Preliminary communication

Alkyl sulfenyl triflate as activator in the thioglycoside-mediated formation of β -glycosidic linkages during oligosaccharide synthesis^{*}

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Thioglycosides have been used in glycosidation reactions and as donors in oligosaccharide synthesis¹⁻¹²: (a) by conversion into glycosyl halides which can be isolated or used *in situ* for reaction, in the presence of a suitable halophilic promoter, with a hydroxylic acceptor; and (b) directly, in the presence of a thiophilic promoter. A participating 2-acyl substituent results in the formation of 1,2-*trans*-glycosides, and a non-participating 2-substituent promotes the formation of 1,2-*cis*-glycosides. In method (b), methyl trifluoromethanesulfonate (MT) and dimethyl(methylthio)-sulfonium trifluoromethanesulfone (DMTST) have emerged as powerful promoters. However, the former is a potential carcinogen, and although methylating agents other than MT may be used in the preparation of DMTST, which otherwise is a crystalline, easily handled compound, they are also carcinogenic.

In searching for alternative promoters more suitable for large-scale synthesis and routine handling, we have considered nitrosonium or alkyl sulfenyl cations. The NO⁺ ion is a border-line acid according to Pearson's description of hard and soft Lewis acids¹³ and AlkS⁺ is a soft acid. The use of NOBF₄ has been reported¹⁴, but, in our hands, NOBF₄ and N₂O₄ did not give consistent yields especially with acceptors that were comparatively unreactive (*e.g.*, the HO-4 hexopyranosides)¹⁵. The reaction of sulfenate esters as glycosyl acceptors with benzylated methyl or phenyl 1-thio- β -D-glucopyranoside carried out in the presence of Lewis acids (CF₃SO₃SiMe₃, TrBF₄, or BF₃·Et₂O) at -35° generally afforded $\alpha\beta$ -mixtures¹⁶. We now report on the use of alkyl sulfenyl halides (triflates) as efficient activators of thioglycosides at room temperature during 1,2-*trans* glycosidation.

The results (Table I) show that alkyl sulfenyl cations, in general, can activate thioglycosides. Thus, methylsulfenyl bromide (MSB) and *o*-nitrobenzenesulfenyl chloride (NSC) were used for the activation of 1, 2, and 4. Both MSB (method A) and NSC (method B) promoted the coupling between 1, 2, or 4 with 9, in the presence of silver triflate (ST), to afford methyl 2-O-benzyl-4,6-O-benzylidene-3-

^{*}Use of Sulfenyl Halides in Carbohydrate Reactions, Part I.

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Entry	D/Aª	Product ^d	Yield (%) ^c /Time (min)			$[\alpha]_{D}^{25f}$ (c)	M.p. (degrees)	N.m.r. data for C-1 and
			Method A	Method B	Method C			C-1' (p.p.m.)
1	1/9	12 ^b	96/45	60-65/70	67/45	-24° (1.0)	179–180	98.8, 100.9
2	2/9	13 ^c	94/40	85/90	79/70	-20° (1.65)	foam	98.86, 101.1
3	4/9	14	95/45	64/120	66/45	$+12^{\circ}$ (1.1) ^g	188–189 ^g	98.68, 98.68
4	2/7	15 ^c -	86/45			-2.5° (1.4)	syrup	102.7, 101.3
5	2/8	16 ^{b, c}	97/45			+18° (1.6)	foam	100.3, 102.3
6	6/10	17 ⁶	77 ^{h,i} /<120			-3° (1.65)	foam	98.5, 100.4
7	2/11	18 ^c	82 ^{h,i} /40			-20° (1.4)	foam	91.4, 101.4
8	3/9	19 ¢	97/40			+3.5° (1.5)	foam	98.99, 101.1
9	5 ^{b.j} /9	20 ^b	88 ^k /45			-11.5° (1.4)	219–220	98.64, 100.9

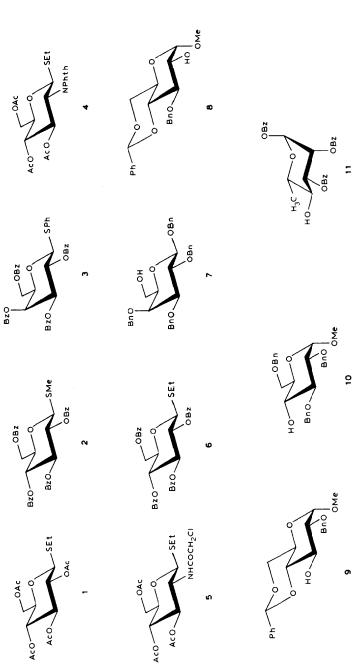
TABLE I

YIELDS AND SELECTED PHYSICAL PROPERTIES OF β -D-LINKED DISACCHARIDES

^aD, thioglycoside; A, acceptor. ^bGave correct elemental analysis. ^cGave correct molecular ion $(M + 1^+)$ and/or $(M + 18^+)$ in f.a.b.-m.s. ^dSmall quantities of D and A could be isolated. ^eBased on the acceptor. ^fMeasured in chloroform. ^gMatched the reported¹⁸ value. ^hBased on the acceptor used. ^fBetter yield on addition of fresh ST, donor, and MSB in quick succession. ^jMade from the corresponding β -D-tetraacetate precursor, using a known method¹⁹, m.p. 197–198°, $[\alpha]_D^{25} - 24^\circ$ (c 1.7, chloroform). ^kSame result on carrying out the reaction at 0°.

O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (12), methyl 2-O-benzyl-4,6-O-benzylidene-3-O-(2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyl)- α -D-glucopyranoside (13), and methyl 2-O-benzyl-4,6-O-benzylidene-3-O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)- α -D-glucopyranoside (14), respectively. The yields were poorer with NSC probably due to its highly hygroscopic nature and lower reactivity. The reactive species in these reactions are methyl sulfenyl triflate (MST) and o-nitrobenzenesulfenyl triflate (NST) since no reaction took place in the absence of ST. Reactions were also carried out after the prior preparation of the triflate (MST, method C), to give 12–14, but 6–10 mol of the reagent were necessary. Dichloromethane was a better solvent than acetonitrile in these reactions.

Comparison of methods A-C (Table I, entries 1-3) indicated that method A was superior in promoting the formation of 1,2-*trans* linkages. Using method A, the reactions between 2 and 7, 2 and 8, 6 and 10, 2 and 11, 3 and 9, and 5 and 9 (ratio of thioglycoside to alcohol, 1.2:1) afforded benzyl 2,3,4-tri-O-benzyl-6-O-(2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl)- β -D-galactopyranoside (15), methyl



3-O-benzyl-4,6-O-benzylidene-2-O-(2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl)- α -D-glucopyranoside (16), methyl 2,3,6-tri-O-benzyl-4-O-(2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl)- α -D-glucopyranoside (17), 1,2,3-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl)- α -L-rhamnopyranose (18), methyl 2-O-benzyl-4,6-O-benzylidene-3-O-(2,3,4,6-tetra-O-benzoyl- β -D-galactopyranosyl)- α -Dglucopyranoside (19), and methyl 2-O-benzyl-4,6-O-benzylidene-3-O-(3,4,6-tri-Oacetyl-2-chloroacetamido-2-deoxy- β -D-glucopyranosyl)- α -D-glucopyranoside (20), respectively (Table I, entries 4–9). Each reaction (except entry 6) was complete within 30–60 min (t.1.c.) and gave β -linked products. The acetylated thioglycoside 1 gave more side products and poorer yields than the benzoylated donors 2, 3, and 6, which corroborates previous observations¹⁷. Although the chloroacetamido donor 5 showed a clean reaction (t.1.c.), a larger proportion was needed than for the corresponding phthalimido donor 4 in order to afford comparable yields of 14 and 20.

MSB was prepared by mixing equimolar quantities of dimethyl disulfide and bromine in 1,2-dichloroethane at room temperature and stirring the solution in the dark under nitrogen for 2–4 h. This solution can be preserved effectively for almost a week. Calculated volumes were used in the coupling reactions.

MST was prepared by injecting MSB (1 equiv.) through a septum into a flask which was protected from light and contained silver triflate (1.05 equiv.) dispersed in dry 1,2-dichloroethane. The solution was stirred for 15–30 min, the precipitate was allowed to settle, and a calculated volume of the supernatant solution was used directly in method C.

Method A: a solution of the donor 1 (80 mg, 0.2 mmol) and the acceptor 9 (63 mg, 0.17 mmol) in dichloromethane (3 mL) was stirred (40 min) in the presence of powdered molecular sieve (4 Å, 0.5 g). ST (71 mg, 0.28 mmol) was added and a septum was attached. MSB (0.24 mL = 30.5 mg, 0.24 mmol) was injected in two equal portions at an interval of 30 min. All operations were carried out under a positive pressure of nitrogen. T.l.c. (15:1 toluene–acetone) after 45 min indicated major conversion into one product. The mixture was neutralized with triethylamine and filtered through Celite, and the flask and Celite were washed with dichloromethane (5 mL). The combined filtrate and the washings were washed with aqueous NaHCO₃ and water, dried (MgSO₄), and concentrated *in vacuo*, and the residue was subjected to chromatography on silica gel.

Method B: the reaction between 1 and 9 was carried out essentially as in method A, but using NSC instead of MSB.

Method C: freshly prepared MST (6–10 molar excess) was injected slowly (20–30 min) through a septum into a flask containing **1**, **9**, ST (same proportion as in method A), and molecular sieve (4 Å) in dichloromethane (kept under N_2). The mixture was stirred for 30 min and then worked-up as in method A.

After completion of this work, benzeneselenyl triflate was described as a promoter for glycosidations using thioglycosides²⁰.

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