Neuraminic Acid and Related Compounds. V. Syntheses of Biologically Active Sialosyl-Glycerol Derivatives and Galactosyl-Glycerol Derivative

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New 1-acyl-sialosyl-glycerol derivatives (1a— $d\alpha$, 1a— $d\beta$, 2α , 2β , which mimic the structure of the capusular polysaccharide of group C meningococcal were synthesized by the use of a chiral glycerol derivative, and were found to have phospholipases A_2 and C inhibitory activities. Furthermore, syntheses of 2-palmitoyl-sialosyl-glycerol derivative (4α , 4β , 5α , 5β), galactosyl-glycerol derivative (6), and sialosyl-galactosyl-glycerol derivative (7) were carried out to examine the difference between these activities. Among these sialosyl derivatives, 3-palmitoyl-sialosyl-glycerol derivatives (1— 3α , 1— 3β) demonstrated the most potent inhibitory activities.

Keywords sialosyl-glycerol derivative; phospholipase A_2 inhibitor; phospholipase C inhibitor; galactosyl-glycerol derivative; sialosyl-galactosyl-glycerol derivative

Capsular polysaccharides are located on the surface of the bacterial cell wall. Therefore, they are important agents in bacterial pathogenesis, and they also interact directly with the host's immune system. The capusular polysaccharide of group C meningococcal, whose structure was determined by Gotshlich and coworkers, 1) includes an α (2 \rightarrow 9) linked homopolymer of sialic acid and phosphoglycerolipid. We conducted synthesis studies on biologically active new compounds by modifying the cell wall structures of gram negative bacteria. 2) As an extension of these studies, we have focused our attention on a polysaccharide

involving sialic acid which plays important roles in various phenomena in living organisms. We synthesized sialosyl derivatives to search into a lead compound for medicines. In a recent communication, we described the novel syntheses of (S)- and (R)-3-O-acyl-1-O-sialosyl glycerol derivatives ($1a-d\alpha$, $1a-d\beta$, 2α , 2β , 3α , 3β : Fig. 1) which were expected to produce phospholipases A_2 and C inhibitory activities. This paper describes these results in detail, as well as the syntheses of new compounds, (S)- and (R)-2-O-palmitoyl-1-O-sialosyl-glycerol (4α , 4β , 5α , 5β : Fig. 1), (S)-1-O-galactosyl-3-O-palmitoyl-glycerol (6: Fig. 1) and

 $1 d\alpha$, $1 d\beta$: R_1 =Me, R_2 =H, R_3 =OH, R_4 =- $C(CH_2)_3CH_3$ CH_2CH_3

 2α , 2β : R₁=Me, R₂=OH, R₃=H, R₄=-(CH₂)₁₄CH₃ 3α , 3β : R₁=H, R₂=H, R₃=OH, R₄=-(CH₂)₁₄CH₃

OAC OAC COOMe

ACOM
H
ACO
$$R_5$$
 $C - R_6$
 $C - R_6$

$$\begin{array}{c} 4\,\alpha,\ 4\,\beta:\ R_5 = -C(CH_2)_{14}CH_3,\ R_6 = H\\ O\\ 5\,a,\ 5\,b:\ R_6 = H,\ R_6 = -C(CH_2)_{14}CH_3\\ O\\ \end{array}$$

Fig. 1

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(S)-3-O-palmitoyl-1-O-sialosylgalactosyl-glycerl derivatives (7: Fig. 1) in order to clarify the influence of such structural changes.

Synthesis of glycosyl acceptors, glycerol derivatives, was carried out in Chart 1. (S)-1-O-Acetyl-2-O-benzylglycerol (8), used as a starting material, 4) was treated with trityl chloride and pyridine at 80°C to produce a tritylated compound (9) in 72.5% yield. Deacetylation of 9 was carried out with NH₄OH-MeOH (1:10) at room temperature to afford the 1-hydroxyl compound (10), as the intermediate of (S)- and (R)-glycosyl acceptor, in 77.4% yield. Acylation of 9 with a variety of acyl groups proceeded smoothly. Compound (9) was acylated with hexadecanoyl chloride, dodecanoyl chloride, octanoyl chloride, and α-ethylhexanovl chloride in the presence of triethylamine at room temperature to produce 11a (85.0%), 11b (93.7%), 11c (88.8%), and 11d (86.0%) respectively. The protective trityl groups of 11a—d were removed by 80% AcOH at 80 °C to obtain glycosyl acceptors of (S)-3-acylsialosyl-(galactosyl)-glycerol derivatives ($1a-d\alpha$, $1a-d\beta$, 3α , 3β , 6, 7), 12a (74.6%), 12b (80.3%), 12c (85.8%), and 12d (73.9%), respectively. The glycosyl acceptors of (R)sialosyl-glycerol derivatives $(2\alpha, 2\beta, 4\alpha, 4\beta)$ were synthesized as follows. The 1-hydroxyl group of 10 was protected with monochloroacetyl chloride and triethylamine at room temperature to afford 13 in a 91.5% yield. The trityl group of 13 was removed with 80% AcOH to produce the 3-hydroxyl compound (14), glycosyl acceptor of (R)-2-acylsialosyl-glycerol derivatives $(4\alpha, 4\beta)$ in a 69.0% yield. Treatment of 14 with hexadecanoyl chloride and triethylamine afforded the acylated compound (15) in a 70.1% yield. Demonochloroacetylation was carried out with disopropylethylamine and thiourea in tetrahydrofuran (THF) to give the (R)-glycosyl acceptor (16) of 2α and 2β in a 92.7% yield. The glycosyl acceptor of (S)-2-acyl-sialosyl-glycerol derivatives (5α , 5β) was obtained in two steps from the starting material (8). The 3-hydroxyl group of 8 was protected with a *tert*-butyldimethylsilyl group by use of *tert*-butyldimethylsilyl chloride and triethylamine to obtain the silylated compound (17) in an 84.0% yield. The 1-acetyl group of 17 was removed by KOH–MeOH (1:10) to give the 1-hydroxyl derivative (18), glycosyl acceptor of 5α and 5β

As a glycosyl donor, 5-acetamido-2-chloro-4,7,8,9-tetra-O-acetyl-D-glycero-D-galacto-2-nonulosonic acid methyl ester (19), prepared from N-acetylneuraminic acid in three steps, ⁵⁾ was used for glycosylation of all sialosyl-glycerol derivatives except 3α and 3β . The glycosyl donor of 3α and 3β was compound (22), the benzyl ester type of 19 which could be removed by hydrogenolysis in the latter step of the synthetic route.

The synthetic route of 3-acyl-sialosyl-glycerol derivatives is shown in Chart 2. Glycosilation of the glycosyl acceptor (12a—d) with the glycosyl donor (19) in the presence of $Hg(CN)_2$ - $HgBr_2$ -Molecular Sieves 4A (MS4A) in CH_2Cl_2 at room temperature for 4d produced the sialosyl-glycerol derivatives (20a—d $\alpha\beta$) as a mixture of anomers. Separation by preparative thin layer chromatography (CHCl₃: MeOH=

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$$\begin{array}{c} O\\ AcO_{m.}\\ AcO_{m$$

Chart 2

20:1) afforded **20a**— $d\alpha$ (**20a** α , 26.9%; **20b** α , 10.8%; **20c** α , 15.6%; **20d** α , 27.0%) and **20a**—**d** β (**20a** β , 32.4%; **20b** β , 11.5%; **20c** β , 17.1%; **20d** β , 23.5%). The anomeric stereochemistry of $20a-d\alpha$ and $20a-d\beta$ was determined by chemical shifts of 3-H_{eq} of the proton nuclear magnetic resonance (${}^{1}H$ -NMR) spectrum. It is known that for α anomers the chemical shift of 3-H_{eq} varies between δ 2.6—2.8, while for β anomers that of 3- \dot{H}_{eq} are δ 2.1—2.5.6) 3- H_{eq} signals of **20a**—**d** α were 2.62, 2.61, 2.61, 2.62 ppm, respectively (ranges of α glycoside). In the case of **20a**—**d** β , these signals overlapped with the methylene proton signals of fatty acids in ¹H-NMR spectrum. Therefore, the anomeric configuration of $20a-d\beta$ as determined by the fact that the 3- H_{eq} signals of 20a— $d\beta$ were not observed downfield from 2.6 ppm, and their other signals supported the structure of $20a-d\beta$. The protective benzyl group of **20a**— $d\alpha$ and **20a**— $d\beta$ was cleaved by hydrogenation in the presence of 30% Pd(OH)₂-C in MeOH to yield (S)sialosyl-glycerol derivatives, $1a-d\alpha$ ($1a\alpha$, 71.2%, $1b\alpha$, 95%; $1c\alpha$, 95%; $1d\alpha$, 82%) and $1a-d\beta$ ($1a\beta$, 74.2%; $1b\beta$, 87%; $1c\beta$, 96%; $1d\beta$, 85%).

Glycosilation of the (R)-glycosyl acceptor (16) and the chloride (19) was carried out as described for 20a— $d\alpha\beta$ to afford (R)-sialosyl-glycerol derivatives (21α , 24.6%; 21β , 20.9%). Each of 21α and 21β were converted to debenzylated compounds (2α and 2β) by usual hydrogenolysis with yields of 86.2% and 84.8%, respectively.

Similarly, the glycerol derivative (12a) was glycosylated with the benzyl ester donor (22) to obtain 23α and 23β (23 α , 22.2%; 23 β , 17.7%). The two protective benzyl

groups of 23α and 23β were removed by hydrogenolysis in the presence of 30% Pd(OH)₂-C to afford the free carboxylic acid derivatives (3α , quant.; 3β , 96.2%).

Syntheses of 2-acyl-sialosyl-glycerol derivatives (4α , 4β , 5α , 5β) were shown in Chart 3. Glycosylation of the silyl derivative (18) with the chloride (19) in the presence of $Hg(CN)_2$ – $HgBr_2$ –MS4A afforded the sialosyl-silylglycerol derivatives (24α , 11.2%; 24β , 8.8%). The benzyl groups of 24α and 24β were smoothly removed by hydrogenolysis with 30% $Pd(OH)_2$. Chemical yields of the debenzylated compounds (25α , 25β) were 77.0% and 85.5%, respectively. The 2-hydroxyl groups of 25α and 25β were acylated with palmitic acid, 1,3-dicyclohexylcarbodiimide (DCC), and 4-dimethylaminopyridine (DMAP), and then the *tert*-butyldimethylsilyl group was selectively removed by aqueous hydrogen fluoride in CH_3CN – $CHCl_3$ to 4α and 4β (4α , 24.7%; 4β , 22.5%), respectively.

The monochloroacetyl glycosyl acceptor (14) and the chloride (19) were converted to sialosyl-glycerol derivatives (27 α and 27 β) via 26 α and 26 β exactly as described for 18+19 \rightarrow 25 α +25 β (14+19 \rightarrow 26 α +26 β : 26 α , 2.8%; 26 β , 16.4%; 26 \rightarrow 27: 26 α , 34.0%; 26 β , 50.4%), respectively. The glycerol-2-hydroxyl group of 27 α and 27 β was acylated with palmitic acid, DCC, and DMAP, and then the monochloroacetyl group was selectively removed with diisopropylethylamine and thiourea to yield 5 α and 5 β (5 α , 16.9%; 5 β , 14.9%; 26 \rightarrow 5).

The synthetic routes of the galactosyl-glycerol derivative (6) and the sialosyl-galactosyl-glycerol derivative (7) are shown in Chart 4. Glycosylation was achieved by use of

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Chart 4

the imidate method. With the glycosyl donor, the galactosyl imidate (28), prepared from D-galactose in three steps, tive (29) in a 53.9% yield. Hydrogenolysis of 29 was was glycosylated from the glycerol derivative (12a) in the performed with a catalyst of Pd(OH)₂-C to obtain 6 in

presence of BF₃-Et₂O to give the galacto-glycerol deriva-

93.6% yield. An α (2 \rightarrow 6)-linked sialosyl-galactosyl derivative (30) was obtained by glycosylation of 1,2,3,4-tetra-O-benzyl-D-galactose, prepared efficiently from D-galactose with chloride (19). The disaccharide was hydrogenated in the presence of Pd(OH)₂-C to afford the tetrahydroxyl compound (31), and then acetylated with acetic anhydride and pyridine to produce a nonaacetyl compound (32). Selective deacetylation at the anomeric position of the compound (32) was achieved with hydrazine acetate to produce the 1-hydroxyl compound (33). Compound (33) was transformed into the imidate (34) in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene(DBU)-trichloroacetonitril as a glycosyl acceptor. Glycosilation of the glycerol derivative (12a) with 34 was performed by treatment of BF₃-Et₂O to yield the sialosyl-galactosyl-glycerol derivative (35) in a 14% yield (33 \rightarrow 35). Hydrogenolysis of 35 was carried out with Pd(OH)₂-C to produce 7 in 81% yield. The structures of all compounds were characterized by ¹H-NMR spectroscopy, as well as infrared (IR) spectroscopy, elemental analyses, and fast-atom bombardment (FAB) mass spectroscopy.

The biological effects⁷⁾ (phospholipases A_2 and C inhibitions) of all compounds ($1\mathbf{a} - \mathbf{d}\alpha$, $1\mathbf{a} - \mathbf{d}\beta$, $2 - 5\alpha$, $2 - 5\beta$, 6, 7) were tested. 3-Palmitoyl-sialosyl-glycerol derivatives ($1 - 3\alpha$, $1 - 3\beta$) and 2-palmitoyl derivative (4α) possessed the strongest activities, while the 2-palmitoyl derivatives (4β , 5α , 5β), the galactosyl-glycerol derivative (6), and the sialosyl-galactosyl-glycerol derivative (7) showed little or no inhibitory activity.

Experimental

All melting points were determined with a micro-melting point apparatus (Yanagimoto) and are uncorrected. Optical rotations were measured on a JASCO-DIP-140 digital polarimeter. IR spectra were measured on JASCO A-202 and JASCO IR-810 infrared spectrophotometers. ¹H-NMR spectra were recorded on JEOL JNM-FX90Q (90 MHz), JEOL JNM-270GX (270 MHz), and JEOL JNM-500GX (500 MHz) spectrophotometers using tetramethylsilane (TMS) as an internal standard. Chemical shifts were recorded in values (δ) downfield from TMS, and the abbreviations of signal patterns are as follows: s, singlet; d, doublet; t, triplet; m, multiplet; br, broad. Thin layer chromatography (TLC) was performed on silica gel (Kieselgel 60F₂₅₄ on aluminium sheet, Merck). All compounds were located by spraying with sulfuric acid and heating on a hot plate. Preparative TLC was performed on the preparative layer chromatography plate (Kieselgel 60F₂₅₄ 2 and 0.5 mm, Merck). Column chromatography was performed on silica gel (Kieselgel 60, 70-230 mesh, Merck).

(S)-1-O-Acetyl-2-O-benzyl-3-O-tritylglycerol (9) Trityl chloride (8.76 g, 3.14×10^{-2} mol) was added to a solution of (S)-1-O-acetyl-2-O-benzylglycerol (8, 4.70 g, 2.09×10^{-3} mol) in dry pyridine (50 ml). The mixture was heated at 80 °C for 3 h, diluted with CHCl₃ (200 ml), and washed with saturated aqueous CuSO₄ and brine. The organic phase was concentrated to dryness and the residue was purified on a column of silica gel (CHCl₃: n-hexane = 10:1) to produced 9 (7.09 g, 72.5%) as colorless oil. $[\alpha]_D + 11.3^\circ$ (c = 1.98, CHCl₃). IR (neat): 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.97 (3H, s, -COCH₃), 4.60 (2H, s, -CH₂Ph), 7.20—7.53 (20H, m, phenyl × 4).

(R)-2-O-Benzyl-1-O-tritylglycerol (10) 9 (753 mg, 1.61×10^{-3} mol) in NH₄OH–MeOH (1:10) (50 ml) was stirred at room temperature for 15 h. The resulting mixture was evaporated to dryness and subjected to column chromatography on silica gel (CHCl₃) to afford 10 (530 mg, 77.4%) as a colorless solid. [α]_D +22.5° (c=0.58). IR (neat): 3450, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 3.24—3.28 (2H, s, -CH₂OH), 4.57 (2H, d, J=4.4 Hz, -CH₂Ph), 7.06—7.54 (20H, m, phenyl × $\overline{4}$).

General Procedure for Syntheses of (S)-1-O-Acyl-2-O-benzyl-3-O-trityl-glycerol (11a—d) Acyl chloride (8.69 \times 10⁻² mol) was added to a solution of 10 (30.8 g, 7.25 \times 10⁻² mol) and triethylamine (11.0 g, 1.09 \times 10⁻¹ mol) in dry CH₂Cl₂ (200 ml) at 0 °C, and then stirred at room temperature for

15 h. The solution was washed with brine, dried (MgSO₄), and purified on silica gel (CHCl₃:n-hexane=10:1) to produce 11a—d (11a, 85.0%; 11b, 93.8%; 11c, 88.8%; 11d, 86.0%) as colorless oils.

11a: $[\alpha]_D + 12.6^\circ$ (c = 0.66, CHCl₃). IR (neat): 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.1 Hz, $-\text{CO(CH}_2)_{14}\text{CH}_3$), 1.26 (26H, s, $-\text{COCH}_2(\text{CH}_2)_{13}\text{CH}_3$), 2.22 (2H, t, J = 7.3 Hz, $-\text{COCH}_2(\text{CH}_2)_{13}\text{CH}_3$), 7.14—7.52 (20H, m, phenyl × 4).

11b: $[\alpha]_D + 11.6^{\circ}$ (c = 1.16, CHCl₃). IR (neat): 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.87 (3H, t, J = 6.1 Hz, $-\text{CO}(\text{CH}_2)_{10}\text{CH}_3$), 1.25 (18H, br s, $-\text{COCH}_2(\text{CH}_2)_9\text{CH}_3$), 2.23 (2H, t, J = 7.3 Hz, $-\text{COC}\overline{\text{H}_2}(\text{CH}_2)_9\text{CH}_3$), 7.12—7.53 (20H, m, phenyl × 4).

11c: $[\alpha]_D + 4.7^\circ$ (c = 1.28, CHCl₃). IR (neat): 1740, 700 cm⁻¹). ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.6 Hz, $-CO(CH_2)_6CH_3$), 1.26 (10H, br s, $-COCH_2(CH_2)_5CH_3$), 2.24 (2H, t, J = 7.3 Hz, $-COC\overline{H_2}(CH_2)_5CH_3$), 7.16—7.55 (20H, m, phenyl × 4).

11d: $[\alpha]_D + 9.3^\circ$ (c = 0.74, CHCl₃). IR (neat): 1735, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.89 (6H, brt, J = 6.8 Hz, $-\text{CH}_2\text{CH}_3 \times 2$), 1.09—1.15 (8H, $-\text{CHC}_2\text{CH}_3$, $-\text{CH}(\text{CH}_2)_3\text{CH}_3$), 7.04—7.55 (20H, m, phenyl × 4).

General Procedure for Syntheses of (S)-1-O-Acyl-2-O-benzylglycerol (12a—d) A solution of 11a—d (6.17× 10^{-2} mol) in 80% acetic acid (300 ml) was heated at 80 °C for 1 h, evaporated to dryness, and purified on a column of silica gel (CHCl₃) to afford 12a—d (12a, 74.6%; 12b, 80.3%; 12c, 85.8%; 12d, 73.9%) as colorless oils.

12a: $[\alpha]_D - 4.8^{\circ}$ (c = 1.16, CHCl₃). IR (neat): 3470, 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.1 Hz, $-\text{CO}(\text{CH}_2)_{14}\text{CH}_3$), 1.25 (26H, s, $-\text{COCH}_2(\text{CH}_2)_{13}\text{CH}_3$), 2.32 (2H, t, J = 7.6 Hz, $-\text{COC}\underline{\text{H}_2}(\text{CH}_2)_{13}\text{CH}_3$), 7.34 (5H, s, phenyl).

12b: $[\alpha]_D - 5.7^\circ$ (c = 0.58, CHCl₃). IR (neat): 3450, 1740, 700 cm⁻¹.

¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.3 Hz, $-\text{CO}(\text{CH}_2)_{10}\text{CH}_3$), 1.25 (18H, $-\text{COCH}_2(\text{CH}_2)_9\text{CH}_3$), 2.30 (2H, t, J = 8.6 Hz, $-\text{COC}\underline{\text{H}_2}(\text{CH}_2)_9\text{CH}_3$), 4.62 (2H, s, $-\text{CH}_2\overline{\text{Ph}}$), 7.30 (5H, s, phenyl).

12c: $[\alpha]_D - 6.4^\circ$ (c = 0.28, CHCl₃). IR (neat): 3450, 1730, 690 cm⁻¹.

¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.3 Hz, $-\text{CO}(\text{CH}_2)_6\text{CH}_3$), 1.27 (10H, s, $-\text{COCH}_2(\text{CH}_2)_6\text{CH}_3$), 2.32 (2H, t, J = 7.6 Hz, $\frac{\text{COCH}_2(\text{CH}_2)_6\text{CH}_3}{\text{CH}_3}$), 7.33 (5H, s, phenyl).

12d: $[\alpha]_D - 8.0^\circ$ (c = 1.00, CHCl₃). IR (neat): 3450, 1740, 700 cm⁻¹.

¹H-NMR (CDCl₃): 0.89 (6H, t, J = 6.8 Hz, $-\text{CH}_2\text{CH}_3 \times 2$), 1.10—1.16 (10H, m, $-\text{CH}(\text{CH}_2)_3\text{CH}_3$, $-\text{CHCH}_2\text{CH}_3$). 7.35 (5H, s, phenyl).

(S)-2-O-Benzyl-I-O-monochloroacetyl-3-O-tritylglycerol (13) To 10 (990 mg, 2.33×10^{-3} mol) and triethylamine (354 mg, 3.50×10^{-3} mol) in dry CH₂Cl₂ (30 ml), monochloroacetyl chloride (316 mg, 2.80×10^{-3} mol) was added at 0 °C under argon. The mixture was stirred at room temperature for 3 h, washed with brine, and dried (MgSO₄). The residue was chromatographed on silica gel (CHCl₃) to produce 13 as a colorless oil (1.07 g, 91.5%). $[\alpha]_D + 10.7^\circ$ (c = 2.26, CHCl₃). IR (neat): 1740, $700 \, \text{cm}^{-1}$. ¹H-NMR (CDCl₃) δ : 3.92 (2H, s, -COCH₂Cl), 4.58 (2H, s, -CH₂Ph), 7.22—7.45 (20H, m, phenyl×4).

(S)-2-O-Benzyl-1-O-monochloroacetylglycerol (14) 13 (1.00 g, 2.00 × 10^{-3} mol) in 80% acetic acid (10 ml) was heated at 80 °C for 1 h, evaporated to dryness, and purified on a column of silica gel (CHCl₃) to afford 14 (356 mg, 69.0%) as a colorless oil. [α]_D -3.1° (c=1.56, CHCl₃). IR (neat): 3450, 1750, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 3.99 (2H, s, -CQCH₂Cl), 4.62 (2H, -CH₂Ph), 7.27 (5H, s, phenyl).

(S)-2-O-Benzyl-3-O-hexadecanoyl-1-O-monochloroacetylglycerol (15) Palmitoyl chloride was added to a solution of 14 (1.34 g, 5.18 × 10^{-3} mol) and triethylamine (1.05 g, 1.04 × 10^{-2} mol) in dry CH₂Cl₂ (30 ml) at 0 °C under argon. The resulting mixture was stirred at room temperature for 15 h, washed with brine, and dried. Purification by the residue was achieved with a column of silica gel (CHCl₃:n-hexane=10:1) to yield 15 (1.81 g, 70.1%) as a colorless oil. [α]_D +2.8° (c=1.60, CHCl₃). IR (neat): 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J=6.1 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 3.99 (2H, s, -COCH₂Cl), $\overline{4.64}$ (2H, s, -CH₂Ph), 7.26 (5H, s, phenyl).

(*R*)-2-*O*-Benzyl-1-*O*-hexadecanoylglycerol (16) A mixture of 15 (1.72 g, 3.47×10^{-3} mol), diisopropylethylamine (538 mg, 4.16×10^{-3} mol), and thiourea (317 mg, 4.16×10^{-3} mol) in dry THF (30 ml) was refluxed for 2 h. The resulting mixture was filtered and the filtrate was concentrated to dryness. The residue was chromatographed on silica gel (CHCl₃) to produce 16 (1.35 g, 92.7%) as a colorless oil. [α]_D +4.7° (c=4.13, CHCl₃). IR (neat): 3445, 1740, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J=6.1 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -CO(CH₂)₁₄CH₃), 2.32 (2H, t, J=7.6 Hz, -COCH₂(CH₂)₁₃CH₃), 7.34 (5H, s, phenyl).

(R)-1-O-Acetyl- $\overline{2}$ -O-benzyl-3-O-tert-butyldimethylsilylglycerol (17) tert-Butyldimethylsilyl chloride was added to a solution of **8** (6.04 g, 2.69×10^{-2} mol) and triethylamine (3.39 g, 3.35×10^{-2} mol) in dry CH₂Cl₂

(50 ml) at 0 °C under argon. The mixture was stirred at room temperature for 3 h, washed with brine, and dried (MgSO₄). The organic phase was evaporated to dryness and the residue was chromatographed on silica gel (CHCl₃) to afford 17 (7.65 g, 84.0%) as a colorless oil. $[\alpha]_D$ +15.6° (c=1.78, CHCl₃). IR (neat): 1745, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.075 (6H, s, -Si(CH₃)₂), 0.89 (9H, s, -SiC(CH₃)₃), 2.04 (3H, s, -CH₂Ph), 4.65 (2H, s, -CH₂Ph), 7.32—7.34 (5H, m, phenyl).

(*R*)-2-*O*-Benzyl-1-*O*-tert-butyldimethylsilylglycerol (18) 17 (6.93 g, 2.05×10^{-2} mol) as described for 10 gave 18 (5.62 g, 92.7%) as a colorless oil. $[\alpha]_D + 17.3^\circ$ (c = 1.00, CHCl₃). IR (neat): 3465, 3400, 1750, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.090 (6H, s, $-\text{Si}(\text{CH}_3)_2$), 0.90 (9H, s, $-\text{Si}(\text{CH}_3)_3$), 4.60, 4.68 (2H, d × 2, J = 11.9 Hz, $-\text{C}_{12}Ph$), 7.29 (5H, s, phenyl).

General Procedure for the Syntheses of 2-O-Benzyl 3-O-acyl-1-O-[methvl(5-acetamido-4.7.8,9-tert-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2nonulopyranosyl)onate]glycerols (20a—d α , 20a—d β , 21 α , 21 β) Glycerol derivatives (12a-d, 16), pulverized MS4A (3g), Hg(CN)₂ (1.26g, 4.99×10^{-3} mol), and HgBr₂ (771 mg, 2.14×10^{-3} mol) were dried by the use of a high vacuum-pump for 2h. The mixture was dissolved in dry CH₂Cl₂ (30 ml) and stirred at room temperature for 1 h under argon. 5-Acetamido-2-chloro-4,7,8,9-tetra-O-acetyl-2,3,5-trideoxy-D-glycero-Dgalacto-2-nonulopyranosonic acid methyl ester (19, $3.64 \,\mathrm{g}$, 7.13×10^{-3} mol) in dry CH₂Cl₂ (20 ml) was added by drops to the mixture within 1 h at room temperature, and the suspension was then stirred for 4d. The resulting mixture was diluted with CH2Cl2, filtered, and the filtrate was washed with aqueous 10% KI and brine. The organic phase was dried (MgSO₄), concentrated to dryness, and purified on a column of silica gel (CHCl₃: MeOH = 40:1) to yield **20a**— $d\alpha\beta$ and **21** $\alpha\beta$. The anomeric mixture was further purified on preparative TLC (CHCl₃: MeOH = 20:1) to afford 20a—da, (20aa, 26.9%; 20ba, 10.8%; 20ca, 15.6%; 20da, 27.0%), **20a**—**d** β (**20a** β , 32.4%; **20b** β , 11.5%; **20c** β , 17.1%; **20d** β , 23.5%), **21** α (24.6%), and 21β (20.9%) as colorless, amorphous powders.

20a α : $[\alpha]_D - 8.5^\circ$ (c = 1.16, CHCl₃). IR (neat): 3220, 1745, 1660, 1540, 709 cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 5.9 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.88—2.14 (15H, m, -COCH₃×5), 2.62 (1H, dd, J = 4.6, 12.4 Hz, 3-H_{eq}), 3.76 (3H, s, -COOCH₃), 7.33 (5H, s, phenyl). Positive FAB-MS (M+H)⁺ m/z: 894.

20bα: [α]_D -7.4° (c=1.24, CHCl₃). IR (neat): 3380, 1740, 1660, 1550, 700 cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.88 (3H, t, J=6.1 Hz, -CO(CH₂)₁₀CH₃), 1.25 (18H, s, -CO(CH₂)₁₀CH₃), 1.88—2.20 (15H, m, -COCH₃ × 5), $\overline{2.61}$ (1H, dd, J=4.6, 12.4 Hz, 3-H_{eq}), 3.79 (3H, s, -COOCH₃), 7.29—7.51 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 838.

20cα: $[α]_D - 18.2^\circ$ (c = 0.18, CHCl₃). IR (neat): 3395, 1745, 1660, 1540, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.87 (3H, t, J = 6.1 Hz, $-CO(CH_2)_6CH_3$), 1.27 (10H, s, $-COCH_2(CH_2)_5CH_3$), 1.98—2.13 (15H, m, $-COCH_3 \times 5$), 2.20—2.48 (2H, m, $-COCH_2(CH_2)_5CH_3$), 2.61 (1H, dd, J = 4.1, $\overline{13.1}$ Hz, 3-H_{eq}), 3.79 (3H, s, $COOCH_3$), 7.21—7.56 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 782.

20d α : $[\alpha]_D - 16.7^\circ$ (c = 0.06). IR (neat): 3270, 3360, 1750, 1660, 1540, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (6H, br t, J = 6.8 Hz, $-CH_2CH_3 \times 2$), 1.26 (8H, br s, $-CHCH_2CH_3$, $-CH(CH_2)_3CH_3$), 1.98—2.13 ($\overline{15H}$, m, $-COCH_3 \times 5$), 2.62 ($\overline{1H}$, dd, J = 4.6, 12.4 Hz, 3-H_{eq}), 3.79 (3H, s, $-COOCH_3$), 7.19—7.57 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 782

20a β : $[\alpha]_D$ - 5.5° (c = 0.36, CHCl $_3$). IR (neat): 3390, 1750, 1660, 1540, 700 cm $^{-1}$. ¹H-NMR (CDCl $_3$) δ : 0.88 (3H, t, J = 5.9 Hz, -CO(CH $_2$) $_{14}$ CH $_3$), 1.26 (26H, s, -COCH $_2$ (CH $_2$) $_{13}$ CH $_3$), 1.82—2.08 (15H, m, -COCH $_3$ ×5), 3.80 (3H, s, -COOCH $_3$), 7.37—7.44 (5H, m, phenyl). Positive FAB-MS (M+H) $^+$ m/z: 894.

20bβ: $[\alpha]_D - 6.6^\circ$ (c = 0.64, CHCl₃). IR (neat): 3395, 1750, 1670, 1590, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J = 5.6 Hz, $-\text{CO}(\text{CH}_2)_{10}\text{CH}_3$), 1.25 (18H, s, $-\text{COCH}_2(\text{CH}_2)_9\text{CH}_3$), 1.98—2.08 (15H, m, $-\text{COCH}_3 \times 5$), 3.79 (3H, s, $-\text{COOC}_{\frac{1}{3}}$), 7.22—7.52 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 838.

20cβ: $[\alpha]_D - 10.7^\circ$ (c = 0.64, CHCl₃). IR (neat): 3400, 1750, 1670, 1540, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.87 (3H, t, J = 6.1 Hz, $-CO(CH_2)_6CH_3$), 1.26 (10H, s, $-COCH_2(CH_2)_5CH_3$), 1.98—2.08 (15H, m, $-COCH_3 \times 5$), 3.80 (3H, s, $-COOCH_3$), 7.28—7.57 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 782.

20dβ: $[\alpha]_D - 8.2^\circ$ (c = 0.44, CHCl₃). IR (neat): 3390, 1740, 1680, 1520, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.89 (6H, br t, J = 7.1 Hz, -CH₂CH₃×2), 1.25 (8H, br s, -CH(CH₂)₃CH₃, -CHCH₂CH₃), 1.98—2.08 (15H, m, -COCH₃×5), 3.79 (3H, s, -COOCH₃), 7.22—7.52 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 782.

21 α : $[\alpha]_D - 10.8^\circ$ (c = 1.00, CHCl₃). IR (neat): 3205, 1740, 1660, 1540, 700 cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.1 Hz, -CO(CH₂)₁₄-

CH₃), 1.25 (26 H, s, $-COCH_2(CH_2)_{13}CH_3$), 1.97—2.19 (15H, m, $-\overline{COCH_3} \times 5$), 2.61 (1H, dd, J = 4.6, 12.7 Hz, $3H_{eq}$), 3.75 (3H, s, $-COC\underline{H_3}$), 7.23—7.46 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 894.

21 β : [α]_D -7.0° (c = 1.42, CHCl₃). IR (neat): 3260, 3350, 1740, 1662, 1540, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 5.9 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.92—2.16 (15H, m, -COCH₃× $\overline{5}$), 2.34 (2H, t, J = 5.6 Hz, -COCH₂(CH₂)₁₃CH₃), 3.75 (3H, s, -COOCH₃, 7.15—7.56 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 894.

General Procedure for the Syntheses of 3-O-Acyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulo-pyranosyl)onate]glycerols (1a— $d\alpha$, 1a— $d\beta$, 2α , 2β) A solution of (20a— $d\alpha$, 20a— $d\beta$, 21α , 21β , 7.16×10^{-5} mol) in methanol (1 ml) was hydrogenated in the presence of 30% Pd(OH)₂ on carbon at room temperature for 2 h. The catalyst was filtered off and the filtrate was concentrated to dryness. The residue was purified on a column of silica gel (CHCl₃: MeOH=20:1) to give 1a— $d\alpha$ ($1a\alpha$, 71.2%; $1b\alpha$, 95%; $1c\alpha$, 95%; $1d\alpha$, 82%), 1a— $d\beta$ ($1a\beta$, 74.2%; $1b\beta$, 87%; $1c\beta$, 96%; $1d\beta$, 85%), 2α (86.2%), and 2β (84.8%) as colorless, amorphous powders.

laa: $[\alpha]_D - 11.3^\circ$ (c = 0.50, CHCl₃). IR (neat): 3400, 1750, 1660, 1560 cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.4 Hz, -CO(CH₂)₁₄CH₃), 1.26 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.89—2.14 (15H, m, -COCH₃×5), 2.59 (1H, dd, J = 4.9, 12.9 Hz, 3-H_{eq}), 3.81 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 804.

1bα: $[\alpha]_D - 8.0^\circ$ (c = 0.64, CHCl₃). IR (neat): 3350, 1740, 1660, 1550 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J = 5.6 Hz, $-\text{CO}(\text{CH}_2)_{10}\text{CH}_3$), 1.26 (18H, s, $-\text{COCH}_2(\text{CH}_2)_9\text{CH}_3$), 1.84—2.19 (15H, m, $-\text{COCH}_3 \times 5$), 2.61 (1H, dd, J = 4.4, 12.4 Hz, 3-H_{eq}), 3.82 (3H, s, $-\text{COOC}_{\underline{\text{H}}_3}$). Positive FAB-MS (M+H)⁺ m/z: 748.

1cα: $[α]_D - 7.8^\circ$ (c = 0.52, CHCl₃). IR (neat): 3400, 1740, 1645, 1550 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J = 5.6 Hz, $-CO(CH_2)_6CH_3$), 1.26 (18H, s, $-COCH_2(CH_2)_5CH_3$), 1.89—2.15 (15H, m, $-COCH_3 \times 5$), 2.61 (1H, dd, J = 4.9, 12.4 Hz, 3-H_{eq}), 3.81 (3H, s, $-COOCH_3$). Positive FAB-MS (M+H)⁺ m/z: 692.

1dα: $[α]_D - 4.9^\circ$ (c = 0.56, CHCl₃). IR (neat): 3350, 1740, 1665, 1550 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.89 (6H, brt, J = 7.3 Hz, $-CH_2CH_3 \times 2$), 1.26 (8H, brs, J = 7.3 Hz, $-CH(CH_2)_3CH$, $-CHCH_2CH_3$), 1.89—2.14 (15H, m, $-COCH_3 \times 5$), 2.61 (1H, dd, J = 4.6, 12.4 Hz, 3-H_{eq}), 3.81 (3H, s, $-COOCH_3$). Positive FAB-MS (M+H)⁺ m/z: 692.

1aβ: [α]_D −12.6° (c=0.18, CHCl₃). IR (neat): 3360, 1750, 1660, 1550 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J=6.4 Hz, −CO(CH₂)₁₄CH₃), 1.26 (26H, s, −COCH₂(CH₂)₁₃CH₃), 1.90—2.16 (15H, m, −COCH₃ × 5), 3.81 (3H, s, −COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 804.

1bβ: $[\alpha]_D - 10.0^\circ$ ($\overline{c} = 0.52$, CHCl₃). IR (neat): 3355, 1740, 1660, 1555 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J = 5.6 Hz, $-\text{CO}(\text{CH}_2)_{10}\text{CH}_3$), 1.26 (18H, s, $-\text{COCH}_2(\text{CH}_2)_9\text{CH}_3$), 1.85—2.22 (15H, m, $-\text{COCH}_3 \times \overline{5}$), 3.80 (3H, s, $-\text{COOCH}_3$). Positive FAB-MS (M+H)⁺ m/z: 748.

1cβ: $[\alpha]_D$ – 5.1° $(\overline{c}$ =1.94, CHCl₃). IR (neat): 3400, 1740, 1650, 1540 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J=6.1 Hz, -CO(CH₂)₆CH₃), 1.26 (10H, s, -COCH₂(CH₂)₅CH₃), 1.90—2.16 (15H, m, -COCH₃×5), 3.81 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 692.

1d β : $[\alpha]_D - 6.0^\circ$ (\overline{c} =1.68, CHCl₃). IR (neat): 0.89 (6H, brt, J=6.8 Hz, -CH₂CH₃×2), 1.26 (8H, brs, -CH(CH₂)₃CH₃, -CHCH₂CH₃), 1.90—2.16 (15H, m, -COCH₃×5), 2.51 (1H, dd, J=4.6, 12.4 Hz, 3-H_{eq}), 3.80 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 692.

2α: $[\alpha]_D$ – 3. $\overline{6}^\circ$ (c = 0.52, CHCl₃). IR (neat): 3390, 1740, 1665, 1540 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.1 Hz, $-\text{CO}(\text{CH}_2)_{14}\text{CH}_3$), 1.25 (26H, s, $-\text{COCH}_2(\text{CH}_2)_{13}\text{CH}_3$), 1.86—2.20 (15H, m, $-\text{COCH}_3 \times 5$), 2.36 (2H, t, J = 7.1 Hz, $-\text{COCH}_2(\text{CH}_2)_{13}\text{CH}_3$), 2.60 (1H, dd, J = 4.4, 12.6 Hz, 3-H_{eq}), 3.81 (3H, s, $-\text{COOC}_{\underline{\textbf{H}}_3}$). Positive FAB-MS (M + H)⁺ m/z: 804.

2 β : [α]_D -4.8° (c=0.30, CHCl₃). IR (neat): 3360, 1740, 1660, 1545 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.87 (3H, t, J=4.4 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.89—2.42 (15H, m, -COCH₃ \times 5), 3.80 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 804.

(S)-2-O-Benzyl-1-O-[benzyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]-3-O-hexadecanoylglycerol (23α, 23β) 12a (242 mg, 5.75×10^{-4} mol), Hg(CN)₂ (204 mg, 8.08×10^{-4} mol), HgBr₂ (124 mg, 3.44×10^{-4} mol), MS4A (1 g), and 5-acetamido-2-chloro-4,7,8,9-tetra-O-acetyl-2,3,5-trideoxy-D-glycero-D-galacto-2-nonulosonic acid benzyl ester (799 mg, 1.19×10^{-3} mol) described for 20α afforded 23α (124 mg, 22.2%) and 23β (99 mg, 17.7%) as colorless, amorphous powders.

23α: $[\alpha]_D$ – 4.1° (c = 1.38, CHCl₃). IR (neat): 3255, 3350, 1740, 1640, 1550, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 5.9 Hz, -CO(CH₂)₁₄CH₃), 1.26 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.91—2.17 (15H, m, -COCH₃ \times 5),

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2.67 (1H, dd, J=4.6 Hz, 3-H_{eq}), 5.18 (2H, s, -COOCH₂Ph), 7.31, 7.34 (10H, s×2, phenyl×2), positive FAB-MS (M+H)⁺ m/z: 970.

23 β : [α]_D -6.4° (c = 1.00, CHCl₃). IR (neat): 3395, 1740, 1680, 1520, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 5.9 Hz, -CO(CH₂)₁₄CH₃), 1.26 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.91—2.17 (15H, m, -COCH₃×5), 5.22 (2H, s, -COOCH₂Ph), 7.22—7.54 (10H, m, phenyl×2), positive FAB-MS (M+H)⁺ m/z: 970.

(S)-1-O-(5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)-3-O-hexadecanoyl-glycerol (3 α , 3 β) A solution of 23 α and 23 β (33 mg, 3.40 × 10⁻⁵ mol) in MeOH (1 ml) was hydrogenated in the presence of 30% Pd(OH)₂-C at room temperature. The catalyst was filtered off and the filtrate was concentrated to dryness. The residue was chromatographed on silica gel (CHCl₃: MeOH = 30:1) to give 3 α (27 mg, quant.) and 3 β (23 mg, 96.2%), respectively.

3α: [α]_D – 13.8° (c=0.26, CHCl₃). IR (neat): 3480, 1740, 1660, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, –CO(CH₂)₁₄CH₃), 1.26 (26H, s, –COCH₂(CH₂)₁₃CH₃), 1.92–-2.16 (15H, m, –COCH₃×5), 2.35 (2H, t, J=7.3 Hz, –COCH₂(CH₂)₁₃CH₃), 5.97 (1H, d, J=9.6 Hz, –NH₋). Positive FAB-MS (M+H)⁺ m/z: 790.

3β: [α]_D –13.3° (c=0.36, CHCl₃). IR (neat): 3480, 1740, 1660, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J=5.9 Hz, -CO(CH₂)₁₄CH₃), 1.26 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.92—2.16 (15H, m, -COCH₃×5), 2.35 (2H, t, J=7.3 Hz, -COCH₂(CH₂)₁₃CH₃), 2.54 (1H, dd, J=4.8, 13.2 Hz, 3-H_{eq}), 5.97 (1H, d, J=9.7 Hz, -NH-).

(R)-2-O-Benzyl-3-O-tert-butyldimethylsilyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-2-nonulopyranosyl)onate]glycerol (24 α , 24 β) Glycerol derivative (18, 1.76 g, 5.94 × 10⁻³ mol), Hg(CN)₂) (1.82 g, 7.21 × 10⁻³ mol), HgBr₂ (1.11 g, 3.09 × 10⁻³ mol), and MS4A (6g) described for 20 α gave 24 α (513 mg, 11.2%) and 24 β (401 mg, 8.8%) as colorless, amorphous powders.

24 α : $[\alpha]_D - 10.0^\circ$ (c = 0.60, CHCl₃). IR (neat); 3375, 1745, 1665, 1550, 700 cm⁻¹. 1 H-NMR (CDCl₃) δ : 0.90 (9H, s, –SiC(CH₃)₃), 1.88—2.17 (15H, m, –COCH₃×2), 2.62 (1H, dd, J = 4.4, 12.4 Hz, $\overline{3}$ -H_{eq}). 3.75 (3H, s, –COOCH₃), $\overline{7.30}$ —7.38 (5H, m, –CH₂Ph). Positive FAB-MS (M+H) + m/z: 770.

24 β : [α]_D -6.1° (c=1.18, CHCl₃). IR (neat): 3400, 1740, 1660, 1550, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.90 (9H, s, $-\text{SiC}(\text{CH}_3)_3$), 1.86 (1H, t, J=12.8 Hz, 3-H_{eq}), 1.78—2.12 (15H, m, $-\text{COCH}_3 \times 5$), 2.45 (1H, dd, J=4.8, 12.8 Hz, 3-H_{eq}), 3.78 (3H, s, $-\text{COOCH}_3$), 7.29—7.44 (5H, m, $-\text{CH}_2$ Ph). Positive FAB-MS (M+H)⁺ m/z: 770.

(R)-3-O-tert-Butyldimethylsilyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]-glycerol (25 α , 25 β) A solution of 24 α and 24 β (3.20 × 10⁻⁴ mol) in EtOH (3 ml) was hydrogenated in the presence of 30% Pd(OH)₂-C (25 mg) at room temperature, the reaction being monitored by TLC (CHCl₃: MeOH = 20:1, Rf=0.40). The catalyst was filtered off and concentrated to dryness. The residue was purified on a column of silica gel (CHCl₃: MeOH = 50:1) and further purified by preparative TLC (CHCl₃: MeOH = 20:1) to afford 25 α (167 mg, 77.0%) and 25 β (185 mg, 85.5%) as colorless, amorphous powders, respectively.

25 α : $[\alpha]_D$ – 10.0° (c = 0.60, CHCl₃). IR (neat): 3430, 1740, 1660, 1540 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.08 (6H, s, -Si(C[H₃)₂), 0.92 (9H, s, -SiC(C[H₃)₃), 1.89—2.15 (15H, m, -COC[H₃) × 5), 2.59 (1H, dd, J = 4.4, 12.8 Hz, $\overline{3}$ -H_{eq}), 3.82 (3H, s, -COOC[H₃). Positive FAB-MS (M+H) + m/z: 680.

25 β : [α]_D -4.5° (c=0.44, CHCl₃). IR (neat): 3400, 1740, 1660, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.10 (6H, s, $-\text{Si}(\text{CH}_3)_2$), 0.92 (9H, s, $-\text{Si}(\text{C(H}_3)_3)$, 1.89—2.16 (15H, m, $-\text{COCH}_3 \times 5$), 2.47 (1H, dd, J=5.1, 12.8 Hz, 3-H_{eq}), 3.82 (3H, s, $-\text{COOC}_{13}$). Positive FAB-MS (M+H)⁺ m/z: 680.

(S)-2-O-Hexadecanoyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]glycerol (4 α , 4 β) DCC (63 mg, 3.07×10^{-4} mol) was added to a solution of 25 α (115 mg, 1.69×10^{-4} mol), palmitic acid (79 mg, 3.07×10^{-4} mol), and DMAP (6 mg, 4.91×10^{-5} mol) in dry CH₂Cl₂ (5 ml) at -15 °C under argon, and then stirred for 3h. The mixture was warmed at room temperature, stirred for 15h, and filtered on Celite. The filtrate was evaporated to dryness and the residue was purified on silica gel (CHCl₃: MeOH = 50:1) to produce a crude acylated compound, to which CH₃CN-CHCl₃ (3:1) involving trace aqueous 46.5% HF was added, and stirred for 1h at room temperature. The resulting mixture was concentrated to dryness and chromatographed on silica gel (CHCl₃: MeOH = 20:1) to afford 4 α (33 mg, 24.7%) as a colorless, amorphous powder.

 4α : [α]_D -15.5° (c=0.66, CHCl₃). IR (neat): 3400, 1740, 1670, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ: 0.88 (3H, t, J=7.0 Hz, -CO(CH₂)₁₄CH₃),

1.26 (26H, s, $-COCH_2(CH_2)_{13}CH_3$), 1.89—2.15 (15H, m, $-COCH_3 \times 5$), 2.35 (2H, t, J=7.5 Hz, $-\overline{COCH_2(CH_2)_{13}CH_3}$), 2.59 (1H, dd, $\overline{J}=4.5$, 12.5 Hz, 3-H_{eq}), 3.81 (3H, s, $-C\overline{OOCH_3}$).

25β (92 mg, 1.35×10^{-4} mol), palmitic acid (67 mg, 2.46×10^{-4} mol), DMAP (8 mg, 6.12×10^{-5} mol), and DCC (54 mg, 2.46×10^{-4} mol) as described for 4α gave 4β (24 mg, 22.5%) as a colorless, amorphous powder.

 4β : [α]_D -8.8° (c=0.48, CHCl₃). IR (neat): 3400, 1750, 1660, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J=7.0 Hz, -CO(CH₂)₁₄CH₃), 1.26 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.89—2.08 (15H, m, -COCH₃×5), 2.35 (2H, t, J=7.7 Hz, -COCH₂(CH₂)₁₃CH₃), 2.47 (1H, dd, \overline{J} =4.8, 12.8 Hz, 3-H_{eq}), 3.81 (3H, s, -COOCH₃).

(S)-2-O-Benzyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]-3-O-monochloro-acetylglycerol (26 α , 26 β) Glycerol derivative (14, 493 mg, 1.91 × 10⁻³ mol), Hg(CN)₂ (1.01 g, 4.00 × 10⁻³ mol), HgBr₂ (1.11 g, 3.09 × 10⁻³ mol), MS4A (3 g), and the chloride (19) as described for 20 α afforded 26 α (41 mg, 2.8%) and 26 β (240 mg, 16.4%) as colorless, amorphous powders.

26 α : $[\alpha]_D - 8.6^\circ$ (c = 0.56, CHCl₃). IR (neat): 3360, 1740, 1660, 1540, 700 cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.95—2.08 (15H, m, -COCH₃×5), 2.61 (1H, dd, J = 4.4, 12.4 Hz, 3-H_{eq}). 3.77 (3H, s, -COCH₃), 4.14 (2H, s, -COCH₂Cl), 4.65 (2H, s, -CH₂Ph), 7.31—7.50 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 732.

26β: [α]_D -10.8° (c=0.48). IR (neat): 3400, 1750, 1670, 1540, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.85—2.09 (15H, m, $-\text{COCH}_3 \times 5$), 2.41 (1H, dd, J=4.9, 12.9 Hz, 3-H_{eq}), 3.77 (3H, s, $-\text{COOCH}_3$), 4.15 (2H, s, $-\text{COCH}_2$ Cl), 7.31—7.50 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 732.

(S)-1-O-[Methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]-3-O-monochloroacetylglycerol (27α, 27β) A solution of 26α (50 mg, 6.51×10^{-5} mol) in MeOH (1 ml) was hydrogenated in the presence of 30% Pd(OH)₂-C (5 mg) at room temperature. The catalyst was filtered off and the filtrate was concentrated to dryness. The residue was purified on a column of silica gel (CHCl₃: MeOH = 20:1) to give 27α (15 mg, 34.0%) as a colorless, amorphous powder. [α]_D -2.7° (c=0.30, CHCl₃). IR (neat): 3325, 1710, 1660, 1510 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.89—2.14 (15H, m, -COCH₃ × 5), 2.63 (1H, dd, J=4.6, 12.4 Hz, 3-H_{eq}), 3.82 (3H, s, -COOCH₃). Positive FAB-MS (M+H)+ m/z: 642.

26β (186 mg, 2.42×10^{-4} mol) and 30% Pd(OH)₂ (19 mg) as described for **27**α gave **27**β (83 mg, 50.4%) as a colorless, amorphous powder. [α]_D -4.5° (c = 0.84, CHCl₃). IR (neat): 3325, 1740, 1660, 1545 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.97—2.17 (15H, m, $-\text{COCH}_3 \times 5$), 2.48 (1H, dd, J = 4.6, 12.7 Hz, 3-H_{eq}), 3.82 (3H, s, $-\text{COOCH}_3$), 4.18 (2H, s, $-\text{COCH}_2$ Cl). Positive FAB-MS (M+H)⁺ m/z: 642.

(R)-2-O-Hexadecanoyl-1-O-[methyl(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosyl)onate]glycerol (5 α , 5 β) DCC (9 mg, 4.4 × 10⁻⁵ mol) was added to a solution of 27 α (15 mg, 2.2 × 10⁻⁵ mol), palmitic acid (11 mg, 4.4 × 10⁻⁵ mol), and DMAP (1.1 mg, 8.8 × 10⁻⁶ mol) in dry CH₂Cl₂ (2 ml) at -15 °C under argon, and then stirred for 3 h. The mixture was warmed at room temperature and stirred for 15 h. The suspension was filtered and the filtrate was washed with brine and dried (MgSO₄). After removal of the solvent, the residue was chromatographed on a flash column of silica gel (CHCl₃: MeOH = 20:1) to afford a crude acylated compound, to which thiourea (2.5 mg, 3.3×10^{-5} mol) and diisopropylethylamine (4.3 mg, 3.32×10^{-5} mol) in dry THF (0.5 ml) was added, and refluxed for 2 h. The resulting mixture was filtered off and the filtrate was concentrated to dryness. The residue was purified by preparative TLC (CHCl₃: MeOH = 20:1) to yield 5 α (3 mg, 16.9%) as a colorless, amorphous powder.

5 α : [α]_D -17.0° (c=0.06, CHCl₃). IR (neat): 3400, 1740, 1660, 1540 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J=7.5 Hz, -CO(CH₂)₁₄CH₃), 1.24—1.28 (26H, m, -COCH₂(CH₂)₁₃CH₃), 1.89—2.16 (15H, m, -COCH₃×5), 2.36 (2H, t, J=8.5 Hz, -COCH₂(CH₂)₁₃CH₃), 2.59 (1H, dd, J=4.5, 12.5 Hz, 3-H_{eq}). 3.79 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺m/z: 804.

DCC (21 mg, 1.0×10^{-4} mol), 27β (34 mg, 5.0×10^{-5} mol), palmitic acid (26 mg, 1.0×10^{-4} mol), DMAP (2.4 mg, 2.0×10^{-5} mol), thiourea (6 mg, 7.5×10^{-5} mol), and diisopropylethylamine (9.7 mg, 7.5×10^{-5} mol) as described for 5α produced 5β (6 mg, 14.9%) as a colorless, amorphous powder.

(S)-2-O-Benzyl-3-O-hexadecanoyl-1-O-(2,3,4,6-tetra-O-acetyl-D-ga-lactopyranosyl)glycerol (29) 2,3,4,6-Tetra-O-acetyl-D-galactopyranosyl trichloroacetimidate (28, 420 mg, 8.52×10^{-4} mol), synthesized from D-galactose in three steps, the glycerol derivative (12a, 4.47×10^{-4} mol), and MS4A (5 g) were dried by a high vacuum-pump for 2 h. The mixture

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was dissolved in dry CH₂Cl₂ (5 ml) and stirred at $-20\,^{\circ}$ C under argon. BF₃–Et₂O (0.08 ml) was added to the mixture $-20\,^{\circ}$ C, stirred for 3 h, and further stirred at room temperature for 15 h. The resulting mixture was filtered and the filtrate was washed with saturated aqueous NaHCO₃ and brine. The organic phase was dried (MgSO₄) and concentrated. The residue was purified on a column of silica gel (CHCl₃: MeOH = 50:1) to afford **29** (184 mg, 53.9%) as a colorless, amorphous powder. [α]_D +5.0° (c= 0.20, CHCl₃). IR (neat): 1740, 745, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.2 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.98—2.16 (12H, m, -COCH₃ × 4), 5.30 (2H, s, -CH₂Ph), 7.33—7.34 (5H, m, phenyl). Positive FAB-MS (M+H)⁺ m/z: 751.

(S)-3-O-Hexadecanoyl-1-O-(2,3,4,6-tetra-O-acetyl-D-galactopyranosyl)glycerol (6) A solution of 29 (295 mg, 3.93×10^{-4} mol) in MeOH (2 ml) was hydrogenated in the presence of Pd(OH)₂–C (30 mg) at room temperature for 2 h. The catalyst was filtered off and the filtrate was chromatographed on a silica gel column to give 6 (248 mg, 93.6%) as a colorless, amorphous powder. [α]_D +18.3° (c=0.24, CHCl₃). IR (neat): 3470, 1750 cm⁻¹. ¹H-NMR (CDCl₃): 0.88 (3H, t, J=6.2 Hz, –CO(CH₂)₁₄CH₃), 1.25 (26H, s, –COCH₂(CH₂)₁₃CH₃), 1.99—2.17 (12H, m, –COCH₃ × 4), 2.34 (2H, t, J=7.8 Hz, –COCH₂(CH₂)₁₃CH₃). Positive FAB-MS (M+H)⁺ m/z: 661.

6-O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosylonate)-D-galactopyranoside (31) A mixture of 6-O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosylonate)-1,2,3,4-tetra-O-benzyl-galactopyranoside (30, 500 mg, 4.93 × 10^{-4} mol) and Pd(OH)₂-C (50 mg) in MeOH (5 ml) was stirred for 3 h at room temperature under H₂. The mixture was filtered, the filtrate was evaporated to dryness, and the residue was purified on a column of silica gel (CHCl₃: MeOH = 10:1) to yield 31 (322 mg, quant.). mp 110-113 °C, $[\alpha]_D -0.01$ ° (c=0.67, CHCl₃). IR (KBr): 3400, 1745, 1660, 1560 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.88-2.15 (15H, m, $-COCH_3$ ×5), 2.62 (1H, dd, J=4.4, 12.9 Hz, $3-H_{eq}$), 3.81 (3H, s, $-COOCH_3$), 5.64 (1H, d, J=8.5 Hz, $-NH_-$). Anal. Calcd for $C_{26}H_{39}NO_{18}$ · H_2O : C, 46.46; H, 6.15; N, 2.08. Found: C, 46.57; H, 5.90; N, 1.85.

6-*O*-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-*glycero* D-*galacto*-2-nonulopyranosylonate)-1,2,3,4-tetra-*O*-acetyl-D-galactopyranoside (32) Acetic anhydrate (94 mg, 9.21 × 10⁻⁴ mol) was added to a solution of 30 (60 mg, 9.18 × 10⁻⁵ mol) in dry pyridine (145 mg, 1.83 × 10⁻³ mol) at 0 °C under argon and stirred for 15 h at room temperature. The mixture was diluted with CHCl₃ (50 ml) and washed 1 N HCl and brine. The organic phase was dried (MgSO₄) and the residue was purified on a column of silica gel (CHCl₃ : MeOH=20:1) to afford 32 (63 mg, 79.6%). mp 91—94 °C, [α]_D –0.5° (*c*=0.76, CHCl₃). IR (KBr): 3360, 1740, 1660, 1540 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.88—2.20 (27H, m, -COCH₃ × 9), 2.30—2.63 (1H, m, 3-H_{eq}). 3.77 (3H, s, -COOCH₃). *Anal.* Calcd for C₃₄H₄₇NO₂₂: C, 48.63; H, 5.88; N, 1.67. Found: C, 48.68; H, 5.64; N, 1.56.

6-*O*-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-*glycero*-D-*galacto*-2-nonulopyranosylonate)-2,3,4-tri-*O*-acetyl-D-galactopyranoside (33) Hydrazine acetate (128 mg, 1.39×10^{-3} mol) was added to a solution of 32 (257 mg, 3.13×10^{-4} mol) in dry *N*,*N*-dimethylformamide (DMF) (1 ml) at 50 °C. The mixture was diluted with AcOEt (30 ml) and cooled to room temperature. The resulting mixture was washed with brine, dried and chromatographed on a silica gel column (CHCl₃: MeOH = 20:1) to give 33 (162 mg, 65.0%). mp 103—106 °C. [α]_D -0.5° (c=0.42, CHCl₃).

IR (KBr): 3260, 1740, 1660, 1550 cm $^{-1}$. $^{1}\text{H-NMR}$ (CDCl₃) δ : 1.89—2.22 (24H, m, –COCH₃ × 5), 2.55 (1H, dd, J = 4.4, 13.2 Hz, 3-Heq), 3.79 (3H, s, –COOCH₃). Anal. Calcd for C₃₂H₄₅NO₂₁ · 2H₂O: C, 47.11; H, 6.06; N, 1.72. Found: C, 47.37; H, 5.56; N, 1.99.

(S)-2-O-Benzyl-3-O-hexadecanoyl-1-O-[6-O-methyl 5-acetamido-4,7,8, 9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosylonate)-2,3,4-tri-O-acetyl-D-galactopyranosyl]glycerol (34) To a solution of 33 (62 mg, 7.79×10^{-5} mol) in dry (CH₂Cl₂ (5 ml) was added Cl₃CCN $(86 \,\mathrm{mg}, 9.35 \times 10^{-4} \,\mathrm{mol})$ and DBU $(6 \,\mathrm{mg}, 3.9 \times 10^{-5} \,\mathrm{mol})$ at $-5 \,^{\circ}\mathrm{C}$. This mixture was directly chromatographed on a column of silica gel (CH₂Cl₂: AcOEt=1:1) to afford the imidate of 33. A mixture of the imidate, 12a (98 mg, 2.34×10^{-4} mol), and MS4A (600 mg) in dry CH₂Cl₂ (2 ml) was treated with BF₃-Et₂O (11 μ l) at -10 °C. The mixture was stirred for 2 h at -10 °C and for 15 h at room temperature. The resulting mixture was filtered and the filtrate was washed with saturated aqueous NaHCO₃ and brine, dried, and evaporated to dryness. The residue was purified on a column of silica gel (CHCl₃: MeOH = 20:1) to give 34 (13 mg, 14%, 2 steps) as a colorless, amorphous powder. [α]_D -7.4° (c = 0.72, CHCl₃). IR (neat): 1750, 1660, 1550, 700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.5 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃),1.88—2.13 (24H, m, $-COCH_3 \times \overline{8}$), 2.58 (1H, dd, J=4.8, 12.8 \overline{Hz} , 3- H_{eq}), 3.82 (3H, s, -COOCH₃), 7.29—7.37 (5H, m, phenyl). Positive FAB-MS $(M+H)^+$ m/z: 1182.

(S)-3-O-Hexadecanoyl-1-O-[6-O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-D-galacto-2-nonulopyranosylonate)-2,3,4-tri-O-acetyl-D-galactopyranosyl]glycerol (7) A soluton of 34 (12 mg, 1.0×10^{-5} mol) in MeOH (0.5 ml) was hydrogenated with Pd(OH)₂-C (6 mg) and stirred for 1 h at room temperature. The catalyst was filtered off and evaporated to dryness. The residue was purified on PTLC (CHCl₃: MeOH = 20: 1) to give 7 (9 mg, 81%) as a colorless, amorphous powder. [α]_D - 5.2° (c = 0.48, CHCl₃). IR (neat): 3370, 1740, 1660 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.88 (3H, t, J = 6.5 Hz, -CO(CH₂)₁₄CH₃), 1.25 (26H, s, -COCH₂(CH₂)₁₃CH₃), 1.86—2.19 (24H, m, -COCH₃ \times 8), 2.58 (1H, dd, J = 4.5, 12.7 Hz, 3-H_{eq}), 3.81 (3H, s, -COOCH₃). Positive FAB-MS (M+H)⁺ m/z: 1092.

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