d, J = 8.7 Hz, 1H), 4.93 – 4.61 (m, 1H), 2.64 (m, 2H), 2.32 – 2.18 (m, 1H), 2.13 (s, 3H), 2.08 (m, 1H), 1.47 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 169.0 , 155.2 , 148.9 , 132.3 , 131.0 , 128.7 , 128.3 , 80.4 , 51.8 , 32.1 , 29.9 , 28.3 , 15.4 . HRMS(ESI) m/z: [M + Na]⁺ Calcd for C₁₇H₂₃ClN₂O₄SNa 409.0959; found 409.0964.

General procedures for decarboxylative Minisci reaction. Condition A: Under argon, [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ (2 mol%) and NHBC esters (0.2 mmol, 1 equiv.), corresponding N-heteroarene (0.4 mmol, 2 equiv.) were placed in a tube with a stirring bar, then dry MeCN (2 mL) was added at room temperature. After that, the tube was exposed to two 60 W blue LEDs at room temperature about 12 h until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired product.

Condition B: The corresponding aminoacid (1.2 mmol, 1.2 equiv.), hydroximoyl chlorides (1 mmol, 1 equiv.), and 4-dimethylaminopyridine (0.05 mmol, 5 mol%) was mixed in a flask with a stirring bar before dichloromethane (8 mL) was added. Then a solution of N,N'-dicyclohexylcarbodiimide (1.2 mmol, 1.2 equiv.) in DCM (2 mL) was added

slowly at 0 °C. The reaction mixture was stirred at room temperature for 4 h. Then Ir[$\{dF(CF_3)ppy\}_2(dtbbpy)]PF_6$ (2 mol%), and corresponding N-heteroarene (2 mmol, 2.0 equiv.) were dissolved in DCM (1 ml), then added to the flask. After that, the flask was exposed to two 60 W blue LEDs at room temperature about 12 h until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in vacuo. The crude products were directly purified by flash chromatography on silica gel to give the desired product.

Large-Scale Synthesis. The reaction took place in flask (100 mL) equipped with a stir bar then NHBC ester **f** (1.21 g, 2.5 mmol, 1.0 eq), 4-methylquinoline (0.75 g, 5 mmol, 2.0 eq) and Ir[$\{dF(CF_3)ppy\}_2(dtbbpy)]PF_6$ (56mg, 2 mol%) was added into bottle. Then 50 mL of MeCN were added into bottle by syringe. The flask was sealed and irradiated with blue LED lamps for 12 h. When the reaction was finished, the solution was removed with a rotary evaporator. The pure product was obtained in 75% yield (0.83 g) by column chromatography on silica gel (PE/EA=10:1).

tert-butyl (1-(4-methylquinolin-2-yl)-2-phenylethyl)carbamate (1) Following the general procedure condition A, **1** was purified by silica gel chromatography (PE/EA=10:1); 61.8 mg (82%); white solid; mp :124-126 °C; 1H NMR (500 MHz, $CDCl_3$) δ 8.05 (d, J = 8.3 Hz, 1H), 7.95 (d, J = 8.2 Hz, 1H), 7.69 (t, J = 7.5 Hz, 2H), 7.53 (t, J = 7.5 Hz, 1H), 7.22 – 7.17 (m, 3H), 7.08 – 6.97 (m, 2H), 6.81 (s, 1H), 6.14 (d, J = 7.8 Hz, 1H), 5.14 – 5.10 (m, 1H), 3.30 (dd, J = 13.4, 5.8 Hz, 1H), 3.16 (dd, J = 13.3, 7.7 Hz, 1H), 2.58 (s, 3H), 1.46 (s, 9H). $^{13}C\{^1H\}$ NMR (126 MHz, $CDCl_3$) δ 159.3, 155.3, 147.3, 144.2, 137.5, 129.7, 129.1, 128.1, 127.3, 126.4, 126.3, 125.9, 123.7, 121.2, 79.2, 56.9, 42.7, 28.4, 18.6. HRMS(ESI) m/z: $[M + H]^+$ Calcd for $C_{23}H_{27}N_2O_2$ 363.2067; found 363.2074.

tert-butyl (1-(4-methylquinolin-2-yl)ethyl)carbamate (2) Following the general procedure condition A, **2** was purified by silica gel chromatography (PE:EA=10:1); 61.8 mg (90%); white solid; mp : 117-120 °C; 1H NMR (500 MHz, $CDCl_3$) δ 8.05 (d, J = 8.4 Hz, 1H), 7.93 (d, J = 8.3 Hz, 1H), 7.67 (t, J = 8.3 Hz, 1H), 7.51 (t, J = 8.1 Hz, 1H), 7.15 (s, 1H), 6.30 (d, J = 7.0 Hz, 1H), 4.98 – 4.92 (m, 1H), 2.66 (s, 3H), 1.53 (d, J = 6.8 Hz, 3H), 1.47 (s, 9H). $^{13}C\{^1H\}$ NMR (126 MHz, $CDCl_3$) δ 161.1, 155.3, 147.0, 144.8, 129.5, 129.1, 127.2, 125.9, 123.6, 120.0, 79.0, 51.2, 28.4, 22.7, 18.7. HRMS(ESI) m/z: $[M + H]^+$ Calcd for $C_{17}H_{23}N_2O_2$ 287.1754; found 287.1765.

benzyl (2-phenyl-1-(4-methylquinolin-2-yl)ethyl) carbamate (3) Following the general procedure condition A, **3** was purified by silica gel chromatography (PE:EA=10:1); 41.6 mg (65%); white solid; mp ; 88-91 °C; 1H NMR (400 MHz, $CDCl_3$) δ 8.04 (d, J = 8.3 Hz, 1H), 7.96 (d, J = 9.2 Hz, 1H), 7.69 (t, J = 7.6 Hz, 1H), 7.54 (t, J = 8.1 Hz, 1H), 7.45 – 7.27 (m, 5H), 7.17 (s, 1H), 6.67 (d, J = 6.9 Hz, 1H), 5.18 – 5.10 (m, 2H), 5.05 – 4.96 (m, 1H), 2.70 (s, 3H), 1.57 (d, J = 6.7 Hz, 3H). $^{13}C\{^1H\}$ NMR (126 MHz, $CDCl_3$) δ 160.4, 155.8, 147.0, 145.1, 136.7, 129.6, 129.3, 128.5, 128.2, 128.0, 127.3, 126.1, 123.7, 119.9, 66.6, 51.6, 22.8, 18.8. HRMS(ESI) m/z: $[M + H]^+$ Calcd for $C_{20}H_{21}N_2O_2$ 321.1598; found 321.1607.

tert-butyl (1-(4-methylquinolin-2-yl)propyl)carbamate (4) Following the general procedure condition A, **4** was purified by silica gel chromatography (PE:EA=10:1); 57.0 mg (95%); white solid; mp : 91-94 °C; 1H NMR (400 MHz, $CDCl_3$) δ 8.06 (d, J = 8.3 Hz, 1H), 7.95 (d, J = 8.3 Hz, 1H), 7.68 (t, J = 7.0 Hz, 1H), 7.52 (t, J = 8.3 Hz, 1H), 7.16 (s, 1H), 6.13 (d, J = 7.8 Hz, 1H), 4.83 (q, J = 6.7 Hz, 1H), 2.68 (s, 3H), 2.08 – 1.95 (m, 1H), 1.91 – 1.76 (m, 1H), 1.46 (s, 9H), 0.87 (t, J = 7.4 Hz, 3H). $^{13}C\{^1H\}$ NMR (126 MHz, $CDCl_3$) δ 160.2, 155.6, 147.2, 144.5, 129.6, 129.1, 127.3, 125.9, 123.6, 120.8, 79.0, 56.5, 29.4, 28.4, 18.7, 9.7. HRMS(ESI) m/z: $[M + H]^+$ Calcd for $C_{18}H_{24}N_2O_2$ 301.1911; found 301.1917.

tert-butyl (2-methyl-1-(4-methylquinolin-2-yl)-2-(3-butyl)carbamate (5) Following the general procedure condition A, **5** was purified by silica gel chromatography (PE:EA=10:1); 44.6 mg (61%) (dr = 1:1); white solid; mp : 78-81 °C; 1H NMR (500 MHz, $CDCl_3$) δ 8.08 (d, J = 8.4 Hz, 1H), 7.97 (d, J = 8.3 Hz, 1H), 7.69 (t, J = 7.7 Hz, 1H), 7.53 (t, J = 7.6 Hz, 1H), 7.16 (d, J = 4.2 Hz, 1H), 6.09 (t, J = 7.3 Hz, 1H), 4.84 (ddd, J = 56.1, 8.7, 5.6 Hz, 1H), 2.70 (s, 3H), 2.12 – 1.91 (m, 1H), 1.67 – 1.52 (m, 1H), 1.48 (d, J = 4.6 Hz, 9H), 1.23 – 1.12 (m, 1H), 0.95 (dt, J = 25.9, 7.4 Hz, 3H), 0.87 (dd, J = 6.9, 3.4 Hz, 3H). $^{13}C\{^1H\}$ NMR (126 MHz, $CDCl_3$) δ 160.2& 159.9, 156.0&155.8, 147.3& 147.1, 144.1&144.0, 129.6, 129.0, 127.2, 125.8, 123.6, 121.7& 121.3, 79.0, 59.8&58.9, 41.0& 40.7, 28.4, 26.6& 25.0, 18.7, 15.6 &14.1, 11.9 &11.6. HRMS(ESI) m/z: $[M + H]^+$ Calcd for $C_{20}H_{29}N_2O_2$ 329.2224; found 329.2228.

tert-butyl 4-((tert-butoxycarbonyl)amino)-4-(4-methylquinolin-2-yl)butanoate (6) Following the general procedure condition A, **6** was purified by silica gel chromatography (PE:EA=5:1) ; 60.8 mg (76%); white solid; mp : 147-149 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.05 (d, J = 8.3 Hz, 1H), 7.96 (d, J = 8.2 Hz, 1H), 7.68 (t, J = 8.2 Hz, 1H), 7.53 (t, J = 7.6 Hz, 1H), 7.19 (s, 1H), 6.16 (d, J = 7.8 Hz, 1H), δ 4.94 (q, J = 6.7 Hz, 1H), 2.69 (s, 3H), 2.40 – 2.15 (m, 3H), 2.12 – 1.95 (m, 1H), 1.46 (s, 9H), 1.40 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 172.6 , 159.7 , 155.6 , 147.2 , 144.9 , 129.6 , 129.2 , 127.4 , 126.0 , 123.6 , 120.5 , 80.2 , 79.2 , 54.7 , 31.5 , 31.5 , 28.4 , 28.0 , 18.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₃₃N₂O₄ 401.2435; found 401.2448.

tert-butyl (1-(4-methylquinolin-2-yl)-3-(methylthio)propyl)carbamate (7) Following the general procedure condition A, **7** was purified by silica gel chromatography (PE:EA=5:1) ; 51.9 mg (75%); white solid; mp : 87-90 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.04 (d, J = 8.4 Hz, 1H), 7.95 (d, J = 8.3 Hz, 1H), 7.68 (t, J = 8.2 Hz, 1H), 7.52 (t, J = 7.6 Hz, 1H), 7.18 (s, 1H), 6.18 (d, J = 7.8 Hz, 1H), 5.01 (d, J = 6.9 Hz, 1H), 2.68 (s, 3H), 2.60 – 2.50 (m, 1H), 2.46 – 2.34 (m, 1H), 2.31 – 2.21 (m, 1H), 2.15 – 2.08 (m, 1H), 2.07 (s, 3H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.4 , 155.5 , 147.2 , 144.9 , 129.6 , 129.2 , 127.3 , 126.0 , 123.6 , 120.6 , 79.2 , 54.6 , 36.1 , 30.0 , 28.4 , 18.7 , 15.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₉H₂₇N₂O₂S 347.1788; found 347.1794.

tert-butyl (2-(4-iodophenyl)-1-(4-methylquinolin-2-yl)ethyl)carbamate (8) Following the general procedure condition A, **8** was purified by silica gel chromatography (PE:EA=5:1) ; 63.5 mg (65%); white solid; mp : 127-130 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, J = 8.4 Hz, 1H), 7.96 (d, J = 8.3 Hz, 1H), 7.70 (t, J = 7.6 Hz, 1H), 7.55 (t, J = 7.6 Hz, 1H), 7.48 (d, J = 7.9 Hz, 2H), 6.86 (s, 1H), 6.74 (d, J = 7.8 Hz, 2H), 6.11 (d, J = 7.7 Hz, 1H), 5.06 (q, J = 7.0 Hz, 1H), 3.25 – 3.16 (m, 2H), 2.62 (s, 3H), 1.45 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 158.8 , 155.3 , 147.3 , 144.5 , 137.1 , 131.8 , 129.6 , 129.2 , 127.3 , 126.1 , 123.7 , 121.1 , 91.7 , 79.4 , 56.6 , 42.0 , 28.4 , 18.7 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₂₆IN₂O₂ 489.1033; found 489.1040.

tert-butyl (2-(4-(tert-butoxy)phenyl)-1-(4-methylquinolin-2-yl)ethyl)carbamate (9) Following the general procedure condition A, **9** was purified by silica gel chromatography (PE:EA=5:1) ; 57.3 mg (66%); white solid; mp : 120-123 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, J = 7.6 Hz, 1H), 7.92 (d, J = 7.1 Hz, 1H), 7.67 (t, J = 8.1 Hz, 1H), 7.51 (t, J = 8.0 Hz, 1H), 6.88 (d, J = 8.0 Hz, 2H), 6.79 (d, J = 8.1 Hz, 2H), 6.73 (s, 1H), 6.21 (d, J = 7.7 Hz, 1H), 5.05 (q, J = 7.3 Hz, 1H), 3.28 (dd, J = 13.3, 5.7 Hz, 1H), 3.05 (dd, J = 13.3, 8.1 Hz, 1H), 2.54 (s, 3H), 1.45 (s, 9H), 1.29 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.3 , 155.3 , 153.6 , 147.2 , 144.0 , 132.5 , 129.9 , 129.5 , 129.0 , 127.2 , 125.9 , 123.9 , 123.6 , 121.3 , 79.1 , 78.1 , 57.1 , 42.2 , 28.7 , 28.4 , 18.6. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₇H₃₅N₂O₃ 435.2648; found 435.2648.

methyl 4-((tert-butoxycarbonyl)amino)-4-(4-methylquinolin-2-yl)butanoate (10) Following the general procedure condition A, **10** was purified by silica gel chromatography (PE:EA=5:1) ; 53.7 mg (75%); white solid; mp : 99-102 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, J = 8.4 Hz, 1H), 7.95 (dd, J = 8.4, 1.3 Hz, 1H), 7.68 (t, J = 8.2 Hz, 1H), 7.53 (t, J = 7.6 Hz, 1H), 7.18 (s, 1H), 6.20 (d, J = 7.7 Hz, 1H), 5.07 – 4.89 (m, 1H), 3.60 (s, 3H), 2.68 (s, 3H), 2.49 – 2.39 (m, 1H), 2.37 – 2.25 (m, 2H), 2.13 – 2.02 (m, 1H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 173.7 , 159.3 , 155.6 , 147.1 , 145.0 , 129.6 , 129.2 , 127.3 , 126.1 , 123.6 , 120.5 , 79.3 , 54.5 , 51.5 , 31.6 , 30.0 , 28.4 , 18.8. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₀H₂₇N₂O₄ 359.1965; found 359.1970.

Di-tert-butyl (1-(4-methylquinolin-2-yl)pentane-1,5-diy)dicarbamate (11) Following the general procedure condition A, **11** was purified by silica gel chromatography (PE:EA=5:1) ; 68.2 mg (77%); white solid; mp : 133-136 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, J = 8.4 Hz, 1H), 7.96 (d, J = 8.3 Hz, 1H), 7.68 (t, J = 7.4 Hz, 1H), 7.53 (t, J = 7.6 Hz, 1H), 7.15 (s, 1H), 6.11 (d, J = 7.5 Hz, 1H), 4.87 (q, J = 7.0 Hz, 1H), 4.60 (s, 1H), 3.07 (t, J = 7.4 Hz, 2H), 2.69 (s, 3H), 2.01 – 1.88 (m, 1H), 1.81 (d, J = 11.9 Hz, 1H), 1.49–1.30 (m, 22H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 160.3 , 156.0 , 155.7 , 147.3 , 144.8 , 129.6 , 129.2 , 127.3 , 126.0 , 123.7 , 120.6 , 79.2 , 78.9 , 55.2 , 40.3 , 36.4 , 29.6 , 28.4 , 28.4 , 22.6 , 18.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₅H₃₈N₃O₄ 444.2857; found 444.2861.

Cyclohexyl 3-((tert-butoxycarbonyl)amino)-3-(4-methylquinolin-2-yl)propanoate (12) Following the general procedure condition A, **12** was purified by silica gel chromatography (PE:EA=5:1) ; 66.1 mg (80%); white solid; mp :

107-110 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, J = 8.4 Hz, 1H), 7.96 (d, J = 8.3 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.53 (t, J = 7.6 Hz, 1H), 7.32 (s, 1H), 6.08 (d, J = 8.6 Hz, 1H), 5.41 – 5.14 (m, 1H), 4.70 (dq, J = 9.0, 4.8, 4.3 Hz, 1H), 3.12 (dd, J = 15.5, 5.3 Hz, 1H), 2.95 (dd, J = 15.5, 6.6 Hz, 1H), 2.68 (s, 3H), 1.80 – 1.68 (m, 2H), 1.63 (d, J = 7.6 Hz, 2H), 1.47 (s, 9H), 1.38 – 1.09 (m, 6H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 170.9, 159.3, 155.4, 147.2, 144.8, 129.7, 129.1, 127.4, 126.1, 123.6, 120.7, 79.5, 72.8, 52.3, 40.0, 31.5, 28.4, 25.3, 23.6, 18.8. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₄H₃₃N₂O₄ 413.2435; found 413.2442.

tert-butyl (2-phenyl-1-(quinolin-2-yl)ethyl)carbamate (13) Following the general procedure condition A, **13** was purified by silica gel chromatography (PE:EA=5:1); 38.3 mg (55%); white solid; mp : 129-132 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.06 (d, J = 8.5 Hz, 1H), 7.96 (d, J = 8.4 Hz, 1H), 7.77 (d, J = 8.1 Hz, 1H), 7.71 (t, J = 7.7 Hz, 1H), 7.52 (t, J = 7.4 Hz, 1H), δ 7.2 – 7.1 (m, 3H), 7.0 – 7.0 (m, 2H), 7.0 (d, J = 8.4 Hz, 1H). 5.16 (q, J = 7.1 Hz, 1H), 3.34 (dd, J = 13.5, 5.6 Hz, 1H), 3.15 (dd, J = 13.1, 8.0 Hz, 1H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.7, 155.3, 147.5, 137.4, 135.9, 129.6, 129.4, 129.1, 128.1, 127.5, 127.3, 126.3, 126.2, 120.6, 79.3, 57.1, 42.8, 28.4. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₅N₂O₂ 349.1911; found 349.1917.

tert-butyl (2-phenyl-1-(4-phenylquinolin-2-yl)ethyl)carbamate (14) Following the general procedure condition A, **14** was purified by silica gel chromatography (PE:EA=5:1); 63.6 mg (75%); white solid; mp : 147-150 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.15 (d, J = 8.4 Hz, 1H), 7.87 (d, J = 8.3 Hz, 1H), 7.72 (t, J = 7.7 Hz, 1H), 7.47 (dt, J = 7.6, 4.8 Hz, 4H), 7.32 (d, J = 6.7 Hz, 2H), 7.20 (d, J = 5.8 Hz, 3H), 7.04 (d, J = 6.6 Hz, 2H), 6.83 (s, 1H), 6.25 (d, J = 7.7 Hz, 1H), 5.19 (q, J = 7.3 Hz, 1H), 3.40 (dd, J = 13.3, 5.7 Hz, 1H), 3.14 (dd, J = 13.2, 7.9 Hz, 1H), 1.48 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.1, 155.3, 148.1, 148.0, 137.9, 137.5, 129.7, 129.5, 129.4, 129.2, 128.4, 128.3, 128.2, 126.3, 126.2, 125.8, 125.7, 120.8, 79.3, 57.2, 43.1, 28.4. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₈H₂₉N₂O₂ 425.2224; found 425.2230.

tert-butyl (1-(3-methylquinolin-2-yl)-2-phenylethyl)carbamate (15) Following the general procedure condition A, **15** was purified by silica gel chromatography (PE:EA=5:1); 55.8 mg (77%); white solid; mp : 129-132 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.06 (d, J = 8.4 Hz, 1H), 7.78 (s, 1H), 7.73 (d, J = 8.1 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.51 (t, J = 7.5 Hz, 1H), 7.20 – 7.15 (m, 3H), 7.00 (dt, J = 7.3, 3.6 Hz, 2H), 6.22 (d, J = 8.5 Hz, 1H), 5.40 (q, J = 7.7 Hz, 1H), 3.30 – 3.16 (m, 2H), 2.19 (s, 3H), 1.48 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.7, 155.2, 146.2, 137.4, 136.0, 129.6, 128.9, 128.7, 128.4, 128.0, 127.4, 126.7, 126.2, 126.2, 79.1, 53.1, 43.0, 28.4, 18.3. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₂₇N₂O₂ 363.2067; found 363.2074.

tert-butyl (1-(6-chloroquinolin-2-yl)-2-phenylethyl)carbamate (16) Following the general procedure condition A, **16** was purified by silica gel chromatography (PE:EA=10:1); 39.0 mg (51%); white solid; mp : 118-121 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, J = 8.9 Hz, 1H), 7.87 (d, J = 8.4 Hz, 1H), 7.75 (d, J = 2.3 Hz, 1H), 7.63 (dd, J = 9.0, 2.4 Hz, 1H), 7.20 – 7.10 (m, 3H), 6.97 (q, J = 3.8 Hz, 3H), 6.07 (d, J = 7.7 Hz, 1H), 5.14 (q, J = 7.3 Hz, 1H), 3.31 (dd, J = 13.3, 5.6 Hz, 1H), 3.13 (dd, J = 13.2, 8.0 Hz, 1H), 1.45 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 160.2, 155.3, 145.9, 137.2, 135.0, 131.9, 130.7, 130.4, 129.6, 128.2, 127.8, 126.4, 126.2, 121.4, 79.4, 57.1, 42.7, 28.4. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₄ClN₂O₂ 383.1521; found 383.1527.

tert-butyl (1-(4-chloroquinolin-2-yl)ethyl)carbamate (17) Following the general procedure condition A, **17** was purified by silica gel chromatography (PE:EA=10:1); 36.1 mg (59%); white solid; mp : 130-135 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, J = 8.4 Hz, 1H), 8.07 (d, J = 8.4 Hz, 1H), 7.75 (t, J = 8.3 Hz, 1H), 7.60 (t, J = 7.6 Hz, 1H), 7.45 (s, 1H), 6.02 (d, J = 6.7 Hz, 1H), 5.03 – 4.91 (m, 1H), 1.54 (d, J = 6.8 Hz, 3H), 1.47 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 161.8, 155.2, 148.2, 143.0, 130.4, 129.4, 127.2, 125.4, 124.0, 119.4, 79.4, 51.4, 28.4, 22.5. HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₆H₂₀ClN₂O₂ 307.1208; found 307.1217.

tert-butyl (1-(4-phenylquinolin-2-yl)ethyl)carbamate (18) Following the general procedure condition A, **18** was purified by silica gel chromatography (PE:EA=10:1); 50.1 mg (72%); white solid; mp : 160-163 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.14 (d, J = 8.4 Hz, 1H), 7.88 (d, J = 9.3 Hz, 1H), 7.71 (t, J = 8.3 Hz, 1H), 7.58 – 7.39 (m, 6H), 7.28 (s, 1H), 7.26 (s, 1H), 6.25 (d, J = 7.1 Hz, 1H), 5.19 – 4.86 (m, 1H), 1.58 (d, J = 6.8 Hz, 3H), 1.49 (s, 9H). ¹³C{¹H} NMR

(126 MHz, CDCl₃) δ 161.12 , 155.41 , 149.25 , 147.86 , 138.04 , 129.53 , 129.44 , 128.59 , 128.47 , 126.30 , 125.75 , 119.57 , 79.29 , 51.53 , 28.50 , 22.95 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₅N₂O₂ 349.1911; found 349.1914.

tert-butyl (1-(isoquinolin-1-yl)-2-phenylethyl)carbamate (19) Following the general procedure condition A, **19** was purified by silica gel chromatography (PE:EA=5:1) ; 28.5 mg (41%); white solid; mp : 166-168 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.42 (d, J = 5.6 Hz, 1H), 8.03 (d, J = 8.5 Hz, 1H), 7.80 (d, J = 8.2 Hz, 1H), 7.64 (t, J = 7.5 Hz, 1H), 7.55 (d, J = 5.6 Hz, 1H), 7.51 (t, J = 7.6 Hz, 1H), 7.16 – 7.05 (m, 3H), 6.88 (dd, J = 6.6, 3.0 Hz, 2H), 6.18 (d, J = 8.3 Hz, 1H), 5.91 (q, J = 6.5 Hz, 1H), 3.30 (dd, J = 13.3, 6.6 Hz, 1H), 3.26 (ddd, J = 43.0, 13.3, 6.3 Hz, 2H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.0 , 155.2 , 141.2 , 137.2 , 136.1 , 129.9 , 129.6 , 127.9 , 127.3 , 127.2 , 126.2 , 125.7 , 124.3 , 120.2 , 79.1 , 52.0 , 42.9 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₅N₂O₂ 349.1911; found 349.1918.

tert-butyl (1-(6-fluoroisoquinolin-1-yl)-2-phenylethyl)carbamate (20) Following the general procedure condition A, **20** was purified by silica gel chromatography (PE:EA=5:1) ; 54.2 mg (74%); liquid ; ¹H NMR (500 MHz, CDCl₃) δ 8.42 (d, J = 5.7 Hz, 1H), 7.98 (dd, J = 9.4, 5.4 Hz, 1H), 7.50 (d, J = 5.7 Hz, 1H), 7.37 (dd, J = 9.1, 2.6 Hz, 1H), 7.21 (td, J = 8.9, 2.6 Hz, 1H), 7.11 – 7.06 (m, 3H), 6.87 (dd, J = 6.3, 3.0 Hz, 2H), 6.16 (d, J = 8.3 Hz, 1H), 5.92 – 5.69 (m, 1H), 3.30 – 3.20 (m, 2H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 163.7 , 161.7 , 159.2 , 155.2 , 142.3 , 137.8 , 137.7 , 137.1 , 129.5 , 128.0 , 127.6 , 127.6 , 126.3 , 123.1 , 119.8 , 119.8 , 117.7 , 117.5 , 110.4 , 110.2 , 79.3 , 52.2 , 43.0 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₄FN₂O₂ 367.1816; found 367.1821.

tert-butyl (1-(6-chloroisoquinolin-1-yl)-2-phenylethyl)carbamate (21) Following the general procedure condition A, **21** was purified by silica gel chromatography (PE:EA=5:1) ; 40.5 mg (53%); liquid; ¹H NMR (500 MHz, CDCl₃) δ 8.46 (d, J = 5.7 Hz, 1H), 7.93 (d, J = 9.1 Hz, 1H), 7.77 (d, J = 2.1 Hz, 1H), 7.47 (d, J = 5.7 Hz, 1H), 7.42 (dd, J = 9.1, 2.2 Hz, 1H), 7.17 – 7.07 (m, 3H), 6.89 (dd, J = 6.5, 3.1 Hz, 2H), 6.17 (d, J = 8.4 Hz, 1H), 5.86 (q, J = 6.6 Hz, 1H), 3.27 (d, J = 6.5 Hz, 2H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.3 , 155.2 , 142.4 , 137.0 , 136.9 , 136.2 , 129.5 , 128.1 , 128.0 , 126.3 , 126.2 , 125.8 , 124.0 , 119.3 , 79.3 , 52.1 , 42.9 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₄ClN₂O₂ 383.1521; found 383.1521.

tert-butyl (1-(6-bromoisoquinolin-1-yl)-2-phenylethyl)carbamate (22) Following the general procedure condition A, **22** was purified by silica gel chromatography (PE:EA=5:1) ; 48.6 mg (57%); white solid; mp : 158-161 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.44 (d, J = 5.6 Hz, 1H), 7.96 (d, J = 2.0 Hz, 1H), 7.83 (d, J = 9.0 Hz, 1H), 7.54 (dd, J = 9.0, 2.0 Hz, 1H), 7.46 (d, J = 5.7 Hz, 1H), 7.11 – 7.08 (m, 3H), 6.86 (dd, J = 6.6, 3.0 Hz, 2H), 6.09 (d, J = 8.4 Hz, 1H), 5.82 (q, J = 6.7 Hz, 1H), 3.23 (d, J = 6.4 Hz, 2H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 159.5 , 155.2 , 142.4 , 137.2 , 137.0 , 130.7 , 129.5 , 129.3 , 128.0 , 126.4 , 126.1 , 124.8 , 124.3 , 119.1 , 79.3 , 52.1 , 43.0 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₄BrN₂O₂ 427.1016; found 427.1023.

tert-butyl (1-(6-methylisoquinolin-1-yl)-2-phenylethyl)carbamate (23) Following the general procedure condition A, **23** was purified by silica gel chromatography (PE:EA=5:1) ; 49.4 mg (68%); white solid; mp : 140-142 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.36 (d, J = 5.6 Hz, 1H), 7.94 (d, J = 8.7 Hz, 1H), 7.55 (s, 1H), 7.45 (d, J = 5.7 Hz, 1H), 7.34 (d, J = 8.6 Hz, 1H), 7.12 – 7.09 (m, 3H), 6.89 (dd, J = 6.6, 2.9 Hz, 2H), 6.21 (d, J = 8.3 Hz, 1H), 5.88 (dt, J = 8.4, 6.3 Hz, 1H), 3.31 (dd, J = 13.3, 6.5 Hz, 1H), 3.21 (dd, J = 13.3, 6.0 Hz, 1H), 2.51 (s, 3H), 1.44 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 158.6 , 155.2 , 141.3 , 140.2 , 137.3 , 136.5 , 129.6 , 129.5 , 127.8 , 126.1 , 126.1 , 124.1 , 124.1 , 119.7 , 79.0 , 52.0 , 42.7 , 28.4 , 21.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₂₇N₂O₂ 363.2067; found 363.2071.

tert-butyl (1-(6-methoxyisoquinolin-1-yl)-2-phenylethyl)carbamate (24) Following the general procedure condition A, **24** was purified by silica gel chromatography (PE:EA=5:1) ; 52.9 mg (70%); white solid; mp : 134-136 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.33 (d, J = 5.7 Hz, 1H), 7.88 (d, J = 9.3 Hz, 1H), 7.43 (d, J = 5.7 Hz, 1H), 7.11 – 7.02 (m, 5H), 6.89 (dd, J = 6.5, 2.9 Hz, 2H), 6.19 (d, J = 8.3 Hz, 1H), 5.81 (q, J = 6.7 Hz, 1H), 3.91 (s, 3H), 3.24 (qd, J = 13.2, 6.4 Hz, 2H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 160.3 , 158.3 , 155.2 , 141.8 , 138.2 , 137.3 , 129.5 , 127.9 , 126.2 , 126.1 , 121.4 , 120.0 , 119.5 , 104.6 , 79.0 , 55.3 , 52.0 , 42.9 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₃H₂₇N₂O₃ 379.2016; found 379.2020.

tert-butyl (1-(5-bromoisoquinolin-1-yl)ethyl)carbamate (25) Following the general procedure condition A, **25** was purified by silica gel chromatography (PE:EA=5:1) ; 21.0 mg (30%) ; white solid; mp : 157-160 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.55 (d, J = 5.9 Hz, 1H), 8.18 (d, J = 8.5 Hz, 1H), 8.03 – 7.85 (m, 2H), 7.52 – 7.43 (m, 1H), 6.32 (d, J = 7.8 Hz, 1H), 5.85 – 5.52 (m, 1H), 1.54 (d, J = 6.7 Hz, 3H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 161.1 , 155.3 , 142.7 , 135.5 , 133.9 , 127.7 , 126.2 , 124.0 , 122.5 , 119.1 , 79.3 , 47.2 , 28.4 , 23.2 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₆H₂₀BrN₂O₂ 351.0703; found 351.0703.

tert-butyl (1-(3-methylquinolin-2-yl)ethyl)carbamate (26) Following the general procedure condition A, **26** was purified by silica gel chromatography (PE:EA=5:1) ; 37.2 mg (65%) ; white solid; mp : 136-138 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.37 (d, J = 5.7 Hz, 1H), 8.05 (d, J = 8.6 Hz, 1H), 7.57 (s, 1H), 7.48 – 7.37 (m, 2H), 6.42 (d, J = 7.7 Hz, 1H), 5.67 (p, J = 6.8 Hz, 1H), 2.51 (s, 3H), 1.53 (d, J = 6.7 Hz, 3H), 1.46 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 160.2 , 155.3 , 141.3 , 140.3 , 136.7 , 129.6 , 126.3 , 124.1 , 123.3 , 119.7 , 79.0 , 47.1 , 28.4 , 23.1 , 21.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₇H₂₃N₂O₂ 287.1754; found 287.1764.

methyl 6-(1-((tert-butoxycarbonyl)amino)-2-phenylethyl)nicotinate (27) Following the general procedure condition A, **27** was purified by silica gel chromatography (PE:EA=5:4) ; 67.6 mg (95%); white solid; mp : 83-85 °C; ¹H NMR (500 MHz, CDCl₃) δ 9.15 (s, 1H), 8.08 (d, J = 7.5 Hz, 1H), 7.19 – 7.14 (m, 3H), 6.95 – 6.92 (m, 3H), 5.75 (d, J = 8.3 Hz, 1H), 5.03 (q, J = 6.8 Hz, 1H), 3.93 (s, 3H), 3.22 (dd, J = 13.4, 6.0 Hz, 1H), 3.05 (dd, J = 13.3, 7.9 Hz, 1H), 1.41 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 165.6 , 163.9 , 155.1 , 150.5 , 137.2 , 136.9 , 129.4 , 128.3 , 126.5 , 124.6 , 121.9 , 79.5 , 56.9 , 52.3 , 42.5 , 28.3 . HRMS(ESI) m/z: [M + Na]⁺ Calcd for C₂₀H₂₄N₂O₄Na 379.1628; found 379.1627.

tert-butyl (2-phenyl-1-(5-(trifluoromethyl)pyridin-2-yl)ethyl)carbamate (28) Following the general procedure condition A, **28** was purified by silica gel chromatography (PE:EA=5:1); 60.1 mg (82%); white solid; mp : 80-83 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.83 (s, 1H), 7.73 (d, J = 7.7 Hz, 1H), 7.22 – 7.16 (m, 3H), 7.05 – 6.86 (m, 3H), 5.72 (d, J = 8.2 Hz, 1H), 5.06 (q, J = 7.1 Hz, 1H), 3.22 (dd, J = 13.5, 6.2 Hz, 1H), 3.06 (dd, J = 13.3, 7.8 Hz, 1H), 1.42 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 163.7 , 155.1 , 146.2 (q, J = 4.1 Hz), 136.8 , 133.2 (q, J = 3.5 Hz), 129.3 , 128.3 , 126.6 , 125.2 (q, J = 33.0 Hz), 123.4 (q, J = 272.2 Hz), 121.9 , 79.7 , 56.8 , 42.5 , 28.3 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₉H₂₂F₃N₂O₂ 367.1628; found 367.1638.

tert-butyl (2-phenyl-1-(5-propionylpyridin-2-yl)ethyl)carbamate (29) Following the general procedure condition A, **29** was purified by silica gel chromatography (PE:EA=5:4) ; 43.2 mg (61%) ; white solid; mp : 84-86 °C; ¹H NMR (500 MHz, CDCl₃) δ 9.10 (d, J = 2.2 Hz, 1H), 8.03 (d, J = 7.6 Hz, 1H), 7.17 (d, J = 7.2 Hz, 3H), 6.96 (d, J = 7.2 Hz, 3H), 5.74 (d, J = 8.1 Hz, 1H), 5.03 (q, J = 7.1 Hz, 1H), 3.21 (dd, J = 13.4, 6.1 Hz, 1H), 3.05 (dd, J = 13.3, 7.9 Hz, 1H), 2.98 (q, J = 7.2 Hz, 2H), 1.41 (s, 9H), 1.22 (t, J = 7.2 Hz, 3H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 199.1 , 163.8 , 155.1 , 149.1 , 136.9 , 135.5 , 130.8 , 129.3 , 128.3 , 126.5 , 122.1 , 79.6 , 56.8 , 42.4 , 32.1 , 28.3 , 7.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₁H₂₇N₂O₃ 355.2016; found 355.2026.

methyl 5-bromo-6-(1-((tert-butoxycarbonyl)amino)ethyl)nicotinate (30) Following the general procedure condition A, **30** was purified by silica gel chromatography (PE:EA=10:1) ; 53.7 mg (75%); white solid; mp : 108-111 °C; ¹H NMR (500 MHz, CDCl₃) δ 9.05 (d, J = 1.8 Hz, 1H), 8.42 (d, J = 1.8 Hz, 1H), 5.88 (d, J = 8.3 Hz, 1H), 5.33 (dd, J = 9.5, 5.2 Hz, 1H), 3.95 (s, 3H), 1.56 – 1.28 (m, 12H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 164.4 , 164.0 , 155.0 , 148.6 , 141.5 , 126.0 , 118.9 , 79.5 , 52.7 , 49.9 , 28.4 , 21.2 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₁₄H₂₀BrN₂O₄ 359.0601; found 359.0603.

methyl 6-(1-((tert-butoxycarbonyl)amino)-2-phenylethyl)-2-methylnicotinate (31) Following the general procedure condition A, **31** was purified by silica gel chromatography (PE:EA=5:4) ; 67.4 mg (91%) ; white solid; mp : 79-82 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 8.0 Hz, 1H), 7.26 – 7.10 (m, 3H), 6.97 (d, J = 6.9 Hz, 2H), 6.71 (d, J = 8.1 Hz, 1H), 5.81 (d, J = 8.0 Hz, 1H), 4.97 (q, J = 7.5 Hz, 1H), 3.89 (s, 3H), 3.22 (dd, J = 13.3, 5.8 Hz, 1H), 3.01 (dd, J = 13.2, 7.9 Hz, 1H), 2.83 (s, 3H), 1.43 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 166.7 , 161.7 , 159.7 , 155.1 , 138.5

, 137.1 , 129.4 , 128.1 , 126.3 , 123.6 , 119.2 , 79.3 , 56.7 , 52.0 , 42.6 , 28.3 , 24.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₀H₂₅N₂O₄ 371.1965; found 371.1970.

tert-butyl (1-(phenanthridin-6-yl)ethyl)carbamate (32) Following the general procedure condition A, **32** was purified by silica gel chromatography (PE:EA=5:1) ; 39.3 mg (61%); white solid; mp : 180-183 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.65 (d, J = 8.3 Hz, 1H), 8.54 (dd, J = 8.1, 1.4 Hz, 1H), 8.25 (d, J = 8.2 Hz, 1H), 8.15 (d, J = 7.8 Hz, 1H), 7.85 (t, J = 8.2 Hz, 1H), 7.72 (q, J = 7.0 Hz, 2H), 7.65 (t, J = 6.9 Hz, 1H), 6.73 (d, J = 7.3 Hz, 1H), 6.31 – 5.56 (m, 1H), 1.63 (d, J = 6.7 Hz, 3H), 1.52 (s, 9H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 160.3 , 155.4 , 142.9 , 133.2 , 130.6 , 129.7 , 128.7 , 127.5 , 126.8 , 125.3 , 123.8 , 123.3 , 122.6 , 122.0 , 79.2 , 47.6 , 28.5 , 23.1 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₀H₂₃N₂O₂ 323.1754; found 323.1768.

tert-butyl (1-(phenanthridin-6-yl)-2-phenylethyl)carbamate (33) Following the general procedure condition A, **33** was purified by silica gel chromatography (PE:EA=5:1) ; 70.1 mg (88%); white solid; mp : 205-207 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.63 (d, J = 8.3 Hz, 1H), 8.55 (d, J = 8.1 Hz, 1H), 8.15 (d, J = 8.3 Hz, 1H), 8.10 (d, J = 8.0 Hz, 1H), 7.81 (t, J = 7.5 Hz, 1H), 7.72 (t, J = 7.5 Hz, 1H), 7.68 – 7.58 (m, 2H), 7.17 – 7.07 (m, 3H), 6.98 (dd, J = 6.6, 3.0 Hz, 2H), 6.35 (d, J = 8.3 Hz, 1H), 5.98 (dt, J = 8.4, 6.3 Hz, 1H), 3.42 (dd, J = 13.3, 6.5 Hz, 1H), 3.25 (dd, J = 13.4, 6.2 Hz, 1H), 1.46 (s, 9H). ¹³C {¹H} NMR (101 MHz, CDCl₃) δ 159.1 , 155.3 , 142.9 , 137.3 , 132.9 , 130.5 , 129.8 , 129.7 , 128.5 , 127.9 , 127.3 , 126.8 , 126.2 , 125.4 , 123.9 , 123.7 , 122.4 , 121.9 , 79.2 , 52.3 , 42.7 , 28.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₆H₂₈N₂O₂ 399.2067; found 399.2072.

2-((2-(4-chlorophenoxy)-2-methylpropanoyl)oxy)ethyl 6-(1-((tert-butoxycarbonyl)amino)ethyl)nicotinate (34) Following the general procedure condition A, **34** was purified by silica gel chromatography (PE:EA=10:1) ; 75.9 mg (75%); colourless oil ; ¹H NMR (500 MHz, CDCl₃) δ 9.00 (dd, J = 2.0, 0.8 Hz, 1H), 8.05 (dd, J = 8.1, 2.2 Hz, 1H), 7.29 (d, J = 8.0 Hz, 1H), 7.12 – 7.01 (m, 2H), 6.80 – 6.68 (m, 2H), 5.68 (d, J = 7.7 Hz, 1H), 4.89 (p, J = 7.0 Hz, 1H), 4.68 – 4.35 (m, 4H), 1.57 (s, 6H), 1.53 – 1.20 (m, 12H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 173.8 , 166.5 , 164.7 , 155.1 , 153.8 , 150.4 , 137.7 , 129.1 , 127.1 , 123.9 , 120.6 , 120.0 , 79.4 , 79.2 , 62.9 , 62.6 , 51.2 , 28.3 , 25.2 , 25.2 , 22.4 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₅H₃₂ClN₂O₇ 507.1893; found 507.1894.

tert-butyl (1-((1-(4-methylquinolin-2-yl)ethyl)amino)-1-oxopropan-2-yl)carbamate (35) Following the general procedure condition B, **35** was purified by silica gel chromatography (PE:EA=5:4) ; 40.0 mg (56%); colourless oil ; ¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, J = 8.2 Hz, 1H), 7.96 (d, J = 8.2 Hz, 1H), 7.88 (s, 1H), 7.69 (t, J = 9.1 Hz, 1H), 7.54 (t, J = 8.1 Hz, 1H), 7.15 (s, 1H), 5.36 – 5.05 (m, 2H), 4.32 (s, 1H), 2.68 (s, 3H), 1.53 (d, J = 6.8 Hz, 3H), 1.43 (s, 9H), 1.40 (d, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 171.8 , 160.1 , 155.3 , 146.9 , 145.3 , 129.5 , 129.4 , 129.3 , 129.3 , 127.3 , 126.1 , 123.7 , 120.0 , 79.7 , 50.3 , 50.2 , 50.0 , 49.9 , 22.4 , 22.2 , 19.0 , 18.8 . HRMS(ESI) m/z: [M + H]⁺ Calcd for C₂₀H₂₉N₃O₃ 358.2125; found 358.2133.

tert-butyl 12-((1-((2-(4-(tert-butoxy)phenyl)-1-(4-methylquinolin-2-yl)ethyl)amino)-3-methyl-1-oxobutan-2-yl)carbamoyl)-9-(4-((tert-butoxycarbonyl)amino)butyl)-2,2-dimethyl-4,7,10-trioxo-6-(3-(3-((2,2,4,6,7-pentamethyl-2,3-dihydrobenzofuran-5-yl)sulfonyl)guanidino)propyl)-3-oxa-5,8,11-triazatetradecan-14-oate (36) Following the general procedure condition B, HRMS(ESI) m/z: [M + H]⁺ Calcd for C₇₀H₁₀₅N₁₀O₁₄S 1341.7527; found 1341.7512.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website.

Optimization, quenching experiment and NMR spectra.

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENT

We gratefully acknowledge the financial support from the National Natural Science Foundation of China (Nos. 21772085 and 21971107).

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