

 Phytochemistry Vol. 49, No. 8, pp. 2375–2378, 1998
© 1998 Elsevier Science Ltd. All rights reserved Printed in Great Britain 0031-9422/98/5 - see front matter

PII: S0031-9422(98)00433-6

THUNALBENE, A STILBENE DERIVATIVE FROM THE ORCHID THUNIA ALBA

P. L. MAJUMDER,* M. ROYCHOWDHURY and S. CHAKRABORTY

Department of Chemistry, University College of Science, 92 Acharya Prafulla Chandra Road, Calcutta 700009, India

(Received in revised form 20 April 1998)

Key Word Index-Thunia alba; Orchidaceae; thunalbene; stilbene derivative.

Abstract—Thunalbene, a new stilbene derivative, was isolated from the orchid *Thunia alba* which also afforded six known stilbenoids: batatasin-III, lusianthridin, 3,7-dihydroxy-2,4-dimethoxyphenanthrene, 3,7-dihydroxy-2,4,8-trimethoxyphenanthrene, cirrhopetalanthrin and flavanthrin. The structure of thunalbene, the first stilbene derivative isolated so far from an Orchidaceae plant, was established as 3,3'-dihydroxy-5-methoxystilbene from spectral and chemical evidence. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

We reported earlier on the isolation of a fairly large number of compounds of diverse structural types from a series of Indian Orchidaceae plants. These compounds encompass a wide variety of stilbenoids, i.e. bibenzyls [1], phenanthrenes [2] and their dimers [3-5], 9,10-dihydrophenanthrenes [6] and their dimers [7], phenanthropyrans and pyrones [8,9] and their 9,10-dihydro derivatives [10], besides a few other polyphenolics [11, 12], simple aromatic compounds [13], several triterpenoids [14] and steroids of biogenetic importance [15]. Our continued search for new phytochemicals from the same source has now resulted in the isolation of a new stilbene derivative, designated thunalbene, from the orchid Thunia alba which also afforded six known stilbenoids, i.e. batatasin-III (3,3'-dihydroxy-5-methoxybibenzyl) [16, 17], lusianthridin (4,7-dihydroxy-2-methoxy-9,10-dihydrophenanthrene) [18], 3,7-dihydroxy-2,4-dimethoxyphenan-threne [19], 3,7dihydroxy-2,4,8-trimethoxyphenanthrene [2], cirrhopetalanthrin (2,2',7,7'-tetrahydroxy-4,4'-dimethoxy-1,1'-biphenanthryl) [3] and flavanthrin (2,2',7,7'tetrahydroxy-4,4'-dimethoxy-9,9',10,10'-tetrahydro-1,1'-biphenanthryl) [7]. While the known compounds were identified by direct comparison with their respective authentic samples, thunalbene was shown to have the structure 1a from the following spectral and chemical evidence.

RESULTS AND DISCUSSION

Thunalbene (1a), $C_{15}H_{14}O_3$ (M⁺ at m/z 242), showed the UV absorptions [λ_{max}^{ErOH} 214 and 303 nm (log ε 4.30 and 4.72)] expected of a *trans*-stilbene derivative. The phenolic nature of the compound was indicated by its characteristic colour reactions with FeCl₃ (violet) and phosphomolybdic acid reagent (intense blue), alkali-induced bathochromic shifts of its UV maxima and by its IR band at 3360 cm⁻¹. The presence of two phenolic hydroxyl groups in **1a** was confirmed by the formation of a diacetyl derivative (**1b**), $C_{19}H_{18}O_5$ (M⁺ at m/z 326), with Ac₂O and pyridine. The IR absorption band at 980 cm⁻¹ of **1a** indicated the presence of a *trans*double bond in the compound.

The ¹H NMR spectrum of **1a** showed signals for an aromatic methoxyl function [δ 3.73 (3H,s)], two phenolic hydroxyl groups [δ 5.28 (2H,s);



^{*}Author to whom correspondence should be addressed.

disappeared on deuterium exchange], seven aromatic protons at δ 6.27–7.14, and a two-proton singlet at δ 6.96 which is typical of the vinylic protons of a trans-stilbene derivative. The relative positions of the methoxyl and hydroxyl groups in the two phenyl rings of 1a were ascertained from the chemical shifts and the splitting patterns of the aromatic protons of the compound and its diacetyl derivative 1b. Thus, the chemical shifts and the splitting patterns of the aromatic protons of 1a resonating at δ 7.14 (1H, appt t; $J_1 = 8.1$ Hz and $J_2 = 7.8$ Hz), 6.97 (1H, br d; J = 7.5 Hz), 6.96 (1H, br signal; obscured in the signal of the olefinic protons), 6.67 (1H, dd; $J_1 = 7.8$ Hz and $J_2 = 1.8$ Hz) and 6.55, 6.51 and 6.27 (each 1H, br signal) are strikingly similar to those of H-5', H-6', H-2', H-4', H-6, H-2 and H-4, respectively, of batatasin-III (1c) [16], except that the signals corresponding to H-2, H-2', H-6 and H-6' of 1a showed characteristic downfield shifts compared to the corresponding protons of 1c due to the diamagnetic anisotropic effect of the olefinic double bond between C_{α} - $C_{\alpha'}$ in **1a**. This was also corroborated by the similarities of the chemical shifts and the splitting patterns of the aromatic proton signals of thunalbene diacetate (1b) and batatasin-III diacetate (1d) exhibiting the same differences in regard to their H-2, H-2', H-6 and H-6' resonances. The above ¹H NMR spectral data of **1a** and 1b, thus, not only indicated identical substitution patterns of the hydroxyl and methoxyl functions in both 1a and 1c, but also implied that the former was the corresponding stilbene derivative of the latter.

The structure of thunalbene (1a) was further supported by the ¹³C NMR spectral data of the compound and its diacetyl derivative 1b (Table 1). The degree of protonation of the carbon atoms of each compound was confirmed by APT experiments and the assignments of the carbon chemical shifts of 1a and 1b were made by comparison with the $\delta_{\rm C}$ values of structurally similar compounds like batatasin-III (1c) [20] and its diacetate 1d [17] taking into consideration the alteration in additive parameters caused by the change of the state of hybridization of C- α and C- α' from sp³ in 1c and 1d to sp²

in 1a and 1b. Thus, the $\delta_{\rm C}$ values of C-3, C-3', C-5 and C-5' of 1a and 1c were virtually identical, while those of C-1 and C-1' of 1a showed upfield shifts of ca. 4-5 ppm compared to the corresponding carbon atoms of 1c due to the change in the state of hybridizations of C- α and C- α' from sp³ in 1c to sp² in 1a. The observed upfield shifts of C-2, C-2', C-6 and C-6' by ca. 2-2.5 ppm and the downfield shifts of C-4 and C-4' by ca. 1.5-1.8 ppm of 1a compared to the corresponding carbon atoms of 1c may also be attributed to the different states of hybridizations of C- α and C- α' of the two compounds, which, as expected appeared at ca. $\delta_{\rm C}$ 129 in **1a** as against ca. $\delta_{\rm C}$ 38 in 1c. The $\delta_{\rm C}$ values of 1b are also compatible with the placement of the two hydroxyl groups at C-3 and C-3' and the methoxyl group at C-5 in 1a and exhibited expected downfield shifts of C-2, C-4, C-6, C-2', C-4' and C-6'. The same trend in the changes of C-1, C-1', C-2, C-2', C-4, C-4', C-6 and C-6' resonances of 1a compared to the corresponding carbon atoms of 1c are also discernible in the $\delta_{\rm C}$ values of the above carbon atoms of **1b**, when compared with the corresponding carbon resonances of 1d.

The structure of 1a was finally confirmed by the conversion of its diacetyl derivative 1b to batatasin-III diacetate (1d) by hydrogenation of 1b over PtO₂.

It is interesting to note that although several stilbene derivatives were reported from a number of plant species [21], i.e. Gnetum ula [22], Alnus elongata virides [23], Virola [24], Cassia roxburghii [25], Diphysia robinioides [26], Phoenix dactylifera [27] and Combretum caffrum [28], all belonging to botanical families other than Orchidaceae, the isolation of thunalbene (1a) from the orchid Thunia alba constitutes the first report of the occurrence of a stilbene derivative in an orchid. This is despite the fact that the large number of orchids so far chemically investigated were shown to elaborate preponderantly a wide range of stilbenoids including a fairly large number of bibenzyl derivatives. In the light of the above observations, the isolation of thunalbene is of considerable biogenetic and chemotaxonomical importance.

С	$1a^{\dagger}$	1b [‡]	1c [‡]	1d [‡]	С	$1a^{\dagger}$	1b [‡]	1c [‡]	1d [‡]
1	139.5 ^a	138.5 ^e	144.3 ⁱ	143.7 ¹	4′	115.4 ^c	121.0 ^g	113.6	119.1 ⁿ
2	106.8	111.9	108.8	113.7	5'	130.2	128.9 ^h	130.0	129.1
3	159.4 ^b	151.8 ^f	159.2 ^j	151.5 ^m	6'	118.1	124.2	120.4	125.8
4	101.6	107.0	99.9	105.2	α	129.3 ^d	129.5 ^h	38.4 ^k	37.3°
5	161.7	160.5	161.9	160.2	α′	129.4 ^d	128.8 ^h	38.1 ^k	37.0°
6	104.1	109.9	106.3	111.8	OMe	55.3	55.4	55.3	55.2
1′	140.1 ^a	139.1 ^e	145.0 ⁱ	143.0 ¹	OAc	-	169.3	_	169.3
2′	113.7 ^c	119.3 ^g	116.2	121.4 ⁿ		-	169.2	-	
3′	158.4 ^b	151.0 ^f	158.2 ^j	150.7 ^m		-	21.0	-	21.0

Table 1. ¹³C NMR spectral data of compounds 1a, 1b, 1c and 1d

[†]Spectra were run in d₆-acetone and chemical shifts were measured with δ (TMS) = δ (d₆-acetone) + 29.6 ppm.

[‡]Spectra were run in CDCl₃ and chemical shifts were measured with δ (TMS) = δ (CDCl₃) + 76.9 ppm.

^{a-o}Values are interchangeable within each column.

EXPERIMENTAL

M.p.'s: Uncorr.; CC: silica gel (100–200 mesh); MPLC: silica gel (230–400 mesh); TLC: silica gel G; UV: 95% aldehyde-free EtOH; IR: KBr discs; ¹H and ¹³C NMR: 300 and 75 MHz, respectively, in CDCl₃ and d₆-acetone using TMS as an int. standard. Chemical shifts are expressed in δ (ppm). MS: direct inlet system, 70 eV. All analyt. samples were routinely dried over P₂O₅ for 24 h *in vacuo* and were tested for purity by TLC and MS. Na₂SO₄ was used for drying organic solvents and the petrol used had b.p. 60–80°. Plant materials were collected from Darjeeling, India in September, 1993. A voucher specimen is deposited in the Herbarium of the Department of Botany, University of Calcutta (CUH).

Isolation of thunalbene (1a), batatasin-III (1c), lusianthridin, cirrhopetalanthrin, flavanthrin, 3,7-dihydroxy-2,4-dimethoxyphenanthrene and 3,7-dihyhydroxy-2,4,8-trimethoxyphenanthrene from Thunia alba

Air-dried powdered whole plants (2 kg) of T. alba were soaked in MeOH (71) for 3 weeks. The MeOH extract was then drained off, concd under red. pres. to ca. 100 ml, diluted with H_2O (500 ml) and the liberated solids exhaustively extracted with Et₂O. The Et₂O extract was fractionated into acidic and non-acidic frs with 2 M NaOH. The aq. alkaline soln was acidified in the cold with conc. HCl and the liberated solids extracted with Et₂O, washed with H₂O, dried and the solvent removed. The residue was chromatographed. The early frs of the petrol-EtOAc (10:1) eluate afforded a mixture of lusianthridin and 1c which on rechromatography using petrol-EtOAc (20:1) as the eluent gave in the early frs pure lusianthridin (0.05 g), crystallized from petrol–EtOAc, m.p. $162^\circ\!,$ and 1c~(0.03~g) as a semisolid mass in the later frs. Elution of the main column with petrol-EtOAc (5:1) afforded a mixture of 3,7-dihydroxy-2,4-dimethoxyphenanthrene, 3,7dihydroxy-2,4,8-trimethoxyphenanthrene and 1a, which was subjected to MPLC using petrol-EtOAc (1:1) as the solvent. The early frs afforded pure 1a (0.04 g) as a semisolid mass (found: C, 74.32; H, 5.73. $C_{15}H_{14}O_3$ requires: C, 74.38; H, 5.78%). UV $\lambda_{max}^{EtOH-0.1 M NaOH}$ nm: 209.0 and 301.5 (log $\epsilon,$ 4.71 and 4.34); IR $\nu_{max}~cm^{-1}\!\!:$ 3360 (OH), 980 (trans-double bond), 1595, 1500, 960, 860, 830 and 780 (phenyl nucleus); MS m/z (rel. int.): 242 [M⁺], (82), 225 (3), 211 (5), 210 (5), 198 (2), 182 (2), 181 (36), 169 (3), 165 (6), 153 (9), 152 (15), 149 (21), 115 (15), 97 (16) and 83 (20).

Compound **1a** was acetylated with Ac₂O and pyridine in the usual manner to give **1b**, crystallized from petrol–EtOAc, m.p. 110° (found: C, 69.90; H, 5.49. C₁₉H₁₈O₅ requires: C, 69.94; H, 5.52%). UV λ_{max} nm: 211 and 296.5 (log ε 4.79 and 4.78); IR v_{max} cm⁻¹: 1230 and 1780 (OAc), 1635, 1390, 930, 915, 890 and 810 (phenyl nucleus) and 990 (*trans*-double bond); ¹H NMR: δ 7.24–7.30 (2H, *m*; H-5' and H-6'), 7.14 (1H, *br* signal; H-2'), 6.90–6.99 (3H, *m*; H-4', H- α and H- α '), 6.80, 6.77 and 6.48 (each 1H, *br* signal; H-2, H-6 and H-4), 3.78 (3H, *s*; ArOMe), 2.22 and 2.23 (each 3H, *s*; 2 × OAc); MS *m*/*z* (rel. int.): 326 [M⁺], (23), 284 (24), 242 (82), 226 (2), 225 (2), 198 (2), 181 (9), 152 (6) and 115 (6).

The later frs in the above MPLC afforded a mixture of 3,7-dihydroxy-2,4-dimethoxyphenanthrene and 3,7-dihydroxy-2,4,8-trimethoxyphenanthrene. Further MPLC of this mixture using petrol–EtOAc (1:1) as the solvent finally gave pure 3,7-dihydroxy-2,4-dimethoxyphenanthrene (0.025 g) as a semisolid mass in the early frs and pure 3,7-dihydroxy-2,4,8trimethoxyphenanthrene (0.015 g) in the later frs. Further elution of the main column with petrol– EtOAc (1:1) eluate gave pure cirrhopetalanthrin (0.015 g), crystallized from petrol–EtOAc mixture, m.p. 296°, in the early frs and pure flavanthrin (0.03 g), crystallized from petrol–EtOAc mixture, m.p. 285°, in the later frs.

Catalytic hydrogenation of 1b

A soln of **1b** (0.02 g) in EtOH (20 ml) containing PtO_2 (0.005 g) was stirred under H_2 atmosphere for 4 h. The catalyst was filtered off and the filtrate on evaporation gave a semisolid residue (0.018 g) which was identical in all respects to batatasin-III diacetate (**1d**).

Acknowledgements—The work was supported by the CSIR and UGC, New Delhi.

REFERENCES

- 1. Majumder, P. L., Roychowdhury, M. and Chakraborty, S., *Phytochemistry*, 1997, **44**, 167.
- Majumder, P. L. and Sen, R. C., *Phytochemistry*, 1991, **30**, 2432.
- Majumder, P. L., Pal, A. and Joardar, M., *Phytochemistry*, 1990, **29**, 271.
- Majumder, P. L. and Lahiri, S., *Tetrahedron*, 1990, 46, 3621.
- 5. Majumder, P. L. and Pal, S., *Phytochemistry*, 1992, **31**, .
- Majumder, P. L., Banerjee, S., Lahiri, S., Mukhoti, N. and Sen, S., *Phytochemistry*, 1998, 47, 855.
- Majumder, P. L. and Banerjee, S., *Tetrahedron*, 1988, 44, 7303.
- Majumder, P. L. and Sabzabadi, E., *Phytochemistry*, 1899, **1988**, 27.
- 9. Majumder, P. L. and Maiti, D. C., *Phytochemistry*, 1991, **30**, 971.
- 10. Majumder, P. L. and Maiti, D. C., *Phytochemistry*, 1988, **27**, 899.

P. L. MAJUMDER et al.

- 11. Majumder, P. L., Lahiri, S. and Mukhoti, N., *Phytochemistry*, 1995, **40**, 271.
- Majumder, P. L., Lahiri, S. and Pal, S., Journal of the Indian Chemical Society, 1994, 71, 645.
- 13. Majumder, P. L. and Lahiri, S., Indian Journal of Chemistry, 1989, 28B, 771.
- 14. Majumder, P. L. and Ghosal, S., Journal of the Indian Chemical Society, 1991, 68, 88.
- 15. Majumder, P. L. and Pal, S., *Phytochemistry*, 1990, **29**, 2717.
- 16. Sachdev, K. and Kulshreshtha, D., *Phytochemistry*, 1986, **25**, 499.
- 17. Majumder, P. L. and Basak, M., *Phytochemistry*, 1991, **30**, 321.
- Majumder, P. L. and Lahiri, S., *Phytochemistry*, 1990, **29**, 621.
- Tuchinda, P., Udchachem, J., Khumtaveeporm, K., Taylor, W. C. and White, A. H., *Phytochemistry*, 1988, 27, 3267.

- Juneja, R. K., Sharma, S. C. and Tandon, J. S., *Phytochemistry*, 1987, 26, 1123.
- 21. Harborne, J. B., *Methods in Plant Biochemistry*, Vol. 1. Academic Press, London, 1989.
- 22. Prakash, S., Khan, M. A., Khan, K. Z. and Zaman, A., *Phytochemistry*, 1985, **24**, 622.
- 23. Bouvin, J. F., Jay, M. and Weber, E. W., *Phytochemistry*, 1978, **17**, 821.
- 24. Macrae, W. D. and Tower, G. H. N., *Phytochemistry*, 1985, **24**, 561.
- 25. Ashok, D. and Sharma, P. N., Journal of the Indian Chemical Society, 1987, 64, 559.
- Castro, O., Lopez, J., Vergara, A., Sternitz, P. R. and Gardener, D. R., *Journal of Natural Products*, 1986, 49, 680.
- 27. Fernendez, M. I., Pedro, J. R. and Seoane, E., *Phytochemistry*, 1983, **22**, 2819.
- Petit, G. R., Singh, S. B., Niven, M. L., Hammel, E. and Schmidt, J. M., *Journal of Natural Products*, 1987, 50, 119.

2378