

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 17 (2007) 4657-4663

Novel tetrahydro-β-carboline-1-carboxylic acids as inhibitors of mitogen activated protein kinase-activated protein kinase 2 (MK-2)

John I. Trujillo,* Marvin J. Meyers,* David R. Anderson, Shridhar Hegde, Matthew W. Mahoney, William F. Vernier, Ingrid P. Buchler, Kun K. Wu, Syaluan Yang, Susan J. Hartmann and David B. Reitz

Department of Medicinal Chemistry, Pfizer Global Research and Development, Chesterfield, MO 63017, USA

Received 16 April 2007; revised 21 May 2007; accepted 22 May 2007 Available online 25 May 2007

Abstract—A structure–activity relationship study was conducted on a series of tetrahydro- β -carboline-1-carboxylic acid analogs in order to identify the key functionality responsible for activity against the mitogen-activated protein kinase-activated protein kinase 2 enzyme (MK-2). The compounds were further evaluated for their ability to inhibit TNF α production in U937 cells and in vivo. These compounds represent a novel structural class of compounds capable of inhibiting MK-2 with remarkable selectivity. © 2007 Elsevier Ltd. All rights reserved.

The proinflammatory cytokine TNF- α was originally described as a product of monocytes that induced tumor lysis¹ and since then has been implicated in several inflammatory diseases in humans.² The development of anti-TNF α biological therapies has been described and been shown to be effective in diseases such as rheumatoid arthritis and psoriasis.³ More recently, many other biological targets have been identified for the inhibition of TNF- α production. The most notable of the targets is the kinase p38 MAPK.^{4,5} Several publications have demonstrated the effect of p38 inhibitors in in vitro and in vivo models, as well as demonstrating efficacy in rheumatoid arthritis (RA) patients in a clinical study.⁶

The mitogen-activated protein kinase-activated protein kinase 2 (MK-2) has only recently received a significant degree of interest for its potential as a target for treatment of RA.⁷ MK-2 is a direct substrate of the p38 α/β kinases and has been linked to TNF- α , IL-6, and IFN- γ expression levels.⁸ A genetic knockout of the MK-2 enzyme in mice has been described^{8a} and shown to be resistant to disease in models of arthritis.^{7b} For example, treatment of

MK-2 knockout mice with LPS resulted in only a small increase of serum TNF α (10–20%) of the wild type control.^{8a} Additionally, the MK-2 knockout mice were observed to be resistant to collagen induced arthritis (CIA),^{7b} a model well established for the study of (RA). Furthermore, while MK-2 knockout mice are healthy and have a normal phenotype, genetic knockout of the p38 gene is embryonic lethal, suggesting an improved safety profile for MK-2 inhibition relative to p38. These observations taken together lend strong support for MK-2 as an attractive target for treatment of TNF- α mediated diseases and rheumatoid arthritis (RA).

Efforts from our laboratories have recently been reported for a series of aminocyanopyridines and a series of pyrrolopyridines as inhibitors of MK-2.⁹ Through our compound screening effort, we also identified commercially available β -carboline derivative **1** as a modest inhibitor of MK-2 (Fig. 1).¹⁰ The compound exhibited



Figure 1. Structure of MK-2 inhibitor and in vitro biological activity.

Keywords: MAPKAP kinase 2; MK-2; TNF- α ; Inflammation; β -Carboline-1; Rheumatoid arthritis.

^{*} Corresponding authors. Tel.: +1 636 244 1937; e-mail: john.i.trujillo@pfizer.com

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2007.05.070

an MK-2 IC₅₀ value¹¹ of 2.5 µM and was identified to be an ATP competitive inhibitor. This structure was of particular interest to us given its low molecular weight and unusually good selectivity versus a number of related kinases¹² (see Fig. 1). In order to elucidate the essential features required for the activity observed and to further improve the potency of compound 1, an investigation of the structure-activity relationship between 1 and the MK-2 enzyme was initiated. It is important to note that no X-ray crystal structure of the MK-2 enzyme existed at the time of this work.¹³ In this report, we describe the synthesis and key structure-activity relationships identified during the investigation of this novel series of tetrahydro-\beta-caroline-1-carboxylic acids as selective inhibitors of MK-2. It is also important to note that SAR for this class of β -carboline carboxylic acids was conducted on the racemates as the α -carbon was observed to be readily epimerizable.

Starting with the A-ring, an investigation of the role of the methoxy group was conducted (see Table 1). The syntheses of these analogs are presented in Schemes 1 and 2.¹⁴ The position of the methoxy group was found to be critical as the 6-position was optimal by more than ten-fold (1, 7, 15, 17). Addition of a second group adjacent to the 6-methoxy group was not tolerated (32, 35), presumably due to twisting of the methoxy group out of the plane. Substitution of the 6-methoxy for other functionalities was subsequently investigated (8-10, 20, 23, 29). Hydroxy analog 20 and methyl ester 29 proved to be well tolerated, while other variations, including deletion, resulted in a 10- to 100-fold loss in potency against MK-2 (8–11, 23, 29). Extension of the methoxy group to benzyloxy was tolerated to some extent (19), but acetophenone derivative 26 was essentially equipotent to methoxy, demonstrating the tolerability for larger groups extending from the A-ring.

Table 1. IC₅₀ values for β -carboline derivatives



Compound	Substitution	MK-2 IC ₅₀ (µM)	
1	6-OMe	2.5	
15	5-OMe	42	
7	7-OMe	>200	
17	8-OMe	>200	
32	6-OMe, 7-OH	53	
35	6-OMe, 5-Me	>200	
20	6-OH	4.5	
29	6-CO ₂ Me	6.6	
23	6-OAc	170	
11	6-H	87	
8	6-F	26	
9	6-Cl	19	
10	6-CONH ₂	28	
19	6-OCH ₂ Ph	12	
26	6-OCH ₂ C(O)Ph	1.1	



Scheme 1. Synthesis of compounds 2–11, and 15, 17. Reagents and conditions: (a) Glyoxylic acid, pH 3–4, H_2O/KOH , rt, 12 h; (b) $Me_2NCH=CHNO_2$, TFA, rt; (c) LAH, THF, rt then reflux.

Having established the importance of the 6-methoxy group in the A-ring for potency, we turned our attention to the C-ring. Oxidation of the tetrahydropyridine to pyridine was investigated but proved to be incompatible with MK-2 activity (Scheme 2; **37** and **38**). The lack of potency of compound **38** in particular suggests a requirement of the acid functionality to exist out of plane with respect to the ring system. The lack of potency may also be a result of a reduction in basicity of the nitrogen in the C-ring (secondary amine to pyridine) (Scheme 3).

Examination of the role of the indole and amino acid functionalities was also conducted (Table 2). The synthesis of compounds **36–63** is shown in Schemes 4 and 5. The presence of the nitrogen was found to be critical for potency as demonstrated by carbon analog **65** and oxygen analog **61**. While the nitrogen itself was required for activity, the hydrogen was not, as demonstrated by N-alkylation with small alkyl groups such as methyl or ethyl (**40** and **49**). Larger groups such as isopropyl and benzyl, however, resulted in significant losses in potency (**50** and **51**), suggesting a sensitive steric environment.

Modification of the carboxylic acid moiety to other functional groups was also investigated. Conversion of the carboxylic acid to a methyl ester (**36**), primary amide (**43**), primary alcohol (**42**) or amine (**56**) all resulted in a dramatic loss in potency. Functional groups capable of mimicking the acid, such as a hydroxamic acid **44** or tetrazole **58**, did not prove to be suitable replacements for the acid. Addition of a methyl group in the α -position (**54**) or incorporation of a methylene spacer between the C-ring and the carboxylic acid (**55**) was not tolerated. The intolerance for any changes to the carboxylic acid suggests a key ionic interaction between the inhibitor and MK-2 is critical for binding affinity in this series. Methylation of the indole NH also proved to be detrimental to activity (**52**), suggesting a potential interaction



Scheme 2. Synthesis of compounds 20, 23, 26, 29, 32, and 35. Reagents and conditions: (a) Glyoxylic acid, pH 3–4, H₂O/KOH, rt, 12 h; rt; (b) 40 psi H₂, 10% Pd/C, MeOH, rt; (c) K₂CO₃ then Boc₂O, THF, DMAP, rt, 2 h; (d) 40 psi H₂, 10% Pd/C, MeOH, rt; (e) Ac₂O, pyridine, CH₂Cl₂, 0 °C to rt, 12 h; (f) 2.0 N HCl in Et₂O, rt, 3 h; (g) Boc₂O, K₂CO₃, H₂O, rt, 14 h; (h) PhC(O)CH₂Br, K₂CO₃, MEK, 80 °C, 14 h; (i) 2.0 N HCl in Et₂O, CH₂Cl₂, rt, 14 h; (j) Me₂NCH=CHNO₂, TFA, rt; (k) NaBH₄, THF:MeOH, rt; (l) 10% Pd/C, NH₄CO₂H, MeOH, reflux, 14, h; (m) 85% H₂NNH₂^{*}H₂O, 75 °C, 14 h; (n) glyoxylic acid, H₂O, pH 4, 80 °C, 30 min.



Scheme 3. Synthesis of compounds 37 and 38. Reagents and conditions: (a) S, m-xylene, reflux, 4 h; (b) LiOH*H₂O, THF:H₂O, rt, 4 h.

of the indole N–H with the enzyme, a limited steric environment, or distortion of the orientation of the carboxylic acid.

In order to further probe the tolerance of MK-2 to substitution of the β -carboline C-ring, analogs were prepared to probe the 3- and 4-positions (Table 3 and Scheme 6). The enzyme exhibited tolerance of only small substituents such as methyl and ethyl in the 4-position (**70–72**), but less tolerant of alkyl substituents, even ethyl, in the 3-position (**88**, **89**). Further exploration of the 3-position demonstrated tolerance for only a select group of small functional groups capable of H-bonding (**76**, **82**, **83**) (Table 4).

Having established the tolerance of 2- and 4-positions in the C-ring for alkylation with small groups such as methyl or ethyl, a series of C-ring bridged analogs were prepared (see Table 5 and Scheme 7). While the threecarbon bridge between N-2 and C-4 was tolerated



Scheme 4. Synthesis of compounds 36, 39–44, 49–52, 54–56, and 58. Reagents and conditions: (a) MeOH/HCl, rt; (b) Na₂CO₃, Mel, THF:H₂O, rt; (c) KOH(aq), THF:MeOH; (d) for 46: AcCl, Et₃N, CH₂Cl₂, rt; for 47: acetone, MeOH, rt; for 48: PhCHO, MeOH, reflux₄, MeOH; (e) for 446: LAH, THF, rt; for 47 and 48: NaBH₄, MeOH, 0 °C; (f) for 49: glyoxylic acid, TFA, CF₃CN, rt; for 51 and 52: glyoxylic acid, H₂O, rt; (g) acetone, Na(OAc)₃BH, CH₂Cl₂, rt; (h) NaOH, EtOH; (i) Boc₂O, THF, rt; (j) NaH, Mel, DMF, rt; (k) TFA, CH₂Cl₂, anisole, rt; (l) glyoxylic acid, H₂O, rt; (m) LAH, Et₂O, 0 °C to rt; (n) conc. NH₄OH(aq); (o) LAH, THF, rt; (p) for 53: (MeO)₂CH₂CO₂Me, CH₃CN, TFA, 80 °C; for 54: Me(C=O)CO₂H, MeOH:H₂O, 50 °C; (q) LiOH, THF:H₂O, rt; (r) 50% NH₂OH(aq), THF:H₂O, K₂CO₃, rt; (s) TFAA, Et₃CN, CH₂Cl₂, 0 °C; (t) Me₃SnN₃, dioxane, reflux, 5 h; (u) K₂CO₃, MeOH/H₂O, rt. See above mentioned references for further information.

(92), the two-carbon bridge resulted in a 6-fold enhancement in potency, representing the first β -carboline we identified to achieve submicromolar potency against

MK-2. Furthermore, compound **92** exhibited remarkable selectivity against a number of structurally related kinases (IC₅₀, μ M: MSK1 > 200, MNK1 > 200,



Scheme 5. Synthesis of compounds 61 and 65. Reagents and conditions: (a) HCOCO₂Et, pTsOH, rt to 80 °C, 20 h; (b) MeOH:H₂O, KOH, rt, 2 h; (c) 63, EtOH, reflux, 24 h; (d) LiOH, THF:H₂O:MeOH, 4 h.



Scheme 6. Synthesis of compounds 70–72, 76, 77, 82, 83, 88, and 89. Reagents and conditions: (a) for 67: MeMgBr (3.0 M in Et₂O), THF, $-50 \degree$ C, 2 h then 0 °C, 1 h; for 68: EtMgBr, THF, $-78 \degree$ C, 3 h; for 69: PhCH₂MgCl (2.0 M in Et₂O), THF, $-50 \degree$ C, 2 h then 0 °C, 1 h; (b) 10% Pd/C, HCO₂NH₄, MeOH, 80 °C; (c) glyoxylic acid, H₂O, pH 4–5, rt, 3 h; (d) for 74: BrCH₂C(=NOH)CO₂Et, Na₂CO₃, CH₂Cl₂, rt, 24 h; for 75: BrCH₂C(–NOH)CF₃, Na₂CO₃, 'BuOMe, rt, 24 h; (e) LAH, THF, $-50 \degree$ C to rt; (f) glyoxylic acid, H₂O, pH 4–5, rt, 24 h; (g) NaOMe, Me₂SO₄, MeNO₂, MeOH, rt, 5 h; (h) for 79: Bn₂NH, H₂CO, H₂O:MeOH, rt then reflux, 3 h; for 80: Me₂NH, H₂CO, H₂O, MeOH, rt; (i) for 81: LAH, THF, 0 °C then reflux; for 83: HCO₂NH₄, 10% Pd/C, MeOH, reflux; (j) glyoxylic acid, MeOH/H₂O, pH 3–4, 55 °C; (k) HCO₂NH₄, 10% Pd/C, MeOH, reflux; (l) for 84: nPrNO₂, NaOMe, Me₂SO₄, MeOH, rt; for 86: EtNO₂, NaOMe, Me₂SO₄, MeOH, rt.



Scheme 7. Synthesis of compounds 92 and 93 and 94–98. Reagents and conditions: (a) For 92: glyoxylic acid, MeOH:H₂O, 60 °C, 10 h; for 93: same as 92 except 45 h; (b) ROH, EDAC, DMAP, rt, 5 h.

JNK2 > 200, p38 α > 100, IKK2 > 200), even close homologs MK-3 and PRAK (IC₅₀, μ M: 90 and 24, respectively).

The potency and selectivity of compound 92 prompted further investigation in cellular models, specifically the inhibition of TNF α production in U937 cells.





Compound	\mathbf{R}^1	\mathbf{R}^2	Х	R^3	MK-2 IC ₅₀ (µM)
1	CO_2H	Н	NH	Н	2.5
65	CO_2H	Н	CH_2	Н	>200
61	CO_2H	Н	0	Н	>200
40	CO_2H	Н	NMe	Н	1.0
49	CO_2H	Н	NEt	Н	5.8
50	CO_2H	Η	N ^{<i>i</i>} Pr	Η	170
51	CO_2H	Н	NBn	Н	130
36	CO ₂ Me	Н	NH	Н	47
43	CONH ₂	Η	NH	Η	>200
42	CH ₂ OH	Н	NH	Н	>200
56	CH_2NH_2	Η	NH	Η	>200
44	CONHOH	Н	NH	Н	88
58	Tetrazole	Н	Н	Н	130
54	CO_2H	Me	NH	Η	>200
55	CH ₂ CO ₂ H	Н	NH	Н	>200
52	CO_2H	Η	NH	Me	>200

Table 3. IC₅₀ values for β -carboline derivatives



Compound	\mathbb{R}^1	\mathbb{R}^2	MK-2 IC ₅₀ (µM)
70	Me	Н	2.2
71	Et	Н	2.2
72	Bn	Н	16
88	Н	Me	7.1
89	Н	Et	64
77	Н	CF_3	>200
76	Н	CH ₂ OH	3.0
82	Н	CH_2NH_2	5.9
83	Н	CH ₂ NMe ₂	>200

Unfortunately, compound **92** did not possess the desired cellular activity. It was thought that the zwitterionic character of the amino and carboxylic acid moieties was limiting the cellular permeability of this compound.

Table 4. IC₅₀ values for β -carboline derivatives 92 and 93



Compound	п	MK-2 IC ₅₀ (µM)	MK-3 IC ₅₀ (µM)	PRAK IC ₅₀ (µM)
92	1	0.29	90	24
93	2	5.1	160	25

Table 5. IC₅₀ values for β -carboline derivatives 92–97



Compound	R	MK-2 IC ₅₀ (µM)	U937 IC ₅₀ (µM)
92	Н	0.29	80.8
94	Et	29.3	5.6
95	Bn	7.09	1.7
96	<i>n</i> -Pr	10.1	0.83
97	H ₂ C=CHCH ₂	3.27	14.4
98	PhCH(Me)	13.0	10.3

Indeed, when ester prodrugs of **92** were synthesized and evaluated in the U937 cell assay, a dramatic enhancement in cell potency was observed, suggesting an intracellular conversion of the ester to the active acid. The most effective compound in the U937 cellular assay was identified to be compound **96**, an *n*-propyl ester (see Table 5). The compound was further evaluated in the rat LPS model (40 mpk, IP) and was shown to inhibit TNF- α production (84%, compound administered 1 h prior to LPS stimulation).

In conclusion, a group of β -carbolines capable of inhibiting MK-2 has been discovered, and the SAR of the series has been described. Key SAR observations include (1) the importance of an ether functionality at the 6-position of the A-ring, (2) the critical sensitivity of the carboxylic acid, and (3) the enhancement in binding affinity observed by bridging the C-ring nitrogen with the 4-position. Compound **92** represents remarkable selectivity for MK-2 over a number of closely related kinases, including MK-3 and PRAK. While the carboxylic acids were inactive in U937 cells, an ester prodrug of **92**, compound **96**, was found to be effective in modulation of TNF α production in both U937 cellular assays and in the rat LPS model.

Acknowledgments

We thank Robert P. Compton, Jeffrey L. Hirsch, Heidi M. Morgan, Matt J. Saabye, and John F. Schindler for enzyme support; Heidi R. Hope, Shelia C. Short, and Jian Zhang for cellular assays; and Gary D. Anderson, Stephen J. Mnich, Mark A. Thiede, and Elizabeth G. Webb for in vivo model support.

References and notes

- 1. Huizinga, T. W. J.; Breedveld, F. C. Rheum. Arthritis 2000, 501.
- 2. Camussi, G.; Lupia, E. Drugs 1998, 55, 613.
- (a) Richard-Miceli, C.; Dougados, M. *BioDrugs* 2001, 15, 251;
 (b) Braun, J.; Sieper, J. *BioDrugs* 2003, 17, 187;
 (c) Olsen, N. J.; Stein, C. M. N. Eng. J. Med. 2004, 350, 2167.
- Chen, Z.; Gibson, T. B.; Robinson, F.; Silvestro, L.; Pearson, G.; Xu, B.; Wright, A.; Vanderbilt, C.; Cobb, M. *Chem. Rev.* 2001, 101, 2449.

- 5. Pargellis, C.; Regan, J. Curr. Opin. Invest. Drugs 2003, 4, 566.
- 6. (a) Lee, M. R.; Dominguez, C. Curr. Med. Chem. 2005, 12, 2979; (b) Haddad, J. Curr. Opin. Invest. Drugs 2001, 2, 1070, and references cited therein.
- (a) Gaestel, M. Nat. Rev. Mol. Cell Biol. 2006, 7, 120; (b) Hegen, M.; Gaestel, M.; Nickerson-Nutter, C. L.; Lin, L.-L.; Telliez, J.-B. J. Immunol. 2006, 177, 1913.
- (a) Kotlyarov, A.; Neininger, A.; Schubert, C.; Eckert, R.; Birchmeier, C.; Volk, H.; Gaestel, M. *Nat. Cell Biol.* **1999**, *1*, 94; (b) Neininger, A.; Kontoyiannis, D.; Kotylarov, A.; Winzen, R.; Eckert, R.; Volk, H. D.; Holtmann, H.; Kollias, G.; Gaestel, M. *J. Biol. Chem.* **2002**, *277*, 3065.
- (a) Anderson, D. R.; Hegde, S.; Reinhard, E.; Gomez, L.; Vernier, W. F.; Lee, L. F.; Liu, S.; Sambandam, A.; Snider, P.; Masih, L. *Bioorg. Med. Chem. Lett.* 2005, *15*, 1587; (b) Anderson, D. R.; Meyers, M. J.; Vernier, W. F.; Mahoney, M. W.; Kurumbail, R. G.; Caspers, N.; Poda, G. I.; Schindler, J. F.; Reitz, D. B.; Mourey, R. J. *J. Med. Chem.* 2007, *50*, 2647.
- Meyers, M. J.; Trujillo, J. I.; Vernier, W. F.; Anderson, D. R.; Reitz, D. B.; Buchler, I. P.; Hegde, S. G.; Mahoney, M. W.; Wu, K. K. WO 2005009370 A2, 2005.
- 11. For description of MK-2 IC50 assay determinations and modulation of TNF α production in both U937 cellular assays and in the rat LPS model, see Ref. 9a.
- 12. For description of kinase selectivity assays, see Ref. 9b.

- For a report on a MAPKAP kinase 2 structure, see: Kurumbail, R. G.; Pawlitz, J. P.; Stegeman, R. A.; Stallings, W. C.; Shieh, H. S.; Mourey, R. M.; Bolten, S. L.; Broadus, R. M., WO2003076333 A2, 2003 and Ref. 9b.
- For a review of the Pictet–Spengler reaction, see: Cox, Eric D.; Cook, James M. Chem. Rev. 1995, 95, 1797.
- Rinehart, K. L.; Kobayashi, J.; Harbour, G. C.; Gilmore, J.; Mascal, M.; Holt, T. G.; Shield, L. S.; Lafargue, F. *J. Am. Chem. Soc.* **1987**, *109*, 3378.
- T, Kurihara; Y, Sakamoto; T, Kimura; H, Ohishi; S, Harusawa; R, Yoneda; T, Suzutani; M, Azuma *Chem. Pharm. Bull.* **1996**, *44*, 900.
- 17. Wilkening, D.; Mundy, B. P. Synthetic Commun. 1984, 14, 227.
- 18. Julia, M.; Lenzi, J. Bull. Soc. Chim. Fr. 1962, 2262.
- 19. Gilchrist, T. L.; Roberts, T. G. J. Chem. Soc., Perkin Trans. 1 1983, 1283.
- (a) Bergman, J. *Tetrahedron* **1971**, *27*, 1167; (b) Hermkens,
 P. H. H.; van Maarseveen, J. H.; Cobben, P. L. H. M.;
 Ottenheijm, H. C. J.; Kruse, C. G.; Scheeren, H. W. *Tetrahedron* **1990**, *46*, 833.
- 21. Macor, J. E.; Blank, D. H.; Ryan, K.; Post, R. J. Synthesis 1997, 443.
- (a) Gremmen, C.; Burm, B. E. A.; Wanner, M. J.; Koomen, G.-J. *Tetrahedron Lett.* **1998**, *39*, 1441; (b) Gharagozloo, P.; Miyauchi, M.; Birdsall, N. J. M. *Tetrahedron* **1996**, *52*, 10185.