Synthesis of the C15–C35 Segment of Chivosazole A

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Abstract: The synthesis of the C15–C35 segment of chivosazole A is reported using a convergent approach that incorporates an *E*-selective Wittig olefination for joining both subunits of this fragment. **Key words:** chivosazole, natural product, antibiotic, myxobacteria

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Chivosazole A (Table 1) belongs to a family of 31-membered macrolide glycosides isolated at the Helmholtz Centre for Infection Research (HZI, formerly GBF) in 1997 from *S. cellulosum* So ce12.¹ They are active against yeasts and filamentous fungi and they are highly cytotoxic against mammalian cell cultures (IC50 9 ng/mL L 929 and HeLa). Except for chivosazole F (7), all natural variants possess a 6-desoxyglucopyranose (chinovose) at C11.

Table 1 Chivosazoles



The stereochemistry of chivosazole A (1) was solved by chemical degradation, NMR studies, and analysis of the polyketide reductase gene cluster.² Finally, an additional confirmation of the configurational assignment of the segment obtained from ozonolysis was obtained by re-

SYNLETT 2007, No. 17, pp 2667–2670 Advanced online publication: 25.09.2007 DOI: 10.1055/s-2007-991049; Art ID: G27407ST © Georg Thieme Verlag Stuttgart · New York synthesizing this segment. In order to independently confirm the proposed stereochemistry of the remaining structural motifs and to access derivatives for structure– activity investigations we initiated a program that aims at the synthesis of chivosazole A (1).



Scheme 1 Retrosynthetic disconnection of chivosazole

We envisioned constructing chivosazole A (1) from the two equally complex fragments 8 and 9 via olefination at carbons C14 and C15 followed by macrolactone formation (Scheme 1). In synthetic direction the northern hemisphere 8 can be constructed from aldehyde 10 and Wittig reagent 11. This segment contains already eight out of the ten stereocenters of the aglycon of chivosazole A (1). The synthesis of 11 commenced with the construction of the C28–C35 segment 12 which was synthesized according to our strategy used for the structural elucidation. An *anti*- Felkin selective Mukaiyama aldol reaction and an *anti*selective reduction of the so-obtained hydroxyl ketone were used as the pivotal transformations for this segment.² Reductive opening of the PMP acetal and subsequent Swern oxidation provided compound **13**. In the subsequent olefination the Ando protocol³ installing the *Z*configured double bond provided higher yields and selectivities compared to the Still–Gennari⁴ reaction (*Z*/*E* = 20:1, 58%). The so-obtained ester **15** was reduced and transformed to the corresponding bromide,⁵ which was subsequently reacted with PBu₃ to generate Wittig salt **16**⁶ (Scheme 2).



Scheme 2 Synthesis of segment 16

The synthesis of the C15–C25 segment 10 began with PMB protection and oxidation of diol 17. For the required enantioselective aldol reaction the Nagao protocol⁷ provided high yields and selectivity (90% ee, 97%). After TBS protection and transformation of the amide to the corresponding aldehyde the *anti*-aldol reaction according to Masamune⁸ gave superior results compared to the Evans or Paterson anti-aldol reactions and provided aldol product 23 with three configurations established. Methylation of the so-generated secondary alcohol was achieved by treatment with MeI and Ag₂O⁹ and for cleaving the chiral auxiliary reduction with LiAlH₄ was mandatory. Re-oxidation to acid 25 was accomplished via a two-step Swern¹⁰ and Pinnick¹¹ oxidation sequence. Finally, the oxazole moiety was introduced under standard conditions.¹² In order to liberate the aldehyde carbonyl group necessary for coupling, the PMB group was removed using DDQ and the alcohol oxidized with MnO₂ to provide segment 27 (Scheme 3).

Even though the Nagao and Masamune protocols are known as reliable and widely applicable methods in aldol chemistry we independently confirmed the relative configuration of compound **25** through analyzing the H–H coupling constants of lactone **28**, obtained by treatment of **25** with HF in pyridine (Scheme 4).¹³



Scheme 3 Synthesis of segment 27



Scheme 4 Configurational assignment of lactone 28

The observed NOE contacts and the coupling constants clearly support the configurations expected from these aldol reactions. The boat conformation on which the configurational analysis was performed was deduced from coupling constants and computational analysis of the lactone. The final coupling of both segments could then be obtained in toluene at 0 °C with KOt-Bu as the base with gratifying yields and the E-configured double bond as the only detectable isomer (Scheme 5).¹⁴ Even though the ¹H NMR spectrum of the C24-C28 segment was of higher order and confirmation of the E-configuration was therefore not accessible through analysis of the coupling constants, a Win-Dyna simulation clearly confirmed the expected *E*-configured double bond.

With a reliable and efficient route for the northern hemisphere in hand we now aim for the construction of the aglycon of chivosazole A (1).



Scheme 5 Coupling of segments 27 and 16 via Wittig reaction

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- (14) Synthesis of Alkene 15 Reagent 14 (14 mg, 44 μ mol) was dissolved in THF (1 mL) and cooled to 0 °C. Then, NaH (2.5 mg, 62 µmol, 60% dispersion on mineral oil) was added, the solution stirred for 15 min at 0 °C, and then cooled to -78 °C. Aldehyde 13 (27 mg, 49 µmol) was added and the mixture was warmed over 2 h to 0 °C and then quenched with sat. NH₄Cl solution (5 mL). The layers were separated and the aqueous layer was extracted with EtOAc $(3 \times 5 \text{ mL})$. The combined organic layers were washed with brine $(1 \times 10 \text{ mL})$ and dried over MgSO₄. The solvent was removed under reduced pressure and the residue was purified via flash chromatography. The reaction yielded 21 mg of the Z-olefin (0.03 mmol, 69%). $R_f = 0.29$ (EtOAc-hexane, 1:30); $[\alpha]_D^{23} + 6.5$ (c 1.07, $CHCl_3$). ¹H NMR (400 MHz, CDCl_3): $\delta = 7.29$ (d, J = 8.5Hz, 2 H), 6.87 (d, J = 8.5 Hz, 2 H), 6.40 (dd, J = 11.6, 10.2 Hz, 1 H), 5.82 (d, J = 11.6 Hz, 1 H), 4.57 (d, J = 10.6 Hz, 1 H), 4.52 (d, J = 10.6 Hz, 1 H), 4.15 (q, J = 7.1 Hz, 2 H), 3.95-3.90 (m, 1 H), 3.88-3.83 (m, 1 H), 3.80 (s, 3 H), 3.23 (dd, J = 5.8, 3.8 Hz, 1 H), 1.77–1.73 (m, 2 H), 1.57 (ddd, J = 14.0, 8.0, 1.9 Hz, 1 H), 1.41 (ddd, J = 13.7, 9.1, 4.1 Hz, 1 H), 1.28 (t, J = 7.2 Hz, 3 H), 1.15 (d, J = 6.2 Hz, 3 H), 1.11 (d, J = 6.8 Hz, 3 H), 0.95 (d, J = 6.8 Hz, 3 H), 0.90 (s, 9 H),0.87 (s, 9 H), 0.10 (s, 3 H), 0.09 (s, 3 H), 0.02 (s, 3 H), -0.01 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃): δ = 195.6, 166.1, 159.1, 156.3, 152.4, 131.4, 129.4, 119.8, 113.8, 83.8, 74.3, 72.2, 66.9, 59.8, 55.4, 43.7, 37.1, 26.2, 26.1, 25.4, 18.2, 17.7, 14.4, 9.5, -3.6, -3.8, -4.3, -4.3. ESI-HRMS: *m/z* calcd for $C_{33}H_{62}O_6NaSi_2$: 645.3983 [M + Na⁺]; found: 645.3975. Synthesis of Lactone 28

Acid 25 (10 mg, 22.1 µmol) was dissolved in THF (0.25 mL) and treated with pyridine (0.25 mL). A HF-pyridine complex (ca. 70% HF, ca. 30% pyridine, 0.4 mL, 15.3 mmol) was added. The reaction mixture was stirred for 16 h at r.t. and then poured into sat. NaHCO₃ solution (10 mL). The layers were separated and the aqueous layer was extracted with MTBE (5×5 mL). The combined organic layers were washed with sat. NH₄Cl solution (5 mL) and brine (5 mL) and dried over MgSO₄. The solvent was removed under reduced pressure and the residue purified via flash chromatography. The reaction yielded 4 mg of lactone **28** (12.5 μ mol, 56%). $R_f = (EtOAc-hexane, 1:4); [\alpha]_D^{23}$ $-14.2 (c 0.30, CHCl_3)$. ¹H NMR (400 MHz, CDCl_3): $\delta = 7.26$ (d, J = 8.5 Hz, 2 H), 6.88 (d, J = 8.7 Hz, 2 H), 5.91 (dt, J = 8.7 Hz, 2 Hz, 2 Hz), 5.91 (dt, J = 8.7 Hz, 2 Hz), 5.91 (dt, J = 8.7 Hz), 5.91 (dt,*J* = 15.5, 5.2 Hz, 1 H), 5.82 (dd, *J* = 15.7, 6.0 Hz, 1 H), 4.75 (ddd, J = 10.6, 5.3, 5.3 Hz, 1 H), 4.46 (s, 2 H), 4.02 (d, J = 5.0 Hz, 2 H), 3.81 (s, 3 H), 3.73 (ddd, J = 6.8, 4.6, 4.6 Hz, 1 H), 3.31 (s, 3 H), 2.75 (dq, J = 6.8, 4.6 Hz, 1 H), 2.25 (ddd, J = 14.5, 6.9, 4.4 Hz, 1 H), 1.87 (ddd, J = 14.6, 11.2, 4.6 Hz, 1 H), 1.28 (d, J = 6.9 Hz, 3 H). ¹³C NMR (100 MHz, CDCl₃): δ = 173.5, 159.4, 130.2, 130.1, 129.6, 129.5, 114.0, 76.2, 75.9, 72.3, 69.4, 56.8, 55.4, 39.6, 34.0, 11.5. ESI-HRMS: m/z calcd for C₁₈H₂₄O₅Na: 343.1521 [M + Na⁺]; found: 343.1533

Synthesis of the C15–C35 Segment (29)

Wittig salt 16 (17 mg, 20 µmol) was dissolved in toluene (1 mL) and cooled to 0 °C. A solution of the aldehyde 27 (10 mg, 24 µmol) in toluene (0.2 mL) was added to the mixture. The reaction mixture was then treated with a KOt-Bu solution (18 µL, 28 µmol, 1 M solution in THF). After 30 min H₂O (2 mL) was added, the layers separated, and the

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aqueous layer was extracted with MTBE $(2 \times 5 \text{ mL})$. The combined organic layers were dried over MgSO₄. The solvent was removed under reduced pressure and the residue was purified via flash chromatography. The reaction yielded 11 mg of compound **29** (15 µmol, 57%). $R_f = 0.32$ (EtOAc-hexane, 1:10); $[\alpha]_D^{23} + 3.9$ (*c* 1.02, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.17$ (s, 1 H), 7.29 (d, J = 8.5 Hz, 2 H), 6.85 (d, J = 8.9 Hz, 2 H), 6.42–6.36 (m, 1 H), 6.18–6.09 (m, 2 H), 6.04 (t, J = 11.1 Hz, 1 H), 5.62 (t, J = 10.4 Hz, 1 H), 5.59 (dd, J = 14.0, 8.2 Hz, 1 H), 5.32–5.54 (m, 2 H), 4.32–4.27 (m, 1 H), 3.97–3.92 (m, 1 H), 3.91 (s, 3 H), 3.84–3.81 (m, 1 H), 3.79 (s, 3 H), 3.78–3.76 (m, 1 H), 3.42

 $(dq, J = 6.8, 5.0 Hz, 1 H), 3.37 (s, 3 H), 3.12 (dd, J = 6.5, 3.8 Hz, 1 H), 2.93-2.84 (m, 1 H), 1.86-1.79 (m, 1 H), 1.43-1.37 (m, 4 H), 1.34 (d, J = 6.8 Hz, 3 H), 1.16 (d, J = 5.8 Hz), 1.06 (d, J = 6.8 Hz, 3 H), 0.95 (d, J = 6.8 Hz, 3 H), 0.92 (s, 9 H), 0.86 (s, 9 H), 0.85 (s, 9 H), 0.10 (s, 6 H), 0.04 (s, 3 H), -0.01 (s, 3 H), -0.02 (s, 6 H). ^{13}C NMR (100 MHz, CDCl_3): \delta = 166.9, 162.0, 159.1, 144.1, 137.2, 134.6, 133.2, 132.6, 131.3, 130.0, 129.5, 128.9, 128.4, 113.8, 84.7, 79.1, 74.6, 71.7, 70.3, 66.7, 57.6, 55.4, 52.3, 43.1, 40.3, 36.2, 32.1, 29.9, 26.2, 26.1, 26.0, 25.6, 22.9, 18.8, 18.2, 14.3, 11.5, 9.2, 7.5, 1.2, -3.3, -3.5, -3.8, -4.1, -4.3, -4.8. ESI-HRMS:$ *m/z*calcd for C₅₂H₉₁NO₉Si₃Na: 980.5899 [M + Na⁺]; found: 980.5900.

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