## New Synthesis of (+)- and (-)-Nojirimycin from myo-Inositol

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A report of a new synthesis of nojirimycin (1a), as well as its antipode (1b), from optically active seven-membered hemiacetal lactones (2a,b) derived from myo-inositol by a five step reaction; the hydrogen sulphite adduct of (1b) shows high inhibitory activity against  $\beta$ -glucosidase and  $\alpha$ -mannosidase, being almost comparable to that of mannojirimycin.

Nojirimycin¹ and mannojirimycin² (nojirimycin B), produced by *Streptomyces lavendulae* SF-425, show a potent inhibitory activity against glucosidases and glucoamylase.³ In particular, much attention has been focused on the corresponding 1-deoxy analogues, <sup>4.5</sup> the inhibitors of trimming glycosidase, <sup>6</sup> which have been shown to interfere with HIV-induced syncytium formation and viral infectivity. <sup>7</sup> Recently, therefore, interest in the structure and enzyme-inhibitory activity relationship and chemical modification<sup>8</sup> of these compounds has increased.

Synthesis of nojirimycin has been achieved successfully by several groups starting from p-glucose<sup>1,9</sup> and L-tartaric acid.<sup>10</sup> However, as yet its enantiomer has not been synthesized. We describe herein a new synthesis of (+)-(1a) and (-)-nojirimycin (1b) from *myo*-inositol *via* the optically active, sevenmembered hemiacetal lactones (2a,b),<sup>11</sup> together with the inhibitory activity of (1b).

Treatment of compound (2a) with trimethyl orthoformate in methanol in the presence of toluene-p-sulphonic acid (reflux, 40 min), and successive reduction with lithium aluminum hydride in tetrahydrofuran (THF), afforded 2,3,4tri-O-benzyl-L-idose dimethyl acetal (3a) in 74% yield. The primary hydroxy group was protected with the methoxymethyl group by treatment of (3a) with chloromethylmethylether and di-isopropylethylamine in  $CH_2Cl_2$  (0 °C, 5 h), giving compound (4a) in 79% yield. The Mitsunobu reaction<sup>12</sup> of (4a) with phthalimide in THF was carried out successfully to introduce the phthalimido function at C-5 via S<sub>N</sub>2 reaction, giving mainly compound (5a) in 75% yield. Removal of the phthaloyl group of (5a) was effected by treatment with hydrazine in methanol and the resulting amine was successively converted into the N-t-butoxycarbonyl derivative (6a) in 74% yield. Hydrogenolysis of (6a) in ethanol in the presence

Table 1. Inhibitory activity of the hydrogen sulphite adducts against three enzymes.

Compounds	α-Glucosidasea	β-Glucosidase <sup>b</sup>	α-Manno- sidase <sup>c</sup>
Nojirimycin hydrogen sulphite adduct	77.4 (14.5) <sup>d</sup>	89.6 (8.0)	9.4 (>100)
Mannojirimycin hydrogen sulphite			
adduct	1.3 (>100)	98.0 (4.4)	55.5 (84.0)
(8a)	76.1 (17.0)	85.8 (9.4)	11.7 (>100)
(8b)	2.1 (>100)	91.7 (4.5)	31.2 (>100)

<sup>&</sup>lt;sup>a</sup> Yeast α-glucosidase, p-nitrophenyl-α-D-glucopyranoside (0.66 mm), PBS (100 mm), pH 6.8. <sup>b</sup> Almonds β-glucosidase, p-nitrophenyl-β-D-glucopyranoside (0.33 mm), acetate buffer (100 mm), pH 5.0. <sup>c</sup> Jack bean α-mannosidase, p-nitrophenyl-α-mannopyranoside (20 mm), acetate buffer (100 mm), pH 4.5. <sup>d</sup> Inhibition (I%) determined at the final concentration of 100 μg/ml; numbers in the parentheses denotes IC<sub>50</sub> (concentrations required to cause 50% inhibition, μg/ml) values.

of Pd(OH)<sub>2</sub> on carbon gave quantitatively the trihydroxy compound (7a), an aqueous solution of which was treated with sulphur dioxide at 40 °C to give, after 3 days, the crystalline

Scheme 1. For convenience, only single enantiomers [(3)—(8)] corresponding to (+)-nojirimycin (a series) are depicted. Reagents and conditions: i, CH(OMe)<sub>3</sub>, TsOH (Ts = OSO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>Me), MeOH, 70 °C, then LiAlH<sub>4</sub>, THF, 0 °C; ii, MOMCl, (Prl)<sub>2</sub>NEt, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C; iii, phthalimide, Ph<sub>3</sub>P, diethylazodicarboxylate, THF, room temp.; iv, H<sub>2</sub>NNH<sub>2</sub>·H<sub>2</sub>O, MeOH, reflux, then (BOC)<sub>2</sub>O, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, room temp.; v, H<sub>2</sub>, 20% Pd(OH)<sub>2</sub>, EtOH; vi, SO<sub>2</sub> gas, H<sub>2</sub>O, 0—40 °C, 3 days.

BOC = t-butoxycarbonyl MOM = methoxymethyl hydrogen sulphite adduct (8a) (58% yield) whose  ${}^{1}H$  and  ${}^{13}C$  n.m.r. (D<sub>2</sub>O) and i.r. (KBr) spectra were superimposable on those of an authentic sample. The adduct was treated with Dowex 1 × 2 (OH<sup>-</sup>) resin to give the free base (1a),  $[\alpha]_{D}^{23}$  + 74° (H<sub>2</sub>O) [lit.  $[\alpha]_{D}^{24}$  + 71° (H<sub>2</sub>O)].

Similarly, starting from the lactone (2b), (-)-nojirimycin (1b),  $[\alpha]_D^{20}$  -74° (H<sub>2</sub>O), was synthesised, which was identical to (1a), except for the sign of the optical rotation.

Biological activity of the hydrogen sulphite adducts (8a,b) of synthetic (1a,b), and authentic samples of (1a) and mannojirimycin are shown in Table 1. Although nojirimycin hydrogen sulphite adducts show somewhat different inhibitory activity  $^{13}$  compared to their parent free nojirimycins, its chemical stability allowed us to compare and evaluate their activity accurately. Judging from the activity of (8b), the synthetic (-)-nojirimycin (1b) would conceivably possess high inhibitory activity against  $\beta$ -glucosidase as well as  $\alpha$ -mannosidase, almost comparable to mannojirimycin, being rather different from its antipode.

The inhibitory activity of (1b) itself, and of the antipodes of mannojirimycin and the corresponding 1-deoxy derivative now become of interest.

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## References

- S. Inouye, T. Tsuruoka, and T. Niida, J. Antibiot. (Tokyo), 1967,
  19A, 288; S. Inouye, T. Tsuruoka, T. Ito, and T. Niida,
  Tetrahedron Lett., 1968, 23, 2124.
- 2 T. Niwa, T. Tsuruoka, H. Goi, Y. Kodama, J. Itoh, S. Inouye, Y. Yamada, T. Niida, M. Nobe, and Y. Ogawa, J. Antibiot., 1984, 37, 1579.
- 3 T. Niwa, S. Inouye, T. Tsuruoka, Y. Koaze, and T. Niida, Agric. Biol. Chem., 1970, 34, 966.
- 4 D. D. Schmidt, W. Frommer, L. Müller, and E. Truscheit, Naturwissenschaften, 1979, 66, 584.
- 5 S. Murao and S. Miyata, Agric. Biol. Chem., 1980, 44, 219.
- 6 U. Fuhrmann, E. Bause, G. Legler, and H. Ploegh, *Nature*, 1984, 307, 755.
- 7 R. A. Gruters, J. J. Neefjes, M. Tersmette, R. E. Y. de Goede, A. Tulp, H. G. Huisman, F. Miedema, and H. L. Ploegh, *Nature*, 1987, 330, 74.
- 8 H. Böshagen, F-R. Heiker, and A. M. Schüller, Carbohydr. Res., 1987, 164, 141.
- H. Saeki and E. Ohki, Chem. Pharm. Bull., 1968, 16, 962; Y. Tsuda, Y. Okuno, and K. Kanemitsu, Heterocycles, 1988, 27, 63;
  B. Rajanikanth and S. Seshadri, Tetrahedron Lett., 1989, 30, 755.
- H. Iida, N. Yamazaki, and C. Kibayashi, J. Org. Chem., 1987, 52,
  3337; A. Vasella and R. Voeffray, Helv. Chim. Acta, 1982, 65,
  1134.
- N. Chida, E. Yamada, and S. Ogawa, J. Carbohydr. Chem., 1988, 7, 555.
- 12 O. Mitsunobu, Synthesis, 1981, 1.
- 13 Y. Kodama, T. Tsuruoka, T. Niwa, and S. Inouye, *J. Antibiot.*, 1985, **38**, 116.