Synthesis and Biological Evaluation of 5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]oxazoles, -thiazoles, and -imidazoles: Novel Dual 5-Lipoxygenase and Cyclooxygenase Inhibitors with Antiinflammatory Activity¹

Paul C. Unangst,* David T. Connor, Wiaczesław A. Cetenko, Roderick J. Sorenson, Catherine R. Kostlan, Jagadish C. Sircar,[†] Clifford D. Wright, Denis J. Schrier, and Richard D. Dyer

Departments of Chemistry, Immunopathology, and Biochemistry, Parke-Davis Pharmaceutical Research, Division of Warner-Lambert Company, 2800 Plymouth Road, Ann Arbor, Michigan 48105

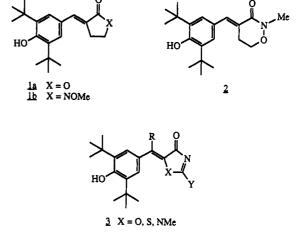
Received September 7, 1993®

A variety of benzylideneoxazoles, -thiazoles, and -imidazoles derived from 2,6-di-*tert*-butylphenol were prepared and evaluated as dual inhibitors of 5-lipoxygenase and cyclooxygenase in rat basophilic leukemia (RBL-1) cells. The target compounds exhibit varying degrees of selectivity toward the two enzymes. Several compounds are orally active in the rat carageenan footpad edema (CFE) and mycobacterium footpad edema (MFE) antiinflammatory models. Structure-activity relationships are discussed. From this work, (Z)-5-[[3,5-bis(1,1-dimethylethyl)-4-hydroxyphenyl]-methylene]-2-imino-4-thiazolidinone methanesulfonate salt (CI-1004) was identified as a potent dual inhibitor of 5-lipoxygenase (IC₅₀ = 0.77 μ M) and cyclooxygenase (IC₅₀ = 0.39 μ M), with oral activity (ID₄₀ = 0.6 mg/kg) in the rat MFE model of inflammation.

Nonsteroidal antiinflammatory drugs (NSAIDS) are widely used in the treatment of rheumatoid arthritis and other inflammatory diseases.² However, long-term NSAID use has been associated with gastrointestinal ulceration, bleeding, and nephrotoxicity.³ The beneficial effects of NSAIDS have been historically attributed to inhibition of the enzyme cyclooxygenase, thereby preventing production of proinflammatory prostaglandins.^{4a} Recently, two related isoenzymes of cyclooxygenase have been identified in mammalian cells.4b,c One isoenzyme is thought to be involved in the conversion of arachidonic acid to regulatory prostaglandins that control normal kidney and gastric function, while the other isoenzyme produces prostaglandins involved in inflammation. The identification of selective isoenzyme inhibitors may eventually lead to safer drugs for inflammation therapy.

In addition to cyclooxygenase (CO), the 5-lipoxygenase (5-LO) enzyme is also associated with the metabolism of cellular arachidonic acid.⁵ Leukotrienes, produced through the 5-lipoxygenase enzyme pathway, may also contribute to both inflammation and NSAID-induced side effects.⁶ For these reasons, compounds that are dual inhibitors of both cyclooxygenase and 5-lipoxygenase are being studied as potential antiinflammatory agents with an improved safety profile in comparison to NSAIDS.

Derivatives of 2,6-di-*tert*-butylphenol have been extensively studied as dual CO/5-LO inhibitors. Earlier reports from our laboratories described a variety of chemical series in which the di-*tert*-butylphenol ring is either directly bonded^{7a,b} or linked by a two-carbon chain^{7c} to various heterocyclic systems. Others⁸ have reported related compounds in which the phenol portion of the molecule is part of a benzylidene moiety attached to a heterocyclic ring. Examples of the latter type include 1a (KME-4),⁹ 1b (E-5110),¹⁰ and 2 (BF-389).¹¹



Many di-*tert*-butylphenol bicyclic derivatives are highly lipophilic compounds, resulting in generally poor aqueous solubility and associated bioavailability. Results from our earlier chemical series^{7a,b} demonstrated that the addition of ionizable functionality to the heterocyclic ring linked to the phenol ring would reduce the lipophilicity and aid in adsorption and elimination of these compounds.

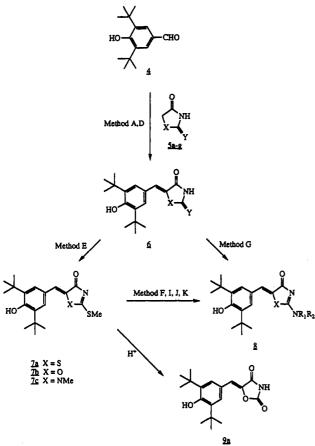
This paper describes the synthesis and biological activity of a series of benzylideneoxazoles, -thiazoles, and -imidazoles, generically represented in 3. Many of these compounds are dual inhibitors of cyclooxygenase and 5-lipoxygenase activity as measured in rat basophilic leukemia (RBL-1) cells. A number of compounds also show oral activity in animal models of inflammation.

Chemistry

A series of benzylidene derivatives 6 (Scheme 1) were prepared by Knoevenagel condensation (methods A, D) of the di-*tert*-butyl aldehyde 4 with heterocycles 5a-c, 5d,¹² 5e, 5f,¹³ and $5g^{14}$ (Table 1). Alkylation of benzylidene thiones 6c,¹⁵ 6g, and 6j with iodomethane (method E) provided the methyl thioether intermediates 7a-c. Reaction of 7a-c with various amines (methods F, I, J, K)

 [†] Current address: Biofor Ltd., P.O. Box 629, Waverly, PA 18471.
 [•] Abstract published in Advance ACS Abstracts, December 15, 1993.

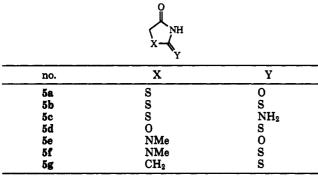
Scheme 1^a



^a Method A: NaOAc, HOAc. Method D: β -alanine, HOAc. Method E: MeI, (i-Pr)₂NEt. Method F: R₁R₂NH. Method G: R₁R₂NH, BaCO₃. Method I: R₁R₂NH, t-BuOK. Method J: guanidine, t-BuOK. Method K: cyanamide, t-BuOK. Variations 5a-g are listed in Table 1, while variations of 6 and 8 are listed in Table 3.

 Table 1. Intermediate Heterocycles for Knoevenagel

 Condensation



gave the amino derivatives 8. Additional examples of 8 were obtained by reaction of amines with thione 6c (method G). A benzylideneoxazoledione 9a was obtained by acidic hydrolysis of thioether 7b. Benzylidene derivatives 6-9 are listed in Table 3.

A sequence of reactions similar to that described in Scheme 1 was used to prepare analogs containing a methyl substituent on the benzylidene double bond. Knoevenagel condensation (method L) of the di-*tert*-butyl ketone 10^{16} (Scheme 2) with heterocycles 5a and 5b gave thiazoles 11a and 11c, respectively. Alkylation of 11c (method E) provided the thioether intermediate 12, and reaction of 12 with cyanamide (method K) or guanidine (method J) gave derivatives 13a and 13b, respectively.

Table 2. Intermediate Aldehydes for Knoevenagel Condensation

	R ₃ CHO	
	R ₄ R ₃	
no.	R ₃	R4
14a	Ph	ОН
14b	i-Pr	OH
14c	I	OH
14d	н	OH
1 4e	t-Bu	н

Benzylidenethiazoles 15 (Scheme 3) without the di-*tert*butylphenol substituent pattern were obtained by Knoevenagel condensation (method D) of heterocycle 5a with aldehydes 14a, 14b,¹⁷ 14c,¹⁸ 14d, and 14e.¹⁹ The aldehydes used are listed in Table 2, while the thiazole products 11, 13, and 15 are listed in Table 4.

Choline (methods B, C), methanesulfonate, and hydrochloride (method H) salts of selected target compounds were prepared as described in the Experimental Section.

In all cases of the Knoevenagel condensation, only a single stereoisomer was obtained for the olefinic product.²⁰

Table 5 summarizes the physicochemical data for the benzylideneoxazoles, -thiazoles, and -imidazoles prepared.

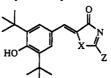
Results and Discussion

Our overall goal was the discovery of a balanced dual inhibitor of cyclooxygenase and 5-lipoxygenase with substantial antiinflammatory activity and a low potential for ulcerogenicity. Test compounds were evaluated for inhibition of PGF_{2 α} (a product of CO) formation and LTB₄ (a product of 5-LO) formation in intact rat basophilic leukemia (RBL-1) cells stimulated with the calcium ionophore A23187. Those compounds with IC_{50} values $\leq 10 \ \mu M$ for both enzymes were then tested in an acute animal model of inflammation, the carrageenan footpad edema (CFE) test. Compounds active in CFE (defined as showing $\geq 40\%$ inhibition at a dose of 30 mg/kg po) were next tested in the mycobacterium footpad edema (MFE) test, a 3-day subacute model more demanding of antiinflammatory efficacy than CFE. Selected compounds with oral activity in both the CFE and MFE tests received additional pharmacological evaluation.

Results from the enzyme assays and in vivo antiinflammatory models for compounds 6-9 are shown in Table 3. Thiazole and oxazole di-*tert*-butylphenols with a 2-hydroxy (6a and 9b), 2-mercapto (6c and 6g), or 2-amino (6f) substituent were found to be potent dual inhibitors and orally active in the animal models. The corresponding imidazoles 6i and 6j, although dual inhibitors, were inactive in vivo, while the pyrrolidine derivative 6k and the thiazole thioether 7a were active in CFE but not in MFE.

In view of the potent activity of thiazoles 6a, 6c, and 6f, a further series (8a-o) of 2-substituted thiazoles was synthesized. Several additional dual inhibitors were identified, including compounds with a methoxymethylamino (8h), cyanoimino (a bioisosteric thione replacement;²¹ 8i), and guanidino (8l) substituent. However, these compounds did not exhibit a satisfactory overall profile after secondary pharmacological evaluation.

The cyanoimino and guanidino substituent patterns were also investigated in the oxazole (8q and 8s) and imidazole (8t and 8v) series. Once again, potent dual inhibitors were obtained in both series.



no.			$\mathrm{IC}_{50},\mu\mathrm{M}^a$		$CFE:^{b}$ inhibn ± SEM at dose in mg/kg po			
	х	Z	5-LO	co	30	10	MFE: ID40, mg/kg p	
6 a	S	OH	1.4	0.35	59 ± 4.6^{d}	54 ± 4.3	2.0 (1.0-3.2) ^d	
6c .	S	SH	0.38	0.012	36 ± 5.0^{e}	32 ± 4.4		
6 f	S	NH ₂ ·MeSO ₃ H	0.77	0.39	61 ± 4.5	52 ± 3.6	0.6 (0.3-0.9)	
6g	0	SH	0.84	1.7	40 ± 2.9	32 ± 6.5	12.4 (8.0-20.2)	
6i	NMe	ОН	0.78	4.6	N ^f			
6j	NMe	SH	0.17	0.79	N			
6k	CH_2	SH	1.7	2.4	39 ± 3.9	19 ± 4.7	N ^g	
7a	s	SMe	0.65	0.60	47 ± 7.1	36 ± 6.2	N	
8a	S	NHMe	\mathbf{N}^{h}	Ν				
8b	8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8	NMe ₂	65 ⁱ	Ν				
8c	S	NHOH	3.7	0.80	\mathbf{N}^{j}			
8e	S	NMeOH	7.3	10.0	N			
8 f	S	NHOMe	Ν	61 ⁱ				
8 h	S	NMeOMe-HCl	1.7	0.33	36 🗭 2.7	41 ± 3.8	6.2 (2.4-11.8)	
8i	S	NHCN	0.63	0.16	49 ± 5.9^{k}	29 ± 4.4	N ^k	
81	S	NHCNHNH2•HCl	0.91	0.083	40 ± 3.1	34 ± 5.4	2.2 (0.8-3.9)	
8m	S	NHCNHNMe ₂	100	Ν				
8 n	S	NH(CH ₂) ₂ NMe ₂	79 ⁱ	Ν				
8o	S	NH(CH ₂) ₃ CO ₂ H	53 ⁱ	N				
8q	0	NHCN-choline	2.6	1.1	N			
8s	0	NHCNHNH2·MeSO3H	1.2	0.34	48 ± 3.3	33 ± 4.0	5.7 (4.1-7.5)	
8t	NMe	NHCN	0.70	3.5	\mathbf{N}^{l}			
8v	NMe	NHCNHNH ₂	1.3	2.1	N			
9b	0	OH-choline	2.8	0.65	51 ± 2.7	35 ± 3.6	3.7 (2.4-5.0)	
1a			2.5	0.15	28 ± 5.1	17 ± 9.5	3.7 (2.0-7.2)	
1b			0.56	0.040	26 ± 4.7	29 ± 3.1	1.9 (1.1-2.9)	
	neclofenamate		24.0	0.10	41 ± 5.3	34 ± 5.5	0.40(0.02-1.0)	

^a The concentration (μ M) of test compound causing 50% inhibition of LTB₄ (5-LO) or PGF_{2a} (CO) formation. The standard errors average 11% of the values shown for 5-LO and 8% for CO. ^b Percent inhibition ±SEM of edema in the carageenan footpad edema test. ^c The dose (mg/kg) of test compound causing 40% inhibition of induced edema in the mycobacterium footpad edema test; 95% confidence limits are in parentheses. ^d Data from the choline salt 6b. ^e Data from the choline salt 6b. ^f Inactive (N) is defined as <25% inhibition at a dose of 30 mg/kg po. ^s Inactive (N) is defined as <30% inhibition at a dose of 10 mg/kg po. ^h Inactive (N) is defined as <50% inhibition at a screening concentration of 10 μ M. ⁱ Percent inhibition at 10 μ M screening concentration. ^j Data from the choline salt 8d. ^k Data from the choline salt 8j. ^l Data from the choline salt 8u.

Biological test results for compounds 11-15 are shown in Table 4. Analogs of the thiazole dual inhibitors 6a, 6c, 8i, and 8l were prepared in which the benzylidene proton was replaced by a methyl group (11b, 11c, 13a, and 13b). In all cases, the methyl derivatives were inactive in the animal models and were generally of lower potency as dual inhibitors when compared to the corresponding nonmethyl analogs.

Thiazole 6a was also used as a model to explore a limited substituent variation in the phenol ring of the molecule. Replacement of both *tert*-butyl groups of 6a with phenyl (15a), isopropyl (15c), iodo (15d), or hydrogen (15e) resulted in compounds with overall activity inferior to that of 6a. Replacement of the phenolic hydroxyl group of 6a with hydrogen (15f) produced a dual inhibitor without activity in the animal models. These results agree with earlier work^{7c,22,23} in a number of diverse chemical series demonstrating the preference for the di-*tert*-butylphenol substructure.

The 2-hydroxythiazole **6a** and the 2-aminothiazole **6f** were selected for additional evaluation. These two compounds exhibited an improved balance in the enzyme inhibitory assays and a similar potency in the in vivo models when compared to the standards **1a**, **1b**, and sodium meclofenamate.

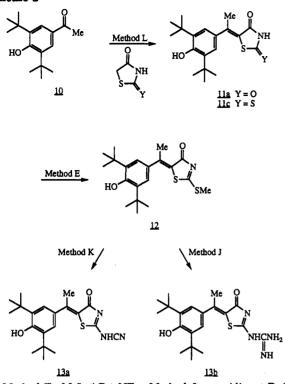
Thiazoles 6a and 6f were evaluated for gastric ulcerogenic potential in a rat stress model.^{7a} When administered orally to fasted rats (N = 10 animals), neither compound caused ulcers at a 200 mg/kg dose in contrast to 1a (30% incidence of ulcers at 30 mg/kg dose) and sodium meclofenamate (50% incidence of ulcers at 36 mg/kg dose). When tested orally in an adjuvant induced polyarthritis assay²⁴ (a chronic model of inflammation in which sodium meclofenamate has $ID_{40} = 1.3$ mg/kg for prophylactic administration), 6a and 6f had ID_{40} values of 2.2 and 1.7 mg/kg, respectively.

We have described the synthesis of novel thiazole, oxazole, and imidazole derivatives of 3,5-di-*tert*-butylphenol. Several of these compounds are relatively balanced dual inhibitors of the 5-lipoxygenase and cyclooxgenase enzyme pathways. Thiazoles **6a** (designated as CI-987) and **6f** (designated as CI-1004) were identified as nonulcerogenic, orally active antiinflammatory agents.

Experimental Section

Melting points were determined on a Thomas-Hoover or Electrothermal capillary apparatus and are uncorrected. Elemental analyses were performed by the Analytical Chemistry staff of Parke-Davis (Ann Arbor, MI) and were within $\pm 0.4\%$ of the theoretical values, unless indicated otherwise. Infrared spectra were recorded as KBr disks on a Nicolet MX-1 FTIR spectrometer. Proton NMR spectra were recorded on a Brüker AM 250 spectrometer, with chemical shifts reported in δ units relative to internal TMS. Reactions were generally run under a nitrogen atmosphere. Organic solutions were concentrated at

Scheme 2^a



^a Method E: MeI, (i-Pr)₂NEt. Method J: guanidine, t-BuOK. Method K: cyanamide, t-BuOK. Method L: NH₄OAc, toluene.

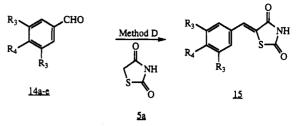
aspirator vacuum on a rotary evaporator. Flash chromatography was performed with E. Merck silica gel 60, 230–400-mesh ASTM, according to the method of Still.²⁵

Method A. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2,4-thiazolidinedione (6a). A suspension of 4 (6.5 g, 28 mmol), 5a (3.0 g, 26 mmol), and NaOAc (7.6 g, 93 mmol) in HOAc (40 mL) was stirred at reflux for 48 h. The cooled reaction mixture was added to H₂O (200 mL), and the precipitated product was filtered and washed with H₂O. Recrystallization from EtOH gave 2.3 g (27%) of 6a: mp 238-240 °C; IR 3619, 1741, 1583, 1214 cm⁻¹; ¹H NMR (DMSO- d_6) δ 1.41 (s, 18H, t-Bu), 7.38 (s, 2H, ArH), 7.77 (s, 1H, olefin), 7.79 (s, 1H, OH), 12.48 (br s, 1H, NH). Anal. (C₁₈H₂₃NO₃S) C, H, N.

Method B. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2,4-thiazolidinedione Choline Salt (6b). A stirred suspension of 6a (10.0 g, 30 mmol) in EtOH (100 mL) was treated dropwise with a 47% aqueous choline bicarbonate solution (10.6 g, 30 mmol). The mixture was heated at reflux for 1 h, cooled, and evaporated. Fresh EtOH (50 mL) was added and the mixture again evaporated two additional times. The final residue was triturated with Et₂O to yield 12.8 g (86%) of 6b: mp 215 °C dec; IR 1675, 1617, 1571, 1206 cm⁻¹; ¹H NMR (DMSO- d_6) δ 1.40 (s, 18H, t-Bu), 3.11 (s, 9H, NMe), 3.40 (m, 2H, CH₂), 3.85 (m, 2H, CH₂), 5.40 (br s, 1H, choline OH), 7.23 (s, 1H olefin), 7.30 (s, 2H, ArH). Anal. (C₁₈H₂₂NO₃S-C₆H₁₄NO) C, H, N.

(Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-imino-4-thiazolidinone Methanesulfonate Salt (6f). A solution of methanesulfonic acid (0.50 mL, 0.74 g, 7.7 mmol) in THF (4.0 mL) was added at room temperature to a stirred solution of 6e (2.3 g, 7.0 mmol) in THF (190 mL). The mixture was stirred for 30 min, and the solvent was evaporated. Trituration of the residue with Et₂O gave a solid which was recrystallized from MeOH/Et₂O to yield 2.7 g (90%) of 6f: mp 252 °C dec; IR 3600, 1734, 1608, 1192 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.42 (s, 18H, t-Bu), 2.43 (s, 3H, Me), 6.67 (br s, 2H, NH₂), 7.39 (s, 2H, ArH), 7.65 (s, 1H, olefin), 7.69 (br s <1H, OH), 9.58 (br s, 1H, NH). Anal. (C₁₈H₂₄N₂O₂S-CH₃SO₃H) C, H, N.

(Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-thioxo-4-oxazolidinone (6g). A mixture of 4 (18.8 g, 80 mmol), $5d^{12}$ (10.0 g, 85 mmol), and β -alanine (1.6 g, 18 mmol) in HOAc (28 mL) and toluene (80 mL) was stirred at Scheme 3^a



^a Method D: β -alanine, HOAc. Variations 14a-e are listed in Table 2. The condensation products 15 are listed in Table 4.

reflux through a Dean–Stark trap for 4 h. The solid that precipitated from the cooled reaction mixture was filtered and washed with hexane. Recrystallization from MeCN/H₂O gave 11.1 g (41%) of **6g**: mp 240 °C dec; IR 3612, 1728, 1653, 1432 cm⁻¹, ¹H NMR (DMSO-*d*₆) δ 1.41 (s, 18H, t-Bu), 6.77 (s, 1H, olefin), 7.72 (s, 2H, ArH), 7.74 (s, 1H, OH), 13.75 (br s, 1H, NH). Anal. (C₁₈H₂₃NO₃S) C, H, N.

Method C. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-thioxo-4-oxazolidinone Choline Salt (6h). A solution of 47% aqueous choline bicarbonate (2.5 g, 7.1 mmol) in MeOH (50 mL) was treated over 5 min with 6g (2.3 g, 6.9 mmol). The mixture was warmed to reflux for 5 min on the steam bath and filtered hot. The cooled filtrate was evaporated, and the residue was redissolved in methanol and reevaporated three times. Recrystallization of the final residue from acetone/ t-BuOMe gave 2.1 g (70%) of salt 6h: mp 167 °C dec; IR 1634, 1653, 1366, 1199 cm⁻¹; ¹H NMR (DMSO- d_{θ}) δ 1.40 (s, 18H, t-Bu), 3.11 (s, 9H, NMe), 3.39 (m, 2H, CH₂), 3.84 (m, 2H, CH₂), 5.32 (br s, 1H, choline OH), 6.03 (s, 1H, olefin), 7.26 (br s, 1H, phenol OH), 7.57 (s, 2H, ArH). Anal. (C₁₈H₂₂NO₃S·C₅H₁₄NO) C, H, N.

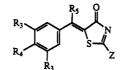
Method D. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-1-methyl-2-thioxo-4-imidazolidinone (6j). A mixture of 4 (19.0 g, 81 mmol), 5f¹³ (10.6 g, 81 mmol), and β -alanine (4.7 g, 53 mmol) in HOAc (150 mL) was stirred at reflux for 24 h. The cooled reaction mixture was added to H₂O (1.5 L) and stirred for 1 h. The precipitated product was filtered and washed with H₂O and then recrystallized from MeCN to yield 17.5 g (62%) of 6j: mp 242-244 °C; IR 3630, 1734, 1497, 1240 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.40 (s, 18H, t-Bu), 3.48 (s, 3H, NMe), 6.74 (s, 1H, olefin), 7.50 (s, 1H, OH), 8.07 (s, 2H, ArH), 12.29 (br s, 1H, NH). Anal. (C₁₉H₂₆N₂O₂S) H, N; C: calcd, 61.17; found, 60.60.

(Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-(methylthio)-4(5H)-oxazolone (7b). A solution of 6g (5.0 g, 15 mmol) in THF (75 mL) was cooled in ice and treated with Et₃N (2.4 mL, 1.7 g, 17 mmol). The mixture was stirred for 30 min, and MeI (4.5 mL, 10.3 g, 72 mmol) was added. The ice bath was removed, and the mixture was stirred for 24 h. The insoluble material was filtered, and the filter cake was washed several times with fresh THF. The filtrates were evaporated, and the residue was purified by flash chromatography (2.5–5% EtOAc in CH₂Cl₂) to yield 4.3g (83%) of 7b. Sample recrystallized from EtOAc/hexane: mp 164–166 °C; IR 3568, 1719, 1656, 1505 cm⁻¹; ¹H NMR (CDCl₃) δ 1.47 (s, 18H, t-Bu), 2.74 (s, 3H, SMe), 5.68 (s, 1H, OH), 6.82 (s, 1H, olefin), 7.60 (1s, 2H, ArH). Anal. (C₁₉H₂₆NO₃S) C, H, N.

Method E. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-1,5-dihydro-1-methyl-2-(methylthio)-4H-imidazol-4-one (7c). A suspension of 6j (4.0 g, 12 mmol) and N,N-diisopropylethylamine (2.4 mL, 1.8 g, 14 mmol) in EtOH (50 mL) was treated with MeI (1.2 mL, 2.7 g, 19 mmol). The mixture was stirred at room temperature for 18 h and then added to H₂O (300 mL). After 1 h the product was filtered, washed with H₂O, and recrystallized from EtOAc to yield 3.0 g (70%) of 7c: mp 177-179 °C; IR 3572, 1677, 1588, 1430 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.42 (s, 18H, t-Bu), 2.63 (s, 3H, SMe), 3.32 (s, 3H, NMe), 6.78 (s, 1H, olefin), 7.47 (br s, 1H, OH), 8.20 (s, 2H, ArH). Anal. (C₂₀H₂₈N₂O₂S-0.25CH₃CN) C, H, N.

Method F. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-(methylamino)-4(5H)-thiazolone (8a). A mixture of 7a (3.6 g, 9.9 mmol) and 40% aqueous methylamine (5.3 g, 68 mmol) in EtOH (300 mL) was stirred at reflux for 22

Table 4. Biochemical and in Vivo Antiinflammatory Activity of Benzylidene Derivatives 11, 13, 15



no. Z			$\mathrm{IC}_{50},\mu\mathbf{M}^{a}$		CFE: ^b % inhibn ± SEM at dose in mg/kg po				
	Z	R_3	R_4	R4 R5	5-LO	CO	30	10	MFE:° ID ₄₀ , mg/kg po
11b	OH-choline	t-Bu	OH	Me	0.48	52 ^d			
11c	SH	t-Bu	OH	Me	2.3	4.5	Nej		
13a	NHCN	t-Bu	OH	Me	1.2	5.1	N		
13b	NHCNHNH ₂	t-Bu	ŎН	Me	1.6	1.9	N		
15a	OH	Ph	OH	H	0.16	9.6	41 ± 5.1^{g}	11 ± 7.6	N ^{g,h}
15c	OH	i-Pr	OH	H	0.22	0.82			Ň
15d	OH	Ī	ŎĦ	H	Ni	60 ^d			
15e	OH	Ĥ	ŎН	Ĥ	N	Ň			
15f	OH	t-Bu	H	Ĥ	3.3	4.6	\mathbf{N}^{j}		

^a The concentration (μ M) of test compound causing 50% inhibition of LTB₄ (5-LO) or PGF_{2a} (CO) formation. The standard errors average 11% of the values shown for 5-LO and 8% for CO. ^b Percent inhibition ±SEM of edema in the carageenan footpad edema test. ^c The dose (mg/kg) of test compound causing 40% inhibition of induced edema in the mycobacterium footpad edema test; 95% confidence limits are in parentheses. ^d Percent inhibition at 10 μ M screening concentration. ^e Data from the choline salt 11d. ^f Inactive (N) is defined as <25% inhibition at a dose of 30 mg/kg po. ^s Data from choline salt 15b. ^h Inactive (N) is defined as <30% inhibition at a dose of 10 mg/kg po. ⁱ Inactive (N) is defined as <50% inhibition at a screening concentration of 10 μ M. ^j Data from the choline salt 15g.

h. After cooling to room temperature, the precipitated product was filtered and washed with EtOH. Recrystallization from DMF/EtOAc gave 2.4 g (69%) of 8a: mp 308-310 °C; IR 3541, 1679, 1591, 1260 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.44 (s, 18H, t-Bu), 3.14 (s, 3H, Me), 7.41 (s, 2H, ArH), 7.71 (s, 1H, olefin). Anal. (C₁₉H₂₆N₂O₂S) C, H, N.

Method G. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-(hydroxymethylamino)-4(5H)-thiazolone (8e). A mixture of N-methylhydroxylamine hydrochloride (12.5 g, 150 mmol), BaCO₃ (14.8 g, 75 mmol), EtOH (50 mL), and H₂O (50 mL) was stirred at room temperature for 10 min. Additional EtOH (600 mL) and 6c (17.5 g, 50 mmol) were added, and the new mixture was stirred at reflux for 8 h. The cooled reaction mixture was filtered and the filtrate evaporated. The residue was stirred in ice H_2O , and the resulting solid was filtered, washed with H₂O, and dissolved in CH₂Cl₂. The organic layer was dried (Na₂SO₄) and evaporated to give 19 g of residue. Purification by flash chromatography (CH₂Cl₂, followed by 9:1 CH₂Cl₂/MeOH), followed by recrystallization from MeOH/EtOAc gave 8.6g (48%) of 8e: mp 199-200 °C; IR 3627, 1613, 1438, 1210 cm^{-1} ; ¹H NMR (DMSO- d_{6}) δ 1.42 (s, 18H, t-Bu), 3.54 (s, 3H, Me), 7.38 (s, 2H, ArH), 7.58 (s, 1H, olefin), 7.63 (br s, <1H, phenol OH), 11.80 (br s, <1H, NOH). Anal. (C₁₉H₂₆N₂O₃S) C, H, N.

Method H. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-(methoxymethylamino)-4(5H)-thiazolone Hydrochloride (8h). A solution of 8g (8.3 g, 22 mmol) in CH₂Cl₂ (50 mL) was treated with excess ethereal HCl. The precipitated salt was filtered and recrystallized from CH₂Cl₂/ MeOH/Et₂O to yield 6.0 g (66%) of salt 8h: mp 177-179 °C; IR 3417, 1734, 1588, 1422 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.42 (s, 18H, t-Bu), 3.56 (s, 3H, NMe), 3.92 (s, 3H, OMe), 7.40 (s, 2H, ArH), 7.67 (s, 1H, olefin). Anal. (C₂₀H₂₈N₂O₃S-HCl) H, N; C: calcd, 58.17; found, 58.60.

Method I. (Z)-N-[5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-4-oxo-2-thiazolyl]-N,N-dimethylguanidine (8m). A mixture of 1,1-dimethylguanidine sulfate (6.7 g, 25 mmol) and t-BuOK (4.8 g, 43 mmol) in EtOH (500 mL) was stirred at room temperature for 15 min. After the addition of 7a (7.3 g, 20 mmol), the new mixture was stirred at reflux for 6 h. The cooled mixture was evaporated, and the residue was partitioned between Et₂O and ice H₂O made slightly acidic with 1.0 N HCl. The organic layer was washed with brine, dried (Na₂SO₄), and evaporated. Purification of the residue by flash chromatography (CH₂Cl₂, then 9:1 CH₂Cl₂/MeOH elution) followed by recrystallization from EtOH/DMF gave 5.1 g (63%) of **8m**: mp 293-295 °C; IR 3620, 1660, 1513, 1261 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.43 (s, 18H, t-Bu), 3.07 (s, 3H, NMe), 3.14 (s, 3H, NMe), 7.41 (s, 2H, ArH), 7.85 (s, 1H, olefin). Anal. (C₂₁H₃₀N₄O₂S) C, H, N. (Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2-[[2-(dimethylamino)ethyl]amino-4(5H)-thiazolone (8n). A mixture of $6c^{15}$ (7.0 g, 20 mmol) and (N,Ndimethylamino)ethylamine (5.3 g, 60 mmol) in EtOH (250 mL) was stirred at reflux for 8 h. The solvent was evaporated and the residue recrystallized first from toluene and then from MeOH/ EtOAc/toluene to give 3.4 g (42%) of 8n: mp 209-211 °C; IR 1684, 1628, 1440, 1315 cm⁻¹; ¹H NMR (DMSO- d_{θ}) δ 1.42 (s, 18H, t-Bu), 2.19 (s, 6H, NMe), 2.46 (t, J = 6 Hz, 2H, CH₂), 3.59 (t, J = 6 Hz, 2H, CH₂), 7.36 (s, 2H, ArH), 7.53 (s, 1H, olefin). Anal. (C₂₂H₃₃N₃O₂S) C, H, N.

(Z)-4-[[[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-4-oxo-2-thiazolyl]amino]butanoic Acid (80). A mixture of 4-aminobutyric acid (1.9 g, 18 mmol) and t-BuOK (1.4 g, 12 mmol) in EtOH (250 mL) was stirred at room temperature for 30 min and then treated with 7a (3.6 g, 9.9 mmol). The new mixture was stirred at reflux for 22 h, cooled, and evaporated. The residue was treated with cold H₂O made slightly acidic with 2.0 N HCl. The insoluble solid was filtered and washed with H_2O . The crude product was dissolved in CH_2 -Cl₂/THF, and the solution was dried (Na₂SO₄) and evaporated. The residue was digested in warm Et₂O and filtered. Recrystallization of the final solid from THF/EtOAc gave 2.7 g (65%) of 80: mp 239-240 °C; IR 3621, 1717, 1570, 1423 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.42 (s, 18H, t-Bu), 1.84 (m, 2H, CH₂CH₂CH₂), 2.33 $(t, J = 7 Hz, 2H, CH_2CO_2H), 3.52 (m, 2H, NHCH_2), 7.36 (s, 2H)$ ArH), 7.56 (s, 1H, olefin), 9.54 (m, 1H, NH), 12.00 (br s, <1, CO_2H). Anal. ($C_{22}H_{30}N_2O_4S$) C, N, H.

(Z)-[5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-4-oxo-2-oxazolyl]cyanamide (8p). A suspension of cyanamide (0.38 g, 9.0 mmol) in EtOH (50 mL) was cooled in ice and treated in small portions with t-BuOK (0.93 g, 8.3 mmol). After the mixture was stirred for 15 min, 7b (2.6 g, 7.5 mmol) was added, followed by additional EtOH (25 mL). The mixture was stirred at reflux for 3 h and then cooled, and the precipitated potassium salt of the product was filtered and washed with Et_2O . The salt was suspended in H_2O (100 mL) and acidified with 1.0 mL of HOAc. The mixture was stirred for 30 min and extracted with EtOAc. The organic layer was washed with brine, dried (Na_2SO_4) , and evaporated to yield 1.0g(38%) of 8p. Sample recrystallized from EtOAc/hexane: mp 220 °C dec; IR 3545, 2219, 1755, 1431 cm⁻¹; ¹H NMR (DMSO- d_6) δ 1.41 (s, 18H, t-Bu), 6.88 (s, 1H, olefin), 7.71 (s, 2H, ArH), 7.72 (s, 1H, OH). Anal. $(C_{19}H_{23}N_3O_3)$ C, H, N.

Method J. (Z)-[5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-4-oxo-2-oxazolyl]guanidine (8r). A solution of guanidine hydrochloride (2.0g, 21 mmol) in EtOH (75 mL) was cooled in ice and treated in portions with t-BuOK (2.2 g, 20 mmol). The mixture was stirred for 15 min

Table 5.	Physicochemical	Data for	Benzylidene	Derivatives
----------	-----------------	----------	-------------	-------------

no.	method	yield, %	mp, °C	recryst solvent	formula	anal.
6a	A	26	238-240	EtOH	C ₁₈ H ₂₈ NO ₈ S	C,H,N
6bª	B	98	215 dec	Ь	$C_{18}H_{22}NO_3S \cdot C_5H_{14}NO$	C,H,N
6c	Α	30	248-249	EtOAc	$C_{18}H_{23}NO_2S_2$	C,H,N
6da	В	93	196 dec	Ь	C ₁₈ H ₂₂ NO ₂ S ₂ ·C ₅ H ₁₄ NO	C,N; H ⁴
6e	Α	38	277-279	EtOAc/MeOH	$C_{18}H_{24}N_2O_2S$	C.H.N
6 f d	е	90	252 dec	MeOH/Et ₂ O	C ₁₈ H ₂₄ N ₂ O ₂ ·CH ₃ SO ₃ H	C,H,N
6g	e	41	240 dec	MeCN/H ₂ O	C ₁₈ H ₂₃ NO ₃ S	C,H,N
6hª	Ċ	70	167 dec	acetone/t-BuOMe	C ₁₈ H ₂₂ NO ₃ S·C ₅ H ₁₄ NO	C.H.N
6i	Ď	10	211-212	EtOH	$C_{19}H_{26}N_2O_3$	Č,H,N
6j	Ď	62	242-244	MeCN	$C_{19}H_{26}N_2O_2S$	H.N; C
6 k	Ă	7	237-239	EtOH	$C_{19}H_{25}NO_2S$	C.H.N
	Ê	55		MeOH	$C_{19}H_{25}NO_2S_2$	C,H,N
7a.			159-160			C, I, N
7b	e	83	164-166	EtOAc/hexane	C ₁₉ H ₂₅ NO ₃ S	C,H,N
7 c	E	70	177-179	EtOAc	$C_{20}H_{28}N_2O_2S \cdot 0.25CH_3CN$	C,H,N
Ba	F	69	308-310	DMF/EtOAc	$C_{19}H_{26}N_2O_2S$	C,H,N
8b	F	77	247-249	MeOH	$C_{20}H_{28}N_2O_2S$	C,H,N
BC	Ğ	30	241 dec	$Et_2O/toluene$	C ₁₈ H ₂₄ N ₂ O ₃ S·CH ₃ OH	C,H,N
3d°	Ċ	27	178-180	MeOH/EtOAc	C ₁₈ H ₂₃ N ₂ O ₃ S·C ₅ H ₁₄ NO	H,N; C
Se .	G	48	1 99– 200	MeOH/EtOAc	$C_{19}H_{26}N_2O_3S$	C,H,N
ßf	I	77	245-246	MeOH	$C_{19}H_{26}N_2O_3S$	C.H.N
Bg	Ğ	27	187-189	CH ₂ Cl ₂ /EtOAc	$C_{20}H_{28}N_2O_3S$	C.H.N
Sh ^h	ň	66	177-179	CH ₂ Cl ₂ /MeOH/Et ₂ O	$C_{20}H_{28}N_2O_3S \cdot HCl$	H,N;C
Si	ĸ	70	240-242	MeOH	C ₁₉ H ₂₃ N ₃ O ₂ S·0.25CH ₃ OH	C,H,N
51 3ja	C	89	189-190			
	Ĩ	89 31		MeOH/EtOAc	$C_{19}H_{22}N_3O_2S \cdot C_5H_{14}NO$	C,H,N
3k	1		267-268	MeOH/EtOAc	$C_{19}H_{28}O_2S \cdot 0.5H_2O$	C,H; N
31 ^h	н	24	217-220	MeOH/EtOAc	C ₁₉ H ₂₈ N ₄ O ₂ S·HCl	C,H,N
3m	I	63	293-295	DMF/EtOH	$C_{21}H_{30}N_4O_2S$	C,H,N
8n	е	42	20 9– 211	MeOH/EtOAc/toluene	$C_{22}H_{33}N_3O_2S$	C,H,N
8 0	е	65	23 9– 240	THF/EtOAc	$C_{22}H_{30}N_2O_4S$	C,H,N
Sp	е	38	220 dec	EtOAc/hexane	C19H23N3O3	C.H.N
Baa	С	53	85 dec	Ъ	C ₁₉ H ₂₂ N ₃ O ₃ ·C ₅ H ₁₄ NO·0.5H ₂ O	C.H.N
Br	Ĵ	55	258 dec	MeCN/H ₂ O	C ₁₉ H ₂₆ N ₄ O ₃	C,H,N
38 ^d	e	83	278 dec	b	C ₁₉ H ₂₆ N ₄ O ₃ ·CH ₃ SO ₃ H	C,H,N
St	ĸ	51	257-259	MeCN	$C_{20}H_{26}N_4O_2$	C.H.N
Su ^a	B	94	185-186	b	$C_{20}H_{25}N_4O_{2*}C_5H_{14}NO$	C.H.N
]∦		288-290	-	$O_{20} I_{125} I_{4} O_{2} O_{5} I_{14} I_{4} O$	C; H,N
8v	-	47		EtOH/DMF	$C_{20}H_{29}N_5O_2$	
Ja	e	53	239 dec	EtOAc/hexane	$C_{18}H_{23}NO_4$	C,H,N
€ba	C	60	165 dec	acetone/t-BuOMe	C ₁₈ H ₂₂ NO ₄ ·C ₅ H ₁₄ NO	C,H,N
11a	L	67	253-254	b	$C_{19}H_{25}NO_{3}S$	C,H,N
11 b ª	В	9 6	191 dec	Ь	$C_{19}H_{24}NO_3S \cdot C_5H_{14}NO$	C,H,N
11c	\mathbf{L}	76	244-246	MeCN	$C_{19}H_{25}NO_2S_2$	C,H,N
l 1da	В	77	213 dec	ь	$C_{19}H_{24}NO_2S_2C_5H_{14}NO$	C,H,N
L 2	Е	93	224-226	MeCN	$C_{20}H_{27}NO_2S_2$	C,H,N
	ĸ	88	229-230	MeCN	$C_{20}H_{25}N_3O_2S$	C,H,N
13b	J	92	277 dec	MeCN/EtOH	$C_{20}H_{23}N_4O_2S$	C.H.N
15a	Ď	54	228-229	MeOH	$C_{22}H_{15}NO_{3}S$	C; H,N'
15 b ª	B	91	113-125	b	$C_{22}H_{14}NO_{3}S C_{5}H_{14}NO 0.5H_{2}O$	C,H,N
	D	91 98		0 b		
15c			201-203		C ₁₆ H ₁₉ NO ₃ S	C,H,N
15d	D	85	>285	b	C ₁₀ H ₅ I ₂ NO ₃ S	C,H,N
15e	D	89	>290	<i>b</i>	C10H7NO38	C,H,N
l 5f	D	56	193-197	MeOH	C ₁₈ H ₂₃ NO ₂ S	C,H,N
15ga	В	88	216 dec	Ь	$C_{18}H_{22}NO_2S \cdot C_5H_{14}NO$	C,H,N

^a This compound is the choline salt of the preceding compound in the table. ^b Compount not recrystallized. ^c H: calcd, 8.02; found, 7.59. ^d This compound is the methanesulfonate salt of the preceding compound in the table. ^e See the Experimental Section. ^f C: calcd, 65.86; found 66.27. ^g C: calcd, 61.17; found, 60.60. ^h This compound is the hydrochloride salt of the preceding compound in the table. ⁱ C: calcd, 58.17; found, 58.60. ^j N: calcd, 14.61; found, 14.12. ^k Heating time increased to 21 h. ⁱ C: calcd, 64.67; found, 65.08. ^m C: calcd, 70.76; found, 70.31.

and then rapidly filtered into a suspension of 7b (4.6 g, 13 mmol) in EtOH (75 mL). The new reaction mixture was stirred at reflux for 3 h, cooled, and filtered. The filtrate was condensed 50% and added to ice and H_2O (500 g). The precipitated solid was filtered, washed with H_2O , and dissolved in EtOAc (200 mL). The solution was washed with brine, dried (Na₂SO₄), and evaporated. Purification of the residue by flash chromatography (10% MeCN in EtOAc elution) gave 2.6 g (55%) of 8r. Sample recrystallized from MeCN/H₂O: mp 258 °C dec; IR 1708, 1653, 1560, 1301 cm⁻¹; ¹H NMR (DMSO- d_6) δ 1.40 (s, 18H, t-Bu), 6.43 (s, 1H, olefin), 7.33 (br s, 2H, NH), 7.40 (s, 1H, OH), 7.56 (s, 2H, ArH), 8.18 (br s, 2H, NH). Anal. (C₁₉H₂₆N₄O₈) C, H, N.

(Z)-N-[5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-4-oxo-2-oxazolyl]guanidinium Methanesulfonate Salt (8s). A solution of 8r (1.9 g, 5.3 mmol) in i-PrOH (100 mL) was filtered warm and treated immediately with a solution of methanesulfonic acid (0.40 mL, 0.59 g, 6.2 mmol) in a few mL of i-PrOH. Upon cooling, the precipitated salt was filtered and washed with Et₂O to yield 2.0 g (83%) of 8s: mp 278 °C dec; IR 1773, 1669, 1432, 1172 cm⁻¹; ¹H NMR (DMSO- d_6) δ 1.40 (s, 18H, t-Bu), 2.34 (s, 3H, Me), 6.83 (s, 1H, olefin), 7.63 (s, 2H, ArH), 7.67 (s, 1H, OH), 8.29 (br s, 2H, NH), 8.55 (br s, 2H, NH). Anal. (C₁₉H₂₆N₄O₃·CH₃SO₃H) C, H, N.

Method K. (Z)-[5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-4,5-dihydro-1-methyl-4-oxo-1*H*-imidazol-2-yl]cyanamide (8t). Cyanamide (0.20 g, 4.8 mmol) was added to a mixture of 7c (1.5 g, 4.2 mmol) and t-BuOK (0.50 g, 4.5 mmol) in EtOH (25 mL). The mixture was stirred at reflux for 2.5 h, cooled, and added to H_2O (200 mL). The mixture was acidified with H_3PO_4 , and the precipitated product was filtered and washed with H_2O . Recrystallization from MeCN gave 0.70 g (51%) of 8t: mp 257-259 °C; IR 3622, 2198, 1745, 1388 cm⁻¹; ¹H NMR (DMSO-d₆) δ 1.41 (s, 18H, t-Bu), 3.21 (s, 3H, NMe), 6.69 (s, 1H, olefin), 7.48 (s, 1H, OH), 7.99 (s, 2H, ArH). Anal. (C₂₀H₂₈₀N₄O₂) C, H, N.

(Z)-5-[[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methylene]-2,4-oxazolidinedione (9a). A solution of 7b (3.0 g, 8.6 mmol) in warm EtOH (25 mL) was diluted with warm H₂O (8 mL). The solution was treated over 10 min with 5.0 mL of concentrated HCl. After the mixture was stirred for 4 h, the precipitated product was filtered and washed with hexane to yield 1.4 g (51%) of 9a. Sample recrystallized from EtOAc/hexane: mp 239 °C dec; IR 3613, 1827, 1664, 1332 cm⁻¹; ¹H NMR (DMSO- d_{6}) δ 1.40 (s, 18H, t-Bu), 6.69 (s, 1H, olefin), 7.58 (s, 1H, OH), 7.61 (s, 2H, ArH), 12.30 (br s, 1H, NH). Anal. (C₁₈H₂₃NO₄) C, H, N.

Method L. (Z)-5-[1-[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]ethylene]-2,4-thiazolidinedione (11a). A mixture of 10¹⁶ (4.0 g, 16 mmol), 5a (3.0 g, 26 mmol), and NH₄OAc (1.9 g, 25 mmol) in toluene (12 mL) was stirred at reflux for 48 h. The reaction mixture was cooled and evaporated. The residue was digested briefly in warm MeOH (20 mL), cooled, and filtered to provide 3.7 g (67%) of 11a: mp 253-254 °C; IR 3613, 1734, 1575, 1434 cm⁻¹; ¹H NMR (DMSO- d_{θ}) δ 1.39 (s, 18H, t-Bu), 2.66 (s, 3H, olefin Me), 7.21 (s, 2H, ArH), 7.46 (br s, 1H, OH), 12.17 (br s, 1H, NH). Anal. (C₁₉H₂₅NO₃S) C, H, N.

3,5-Diphenyl-4-hydroxybenzaldehyde (14a). A stirred mixture of 2,6-diphenylphenol (5.0 g, 20 mmol) and hexamethylenetetramine (5.5 g, 39 mmol) was treated dropwise with HOAc (45 mL). After 20 min, H₂O (10 mL) was added, and the mixture heated to reflux. Distillate was removed until the reaction pot temperature was 114 °C; reflux was then maintained for 6 h. The cooled mixture was treated with H₂O (20 mL), and the precipitated product was filtered and washed with H₂O. Recrystallization from EtOH gave 3.5 g (64%) of 14a: mp 168-170 °C; IR 1673, 1583, 1429, 1325 cm⁻¹; ¹H NMR (DMSO-d₆) δ 7.37-7.61 (m, 10H, ArH), 7.76 (s, 2H, ArH), 9.53 (br s, 1H, OH), 9.93 (s, 1H, CHO). Anal. (C₁₉H₁₄O₂) C, H.

Pharmacology. The whole cell 5-lipoxygenase and cyclooxygenase, carrageenan footpad edema, microbacterium footpad edema, and gastric ulcerogenicity assays were performed as previously described.^{7a}

Acknowledgment. We thank Dr. Gary McCluskey and staff for microanalytical and spectral data, Paul Kuipers and Dirk Bornemeier for the enzyme assays, and Godwin Okonkwo and Mark Lesch for animal testing.

References

- Presented in part in: (a) Kostlan, C. R.; Cetenko, W. J.; Connor, D. T.; Sorenson, R. J.; Sircar, J. C.; Bornemeier, D. A.; Dyer, R. D.; Kuipers, P. J.; Okonkwo, G. C.; Schrier, D. J., Wright, C. D. Structure-Activity Relationships of Thiazolidinone Derivatives as Dual Inhibitors of 5-Lipoxygenase and Cyclooxygenase. 6th International Conference of the Inflammation Research Association, White Haven, PA, September 20-24, 1992; abstract P113. (b) Sorenson, R. J.; Cetenko, W. A.; Connor, D. T.; Dyer, R. D.; Bornemeier, D. A.; Kuipers, P. J. Benzylidenethiazolidinones as Dual Inhibitors of Cyclooxygenase and 5-Lipoxygenase: Potential Anttiinflammatory Agents. 205th National ACS Meeting, Denver, CO, March 28-April 2, 1993; MEDI 141. (c) Unangst, P. C.; Connor, D. T.; Cetenko, W. A.; Sorenson, R. J.; Sircar, J. C.; Wright, C. D.; Schrier, D. J.; Dyer, R. D. Oxazole, Thiazole, and Imidazole Derivatives of 2,6-Di-tert-butylphenol as Dual 5-Lipoxygenase and Cyclooxygenase Inhibitors. *Bio. Med. Chem. Lett.* 1993, 3, 1729-1734.
- (2) Brooks, P. M.; Day, R. O. Nonsteroidal Antiinflammatory Drugs-Differences and Similarities. N. Engl. J. Med. 1991, 324, 1716– 1725.
- (3) (a) Kimmey, M. B. NSAIDS, Ulcers and Prostaglandins. J. Rheumatol. 1992, 19 (Suppl. 36), 68-73. (b) Peskar, B. M. On the Synthesis of Prostaglandins by Human Gastric Mucosa and its Modification by Drugs. Biochem. Biophys. Acta. 1977, 487, 307-314.
- (4) (a) Vane, J. R. Inhibition of Prostaglandin Synthesis as a Mechanism of Action of Aspirin-like Drugs. Nature (New Biol.) 1971, 231, 232-235. (b) Xie, W.; Robertson, D. L.; Simmons, D. L. Mitogen-Inducible Prostaglandin G/H Synthase: A New Target for Nonsteroidal Antiinflammatory Drugs. Drug Develop. Res. 1992, 25, 249-265. (c) DeWitt, D. L.; Meade, E. A.; Smith, W. L. PGH Synthase Isoenzyme Selectivity: The Potential for Safer Nonsteroidal Antiinflammatory Drugs. Am. J. Med. 1993, 95 (Suppl. 2A), 2A-40S-2A-44S.

- (5) Samuelsson, B. Arachidonic Acid Metabolism: Role in Inflammation. Z. Rheumatol. 1991, 50 (Suppl. 1), 3-6.
 (6) (a) Brain, S. D.; Williams, T. J. Leukotrienes and Inflammation.
- (6) (a) Brain, S. D.; Williams, T. J. Leukotrienes and Inflammation. Pharm. Ther. 1990, 46, 57-66. (b) Vaananen, P. M.; Keenan, C. M.; Grisham, M. B.; Wallace, J. L. Pharmacological Investigation of the Role of Leukotrienes in the Pathogenesis of Experimental NSAID Gastropathy. Inflammation 1992, 16, 227-240. (c) Guslandi, M. Gastric Effects of Leukotrienes. Prostaglandins, Leukotrienes Med. 1987, 26, 203-208.
- (a) Unangst, P. C.; Shrum, G. P.; Connor, D. T.; Dyer, R. D.; Schrier, D. J. Novel 1,2,4-Oxadioazoles and 1,2,4-Thiadiazoles as Dual 5-Lipoxygenase and Cyclooxygenase Inhibitors. J. Med. 1992, 36, 3691-3698.
 (b) Mullican, M. D.; Wilson, M. W.; Connor, D. T.; Kostlan, C. R.; Schrier, D. J.; Dyer, R. D. Design of 5-(3,5-Di-tertbutyl-4-hydroxyphenyl)-1,3,4-thiadiazoles, 1,3,4-oxadiazoles, and -1,2,4-triazoles as Orally-Active, Nonulcerogenic Antiinflammatory Agents. J. Med. Chem. 1993, 36, 1090-1099.
 (c) Flynn, D. L.; Belliotti, T. R.; Boctor, A. M.; Connor, D. T.; Kostlan, C. R.; Nies, D. E.; Ortwine, D. F.; Schrier, D. J.; Sircar, J. C. Styrylpyrazoles, Styrylisoxazoles, and Styrylisothiazoles. Novel 5-Lipoxygenase and Cvclooxygenase Inhibitors. J. Med. Chem. 1991. 34, 518-525.
- Styl II.Stazoles, and Styl ynstillazoles. Toto Ell polygenase and Cyclooxygenase Inhibitors. J. Med. Chem. 1991, 34, 518-525.
 (a) Kataumi, I.; Kondo, H.; Fuse, Y.; Yamashita, K.; Hidaka, T.; Hosoe, K.; Takeo, K.; Yamashita, T.; Watanabe, K. Studies on Styrene Derivatives. II. Synthesis and Antiinflammatory Activity of 3,5-Di-tert-butyl-4-hydroxystyrenes. Chem. Pharm. Bull. 1986, 34, 1619-1627. (b) Panetta, J. A. 5-[3,5-Bis(1,1-dimethylethyl)-4-hydroxyphenyl]methyl-4-thiazolidinones for Treatment of Inflammation and Ischemia. Eur. Pat. Appl. EP 211670, 1987.
 (9) Hidaka, T.; Hosoe, K.; Ariki, Y.; Takeo, K.; Yamashita, T.; Katsumi, I.; Kondo, H.; Yamashita, K.; Watanabe, K. Pharmacological
- (9) Hidaka, T.; Hosoe, K.; Ariki, Y.; Takeo, K.; Yamashita, T.; Katsumi, I.; Kondo, H.; Yamashita, K.; Watanabe, K. Pharmacological Properties of a New Anti-inflammatory Compound, α-(3,5-Di-tertbutyl-4-hydroxybenzylidene)-γ-butyrolactone (KME-4), and its Inhibitory Effects on Prostaglandin Synthetase and 5-Lipoxygenase. Jpn. J. Pharmacol. 1984, 36, 77-85.
- (10) Serradell, M. N.; Castăner, J.; Castăner, R. M. E-5110. Drugs Future 1989, 14, 307–310.
- (11) Wong, S.; Lee, S. J.; Frierson, M. R. III; Proch, J.; Miskowski, T. A.; Rigby, B. S.; Schmolka, S. J.; Naismith, R. W.; Kreutzer, D. C.; Lindquist, R. Antiarthritic Profile of BF-389 A Novel Anti-inflammatory Agent With Low Ulcerogenic Liability. Agents Actions 1992, 37, 90-98.
- (12) Ushenko, N. K.; Gorizdra, T. E. Substitution in the Azolidine Ring. V. Preparation of 2-Thiono-4-oxazolidinone and its Transformations. Ukrain. Khim. Zhur. 1950, 16, 545-551.
- (13) Rowley, G. L.; Greenleaf, A. L.; Kenyon, G. L. On the Specificity of Creatine Kinase. New Glycocyamine and Glycocyamine Analogs Related to Creatine. J. Am. Chem. Soc. 1971, 93, 5542-5551.
- (14) Berg, U.; Sandström, J. The Electronic Spectra of Thioamides and Thiohydrazides. Acta Chem. Scand. 1966, 20, 689–697.
- (15) Teuber, H. J.; Krause, H.; Berariu, V. Syntheses von 3,5-Di-tertbutyltyrosin und (3,5-Di-tert-butyl-4-hydroxyphenyl)glycine. (Synthesis of 3,5-di-tert-butyltyrosine and (3,5-di-tert-butyl-4-hydroxyphenyl)glycine.) Justus Liebigs Ann. Chem. 1978, 757-770.
- (16) Nishinaga, A.; Shimizu, T.; Matsuura, T. Oxygenation of 2,6-Dit-butylphenols Bearing Electron-Withdrawing Group at 4-Position Mediated by Co(II)-Schiff Base Complex. *Tetrahedron Lett.* 1981, 22, 5293-5296.
- (17) Cahoy, R. P. 3,5-Disubstituted 4-Hydroxybenzaldehydes. Ger. Offen. DE 2617585, 1976.
- (18) Brühl, J. W. Spectrochemie des Stickstoffs. Chem. Ber. 1895, 28, 2399-2414.
- (19) Newman, M. S.; Lee, L. F. The Synthesis of Arylacetylenes. 3,5-Di-tert-butylphenylacetylene. J. Org. Chem. 1972, 37, 4468-4469.
- (20) An X-ray structural determination for 6b and 6f (data not shown) confirmed the Z-configuration about the benzylidene double bond, as depicted in Scheme 1.
- (21) Durant, G. J.; Emmett, J. C.; Ganellin, C. R.; Miles, P. D.; Parsons, M. E.; Prain, H. D.; White, G. R. Cyanoguanidine-Thiourea Equivalence in the Development of the Histamine H₂-Receptor Antagonist, Cimetidine. J. Med. Chem. 1977, 20, 901-906.
 (22) Lazer, E. S.; Wong, H.-C.; Possanza, G. J.; Graham, A. G.; Farina,
- (22) Lazer, E. S.; Wong, H.-C.; Possanza, G. J.; Graham, A. G.; Farina, P. R. Antiinflammatory 2,6-Di-tert-butyl-4-(2-arylethenyl)phenols. J. Med. Chem. 1989, 32, 100-104.
 (23) Katsumi, I.; Kondo, H.; Yamashita, K.; Hidaka, T.; Hosoe, K.;
- (23) Katsumi, I.; Kondo, H.; Yamashita, K.; Hidaka, T.; Hosoe, K.; Yamashita, T.; Watanabe, K. Studies on Styrene Derivatives. I. Synthesis and Antiinflammatory Activities of α-Benzylidene-γbutyrolactone Derivatives. Chem. Pharm. Bull. 1986, 34, 121-129.
- (24) Newbould, B. B. Chemotherapy of Arthritis Induced in Rats by Mycobacterial Adjuvant. Br. J. Pharmacol. 1963, 21, 127–136.
 (25) Still, W. C.; Kahn, M.; Mitra, A. Rapid Chromatographic Technique
- (25) Still, W. C.; Kahn, M.; Mitra, A. Rapid Chromatographic Technique for Preparative Separations with Moderate Resolution. J. Org. Chem. 1978, 43, 2923-2925.