## An Efficient Asymmetric Synthesis of Cascarillic Acid

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**Abstract:** An efficient six-step asymmetric synthesis of the cyclopropane containing natural product cascarillic acid in 41% overall yield is described. The key synthetic steps involve the use of a temporary stereogenic hydroxyl group to control the facial selectivity of a directed cyclopropanation reaction and its subsequent removal via a retro-aldol reaction.

Key words: cascarillic acid, cyclopropane, aldol, retro-aldol, temporary stereocentre

Cascarillic acid [(3S,4R)-2] is a major component of cascarilla essential oil that has been used for many years in the treatment of colds and bronchitis.<sup>1</sup> It contains a *trans*cyclopropane ring within its fatty acid chain, which contrasts with the majority of naturally occurring cyclopropane fatty acids that are *cis* in orientation.<sup>2</sup> Baird and co-workers have recently confirmed the absolute stereochemistry of this natural product to be 3S,4R through its total synthesis in 11 steps from meso-cis-1,2-dihydroxymethylcyclopropane (1, Scheme 1).<sup>3</sup> This synthesis included an enzymatic desymmetrisation step to introduce the stereogenic centres of the cyclopropane ring, and an epimerisation step to invert a *cis*-cyclopropane ring into its corresponding trans-isomer. Consequently, we now report on an alternative synthetic strategy that affords cascarillic acid 2 in six steps from the chiral auxiliary (R)-N-isovaleroyl-4-benzyl-5,5-dimethyl-oxazolidin-2-one (11).



Scheme 1 A previous asymmetric synthesis of cascarillic acid (2).

We have recently reported the development of novel synthetic strategies to reversibly generate temporary stereogenic centres that may be used to create remote stereocentres using substrate directable reactions.<sup>4–6</sup> In one of these reports, a novel three-step aldol–directed cyclopropanation–retro-aldol protocol was demonstrated for the asymmetric synthesis of chiral cyclopropane carboxaldehydes in high ee (Scheme 2).<sup>4</sup> In this approach, chiral auxiliary fragment **3** reacts with an  $\alpha$ , $\beta$ -unsaturated aldehyde **4** to give *syn*-aldol product **5** (step 1), whose

SYNLETT 2006, No. 7, pp 1119–1121 Advanced online publication: 24.04.2006 DOI: 10.1055/s-2006-939690; Art ID: D02106ST © Georg Thieme Verlag Stuttgart · New York 'temporary'  $\beta$ -hydroxyl functionality is used to control facial selectivity in a directed cyclopropanation reaction to afford cyclopropane **6** in very high de (step 2). Retroaldol cleavage of cyclopropane **6** results in destruction of the 'temporary'  $\beta$ -hydroxyl stereocentre, affording the chiral auxiliary fragment **3** and the desired enantiopure cyclopropane carboxaldehyde **7** in very high ee (step 3).



**Scheme 2** Three-step *syn*-aldol–cyclopropanation–retro-aldol protocol for the asymmetric synthesis of cyclopropane carboxaldehydes.

We anticipated that this cyclopropanation methodology was ideally suited to the preparation of cascarillic acid (3*S*,4*R*)-**2**, since employing (*E*)-non-2-enal **8** as an aldehyde substrate in this three-step protocol would result in formation of enantiopure cyclopropane carboxaldehyde (*R*,*R*)-**9**. This aldehyde **9** could then be converted to cascarillic acid (3*S*,4*R*)-**2** via an oxidative one-carbon homologation reaction using well established dithiane– Peterson elimination methodology<sup>7,8</sup> (Scheme 3).



Scheme 3 Proposed asymmetric synthesis of cascarillic acid (2).

Therefore, our first goal was to employ the syn-aldol-directed cyclopropanation-retro-aldol methodology described in Scheme 2 for the asymmetric synthesis of cyclopropane (R,R)-9 in high de (Scheme 4). Research within our group investigating steric factors that influence the retro-aldol reaction of α-alkyl-β-hydroxy-N-acyl-oxazolidin-2-ones had revealed that N-isovaleroyl-oxazolidin-2-one (11) was particularly well suited as a chiral auxiliary for this methodology. Therefore, (R)-4-benzyl-5,5-dimethyl-oxazolidin-2-one  $(10)^9$  was first prepared in 72% yield from unnatural D-phenylalanine using a previously reported procedure.<sup>10</sup> Treatment of 5,5-dimethyloxazolidin-2-one [(R)-10] in THF at -78 °C with 1.1 equivalents of n-BuLi, followed by addition of 1.1 equivalents of isovaleroyl chloride gave N-isovaleroyl-oxazolidin-2-one [(R)-11] in 87% yield. Treatment of N-acyloxazolidin-2-one [(R)-11] with 1.1 equivalents of 9-BBN-OTf, *i*-Pr<sub>2</sub>NEt in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C, followed by cooling to -78 °C and addition of (E)-non-2-enal (8) resulted in synaldol 12 in 93% de, which was purified to >95% de in 82% yield via silica gel chromatography.<sup>11</sup> The syn-stereochemistry of aldol 12 was confirmed from the  $J_{(2',3')}$  coupling constant of 6.5 Hz observed in its <sup>1</sup>H NMR spectra.<sup>12</sup> Reaction of syn-aldol 12 with 5 equivalents of Et<sub>2</sub>Zn and CH<sub>2</sub>I<sub>2</sub> in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C, resulted in a highly stereoselective cyclopropanation reaction to afford syn-cyclopropyl aldol **13** in >95% de and 94% yield.<sup>13</sup> Cyclopropanation under under modified Furukawa conditions are normally syn-selective due to minimisation of  $A^{1,3}$  strain in the transition state, and as a consequence the absolute configuration of cyclopropane 13 was assigned accordingly.<sup>14</sup> Treatment of cyclopropane 13 with 1.1 equivalents of KHMDS in THF at -40 °C afforded a potassium alkoxide that underwent a clean retro-aldol reaction to give the parent chiral auxiliary (R)-11 and the desired 2-hexylcyclopropanecarboxaldehyde (R,R)-9 in 85% yield.



Scheme 4 Reagents and conditions: (i) n-BuLi, isovaleroyl chloride, THF, -78 to 0 °C; (ii) 9-BBN-OTf, *i*-Pr<sub>2</sub>NEt, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C; (*E*)-non-2-enal (8), -78 °C to r.t.; (iii) Et<sub>2</sub>Zn, CH<sub>2</sub>I<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -10 °C to 0 °C, 2 h; (iv) KHMDS, THF, -40 °C, 3 h.

With (R,R)-2-hexylcyclopropanecarbaldehyde (9) in hand, treatment with the lithium anion of (1,3-dithian-2yl)trimethylsilane in THF at -30 °C resulted in nucleophilic addition of the dithiane anion to the aldehyde functionality, followed by subsequent Peterson elimination to afford  $2-\{[(R,R)-2-hexylcyclopropyl]methylene\}-1,3-di$ thiane (14) in >95% de and 93% yield. Therefore, this reaction was successful in establishing the correct oxidation state of the homologated carbon atom, with no epimerisation of the cyclopropane ring being observed. Treatment of ketene thioacetal 14 under sequential acid and base-catalysed hydrolysis conditions gave cascarillic acid 2 in >95% de and 78% yield, whose spectroscopic data was identical to that previously published for this compound (Scheme 5). The negative sign obtained for the specific rotation of this synthetic sample of cascarillic acid **2** of  $[\alpha]_D^{25}$  –11.0 (*c* 0.41, CHCl<sub>3</sub>); {lit. 1a:  $[\alpha]_{D}^{25}$  -10.5 (c 0.553, CHCl<sub>3</sub>)}, confirmed that we had synthesised the correct enantiomer of this natural product.3,15



Scheme 5 *Reagents and conditions*: (i) *n*-BuLi, (1,3-dithian-2-yl)trimethylsilane, THF, 0 °C, 1 h, then (R,R)-9, -30 °C, 2 h; (ii) p-TSA, THF–H<sub>2</sub>O, 6 h, reflux; (iii) KOH, acetone–H<sub>2</sub>O, 2 h, reflux.

In conclusion, we have described an efficient six-step asymmetric synthesis of the cyclopropane containing natural product cascarillic acid in 41% overall yield. The key synthetic steps employed involve the use of a temporary stereogenic hydroxyl group to control the facial selectivity of a directed cyclopropanation reaction, and its subsequent removal via a retro-aldol reaction. The application of this methodology to the asymmetric synthesis of other cyclopropane containing natural products will be reported in due course.

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(*R*)-4-Benzyl-3-[(*E*)-(2*R*,3*S*)-3-hydroxy-2-isopropylundec-4-enoyl]-5,5-dimethyl-1,3-oxazolidin-2-one (12):  $[a]_D^{25}$ +22.0 (*c* 0.85, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.27–7.11 (5 H, m, Ph), 5.54–5.71 (2 H, m, CH=CHCH<sub>2</sub> and CH=CHCH<sub>2</sub>), 4.53 (1 H, dd, *J* = 10.0, 4.0 Hz, CHN), 4.36 (1 H, app. t, *J* = 6.5 Hz, CHOH), 4.09 (1 H, dd, *J* = 9.0, 6.5 Hz, COC*H*), 3.09 (1 H, dd, J = 14.5, 4.0 Hz,  $CH_AH_BPh$ ), 2.81 (1 H, dd, J = 14.5, 10.0 Hz,  $CH_AH_BPh$ ), 2.04–1.88 [4 H, obs. m, OH, CH=CHCH<sub>2</sub> and CH(CH<sub>3</sub>)<sub>2</sub>], 1.35–1.12 [8 H, m, (CH<sub>2</sub>)<sub>4</sub>], 1.24 [6 H, app. s, (CH<sub>3</sub>)<sub>2</sub>C], 0.90 [3 H, d, J = 7.0 Hz, CH(CH<sub>3</sub>)CH<sub>3</sub>], 0.82 [3 H, obs. d, J = 7.0 Hz, CH(CH<sub>3</sub>)CH<sub>3</sub>], 0.80 (3 H, obs. t, J = 7.0 Hz, CH(CH<sub>3</sub>)CH<sub>3</sub>], 0.80 (3 H, obs. t, J = 7.0 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 174.7$ , 153.9, 137.4, 135.8, 129.5, 129.1, 128.9, 127.2, 82.4, 73.8, 64.3, 54.1, 35.9, 32.7, 32.1, 29.5, 29.3, 28.7, 28.6, 23.0, 22.6, 21.0, 20.4, 14.5. IR (film): 3501 (br OH), 1778 (C=O<sub>ox</sub>), 1693 (C=O) cm<sup>-1</sup>. HRMS (ES): m/z calcd [M + NH<sub>4</sub>]\*: 447.3217; found: 447.3213.

(R)-4-Benzyl-3-{(R)-2-[(S)-[(1R,2R)-2-hexylcyclopropyl](hydroxy)methyl]-3-methylbutanoyl}-5,5**dimethyl-1,3-oxazolidin-2-one** (13):  $[\alpha]_D^{25}$  –21.0 (*c* 0.62, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 7.34-7.18$  (5 H, m, Ph), 4.56 (1 H, dd, J = 10.0, 3.5 Hz, CHN), 4.22 (1 H, dd, J = 8.5, 6.0 Hz, COCH), 3.39 (1 H, dd, J = 8.5, 6.0 Hz, CHOH), 3.23 (1 H, dd, *J* = 14.5, 3.5 Hz, CH<sub>A</sub>H<sub>B</sub>Ph), 2.86  $(1 \text{ H}, \text{ dd}, J = 14.5, 10.0 \text{ Hz}, \text{CH}_{A}\text{CH}_{B}\text{Ph}), 2.31 [1 \text{ H}, \text{m},$ (CH<sub>3</sub>)<sub>2</sub>CH], 1.85 (1 H, br s, OH), 1.44–1.20 [10 H m, (CH<sub>2</sub>)<sub>5</sub>], 1.34 [3 H, s, (CH<sub>3</sub>)C(CH<sub>3</sub>)], 1.33 [3 H, s,  $(CH_3)C(CH_3)$ ], 1.02 [3 H, d, J = 7.0 Hz,  $CH(CH_3)CH_3$ ], 1.00 (1 H, obs. m, cyc-CH), 0.93 [3 H, d, J = 7.0 Hz,  $CH(CH_3)CH_3$ , 0.88 (3 H, t, J = 7.0 Hz,  $CH_2CH_3$ ), 0.76 (1 H, m, cyc-CH), 0.43 (1 H, app. dt, J = 8.5, 4.5 Hz, cyc-CH<sub>A</sub>H<sub>B</sub>), 0.28 (1 H, app. dt, J = 8.5, 5.0 Hz, cyc-CH<sub>A</sub>H<sub>B</sub>). <sup>13</sup>C NMR  $(75 \text{ MHz}, \text{CDCl}_3): \delta = 175.1, 153.7, 137.5, 129.4, 129.1,$ 127.2, 82.2, 75.5, 64.4, 54.5, 35.8, 34.2, 32.3, 29.6, 29.5, 28.8, 28.6, 23.1, 22.8, 22.2, 21.4, 21.1, 18.8, 14.5, 9.6. IR (film): 3516 (br OH), 1778 (C=O<sub>ox</sub>), 1693 (C=O) cm<sup>-1</sup>. HRMS (ES): m/z calcd  $[M + NH_4]^+$ : 461.3374; found: 461.3370.

(*R*,*R*)-2-Hexylcyclopropanecarbaldehyde (9):  $[\alpha]_D^{25}$ -26.0 (*c* 0.35, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.98 (1 H, d, *J* = 5.5 Hz, CHO), 1.61 (1 H, m, C<sub>1</sub>H), 1.51– 1.20 [11 H, m, C<sub>2</sub>H and (CH<sub>2</sub>)<sub>5</sub>], 0.96–0.83 (5 H, m, cyc-CH<sub>2</sub> and CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 201.2, 32.6, 31.7, 30.6, 29.0, 28.9, 22.7, 22.6, 14.9, 14.1. IR (film): 1713 (C=O) cm<sup>-1</sup>. HRMS (ES): *m*/*z* calcd [M + NH<sub>4</sub>]<sup>+</sup>: 172.1696; found: 172.1696.

**2-{[(***R,R***)-2-Hexylcyclopropyl]methylene}-1,3-dithiane (14):**  $[\alpha]_D^{25}$  -20.0 (*c* 0.30, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 5.42 (1 H, d, *J* = 10.0 Hz, C=*CH*), 2.91 (4 H, m, 2 × SCH<sub>2</sub>), 2.22–2.13 (2 H, m, SCH<sub>2</sub>CH<sub>2</sub>), 1.58 (1 H, m, C=CHC*H*), 1.41–1.20 [10 H, m, (CH<sub>2</sub>)<sub>5</sub>], 0.88 (3 H, t, *J* = 7.0 Hz, CH<sub>2</sub>CH<sub>3</sub>), 0.79 (1 H, m, cyc-*CH*), 0.65–0.56 (2 H, m, cyc-*CH*<sub>2</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 140.4, 121.6, 34.1, 32.3, 31.3, 30.5, 29.7, 29.5, 26.0, 23.1, 22.2, 20.3, 15.2, 14.5. IR (film):1678 (C=C) cm<sup>-1</sup>. HRMS (ES): *m/z* calcd [M + H]<sup>+</sup>: 257.1392; found: 257.1393.

**2-[(15,2***R***)-2-Hexylcyclopropyl]acetic acid [(3***S***,4***R***)-cascarillic acid] (2): [\alpha]\_D^{25}-11.0 (***c* **0.41, CHCl<sub>3</sub>); lit. 1a: [\alpha]\_D^{25}-10.5 (***c* **0.553, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): \delta = 2.26 (2 H, app. d,** *J* **= 7.0 Hz, CH<sub>2</sub>CO<sub>2</sub>H), 1.41–1.18 [10 H, m, (CH<sub>2</sub>)<sub>5</sub>], 0.88 (3 H, t,** *J* **= 7.0 Hz, CH<sub>2</sub>CH<sub>3</sub>), 0.77 (1 H, m, C<sub>1</sub>H), 0.56 (1 H, m, C<sub>2</sub>H), 0.33 (2 H, m, cyc-CH<sub>2</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): \delta = 176.6, 37.5, 32.8, 30.9, 28.3, 28.1, 21.6, 17.7, 13.1, 13.0, 10.6. IR (film): 1711 (C=O) cm<sup>-1</sup>. HRMS (EI):** *m/z* **calcd [M]<sup>+</sup>: 184.1458; found: 184.1458.** 

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