Selective Synthesis of ent-15-epi-F_{2t}-Isoprostane and a Deuterated Derivative

Manami Shizuka, Marc L. Snapper*

Department of Chemistry, Eugene F. Merkert Chemistry Center, Boston College, 2609 Beacon Street, Chestnut Hill, MA 02467, USA Fax +1(617)5521442; E-mail: marc.snapper@bc.edu

Received 8 March 2007

Dedicated to Professor Paul A. Wender on the occasion of his 60th birthday

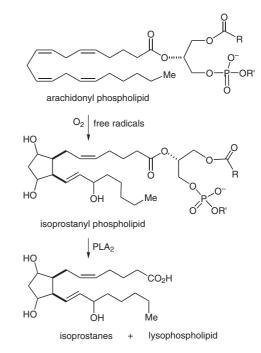
Abstract: Isoprostanes are an emerging class of lipid metabolites whose physiological properties are not well understood. The selective synthesis of *ent*-15-*epi*- F_{2t} -isoprostane, an isomer active in a preliminary screening assay is described. The synthesis features a regioselective cross-metathesis on an enantiomerically enriched divinyl cyclopentyl intermediate to selectively differentiate the sidechains of the target. The route provides the isoprostane, as well as a d_4 -labeled analogue, in 14 steps from readily available starting materials.

Key words: asymmetric synthesis, metathesis, olefination, lipids

Isoprostanes are produced upon non-enzymatic, free radiperoxidation of phosphatidyl arachidoniates cal (Scheme 1).¹ In addition to possible activities within the lipid bilayer, hydrolytic release of these soluble lipid oxidation products may also lead to a variety of physiological responses. The biological activities of the isoprostanes that have been identified to date typically involve inflammatory responses, as well as activities related to smooth muscle growth factors,² and platelet aggregation factors.³ These lipid metabolites are also recognized as important indicators of oxidative stress,4 particularly in human maladies such as Alzheimer's disease,⁵ diabetes,⁶ and cancer.⁷ While progress has been made in understanding the formation and distribution of the isoprostanes, significant questions remain regarding the physiological roles for these endogenous metabolites.

Total synthesis has begun to provide useful quantities of isoprostanes and derivatives required for their further study.⁸ In this regard, we disclosed the preparation of all eight 15-F₂-isoprostane isomers via a stereodivergent strategy (Figure 1).⁹ The key intermediate in the synthesis was obtained from a [2+2] photocycloaddition between a functionalized cyclopentenone and acetylene, followed by a ring-opening cross-metathesis.¹⁰ Subsequent resolution of the racemic diastereomers by an asymmetric CBS-reduction provided the individual isoprostanes isomers in an enantiomerically enriched manner. This sequence offered simultaneous access to the complete library of 15-F₂-isoprostanes from a common synthetic precursor **1**.

Preliminary screening of the eight diastereomeric 15- F_2 isoprostanes in a whole blood platelet aggregation inhibition assay indicated that the unknown *ent*-15-*epi*- F_2 -iso-



Scheme 1 Formation of the 15-F₂-isoprostanes

prostane was more active than the known inhibitor, 15- F_{2t} -isoprostane.¹¹ To further study this particular isoprostane isomer, we modified our synthetic strategy toward the 15- F_2 -isoprostanes to provide access to any specific isoprostane isomer. Herein, we describe how a regioselective olefin cross-metathesis of an enantiomerically enriched divinyl intermediate provides *ent*-15-*epi*- F_{2t} isoprostane and a mass-labeled derivative in an enantioselective manner.

Our synthesis of *ent*-15-*epi*- F_{2t} -isoprostane begins with enone **4**, generated efficiently and selectively from an enzymatic desymmetrization of *meso*-diol **2** (with pancreat-in lipase).^{12,13} Enone **4** can be accessed by functional group manipulations from **3** in 80% overall yield (Scheme 2).¹⁴

A photochemical [2+2] cycloaddition between enone 4 and acetylene generates a mixture of bicyclo[3.2.0]heptenones 5 and 6 (5:6 = 2:1, Scheme 3). DIBAL-H reduction of the mixture yields isomers 7, 8, 9, and 10, which are separable by silica gel chromatography. The reduction favors the desired hydroxyl stereochemistry by an 84:16 ratio. The major product 7 could be isolated and the remaining fractions could be oxidized with PCC back to a mixture of 5 and 6. Preliminary results show that the un-

SYNTHESIS 2007, No. 15, pp 2397–2403 Advanced online publication: 12.07.2007 DOI: 10.1055/s-2007-983768; Art ID: C01807SS © Georg Thieme Verlag Stuttgart · New York

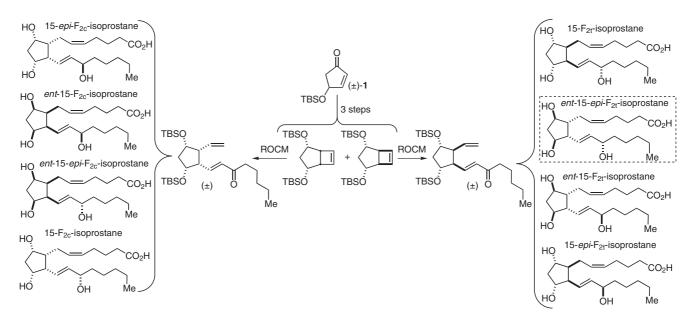
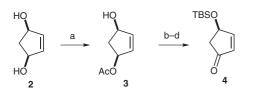


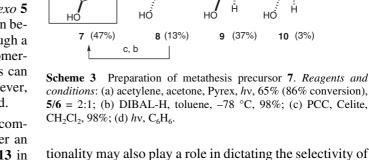
Figure 1 Stereodivergent synthesis of the 15-F₂-isoprostanes



Scheme 2 Preparation of optically enriched enone 4. *Reagents and conditions*: (a) pancreatin lipase, vinyl acetate, THF, >99% ee, 55% yield after recryst.; (b) TBSCl, Et_3N , DMAP, CH_2Cl_2 , 99%; (c) K₂CO₃, MeOH, 90%; (d) PCC, Celite, CH_2Cl_2 , 90%.

desired *syn-endo* photoadduct **6** can be photoisomerized using a Vycor filter to a 1:1 ratio of the desired *syn-exo* **5** isomer and the starting *syn-endo* **6**. This isomerization between the two diastereomers is likely occurring through a 1,3-acyl migration.¹⁵ Given the reoxidation and isomerization pathways, in principle, all undesired isomers can be used to increase the overall yield of adduct **7**; however, in practice only cyclobutene **8** was routinely recycled.

Ring-opening cross-metathesis (ROCM) of syn-exo compound 7 in benzene using Grubbs' catalyst 11 under an ethylene atmosphere generates the divinyl species 13 in 87% yield (Scheme 4).^{10,16} High dilution is required to avoid ring-opening metathesis polymerization (ROMP). In contrast, cross-metathesis (CM) of divinyl compound 13 with oct-1-en-3-one finds optimal conditions under higher concentration in CH₂Cl₂ at 40 °C with 5 mol% Hoveyda-Grubbs catalyst 12. This leads to the desired mono-CM product in high selectivity; none of the other regioisomeric cross-metathesis product is observed and only trace amount of the double cross-metathesis product is obtained.¹⁷ The newly formed enone in the desired product is formed exclusively with *E*-stereoselectivity. The high regioselectivity of the cross-metathesis could arise through ruthenium coordination by the free hydroxyl group on the cyclopentyl ring, although steric hindrance of the silyl protecting group on the other hydroxyl func-



TBSC

TBSQ

tionality may also play a role in dictating the selectivity of the reaction.

d

5

TBSO

TBSC

С

Н

TBSO

н

b

TBSC

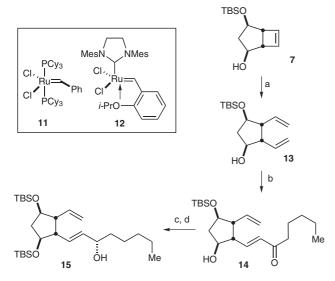
а

TBSO

When cyclobutene 7 was subjected to a direct ROCM with oct-1-en-3-one using Hoveyda–Grubbs catalyst 12, the desired compound 14 was not obtained; mainly ROMP of the cyclobutene was observed. Furthermore, Grubbs' catalyst 11 was unreactive in a ROCM with oct-1-en-3-one; only starting materials were recovered. Since the two metatheses require different reaction conditions, it was necessary to run these transformations in a stepwise fashion.

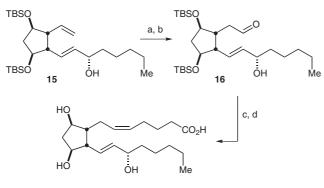
Silyl protection of the free hydroxyl group, followed by an asymmetric catalytic reduction of enone **14** with (*R*)-2-methyl-CBS-oxazaborolidine¹⁸ and catecholborane led to alcohol **15** in high yield and diastereometric excess (>95:5

Downloaded by: University of Arizona Library. Copyrighted material



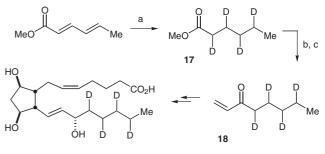
Scheme 4 Installation of the isoprostanyl side-chains through a regio- and stereoselective cross-metathesis. *Reagents and conditions*: (a) ethylene, 5 mol% 11, benzene (0.01 M), 87%; (b) oct-1-en-3-one, 5 mol% 12, CH_2Cl_2 (0.2 M), 40 °C, 70–90%; (c) TBSCl, Et_3N , DMAP, CH_2Cl_2 , 99%; (d) (*R*)-2-methyl-CBS-oxazaborolidine, catecholborane, toluene, -78 °C; 99%.

dr) (Scheme 4).¹⁹ Hydroboration of the terminal olefin with 9-BBN followed by an oxidative workup yields the primary alcohol, which was oxidized with catalytic TEMPO/NCS to generate aldehyde **16** (Scheme 5).²⁰ Wittig olefination forms the *Z*-olefin exclusively to complete the overall isoprostane structure. Removal of the two silyl protecting groups with TBAF delivered the desired *ent*-15-*epi*-F_{2t}-isoprostane in nearly quantitative yield. Of particular note, the efficiency of the synthetic route allows for sufficient quantity of *ent*-15-*epi*-F_{2t}-isoprostane to be prepared for further biological and chemical studies (i.e., > 0.1 g).



ent-15-epi-F_{2t}-isoprostane

In addition, this route also allowed for the preparation of a tetradeuterated analogue of *ent*-15-*epi*- F_{2t} -isoprostane.²¹ Exhaustive reduction of methyl sorbate under D₂ gave **17** in 60% yield (Scheme 6). Weinreb amide²² formation, fol-



d₄-ent-15-epi-F_{2t}-isoprostane

Scheme 6 Synthesis of cross-metathesis partner 18 for the internal standard d_4 -ent-15-epi-F_{2t}-isoprostane. *Reagents and conditions*: (a) D₂ (1 atm), Pd/C, MeOH, 60%; (b) MeNH(OMe)·HCl, *i*-PrMgCl, THF, -10 °C, 88%; (c) vinylmagnesium bromide, THF, 0 °C; 90%.

lowed by vinyl group addition provided tetradeuteroenone **18**. Enone **18** was then used for the cross-metathesis partner with **13** as described in Scheme 4. The rest of the synthesis was carried out in an analogous manner to that shown in Schemes 4 and 5 to yield d_4 -ent-15-epi-F_{2t}isoprostane.

 d_4 -ent-15-epi-F_{2t}-Isoprostane was used as an internal standard in a GC/MS assay to detect isoprostanes in human urine.²³ Four diastereomeric 15-F₂-isoprostanes were separated on an achiral GC column.²⁴ Although the known 15-F_{2t}-isoprostane was not detected in this assay, preliminary results revealed that the ent-15-epi-F_{2t}-isoprostane (or its enantiomer) was present in the urine sample.

In summary, we have achieved a stereoselective synthesis of *ent*-15-*epi*-F_{2t}-isoprostane through a regio- and stereoselective ring-opening cross-metathesis/cross-metathesis sequence. Enzymatic acylation generates the starting enone 4 as a single enantiomer. Photocycloaddition with acetylene followed by functional group manipulations provides an enantiomerically enriched precursor for the key metatheses. A catalyst-controlled asymmetric reduction completes one of the 15-F₂-isoprostanyl side-chains and allows for the installation of the second side-chain. The overall sequence provides efficient access to ent-15epi-F_{2t}-isoprostane, as well as an isotopically labeled ent-15-epi-F_{2t}-isoprostane to be used for in vivo studies. Moreover, it is expected that the synthetic strategy can be readily modified to selectively access any of the 15-series isoprostanes. Further biological and chemical studies of *ent*-15-*epi*-F_{2t}-isoprostane are underway.

Starting materials and reagents purchased from commercial suppliers were used without further purification with the following exceptions: CH_2Cl_2 , THF, benzene, Et_2O , and toluene were dried on alumina columns using a solvent dispensing system. *N*-Chlorosuccinimide was recrystallized from benzene/H₂O, and Bu₄NCl was recrystallized from acetone–Et₂O prior to use. 2,2,6,6-Tetramethyl-1piperdinyloxy free radical (TEMPO) was purified by sublimation prior to use. The Hoveyda–Grubbs catalyst was purified according to published procedure.²⁵

Oxygen- and/or moisture-sensitive reactions were carried out under N_2 in Schlenk glassware that was flame dried under high vacuum (~0.5 mmHg) and purged with N_2 prior to use. Air and/or moisture

sensitive solids were transferred in a glove box. Unless otherwise stated, reactions were stirred with a Teflon-covered stir bar. Concentration refers to the removal of solvent under reduced pressure using a Büchi rotary evaporator. Silica gel column chromatography was performed using Baxter brand silica gel 60 Å (230-400 mesh ASTM). Cerium ammonium molybdate was used as the TLC staining reagent. IR spectra (FTIR) were recorded on a Nicolet 210 FT-IR spectrometer, and reported in wave numbers (cm⁻¹). ¹H NMR and ¹³C NMR spectra were measured on a Varian Gemini-400 instrument at 400 MHz (1H NMR) and 100 MHz (13C NMR). Chemical shifts are reported with the solvent as the internal standard $[CDCl_3: \delta = 7.26 (^{1}H NMR), \delta = 77.0 (^{13}C NMR)]$. Enantiomer ratio was determined by chiral GLC analysis (Supelco Betadex 120 column (30 m \times 0.25 mm). Optical rotations were measured on a Rudolph Research Analytical Autopol IV polarimeter. High-resolution mass spectral (HRMS) analyses were performed on a Micromass LCT ESI-MS (positive mode) at the Mass Spectrometry Laboratory at Boston College.

4-(tert-Butyldimethylsilyloxy)cyclopent-2-enone (4)

Monoacetate compound 3 was obtained from literature procedure²⁶ after recrystallization with hexane-benzene as white needle-like crystals (55% yield, >99% ee). The optical purity was established by chiral GLC analysis. Acetate 3 (957 mg, 6.74 mmol) and DMAP (411 mg, 3.37 mmol) were dissolved in CH₂Cl₂ (67.0 mL) at 0 °C. Et₃N (2.80 mL, 20.2 mmol) and TBSCI (2.03 g, 13.5 mmol) were added to the mixture at 0 °C. The solution was warmed to 23 °C and stirred for 12 h. The mixture was diluted with CH2Cl2 (60 mL) and washed with aq 0.5 M HCl (2×40 mL), aq sat. NaHCO₃ (1×50 mL) and brine $(1 \times 50 \text{ mL})$. The organic layer was dried (MgSO₄), filtered, and solvent was removed under reduced pressure to give TBS-protected cyclopentenyl acetate I (not shown) as a colorless oil, which was pure by ¹H NMR spectroscopy (1.71 g, 99% yield). An aqueous solution of 1 M K₂CO₃ (6.37 mL, 6.37 mmol) was added to a stirred solution of I (1.63 g, 6.37 mmol) in MeOH (27.0 mL) and was stirred for 3 h at 23 °C. The mixture was diluted with CH_2Cl_2 (20 mL) and washed with brine (2 × 50 mL). The aqueous layer was back-extracted with CH_2Cl_2 (3 × 25 mL) and the combined organic layers were dried (MgSO₄) and filtered. The solvent was removed under reduced pressure and the residue was purified by silica gel chromatography (hexanes-EtOAc, 3:1) to give hydroxy cyclopentenyl compound II (not shown) as a colorless oil (1.30 g, 96% yield). PCC (1.96 g, 9.11 mmol) and Celite (2.90 g) were stirred in CH₂Cl₂ (63.0 mL) at 23 °C. Intermediate II (1.30 g, 6.07 mmol) in CH₂Cl₂ (5.0 mL) was added to the mixture. After 2 h, the solution was diluted with Et_2O (15 mL) and was filtered through a pad of Celite (4 cm thick) over silica gel (4 cm thick) and washed with hexanes-Et₂O (1:1, 50 mL). Solvent was removed under reduced pressure and the residue was purified by silica gel chromatography (hexanes- Et_2O , 5:1) to give enone 4 as a colorless oil $(1.18 \text{ g}, 92\%); [\alpha]_{D}^{20} + 43.5 (c \ 1.08, \text{CHCl}_3).$

IR (film): 2954 (m), 2928 (m), 2887 (w), 2857 (m), 1742 (s), 1471 (w), 1354 (m), 1182 (w), 1109 (m), 1071 (m), 900 (m), 838 (m), 777 cm⁻¹ (m).

¹H NMR (CDCl₃, 400 MHz): δ = 7.50 (1 H, dd, *J* = 6.0, 2.4 Hz), 6.23 (1 H, dd, *J* = 5.6, 1.2 Hz), 5.04–5.02 (1 H, m), 2.75 (1 H, dd, *J* = 18.0, 5.6 Hz), 2.29 (1 H, dd, *J* = 18.0, 2.0 Hz), 0.957 (9 H, s), 0.17 (6 H, s).

¹³C NMR (CDCl₃, 100 MHz): δ = 206.3, 163.8, 134.5, 71.1, 45.3, 26.1, 18.5, -4.27, -4.30.

HRMS (ESI⁺): m/z calcd for $C_{11}H_{20}O_2Si + Na (M + Na)$: 235.1130; found: 235.1126.

4-(*tert*-Butyldimethylsilyloxy)bicyclo[3.2.0]hept-6-en-2-ones (5, 6)

In a 3.0 L-photochemical vessel, enone **4** (2.40 g, 11.3 mmol) was dissolved in acetone (2.80 L). A Pyrex immersion well containing a Pyrex sleeve (~3.5 mm thickness) was placed into the vessel and the solution was sparged with acetylene for 5 min. A 100 W medium-pressure mercury vapor lamp was placed in the immersion well and the stirred solution was irradiated with constant bubbling of acetylene for a total of 42 h (reaction progress monitored by GC). Solvent was removed under reduced pressure and the residue was purified by silica gel chromatography (hexanes–Et₂O, gradient 20:1, 10:1, 5:1) to give a mixture of **5** and **6** [**5**:6 = 2:1, 86% conv., 1.73 g, 74% yield, based on recovered starting material (BRSM)]. The ¹H NMR and ¹³C NMR were identical to published results.⁹

4-(tert-Butyldimethylsilyloxy)bicyclo[3.2.0]hept-6-en-2-ol (7)

In a flame-dried Schlenk flask, a mixture of 5 and 6 (1.00 g, 4.20 mmol) was dissolved in toluene (17.4 mL) and cooled to 78 °C. DIBAL-H (823 µL, 4.62 mmol) was added to the mixture and stirred for 3 h, after which the reaction was quenched carefully with MeOH (3.0 mL). The solution was warmed to 23 °C and diluted with Et₂O (10 mL). The reaction was opened to air and aq sat. solution of Rochelle's salt (sodium potassium tartrate, 20 mL) was added to the mixture and stirred for 1 h. The solution was extracted with Et_2O (2 × 30 mL), and the combined organic layers were washed with aq 1 M HCl $(2 \times 30 \text{ mL})$ and brine $(1 \times 30 \text{ mL})$. The combined organic layers were dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel chromatography (hexanes-Et₂O; gradient, 10:1, 5:1, 3:1) to obtain 7, 8, 9, 10 (467 mg, 47%; 129 mg, 13%; 367 mg, 37%; 30 mg, 3% respectively; total yield: 993 mg, 98% yield). Data shown only for cyclobutene 7; $\left[\alpha\right]_{D}{}^{20}$ –0.74 $(c = 0.27, \text{CHCl}_3).$

IR (film): 3530 (b), 3040 (w), 2953 (s), 2929 (s), 2894 (m), 2855 (m), 1469 (m), 1408 (m), 1253 (m), 1088 (s), 1053 (s), 1002 (s), 835 (s), 773 (s), 697 cm⁻¹ (m).

¹H NMR (CDCl₃, 400 MHz): $\delta = 6.05$ (1 H, d, J = 2.8 Hz), 5.99, (1 H, d, J = 2.8 Hz), 4.18 (1 H, d, J = 3.6 Hz), 4.00 (1 H, dd, J = 11.2, 4.4 Hz), 3.36 (1 H, s), 3.23, (1 H, s), 3.10 (1 H, d, J = 11.2 Hz), 2.31 (1 H, dt, J = 14.0, 4.4 Hz), 1.87 (1 H, d, J = 14.4 Hz), 0.89 (9 H, s), 0.09 (6 H, s).

¹³C NMR (CDCl₃, 100 MHz): δ = 140.3, 138.7, 73.8, 72.9, 57.2, 56.5, 40.4, 26.2, 18.4, -4.38, -4.50.

HRMS (ESI⁺): m/z calcd for $C_{13}H_{24}O_2Si + Na (M + Na)$: 263.1443; found: 263.1445.

4-(*tert*-Butyldimethylsilyloxy)-2,3-divinylcyclopentanol (13)

Grubbs catalyst **11** (74.0 mg, 89.8 µmol) was stirred in benzene (87.9 mL) and sparged with ethylene (balloon) for 5 min. Cyclobutene **7** (432 mg, 1.80 mmol) in benzene (2.0 mL) was added to the mixture and stirred under an ethylene atmosphere (balloon) at 23 °C for 15 h. The mixture was opened to air and ethyl vinyl ether (1.0 mL) was added and stirred for 5 min. Silica gel (4.20 g) was then added and stirred for another 15 min. The slurry was filtered through a plug of silica gel and washed with hexanes–EtOAc (3:1, 30 mL). The filtrate was concentrated and purified by silica gel chromatography (hexanes–Et₂O, 5:1) to obtain **13** as a clear yellow oil (426 mg, 88%); $[\alpha]_D^{20}$ +1.6 (*c* 0.37, CHCl₃).

IR (film): 3359 (b), 2955 (s), 2928 (s), 2857 (s), 1639 (w), 1471 (m), 1256 (s), 1089 (b), 912 (s), 836 (s), 775 cm⁻¹ (s).

¹H NMR (CDCl₃, 400 MHz): $\delta = 5.66$ (1 H, ddd, J = 17.1, 10.2, 8.8 Hz), 5.60–5.51 (1 H, m), 5.14–5.06 (3 H, m), 5.03 (1 H, dd, J = 6.0, 1.7 Hz), 4.06–4.01 (2 H, m), 2.84 (1 H, ddd, J = 8.5, 8.3, 5.4 Hz), 2.77 (1 H, ddd, J = 8.8, 8.3, 3.1 Hz), 2.36 (1 H, dt, J = 14.1, 6.5 Hz), 2.00 (1 H, d, J = 6.7 Hz), 1.69 (1 H, dt, J = 14.1, 4.0 Hz), 0.88 (9 H, s), 0.05 (6 H, s).

¹³C NMR (CDCl₃, 100 MHz): δ = 137.3, 137.0, 116.8, 116.7, 77.4, 76.7, 56.6, 55.7, 43.4, 26.2, 18.4, -4.1, -4.3.

HRMS (ESI⁺): m/z calcd for $C_{15}H_{28}O_2Si + Na (M + Na)$: 291.1756; found: 291.1753.

1-[3-(*tert*-Butyldimethylsilyloxy)-5-hydroxy-2-vinylcyclopentyl]oct-1-en-3-one (14)

Divinyl compound **13** (425 mg, 1.58 mmol) and oct-1-en-3-one (300 mg, 2.37 mmol) were dissolved in CH₂Cl₂ (8.0 mL). Hoveyda–Grubbs catalyst **12** (50.0 mg, 79.0 µmol) was added and the mixture was heated to 40 °C. The mixture was stirred for 5 h, then cooled to 23 °C and opened to air. Ethyl vinyl ether (2.0 mL) was added and the mixture was stirred for 5 min. The solvent was removed under reduced pressure and the residue was purified by silica gel chromatography (hexanes–EtOAc, 5:1) to obtain **14** as a light brown oil (429 mg, 74%²⁷); $[\alpha]_D^{20}$ –11 (*c* 0.30, CHCl₃).

IR (film): 3425 (b), 2956 (s), 2928 (s), 2857 (s), 1669 (m), 1625 (m), 1459 (m), 1359 (m), 1248 (m), 839 cm⁻¹ (s).

¹H NMR (CDCl₃, 400 MHz): $\delta = 6.67$ (1 H, dd, J = 15.8, 9.0 Hz), 6.19 (1 H, dd, J = 15.8, 1.1 Hz), 5.49 (1 H, ddd, J = 16.4, 10.8, 9.6 Hz), 5.09–5.04 (2 H, m), 4.13 (1 H, tt, J = 7.3, 4.7 Hz), 4.08 (1 H, dt, J = 5.9, 3.1 Hz), 3.01 (1 H, td, J = 8.3, 5.5 Hz), 2.88–2.83 (1 H, m), 2.50 (2 H, t, J = 7.3 Hz), 2.39 (1 H, ddd, J = 14.3, 7.2, 5.7 Hz), 2.12 (1 H, d, J = 7.5 Hz), 1.74 (1 H, dt, J = 13.8, 3.3 Hz), 1.64–1.56 (2 H, m), 1.35–1.24 (4 H, m), 0.91–0.87 (3 H, m), 0.89 (9 H, s), 0.06 (6 H, s).

¹³C NMR (CDCl₃, 100 MHz): δ = 200.3, 145.1, 135.9, 131.3, 117.8, 77.3, 76.6, 57.1, 54.2, 43.6, 41.0, 31.8, 26.2, 24.3, 22.8, 18.4, 14.3, -4.1, -4.2.

HRMS (ESI⁺): m/z calcd for $C_{21}H_{38}O_3Si + Na (M + Na)$: 389.2488; found: 389.2489.

1-[3,5-Bis(*tert*-butyldimethylsilyloxy)-2-vinylcyclopentyl]oct-1en-3-ol (15)

Enone 14 (362 mg, 0.99 mmol) and DMAP (60.2 mg, 0.49 mmol) were dissolved in CH2Cl2 (9.9 mL). Et3N (412 µL, 2.96 mmol) and TBSCl (297 mg, 1.97 mmol) were added to the mixture. The solution was stirred at 23 °C for 12 h. The mixture was diluted with CH_2Cl_2 (40 mL) and washed with 0.5 M HCl (2×15 mL), aq sat. NaHCO₃ (1 \times 20 mL), and brine (1 \times 20 mL). The organic layer was dried (MgSO₄), filtered, and solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (hexanes-Et₂O, 15:1) to obtain bis(TBS-protected) enone III (not shown) as a colorless oil (435 mg, 92%). To a flame-dried Schlenk flask, intermediate III (300 mg, 0.62 mmol) and toluene (3.12 mL) were added. The solution was cooled to -78 °C and (R)-methyl-CBS-oxazoborilidine catalyst (1 M in toluene, 312 µL, 0.31 mmol) was added followed by catecholborane (266 µL, 2.50 mmol). The mixture was stirred at -78 °C for 15 h. MeOH (2.0 mL) was added to quench the reaction and the mixture was gradually warmed to 23 °C. The solution was diluted with EtOAc (20 mL) and the combined organic layers were washed with aq 1 M NaOH (1×20 mL), aq 1 M HCl (1×20 mL), and brine (1×20 mL). The organic layer was dried (MgSO₄), filtered, and solvent was removed under reduce pressure. The residue was purified by silica gel chromatography (hexanes-Et₂O, 8:1) to afford **15** as a colorless oil (303 mg, 99%); $[\alpha]_D^{20}$ +1.9 (*c* 1.3, CHCl₃).

IR (film): 3342 (b), 3070 (w), 2955 (s), 2925 (s), 2853 (s), 1638 (w), 1469 (m), 1361 (m), 1258 (s), 1095 (b), 835 (s), 775 cm⁻¹ (s).

¹H NMR (CDCl₃, 400 MHz): $\delta = 5.63-5.41$ (3 H, m), 5.06-5.04 (1 H, m), 5.03-5.01 (1 H, m), 4.05 (1 H, qd, J = 10.2, 6.5 Hz), 3.95-3.89 (2 H, m), 2.74-2.65 (2 H, m), 2.36 (1 H, dt, J = 14.0, 7.1 Hz), 1.59 (1 H, dt, J = 13.7, 5.6 Hz), 1.54-1.43 (2 H, m), 1.36-1.25 (7 H, m), 0.91-0.86 (3 H, m), 0.87 (18 H, s), 0.018 (6 H, s), 0.013 (3 H, s), 0.007 (3 H, s). ¹³C NMR (CDCl₃, 100 MHz): δ = 135.5, 129.5, 76.2, 72.9, 61.6, 53.5, 46.0, 44.4, 37.4, 32.4, 31.8, 25.9, 25.2, 22.7, 18.2, 18.1, 14.2, -3.9, -4.4, -4.5.

Diastereomeric ratio was determined by comparison of the ¹H NMR spectrum (C_6D_6) with the authentic 1:1 diastereomeric mixture (estimated detection limits of 20:1).

ent-15-epi-F2t-Isoprostane

9-BBN (0.5 M in THF, 1.0 mL, 0.50 mmol) was added to neat **15** (111 mg, 0.23 mmol) at 0 °C under N₂ and stirred for 5 min. The mixture was warmed to 23 °C and stirred for 5.5 h. The reaction was opened to air and diluted with EtOAc (1.5 mL). The solution was cooled to 0 °C and aq 3 M NaOH (545 μ L, 1.64 mmol) and 30% H₂O₂ (449 μ L, 4.67 mmol) were added. After 1 h, the mixture was extracted with EtOAc (3 × 5 mL) and the combined organic layers were washed with aq sat. NaHCO₃ (2 × 10 mL) and brine (1 × 10 mL). The organic layer was dried (MgSO₄), filtered, and solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (hexanes–EtOAc, gradient 3:1, 1:1) to obtain 1-[3,5-bis(*tert*-butyldimethylsilyloxy)-2-(2-hydroxyethyl)cyclopentyl]oct-1-en-3-ol (**IV**; not shown) as a colorless oil (96.4 mg, 85%); [α]_D²⁰–15.6 (*c* 2.40, CHCl₃).

IR (film): 3388 (b), 2956 (s), 2925 (s), 2857 (s), 1647 (m), 1468 (m), 1363 (m), 1258 (m), 1061 (m), 851 (m), 770 (m), 666 cm⁻¹ (m).

¹H NMR (CDCl₃, 400 MHz): δ = 5.51 (1 H, dd, *J* = 15.3, 6.5 Hz), 5.37 (1 H, dd, *J* = 15.4, 9.9 Hz), 4.06 (1 H, q, *J* = 6.4 Hz), 3.90–3.84 (2 H, m), 3.72–3.60 (2 H, m), 2.56–2.51 (1 H, m), 2.35 (1 H, dt, *J* = 13.7, 6.9 Hz), 2.24 (1 H, dt, *J* = 14.7, 8.1 Hz), 1.67–1.44 (6 H, m), 1.39–1.24 (7 H, m), 0.91–0.89 (2 H, m), 0.89 (9 H, s), 0.88–0.86 (1 H, m), 0.87 (9 H, s), 0.07 (6 H, s), 0.02 (6 H, s).

¹³C NMR (CDCl₃, 100 MHz): δ = 135.5, 129.5, 76.7, 76.2, 72.9, 61.7, 53.6, 46.0, 44.4, 37.4, 32.4, 31.9, 25.9, 25.2, 22.7, 18.2, 18.1, 14.2, -3.9, -4.4, -4.5.

HRMS (ESI⁺): m/z calcd for $C_{27}H_{56}O_4Si_2$ + Na (M + Na): 523.3615; found: 523.3625.

Diol IV (232 mg, 0.463 mmol) and Bu₄NCl (25.7 mg, 92.6 µmol) were stirred in CH₂Cl₂ (3.8 mL) and H₂O (carbonate buffer, pH 8.6, 3.8 mL) at 23 °C. NCS (68.0 mg, 0.509 mmol) and TEMPO (14.5 mg, 92.6 µmol) was added to the mixture and stirred for 3 h. The mixture was diluted with CH₂Cl₂ (10 mL) and the CH₂Cl₂ layer washed with brine $(2 \times 15 \text{ mL})$. The aqueous layer was back-extracted with CH_2Cl_2 (3 × 15 mL) and the combined organic layers were dried (MgSO₄), filtered, and concentrated. The residue was purified by silica gel chromatography (hexanes-EtOAc, 4:1) to obtain the aldehyde 16 as a thick orange oil (184 mg, 99%, BSRM) and starting material (49 mg, 80% conv.). t-BuOK (329 mg, 2.94 mmol) in THF (4.9 mL) was added to a stirred solution of carboxybutyltriphenylphosphonium bromide (651 mg, 1.47 mmol) in THF (3.1 mL) at 23 °C. After this bright orange ylide solution was stirred for 20 min, it was syringed into a stirred solution of the aldehyde (183 mg, 0.37 mmol) in THF (1.1 mL). After the reaction was stirred for 3.5 h, aq sat. NH₄Cl (15 mL) and glacial AcOH (1 mL) were added. The solution was extracted with EtOAc $(3 \times 10 \text{ mL})$, dried (MgSO₄), filtered, and concentrated. The residue was purified by silica gel chromatography (hexanes-EtOAc-AcOH, 5:1:0.1) to obtain TBS-protected-isoprostane V (not shown) as a colorless oil (200 mg, 93%). TBAF (1 M in THF, 2.69 mL, 2.69 mmol) was added to neat V (196 mg, 0.37 mmol) at 23 °C and stirred overnight. The mixture was opened to air and diluted with EtOAc (5.0 mL) and washed with aq sat. NH_4Cl (2 × 5 mL) solution. The aqueous layer was back-extracted with EtOAc (3×10 mL) and the combined organic layers were washed with brine $(1 \times 25 \text{ mL})$. The solution was dried (MgSO₄), filtered, and solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (EtOAc-MeOH-AcOH, 20:1:0.1) to obtain ent-15-epi-F_{2t}-isopros-

Synthesis 2007, No. 15, 2397-2403 © Thieme Stuttgart · New York

tane as a pale yellow oil (104 mg, 87%); $[\alpha]_{\rm D}^{\ 20}$ –1.1 (c 0.55, MeOH).

IR (film): 3363 (b), 2999 (w), 2923 (m), 2847 (m), 1698 (s), 1405 (w), 1235 (w), 1052 (m), 965 cm⁻¹ (m).

¹H NMR (CDCl₃, 400 MHz): $\delta = 5.57$ (1 H, dd, J = 15.3, 6.6 Hz), 5.52–5.40 (3 H, m), 4.12 (1 H, q, J = 6.4 Hz), 4.04–4.00 (2 H, m), 3.94–3.52 (3 H, br s), 2.78 (1 H, dq, J = 8.6, 3.9 Hz), 2.44 (1 H, dt, J = 14.7, 6.9 Hz), 2.32 (2 H, t, J = 6.4 Hz), 2.21–2.04 (4 H, m), 1.99–1.90 (1 H, m), 1.74–1.61 (3 H, m), 1.60–1.45 (2 H, m), 1.39– 1.25 (6 H, m), 0.88 (3 H, t, J = 6.5 Hz).

¹³C NMR (CDCl₃, 100 MHz): δ = 176.5, 135.2, 129.7, 129.4, 129.2, 76.5, 76.4, 73.1, 53.8, 50.8, 42.3, 37.3, 32.6, 31.8, 27.0, 26.3, 25.2, 24.4, 22.7, 14.2.

HRMS (ESI⁺): m/z calcd for $C_{20}H_{34}O_5$ + Na (M + Na): 377.2304; found: 377.2290.

d_4 -ent-15-epi-F_{2t}-Isoprostane

The synthesis was carried out analogously as above, starting with cross-metathesis of the appropriate d_4 -oct-1-en-3-one and divinyl cyclopentane **13**;²⁸ [α]_D²⁰ –0.6 (*c* 0.33, MeOH).

IR (film): 3420 (b), 2930 (m), 2873 (m), 1709 (s), 1413 (w), 1255 (w), 1193 (w), 1060 (m), 966 cm⁻¹ (w).

¹H NMR (CDCl₃, 400 MHz): $\delta = 5.56$ (1 H, dd, J = 15.2, 6.8 Hz), 5.48–5.40 (3 H, m), 4.18 (1 H, br s), 4.14 (1 H, q, J = 6.0 Hz), 4.00– 4.01 (2 H, m), 2.81–2.76 (1 H, m), 2.43 (1 H, dt, J = 14.8, 6.8 Hz), 2.32 (2 H, t, J = 6.5 Hz), 2.21–2.04 (4 H, m), 1.98–1.93 (1 H, m), 1.70–1.61 (3 H, m), 1.54–1.46 (2 H, m), 1.34–1.24 (4 H, m), 0.89–0.84 (3 H, m).

 ^{13}C NMR (CDCl₃, 100 MHz): δ = 175.8, 135.6, 130.1, 129.7, 129.4, 76.6, 76.5, 73.2, 53.8, 50.9, 42.4, 32.7, 27.0, 26.4, 26.4, 24.5, 14.1.

HRMS (ESI⁺): m/z calcd for $C_{20}H_{30}D_4O_5$ + Na (M + Na): 381.2550; found: 381.2555.

Acknowledgment

We thank the National Institutes of Health (R01-CA66617) for financial support. We are also grateful to Materia for the gift of metathesis catalyst.

References

- (1) For free radical peroxidation of arachidonic acid, see: (a) Morrow, J. D.; Hill, K. E.; Burk, R. F.; Nammour, T. M.; Badr, K. F.; Roberts, L. J. II Proc. Nat. Acad. Sci. U.S.A. 1990, 87, 9383. (b) Morrow, J. D.; Awad, J. A.; Boss, H. J.; Blair, I. A.; Roberts, L. J. II Proc. Natl. Acad. Sci. U.S.A. 1992, 89, 10721. (c) Morrow, J. D.; Minton, T. A.; Mukundan, C. R.; Campbell, M. D.; Zackert, W. E.; Daniel, V. C.; Badr, K. F.; Blair, I. A.; Roberts, L. J. II J. Biol. Chem. 1994, 269, 4317. (d) Harrison, K. A.; Murphy, R. C. J. Biol. Chem. 1995, 270, 17273. (e) Morrow, J. D.; Awad, J. A.; Wu, A.; Zackert, W. E.; Daniel, V. C.; Roberts, L. J. II J. Biol. Chem. 1996, 271, 23185. (f) Basu, S. Prostaglandins, Leukotrienes Essent. Fatty Acids 1998, 58, 319. (g) Roberts, L. J.; Fessel, J. P. Chem. Phys. Lipids 2004, 128, 173. (h) See also: Wang, D.; DuBois, R. N. Proc. Natl. Acad. Sci. U.S.A. 2004, 101, 415. (2) (a) Natarajan, R.; Lanting, L.; Gonzales, N.; Nadler, J. Am.
- (2) (a) Natarajan, R.; Lanting, L.; Gonzales, N.; Nadler, J. Am. J. Physiol. **1996**, 271, E159. (b) Kunapuli, P.; Lawson, J. A.; Rokach, J. A.; Meinkoth, J. L.; FitzGerald, G. A. J. Biol. Chem. **1998**, 273, 22442.

- (3) (a) Leitinger, N.; Blazek, I.; Sinzinger, H. *Thrombosis Res.* 1997, 86, 337. (b) Cranshaw, J. H.; Evans, T. W.; Mitchell, J. A. *Brit. J. Pharm.* 2001, 132, 1699. (c) For review, see: Csiszar, A.; Stef, G.; Pacher, P.; Ungvari, Z. *Prostaglandins, Leukotrienes Essent. Fatty Acids* 2002, 66, 557.
- (4) (a) Morrow, J. D.; Zackert, W. E.; Van Der Ende, D. S.; Reich, E. E.; Terry, E. S.; Cox, B.; Sanchez, S. C.; Montine, T. J.; Roberts, L. J. Oxidative Stress Disease 2002, 8, 57.
 (b) For a recent review, see: Milne, G. L.; Morrow, J. D. Antioxid. Redox Signal. 2006, 8, 1379. (c) Yin, H.; Musiek, E. S.; Morrow, J. D. J. Biol. Sci. 2006, 6, 469. (d) Morrow, J. D. Curr. Pharmaceut. Design 2006, 12, 895.
- (5) (a) Quinn, J. F.; Montine, K. S.; Moore, M.; Morrow, J. D.; Kaye, J. A.; Montine, T. J. J. Alzheimer's Dis. 2004, 6, 93.
 (b) Nishio, T.; Miyadera, R.; Sakai, R.; Abe, K.; Kanazawa, H.; Fukui, K.; Urano, S. J. Clin. Biochem. Nutr. 2006, 38, 161. (c) For a review, see: Montine, T. J.; Quinn, J. F.; Kaye, J. A.; Morrow, J. D. Oxidative Stress Disease 2006, 22, 147.
- (6) (a) For recent review, see: Giovanni, D.; Falco, A.; Patrono, C. *Chem. Phys. Lipids* 2004, *128*, 149. (b) See also: Boyne, M. S.; Sargeant, L. A.; Bennett, F. I.; Wilks, R. J.; Cooper, R. S.; Forrester, T. E. *Diabetes Res. Clin. Pract.* 2007, *76*, 149.
- (7) (a) Camphausen, K.; Menard, C.; Sproull, M.; Goley, E.; Basu, S.; Coleman, C. N. *Int. J. Radiat. Oncol. Biol. Phys.* 2004, 58, 1536. (b) Rossner, P. Jr.; Gammon, M. D.; Terry, M. B.; Agrawal, M.; Zhang, F. F.; Teitelbaum, S. L.; Eng, S. M.; Gaudet, M. M.; Neugut, A. I.; Santella, R. M. *Cancer Epidem. Biomar.* 2006, *15*, 639.
- (8) For reviews, see: (a) Taber, D. F.; Hoerner, S. R.; Herr, J. R.; Gleave, M. D.; Kanai, K.; Pina, R.; Jiang, Q.; Xu, M. Chem. Phys. Lipids 2004, 128, 57. (b) Quan, L. G.; Cha, J. K. Chem. Phys. Lipids 2004, 128, 3. (c) Rokach, J.; Kim, S.; Bellone, S.; Lawson, J. A.; Pratico, D.; Powell, W. S.; FitzGerald, G. A. Chem. Phys. Lipids 2004, 128, 35. For early references, see: (d) Corey, E. J.; Shih, N. Y.; Shimoji, K. Tetrahedron Lett. 1984, 25, 5013. (e) O'Connor, D. E.; Mihelich, E. D.; Coleman, M. C. J. Am. Chem. Soc. 1984, 106, 3577. (f) Rondot, B.; Durand, T.; Girard, J. P.; Rossi, J. C.; Schio, L.; Khanapure, S. P.; Rokach, J. Tetrahedron Lett. 1993, 34, 8245. (g) Hwang, S. W.; Adiyama, M.; Khanapure, S.; Schio, L.; Rokach, J. J. Am. Chem. Soc. 1994, 116, 10829. (h) Larock, R. C.; Lee, N. H. J. Am. Chem. Soc. 1991, 113, 7815. (i) See also: Roland, A.; Durand, T.; Egron, D.; Vidal, J. P.; Rossi, J. C. J. Chem. Soc., Perkin Trans. 1 2000, 245. (j) Durand, T.; Guy, A.; Vidal, J. P.; Rossi, J. C. J. Org. Chem. 2002, 67, 3615. (k) Jacobo, S. H.; Chang, C.-T.; Lee, G.-J.; Lawson, J. A.; Powell, W. S.; Pratico, D.; FitzGerald, G. A.; Rokach, J. J. Org. Chem. 2006, 71, 1370. (1) Jung, M. E.; Berliner, A.; Angst, D.; Yue, D.; Koroniak, L.; Watson, A. D.; Li, R. Org. Lett. 2005, 7, 3933. (m) Pinot, E.; Guy, A.; Guyon, A.-L.; Rossi, J.-C.; Durand, T. Tetrahedron: Asymmetry 2005, 16, 1893.
- (9) Schrader, T. O.; Snapper, M. L. J. Am. Chem. Soc. 2002, 124, 10998.
- (10) (a) Schrader, T. O.; Snapper, M. L. *Tetrahedron Lett.* 2000, *41*, 9685. For earlier studies on ring-opening cross-metathesis of cyclobutenes, see: (b) Randall, M. L.; Tallarico, J. A.; Snapper, M. L. *J. Am. Chem. Soc.* 1995, *117*, 9610. (c) Tallarico, J. A.; Bonitatebus, P. J. Jr.; Snapper, M. L. *J. Am. Chem. Soc.* 1997, *119*, 7157. (d) Snapper, M. L.; Tallarico, J. A.; Randall, M. L. *J. Am. Chem. Soc.* 1997, *119*, 1478. (e) Tallarico, J. A.; Randall, M. L.; Snapper, M. L. *Tetrahedron* 1997, *53*, 16511.
- (11) Schrader, T. O. *Ph.D. Thesis*; Boston College: Chestnut Hill Massachusetts, **2002**.

- (12) Theil, F.; Schick, H.; Winter, G.; Reck, G. *Tetrahedron* **1991**, *47*, 7569.
- (13) Gosh, A. K.; Liu, W. J. Org. Chem. 1997, 62, 7908.
- (14) For a more recent and convenient approach to this intermediate, see: Zhao, Y.; Rodrigo, J.; Hoveyda, A. H.; Snapper, M. L. *Nature (London)* **2006**, *443*, 67.
- (15) (a) Buchi, G.; Burgess, E. M. J. Am. Chem. Soc. 1960, 82, 4333. (b) Eaton, P. E. Tetrahedron Lett. 1964, 3695.
 (c) Serebryakov, E. P.; Kulomzina-Pletneva, S. D.; Margaryan, A. K. Tetrahedron 1979, 35, 77.
- (16) Cho, J. H.; Kim, B. M. Org. Lett. 2003, 5, 531.
- (17) Regioselectivity was determined by COSY NMR.
- (18) Corey, E. J.; Helal, C. J. Angew. Chem. Int. Ed. **1998**, 37, 1986.
- (19) Selectivity was determined by ¹H NMR (estimated detection limits of 20:1) by comparison with authentic samples of each diastereomer.
- (20) Einhorn, J.; Einhorn, C.; Ratajczak, F.; Pierre, J.-L. J. Org. Chem. **1996**, *61*, 7452.
- (21) This isotopically labeled isoprostane could be used as an alternative standard for metabolic studies as shown in: Kim, S.; Powell, W. S.; Lawson, J. A.; Jacobo, S. H.; Pratico, D.; FitzGerald, G. A.; Maxey, K.; Rokach, J. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 1613.
- (22) Williams, J. M.; Jobson, R. B.; Yasuda, N.; Marchesini, G.; Dolling, U.-J.; Grabowski, E. J. J. *Tetrahedron Lett.* **1995**, *36*, 5461.

- (23) For selected examples of GC/MS studies for the analysis of isoprostanes, see: (a) Roberts, L. J. II; Morrow, J. D. In *Methods in Biological Oxidative Stress*; Hensley, K.; Floyd, R. A., Eds.; Humana Press: Totawa NJ, **2003**, Chap. 4, 33–39. (b) Morrow, J. D.; Harris, T. M.; Roberts, L. J. II *Anal. Biochem.* **1990**, *184*, 1. (c) Wang, Z.; Ciabattoni, G.; Creminon, C.; Lawson, J.; FitzGerald, G. A.; Patrono, C.; Maclouf, J. J. Pharmacol. Exp. Ther. **1995**, *275*, 94. (d) Bachi, A.; Zuccato, E.; Baraldi, M.; Fanelli, R.; Chiabrando, C. Free Radic. Biol. Med. **1996**, *20*, 619. (e) Waugh, R. J.; Murphy, R. C. J. Am. Soc. Mass Spectrom. **1996**, *7*, 490. (f) Patrignani, P.; Santini, G.; Panara, M. R.; Sciulli, M.; Greco, A.; Rotondo, M. T.; diGiamberardino, M.; Maclouf, J.; Ciabattoni, G.; Patrono, C. Br. J. Pharmacol. **1996**, *118*, 1285.
- (24) More details on the setup and results of the GC/MS detection assay may be obtained by contacting the authors.
- (25) Garber, S. B.; Kingsbury, J. S.; Gray, B. L.; Hoveyda, A. H. J. Am. Chem. Soc. 2000, 122, 8168.
- (26) Theil, F.; Schick, H.; Winter, G.; Reck, G. *Tetrahedron* 1991, 47, 7569.
- (27) Data shown for a 74% yield reaction. Yields range from 70–90%, depending on catalyst purity.
- (28) This compound is the major isomer; however, it is in a mixture of isomers of lower deuterium incorporation.