



Aminocyclitol Synthesis

Chemoenzymatic Synthesis of Hygromycin Aminocyclitol Moiety and its C2 Epimer

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Abstract: This manuscript describes the enantioselective synthesis of the aminocyclitol moiety of the antibiotic hygromycin A in eight steps and 39 % overall yield from a non-chiral starting material. The sequence made use of an initial enzymatic step to transfer chirality to an aromatic ring and was followed by selective organic chemistry transformations (oxidation, pro-

tection, azidation, hydrolysis) of the six-membered ring in order to achieve the target. The approach is also amenable to the synthesis of other related unnatural analogues as exemplified by the synthesis of the C2 epimer of the natural aminocyclitol. All the intermediates were fully characterized, and the absolute stereochemistry assigned by spectrometric methods.

Introduction

Hygromycin A (Figure 1) is an antibiotic first isolated in 1953 from *Streptomyces hygroscopicus*,^[1] that inhibits peptidyl transferase activity.^[2] It exerts moderate activity against gram-posi-

tive and some gram-negative bacteria,^[3] but exhibits potent activity both in vitro^[4] and in vivo^[4,5] against the spirochete responsible for swine dysentery. This fact encouraged a research group from Pfizer to synthesize several analogues by a semi-synthetic approach to develop new antibiotics of potential



Figure 1. Structure of antibiotics: hygromycin A, A201A, and puromycin.

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- local available on the WWW under https://doi.org/10.1002/ejoc.201801424.

veterinary use (see^[6] and references cited therein). To date, two total syntheses of hygromycin A have been described, the first one by Ogawa group,^[7] and the second one by Donohoe et al.^[8] One formal synthesis was also accomplished by Yan group,^[9] and Trost group has synthesized C-2"*-epi*-hygromycin A.^[10]

Hygromycin A structure is originated from the assembling of three independent subunits: a ketoaldose, a substituted caffeic acid residue, and an aminocyclitol moiety.^[11] Both sugar and caffeic acid residues resemble those of A201A antibiotic (Fig-





ure 1), and to a lesser extent, puromycin. However, the aminocyclitol moiety is unique and does not resemble any cyclitol of the aminoglycoside antibiotics family. Hygromycin particular structure encouraged several groups to study its biosynthesis and embrace efficient synthetic approaches. To date, the literature has described five synthetic approaches towards hygromycin A aminocyclitol moiety (Table 1). The Yan group accomplished the shortest sequence to obtain the aminocyclitol core in 2012,^[9] although Shashidhar group reported the sequence with best global yield.^[12]

Table 1. Reported syntheses of the aminocyclitol moiety associated with hygromycin $A.^{[7,-10,12-15]}$

Year	Research group	Starting material	Steps	Global yield
1989 ^[7,13]	Ogawa	Methyl α-D- glucopyranoside	18	2%
		Benzoquinone	13	10%
2001 ^[10,14]	Trost	(±)-conduritol B tetraacetate	10	23%
2005 ^[15]	Develop		14	12%
2009 ^[8a]	Donohoe		15	20%
2012 ^[12] 2012 ^[9]	Shashidhar Yan	<i>myo</i> -inositol L-tartaric acid	11 9	31% 21%

Herein we describe a chemoenzymatic approach towards hygromycin A aminocyclitol moiety, as well as its C2 epimer. The development of efficient synthetic routes, with good diastereomeric control while introducing new stereogenic centers, could be of great value for synthesizing hygromycin A analogues. This approach would allow exploring new compounds that cannot be synthesized by a semi-synthetic approach.

We have made use of enzymatic transformations, particularly microbial dihydroxylations^[16] and, to a minor extent, yeast reductions^[17] to prepare a diverse array of optically pure chiral molecules. Aminocyclitols have been preferred targets in chemoenzymatic synthesis. In particular, the dioxygenase catalyzed oxidation of aromatics have resulted in a very efficient approach to optically pure aminoinositols,^[18] conduritols,^[19] as well as their deoxygenated analogues and conjugates.^[20] Initial efforts

of Hudlicky et al. since the early nineties of the last century have developed into a full stereodivergent strategy that has been exploited around the world to render a wide diversity of this nitrogen containing polyols.^[21] Recently, we have targeted the synthesis of more complicated structures bearing a cyclitol core such as pancratistatin analogues.^[22] and linear molecules inspired in the hygromycin A skeleton.^[23]

Results and Discussion

Synthesis of L-4,5-O-Methylene-2-neo-inosamine (25)

Bromobenzene was enzymatically dihydroxylated by means of a recombinant strain of *E. coli* (JM109 (pDTG601)), and a final concentration of 40 g/L of culture medium of enantiomerically pure (15,25)-3-bromo-3,5-cyclohexadiene-1,2-diol (1) was achieved (Scheme 1).^[24] Our initial approach was to introduce the methylenedioxy group directly on compound 1, not only to avoid aromatization but also to sterically direct the following reactions. Unfortunately, all attempts to protect the diol group led only to aromatization products and we were not able to obtain compound 2. We, therefore, decided to modify our strategy and protect the diol function as an isopropylidene (Scheme 2).



Scheme 1. Attempts to introduce the methylenedioxy group on diol 1.

The stereoselective transformation of epoxide **3** into hydroxyazide **4** was achieved by means of a two-step sequence first described by Hudlicky et al. for the corresponding chloro-analogue of compound **3** (Scheme 2).^[25] The epoxide ring was initially opened by lithium chloride. In this reaction, the presence of ethyl acetoacetate provided the optimal acidity to catalyze the reaction.^[26] In a second stage, the chloride was displaced to ren-



Scheme 2. Functionalization of the electron-richer olefin.





der the desired azide **4** with overall retention of configuration. Then the remaining hydroxyl group was protected as the benzyl ether **5** and the acetonide group was deprotected to render diol **6**, which is ready to introduce the methylenedioxy group (Scheme 2).

Conduritol **6** was completely resistant to acetalization under basic conditions. The diol was unreactive from 0 °C to room temperature and completely decomposed upon warming up the reaction (Table 2, entries 1-5). On the other hand, the use of silver oxide under neutral conditions to enhance the electrophilicity of the dihalomethane (entries 6-8) led to the isolation of the desired product, but only in 10 % yield (entry 6). Much better results were obtained for the acid-catalyzed acetalization with dimethoxymethane (DMM) as electrophile (entries 9-14). Although *p*TsOH did not promote the reaction at room temperature even using a vast excess of DMM (entry 9), the reaction did take place under reflux in dichloromethane and yielded the desired ketal **7** in 91 % yield as the only product (entry 11). Along the study we observed that ketals **8**, **9** and **10** were formed if only moderate excess (1.5-fold excess) of DMM was used, but they were all converted to compound **7** under refluxing temperature (entries 13 and 14).

At this point, the introduction of the methylenedioxy group was successfully accomplished, however, it was still necessary to introduce the acetonide first as a transient protective group, then operate on the electron-richer olefin, remove the acetonide and finally introduce the methylenedioxy group. Since the acidic conditions used to introduce the methylenedioxy group may allow the deprotection of the acetonide, we envisioned that a transacetalization reaction over compound **5** might lead directly to compound **7**, without isolating diol **6**. We gladly observed that after 15 hours the reaction was completed and the desired ketal **7** was obtained in 79 % yield (Scheme 3).

Attempts to oxidize the bromoalkene either with m-CPBA or with OsO_4 led only to recovered starting material (Scheme 3). We assayed the use of the more reactive RuO_4 (generated in



Scheme 3. Transacetalization reaction on 5 and functionalization of the electron-poorer olefin on the dioxolane 7.

	N ₃ ^{wir}	Br OH OH Table 2	N3 ^{WT} OBn N	Br OMC OBn	Br N ₃ ^w Of OBn	H MOM N ₃ ^w	омом омом Зп
	(6	7	8	9	1	10
Entry	Reagent (eq)	Catalyst (Eq)	Reaction temp	erature (°C)	Solvent	Reaction time (h)	Product (yield)
1	$CH_{2}I_{2}$ (5)	NaH	0 - r.t.		DMF	7	7 (trace)
2	CH ₂ Br ₂ (3.4)	NaH	0 - r.t.		DMF	19	Recovered sm
3	CH ₂ Br ₂ (3.4)	NaH	40 - 85		DMF	11	Decomposition
4	CH ₂ Br ₂ (3.4)	K ₂ CO ₃ , 18-c-6	r.t.		CH_2CI_2	96	Recovered sm (70 %)
5	CH ₂ Br ₂ (3.4)	K ₂ CO ₃ , 18-c-6	reflux		CH ₂ Cl ₂	4	Decomposition
6	$CH_{2}I_{2}$ (15)	Ag ₂ O, TBAI	r.t.		THE	24	7 (10 %)
7	CH ₂ I ₂ (15)	Ag ₂ O, TBAI	reflux		THF	4	7 (trace)
8	CH ₂ I ₂ (100)	Ag₂O, TBAI	r.t.		CH_2I_2	10	Decomposition
9	DMM (100)	<i>p</i> TsOH	r.t.		CH_2CI_2	48	7, 8, 9, 10
10	DMM (10)	<i>p</i> TsOH	reflux		THF	24	7 (trace)
11	DMM (10)	<i>p</i> TsOH	reflux		CH_2CI_2	19	7 (91 %)
12	DMM (100)	P ₂ O ₅	r.t.		CH_2CI_2	3	10 (quant.)
13	DMM (1.5)	P_2O_5	r.t.		CH_2CI_2	19	8, 9, 10
14	DMM (1.5)	P_2O_5	reflux		CH_2CI_2	2	7 (40 %)

Table 2. Protection of azido conduritol derivative 6.







Scheme 4. Proposed mechanism for the transformation of 7 into 11.

situ) and observed complete consumption of the starting material. However, we did not obtain the desired hydroxy ketone, but a compound identified as **11**. We assumed that unsaturated ketone **11** was derived from hydroxy ketone **12** by hydrazoic acid loss followed by a 1,2-hydride shift as proposed in Scheme 4.

As we were not able to isolate the desired hydroxy ketone **12**, we decided to change the synthetic steps sequence. Attempts to functionalize the bromoalkene on epoxide **3** with RuO₄ (Scheme 5) led to a mixture of the hydroxy ketone **13** and the rearranged product **14**. The stereochemistry of the new stereogenic center in compound **13** was suggested by coupling constant measurement ($J_{2-3} = 1.4$ Hz indicates a *cis* stereochemical relationship) and NOE experiments (Figure 2). Crystallization of compound **13** followed by X-ray diffraction analysis confirmed the assigned stereochemistry (Figure 2). Several attempts were made to maximize the formation of epoxide **13**,

and it was noticed that **14** was formed during the purification stage by column chromatography. Consequently, we decided to perform a one-pot sequence adding the NaBH₄ directly to reaction flask once the consumption of epoxide **3** was verified. This approach rendered diol **15** in 60 % yield for both steps. In the same reaction, bromodiol **16** was obtained as a by-product



Figure 2. NOE correlations of hydroxyl-ketone 13 and ORTEP diagram.



Scheme 5. Functionalization of the bromo-olefin on the epoxide 3.



with 22 % yield. The latter probably formed by nucleophilic ring opening of the oxirane by water over the allylic position in a *trans*-diaxial fashion.

The stereochemistry of **15** could not be derived from the ¹H-NMR spectra since the coupling constants could not be determined due to second order splitting. Therefore, the hydroxyl groups were protected to render diacetate 17 (Scheme 5). The coupling constant of H₃₋₄ (Figure 3) was determined to be 4.6 Hz, an intermediate value that could arise either from a cis or a trans relationship between the involved protons. Data from a 1D NOE experiment suggested a cis relationship because when irradiating the endo methyl group of the isopropylidene an enhancement was observed in all β protons (including H₄). In order to confirm the assignment, the epoxide was opened with azide anion. The facial approach of the azide group is governated by the Fürst-Plattner rule, and the regiochemistry of the attack was determined by COSY experiment. The ¹H-NMR spectrum showed a triplet signal (J = 10.0 Hz) at 3.52 ppm that was assigned to H₄ (Figure 4). This suggested a chair conformation with H₁-H₂ and H₂-H₃ in a trans-diaxial relationship. If OH₆ was on the β face, then H₅-H₆ should also have a *trans*-diaxial disposition, but this is not what coupling constant suggested. A measured J₁₋₆ value of 3.0 Hz hinted two pseudo-equatorial protons, confirming a *cis* relationship between H₅ and H₆. We were also able to crystallize compound 18, and its structure was confirmed by single-crystal X-ray crystallography (Figure 4).



Figure 3. Coupling constants and NOE correlations of diacetate 17.





Figure 4. Coupling constant of compound 18 and its ORTEP diagram.

As the compound synthesized by the hydroxy-carbonylation/ reduction sequence over the epoxide was not a *trans* diol as we expected, we decided to explore the reaction over the less strained substrate **4** (Scheme 6). When we performed only the oxidation stage, the main isolated product was **14** (\approx 60 %). During the reaction, another product was observed by TLC (possibly the hydroxy ketone) but it decomposed during isolation as the amount of **14** increased, so we returned to the one pot dihydroxylation/reduction procedure. In this case, a mixture of *trans*(**19**)/*cis*(**20**) diols was obtained in ratio 7.6/1. The diastereomers could be separated by column chromatography, and the desired product was obtained in 61 % yield for both steps.

Structure confirmation of the *cis* isomer **20** was achieved by single-crystal X-ray diffraction analysis (Figure 5). On the other hand, to verify that **19** had the assigned stereochemistry, the compound was acetylated (**21**, Scheme 7), the acetonide was



Figure 5. Crystal structure of triol 20.



Scheme 6. Functionalization of the bromo-olefin on compound 4.



Scheme 7. Confirmation of stereochemistry of compound 19.





removed and the product was acetylated again. This rendered *meso* compound **22**, corroborating the assigned stereochemistry.

Once the stereochemistry of **19** was confirmed, the three hydroxyl groups were protected as benzyl ethers (**23**, Scheme 8), and the methylenedioxy group was introduced by means of a transacetalization reaction. Finally, simultaneous removal of the benzyl groups and reduction of the azide moiety by hydrogenolysis rendered the natural aminocyclitol **25** in 95 % yield (Scheme 8). The recorded NMR spectroscopic data of compound **25** is in full accordance with the reported by Shashidhar.^[12]



Scheme 8. Synthesis of natural aminocyclitol 25.

Synthesis of D-1,2-O-Methylene-5-myo-inosamine [2-epi-(L- 4,5-O-Methylene-2-neo-inosamine)] (30)

In order to synthesize the C2 epimer, the same strategy was applied but starting from the *trans*-azidoalcohol **26** (obtained from epoxide **3**) instead of compound **4** (Scheme 9). The bromo-olefin was functionalized by the same oxidation/ reduction sequence rendering the *trans* diol (**27**). The stereo-chemistry was confirmed by coupling constant measurement: $J_{3-4} \approx J_{4-5} \approx J_{5-6} \approx 10$ Hz indicating a *trans*-diaxial relationship between those protons. The hydroxyl groups were protected as benzyl ethers (**28**) and then the acetonide was interconverted

into the methylenedioxy group to deliver **29**. Finally, a hydrogenolysis reaction rendered aminocyclitol **30** (C2-epimer).

Conclusions

The aminocyclitol core present in hygromycin A was prepared in eight steps with an overall yield of 39 % from cyclohexadienediol **1**. The latter is commercially available or can be prepared in high yield by whole-cell enzymatic dihydroxylation of bromobenzene. The process has now been optimized in our 5liter fermenter to render optically pure diol in up to 40 g/L of cell culture. Comparable yields have been described by the Hudlicky group in Canada that has pioneered the technique for over two decades. To the best of our knowledge, this is the shortest synthesis of the hygromycin A core reported to date and provides the highest overall yield.

The preparation of the C2 epimer in an even shorter sequence provided proof of concept that this approach is suitable for the preparation of hygromycin A analogues of unnatural stereochemistry. Our results in the synthesis of a series of analogues are on development and will be reported in due course.

Experimental Section

General methods

Commercially available reagents were purchased from Sigma-Aldrich and used without further purification. Melting pionts were determined either on a capilar Gallenkamp apparatus or on a capilar Electrothermal IA 9100 MK2 apparatus and are uncorrected. Optical rotation was determined on a Kruss Optronic GmbH P8000 on a 0.5 dm cell (concentration is expressed as W/V percentage). Nuclear Magnetic Resonance spectra were recorded either on a Bruker AVANCE DPX-400 instrument or on a Bruker AVANCE III 500 spectrometer. Chemical shifts (δ) are given in parts per million downfield from tetramethylsilane, and coupling constants (J) are reported in Hertz. 1D-NOESY experiments were carried out at 25 °C using the DPFGSE-NOE pulse sequence of Stott et al. and a mixing time of 300 ms.^[27] Selective excitation of specific protons was achieved with Gaussian shaped pulses. Infrared spectra (IR) were recorded on a Shimadzu FT-IR model IRprestige-21. Low-resolution mass spectra (MS) were recorded on a Shimadzu GC-MS-QP2010 ultra instrument (70 eV electron impact mode), using either Direct Injection (DI) or



Scheme 9. Synthesis of C2 epimer of the natural aminocyclitol.





by means of GC (column DB-WAX: Length 30m, diameter 0.25 mm and Film 0.25 μ m). High-resolution mass spectra were performed on a Bruker Daltonics model ToF_LC (ESI $^+$ mode).

Analytical TLC was performed on silica gel 60F-254 plates and visualized with UV light (254 nm) and/or anisaldehyde– H_2SO_4 –EtOH or vanillin– H_2SO_4 –EtOH as detecting agent. Purifications by column chromatography were performed on Merck 60 silica gel (0,040–0.063 mm).

Single crystal X-ray diffraction data was collected on a Bruker D8 Venture diffractometer using Siemens Sealed-tube Mo- K_{α} radiation monochromatized with a graphite single crystal (Compounds 13 and 18) and Incoatec I microfocus source 1.0 Cu- K_a radiation focused and monochromatized with a Quazar multilayer mirror (Compound 20). A single crystal of each compounds was mounted on the four-circle kappa goniometer and data was collected in all cases in shutterless mode using a PHOTON100 CMOS detector. The diffractometer was operated using APEX2 control software providing cell determination, data collection and processing and structure report routines. Data scaling and absorption correction was performed with SADABS.^[28] Structure determination was performed by modern direct methods with SHELXT program^[29] and least-squares structure refinement with SHELXL^[30] in shelXle environment.^[31] CIF preparation and molecular graphics were prepared with enCIFer (v 1.6.2) and Mercury CSD 3.10.1 respectively. Figures S1, S2 and S3 show ORTEP representations of compounds 13, 18 and 20 and Tables S1, S2 and S3 the crystallographic data, processing and refinement details respectively. CCDC 1535525 (for 13), 1535526 (for 18), and 1535562 (for 20) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre.

(1R,2R,3S,4S)-5-bromo-1,2-epoxy-3,4-isopropylidenedioxy-5cyclohexene (3)^[32]



The ¹H- and ¹³C-NMR spectra data of the compound were found to be in agreement with previous reports.^[32] $\alpha_D^{21,0} = +101$ (c = 1.55, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 6.48 (dd, $J_{1-6} = 4.5$ Hz, $J_{4-6} = 1.3$ Hz, 1H, H₆), 4.87 (ddd, $J_{3-4} = 6.7$ Hz, $J_{2-3} = 1.9$ Hz, $J_{1-3} = 1.1$ Hz, 1H, H₃), 4.42 (dd, $J_{3-4} = 6.7$ Hz, $J_{4-6} = 1.3$ Hz, 1H, H₄), 3.60 (dd, $J_{1-2} = 3.7$ Hz, $J_{2-3} = 1.9$ Hz, 1H, H₂), 3.34 (ddd, $J_{1-6} = 4.6$ Hz, $J_{1-2} = 3.7$ Hz, $J_{1-3} = 1.1$ Hz, 1H, H₁), 1.46 (s, 3H, CH₃), 1.44 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 130.0 (C₅), 126.6 (C₆), 111.5 (C_{isopropylidene}), 74.2 (C₄), 72.7 (C₃), 49.6 (C₂), 48.4 (C₁), 27.6 (CH₃), 26.1 (CH₃).

(1R,2R,5S,6S)-2-azido-4-bromo-5,6-isopropylidenedioxycyclohexa-3-ene-1-ol (4).^[33]



The ¹H- and ¹³C-NMR spectra data of the compound were found to be in agreement with previous reports.^[33] ¹H NMR (400 MHz, CDCl₃) δ (ppm): 6.15 (d, $J_{3-2} = 3.1$ Hz, 1H, H₃), 4.66 (d, $J_{5-6} = 5.3$ Hz, 1H, H₅), 4.40 (t, $J_{1-6} = J_{5-6} = 5.3$ Hz, 1H, H₆), 4.25-4.17 (m, 2H, H₁ and H₂), 2.49 (bs, 1H, OH₁), 1.44 (s, 3H, CH₃), 1.40 (s, 3H, CH₃). ¹³C NMR (100 MHz,

CDCl₃) δ ppm: 126.3 (C₃), 126.4 (C₄), 110.4 (C_{isopropylidene}), 76.1 (C₆), 76.0 (C₅), 69.2 (C₁), 59.5 (C₂), 27.6 (CH₃), 26.0 (CH₃).

(1R,2R,5S,6S)-2-Azido-1-benzyloxy-4-bromo-5,6-isopropylidenedioxy-3-cyclohexene (5).



122 mg of NaH 55 % dispersion in mineral oil (2.8 mmol, 1.6 eq.) were added to a solution of 511 mg of compound 4 (1.7 mmol, 1 eq.) in 8 mL of dry DMF at 0 °C under N₂ atmosphere. The reaction mixture was left to stir during 15 minutes, and then 0.67 mL of BnBr (5.6 mmol, 3.2 eq.) were added. The reaction was kept at 0°C for 10 minutes, and then it was warmed up to room temperature. The reaction was monitored by TLC, and after completion of the reaction, a saturated aqueous solution of NH₄Cl was added and the system was extracted with three portions of AcOEt. The combined organic phases were washed with three portions of saturated agueous solution of CuSO₄, then washed with brine, dried with Na₂SO₄, filtered and the solvent was evaporated under reduced pressure to render an oily crude. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 0 %, then 5 % and finally 10 % (v/v of AcOEt in Hexane). 663 mg of 5 were obtained as a colorless oil (99 % yield). $\alpha_D^{22} = -36$ (c = 3.4, AcOEt). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.41-7.28 (m, 5H, H arom), 6.17 (d, $J_{2-3} = 3.6$ Hz, 1H, H₃), 4.80 (d, J_{gem} = 12.0 Hz, 1H, -OCH₂Ph), 4.71 (d, J_{gem} = 12.0 Hz, 1H, -OCH₂Ph), 4.64 (dd, $J_{5-6} = 5.5$ Hz, $J_{1-5} = 1.4$ Hz, 1H, H₅), 4.39 (t, $J_{1-6} = J_{5-6} = 5.5$ Hz, 1H, H₆), 4.03-3.98 (m, 1H, H₁), 3.97 (dt, $J_{1-2} =$ $J_{2-3} = 3.6$ Hz, $J_{2-6} = 1.5$ Hz, 1H, H₂), 1.40 (s, 3H, CH₃), 1.37 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 137.4 (C_{Ph-quaternary}), 128.7 (C_{Ph-m}), 128.3 (C_{Ph-p}), 128.1 (C_{Ph-o}), 127.3 (C₃), 125.5 (C₄), 110.5 $(C_{isopropylidene}),\ 76.5\ (C_{1or5}),\ 76.5\ (C_{5or1}),\ 75.0\ (C_6),\ 73.6\ (-OCH_2Ph),$ 57.9 (C2), 27.7 (CH3), 26.2 (CH3). FT-IR (vmax/cm): 3032 (=C-H arom), 2988 (CH₂), 2934 (CH₃), 2899 (CH₃), 2102 (N₃), 1645 (C=C), 1227 (C-O-C), 1074 (O-C-O), 739 (=C-H arom), 698 (=C-H arom). DI-MS m/z (Rel. Int): 352 (1), 350 (1), 324 (0.6), 322 (0.7), 191 (22), 91 (100). HRMS (ESI/Q-TOF) m/z: $[M + Na]^+$ Calcd for $[C_{16}H_{18}BrN_3O_3Na]^+$ 402.0424, found 402.0443.

(1R,2S,5R,6R)-5-Azido-3-bromo-6-benzyloxy-cyclohex-3-ene-1,2-diol (6).



7.06 g of DOWEX-H⁺ 50WX8-100 resin were added to a stirring solution of 706 mg of compound **5** in 18 mL of a mixture MeOH:H₂O (8:2, V/V) at room temperature, and the mixture was warmed up to 45 °C by means of an oil bath. The reaction was monitored by TLC, and once the starting material was consumed (approximately 48 h), the resin was filtered off and washed several times with methanol.





The solvent was removed under reduced pressure, and a brownish solid was obtained. The product was purified by column chromatography, using SiO₂ as stationary phase, and eluting with a solution 20 % (v/v) of AcOEt in Hexane. 516 mg of 6 were obtained as a white solid (82 % yield). M.P. = 103-104 °C. $\alpha_D^{22} = -182$ (c = 1.05, MeOH). ¹H NMR ((CD₃)₂CO, 400 MHz) δ (ppm): 7.47-7.39 (m, 2H, H_{Ph}), 7.39-7.25 (m, 3H, H_{Ph}), 6.18 (d, J_{4-5} = 4.1 Hz, 1H, H_4), 4.79 (s, 2H, -OCH₂Ph), 4.48 (bs, 1H, OH₁), 4.45 (bs, 1H, OH₂), 4.31 (bd, J₅₋₆ = 4.1 Hz, 1H, H₆), 4.27 (dt, J₄₋₅ = J₅₋₆ = 4.1 Hz, J = 1.6 Hz, 1H, H₅), 4.16 (dd, $J_{1-2} = 7.4$ Hz, $J_{1-OH} = 4.0$ Hz, 1H, H₁), 4.03 (dd, $J_{1-2} = 7.7$ Hz, $J_{2-1} = 7.7$ Hz $_{OH}$ = 4.0 Hz, 1H, H₂). ¹³C NMR ((CD₃)₂CO, 101 MHz) δ (ppm): 139.3 (C_{Ph-quaternary}), 129.8 (C₃), 129.1 (C_{Ph-m}), 128.7 (C_{Ph-o}), 128.5 (C_{Ph-p}), 128.0 (C₄), 78.1 (C₂), 73.7 (OCH₂Ph), 71.8 (C₆), 69.9 (C₁), 59.5 (C₅). FT-IR ($\tilde{\nu}_{max}$ /cm): 3296 (OH), 2124 (N₃), 1651 (C=C), 731 (=C-H arom), 696 (=C-H arom). DI-MS m/z (Rel. Int): 312 (4), 310 (4), 254 (1), 252 (1), 150 (13), 91 (100). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₁₃H₁₄BrN₃O₃Na]⁺ 362.01108, found 362.01195.

General procedure for the synthesis of dioxolane under basic media.

14 mg of NaH 55 % dispersion in mineral oil (0.32 mmol, 2.1 eq.) were added to a solution of 50 mg of diol **6** (0.15 mmol, 1 eq.) in 1 mL of dry DMF at 0 °C under N₂ atmosphere. The reaction mixture was left to stir during 45 minutes, and then 35 μ L of CH₂Br₂ (0.50 mmol, 3.4 eq.) were added in the absence of light. The reaction was taken up to the temperature listed on Table 2. After completion of the reaction, a saturated aqueous solution of NH₄Cl was added and the system was extracted with three portions of AcOEt. The combined organic phases were treated with four portions of a saturated aqueous solution of Drine, dried with Na₂SO₄, filtered and the solvent was evaporated under reduced pressure to render an oily crude. Purification was accomplished by flash chromatography using SiO₂ as stationary phase.

General procedure for the synthesis of dioxolane under neutral conditions.

41 mg of diol **6** (0.12 mmol, 1 eq.) were dissolved in 1 mL of dry THF at room temperature under nitrogen atmosphere, and 145 μ L of CH₂I₂ (1.81 mmol, 15 eq.) and 9.0 mg of TBAI (tetrabutilamonium iodide, 0.024 mmol, 0.2 eq.) were added in the absence of light. After stirring for 15 minutes, 224 mg of Ag₂O (0.97 mmol, 8eq.) were added and the reaction was stirred until consumption of the starting material (conditions listed in Table 2). Et₂O was added and Ag₂O was filtered off through a pad of celite. The solvent was evaporated to render an oily crude product that was subjected to flash chromatography on silica gel.

General procedure for the synthesis of dioxolane under acidic conditions using *p*-toluenesulfonic acid as catalyst.

A solution of 516 mg of diol **6** (1.52 mmol, 1 eq.) in 80 mL of dry CH_2CI_2 was treated with the corresponding amount of DMM (Table 2) under N₂ atmosphere and 72 mg of *p*TsOH (0.46 mmol, 0.3 eq.) were added. The reaction mixture was refluxed until consumption of the starting material, then solid NaHCO₃ was added and the resulting system was left to stand at room temperature for 5 minutes. Saturated aqueous solution of NaHCO₃ was added, and the system was extracted with three portions of CH_2CI_2 . The combined organic layers were washed with one portion of brine, dried with Na₂SO₄, filtered and the solvent was removed under reduced pressure rendering an oily crude that was purified over silica flash.

General procedure for the synthesis of dioxolane under acidic conditions using P_2O_5 as catalyst.

A solution of 25 mg of diol **6** (0.074 mmol, 1 eq.) in 1 mL of dry CH_2Cl_2 , under N_2 atmosphere, was treated with the corresponding amount of DMM (Table 2) and 17 equiv. of P_2O_5 were added. The reaction mixture was stirred at room temperature until consumption of the starting material, and then the reaction was cooled by means of an ice bath. Cooled CH_2Cl_2 was added, and P_2O_5 excess was destroyed with cooled saturated aqueous solution of NaHCO₃. Then, the mixture was extracted with sat. NaHCO₃, washed with sat. NaCl and dried with Na₂SO₄. The solvent was filtered off, and the product was then purified by flash chromatography.



Compound 7 was synthesized from 5, following the general procedure for the synthesis of dioxolane under acidic conditions using ptoluenesulfonic acid as catalyst. 10 eq. of DMM were used, and the reaction was kept at reflux temperature for 19h. The crude was purified by flash chromatography, performing a gradient elution: first hexane, and then a 5 % v/v solution of AcOEt in hexane. Colorless oil. α_D^{22} = -93 (c = 1.24, AcOEt). ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 7.39-7.30 (m, 5H, H_{Ph}), 6.26 (dd, J₂₋₃ = 3.9 Hz, J₃₋₅ = 0.7 Hz,1H, H₃), 5.00 (s, 1H, -OCH₂O-), 4.98 (s, 1H, -OCH₂O-), 4.80 (d, J_{aem} = 11.9 Hz, 1H, -OCH₂Ph), 4.72 (d, J_{qem} = 11.9 Hz, 1H, -OCH₂Ph), 4.65 (bd, $J_{5-6} = 5.7$ Hz, 1H, H₅), 4.31 (t, $J_{1-6} = J_{5-6} = 5.7$ Hz, 1H, H₆), 4.01-3.94 (m, 2H, H₁ and H₂). ¹³C NMR (CDCl₃, 101 MHz) δ (ppm): 137.2 (C_{Ph-quaternary}), 128.7 (C_{Ph-p}), 128.6 (C_{Ph-m}), 128.2 (C₃), 128.0 (C_{Ph-o}), 123.6 (C₄), 94.4 (-OCH₂O-), 76.4 (C₅), 75.9 (C₁), 74.9 (C₆), 73.5 (-OCH2Ph), 57.7 (C2). FT-IR (vmax/cm): 2870 (CH2), 2104 (N3), 1645 (C=C), 1088 (O-C-O), 926 (O-C-O), 739 (=C-H arom), 698 (=C-H arom). DI-MS m/z (Rel. Int): 324 (3), 322 (3), 296 (1), 294 (1), 91 (100). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₁₄H₁₄BrN₃O₃Na]⁺: 374.01108, found 374.01139.

(1S,2S,5R,6R)-5-Azido-6-benzyloxy-3-bromo-2-(methoxymethoxyl)-cyclohex-3-ene-1-ol (8).



Colorless oil. $\alpha_D^{21} = -98$ (c = 1.35, AcOEt). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.41 – 7.30 (m, 5H, H_{Ph}), 6.16 (dt, J₄₋₅ = 3.9 Hz, J₂₋₄ = J₄₋₆ = 1.0 Hz, 1H, H₄), 4.84 (d, J_{gem} = 6.8 Hz, 1H, -OCH₂OMe), 4.80 (d, J_{gem} = 6.8 Hz, 1H, -OCH₂OMe), 4.79 (d, J_{gem} = 11.8 Hz, 1H, -OCH₂Ph), 4.68 (d, J_{gem} = 11.8 Hz, 1H, -OCH₂Ph), 4.36 (bd, J₁₋₂ = 3.9 Hz, 1H, H₂), 4.13 (dt, J₁₋₆ = 7.6 Hz, J₁₋₂ = J_{1-OH} = 3.9 Hz, 1H, H₁), 4.09 (td, J₄₋₅ =





(1S,4R,5R,6R)-4-Azido-5-benzyloxy-2-bromo-6-(methoxymethoxyl)-cyclohex-2-ene-1-ol (9)



Colourless oil. $\alpha_D^{21} = -54$ (c = 0.85, AcOEt). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.38 – 7.29 (m, 5H, H_{Ph}), 6.15 (dt, J₃₋₄ = 3.8 Hz, $J_{1-3} = J_{3-5} = 0.7$ Hz, 1H, H₃), 4.78 (d, $J_{gem} = 6.7$ Hz, 1H, -OCH₂OMe), 4.75 (d, $J_{gem} = 11.8$ Hz, 1H, -OCH₂Oh), 7.71 (d, $J_{gem} = 6.7$ Hz, 1H, -OCH₂OMe), 4.71 (d, J_{gem} = 11.8 Hz, 1H, -OCH₂Ph), 4.45-4.41 (m, 1H, H₁), 4.08-4.03 (m, 2H, H₅ + H₆), 4.00 (td, $J_{3-4} = J_{4-5} = 3.8$ Hz, J =1.5 Hz, 1H, H₄), 3.39 (s, 3H, -OCH₃), 2.99 (d, J_{1-OH1} = 6.6 Hz, 1H, OH). ^{13}C NMR (101 MHz, CDCl_3) δ (ppm): 137.5 (C_{Ph-quaternary}), 128.7 (C_{Ph-m}), 128.3 (C_{Ph-p}), 128.1 (C_{Ph-o}), 128.0 (C₃), 127.4 (C₂), 98.0 (-OCH₂OMe), 76.5 (C₅ or C₆), 76.1 (C₆ or C₅), 73.9 (-OCH₂Ph), 70.6 (C1), 58.9 (C4), 56.2 (-OCH3). FT-IR (vmax/cm): 3447 (OH), 2928 (CH2), 2897 (CH₂), 2102 (N₃), 1643 (C=C), 1115 (C-O-C), 1034 (C-O-C), 749 (=C-H arom), 698 (=C-H arom). DI -MS m/z (Rel. Int): 356 (0.4), 354 (0.6), 312 (1.0), 310 (1), 194 (11), 91 (100), 45 (54). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₁₅H₁₈BrN₃O₄Na]⁺ 406.0373, found 406.0371.

(1R,4R,5S,6R)-2-Azido-1-benzyloxy-4-bromo-5,6-bis(methoxymethoxyl)-3-cyclohexene (10)



Compound **10** was synthesized from **6**, following the general procedure for the synthesis of dioxolane under acidic conditions using P₂O₅ acid as catalyst. 100 eq. of DMM were used, and the reaction was stirred at 0°C for 3h. The crude was purified by flash chromatography, performing a gradient elution: first hexane, and then a 5 % v/v solution of AcOEt in hexane. Colorless oil (quantitative yield). $\alpha_D^{22} = -74$ (c = 0.50, AcOEt). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.41-7.29 (m, 5H, H_{arom}), 6.13 (d, J₂₋₃ = 3.8 Hz, 1H, H₃), 4.86 (d, J_{gem} = 6.8 Hz, 1H, -OCH₂OMe), 4.82 (d, J_{gem} = 6.8 Hz, 1H, -OCH₂OMe), 4.74 (d, J = 0.9 Hz, 2H, -OCH₂Ph), 4.71 (d, J_{gem} = 6.7 Hz, 1H, -OCH₂OMe), 4.73 (d, J₅₋₆ = 3.1 Hz, 1H, H₅), 4.12-4.08 (m, 1H, H₆), 4.08-4.03 (m, 2H, H₁ y H₂), 3.47 (s, 3H, -OCH₃), 3.37 (s, 3H, -OCH₃). ¹³C NMR (101 MHz,

 $\begin{array}{l} {\sf CDCI}_3) \; \delta \; (ppm): 137.5 \; ({\sf C}_{{\sf Ph-quaternary}}), \; 128.5 \; ({\sf C}_{{\sf Ph-}m}), \; 128.1 \; ({\sf C}_{{\sf Ph-}p}), \\ 128.0 \; ({\sf C}_3), \; 127.9 \; ({\sf C}_{{\sf Ph-}o}), \; 126.0 \; ({\sf C}_4), \; 97.5 \; (-{\sf OCH}_2{\sf OMe}), \; 97.0 \\ (-{\sf OCH}_2{\sf OMe}), \; 76.0 \; ({\sf C}_1), \; 75.9 \; ({\sf C}_5), \; 74.4 \; ({\sf C}_6), \; 73.7 \; (-{\sf OCH}_2{\sf Ph}), \; 59.0 \; ({\sf C2}), \\ 56.4 \; (-{\sf OCH}_3), \; 55.9 \; (-{\sf OCH}_3). \; {\sf FT-IR} \; (\tilde{v}_{max}/{\sf cm}): \; 2930 \; ({\sf CH}_2), \; 2893 \; ({\sf CH}_2), \\ 2824 \; (-{\sf OCH}_3), \; 2102 \; ({\sf N}_3), \; 1641 \; ({\sf C=C}), \; 1030 \; ({\sf O-C-O}), \; 920 \; ({\sf O-C-O}), \; 739 \\ (={\sf C-H} \; arom), \; 698 \; (={\sf C-H} \; arom). \; {\sf DI-MS} \; m/z \; ({\sf Rel. Int}): \; 296 \; (1), \; 294 \; (1), \\ 250 \; (1), \; 248 \; (2), \; 194.1 \; (11), \; 91 \; (100). \; {\sf HRMS} \; ({\sf ESI}/{\sf Q-TOF}) \; m/z: \; [{\sf M} \; + \\ {\sf Na}]^+ \; {\sf Calcd} \; for \; [{\sf C}_{17}{\sf H}_2{\sf B}{\sf RN}_3{\sf O}_5{\sf Na}]^+ \; 450.06350, \; found \; 450.06519. \end{array}$

(5S,6S)-3-benzyloxy-2-hydroxy-5,6-methylendioxycyclohexa-2ene-1-one (11).



59 mg of compound 7 (0.17 mmol, 1 eq.) were dissolved in 3 mL of CH₃CN, and the system was cooled to 0 °C. 43 mg of NalO₄ (0.20 mmol, 1.2 eg.) and 2.4 mg of RuCl₃•2H₂O (0.010 mmol, 0.06 eq) were dissolved in 0.5 mL of water on a separated vial, and were also cooled to 0 °C. Once both solutions were at 0°C, the aqueous solution was added all at once over de organic solution, and the reaction was kept at 0°C under vigorous stirring for 5 minutes, and then it was guenched by the addition of 0.5 mL of an agueous solution of Na₂S₂O₃ (20 % W/V). The reaction was filtered through a SiO₂ pad, washed with abundant AcOEt and the eluted was evaporated rendering 50 mg of a brownish oily crude. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 10 % followed by 20 %, 30 % and finally 40 % (v/v of AcOEt in Hexane). 33 mg of 11 were obtained as white solid (67 % yield). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.41 - 7.29 (m, 5H, H_{Pb}), 5.73 (s, 1H, -OCH₂O-), 5.49 (bs, 1H, OH₂), 5.43 (s, 1H, -OCH₂O-), 4.84 (d, J_{gem} = 11.7 Hz, 1H, -OCH₂Ph), 4.80 (d, J₅₋₆ = 8.2 Hz, 1H, H₆), 4.69 (d, J_{gem} = 11.7 Hz, 1H, -OCH₂Ph), 3.99 (ddd, J_{4'-5} = 11.7 Hz, J₅₋₆ = 8.2 Hz, J₄₋₅ = 5.4 Hz, 1H, H₅), 2.92 (dd, $J_{gem} =$ 16.9 Hz, $J_{4-5} =$ 5.4 Hz, 1H, H₄), 2.52 (dd, $J_{\text{qem}} = 16.9 \text{ Hz}$, $J_{4'-5} = 11.7 \text{ Hz}$, 1H, $H_{4'}$). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 190.0 (C₁), 146.7 (C₃), 137.3 (C₂), 128.6 (C_{Ph-m}), 128.1 (C_{Ph-p}), 127.8 (C_{Ph-o}), 126.9 (C_{Ph-quaternary}), 99.9 (OCH₂O), 81.5 (OCH₂Ph), 75.5 (C₆), 72.2 (C₅), 39.2 (C₄). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₁₄H₁₄O₅Na]⁺ 285.0733, found 285.0735.

(2S,3R,4R,5S,6S)-3,4-epoxy-2-hydroxy-5,6-isopropylidenedioxy-1-cyclohexanone (13).^[34]



The same procedure described for compound **11** was applied to synthesize **13**, but the system was left to react for 2 minutes. Purification was accomplished by flash chromatography, using deactivated SiO₂ as stationary phase (deactivation was achieved by adding 10 mL of distilled water to 100 g of SiO₂ and stirring several hours until it is homogeneous), and performing the following gradient elution: 20 % followed by 25 %, 30 %, 35 % and finally 40 % (v/v of AcOEt in Hexane) (*NOTE*: The column should be done as quickly as possible to avoid decomposition of the product, and avoid absorbing the crude in SiO₂; load it in the less possible amount of DCM). White solid. The spectroscopic data of the compound were found to be in agreement with previous reports.^[34] M.P = 125 - 126 °C. $\alpha_D^{21.5} = +77$ (c = 0.60, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 5.11 (d, J_{2-OH2} = 5.0 Hz, 1H, H₂), 4.85 (dt,





$$\begin{split} J_{5-6} &= 6.0 \text{ Hz}, \ J_{4-6} &= J_{3-6} &= 1.4 \text{ Hz}, \ 1\text{H}, \ \text{H}_6\text{)}, \ 4.41 \ (\text{dd}, \ J_{5-6} &= 5.9 \text{ Hz}, \\ J_{4-5} &= 1.5 \text{ Hz}, \ 1\text{H}, \ \text{H}_5\text{)}, \ 3.66 \ (\text{dt}, \ J_{3-4} &= 3.8 \text{ Hz}, \ J_{2-3} &= J_{3-6} &= 1.4 \text{ Hz}, \ 1\text{H}, \\ \text{H}_3\text{)}, \ 3.38 \ (\text{dt}, \ J_{3-4} &= 3.8 \text{ Hz}, \ J_{4-5} &= J_{4-6} &= 1.4 \text{ Hz}, \ 1\text{H}, \ \text{H}_4\text{)}, \ 3.34 \ (\text{d}, \ J &= 5.0 \text{ Hz}, \ 1\text{H}, \ \text{OH}_2\text{)}, \ 1.59 \ (\text{s}, \ 3\text{H}, \ \text{CH}_3\text{)}, \ 1.38 \ (\text{s}, \ 3\text{H}, \ \text{CH}_3\text{)}, \ 1^3\text{C} \text{ NMR} \\ (101 \text{ MHz}, \ \text{CDCl}_3\text{)} \ \delta \ (\text{ppm}\text{)}: \ 202.6 \ (\text{C}_1\text{)}, \ 113.5 \ (\text{C}_{\text{isopropylidene}}\text{)}, \ 78.3 \ (\text{C}_5\text{)}, \\ 77.5 \ (\text{C}_6\text{)}, \ 70.2 \ (\text{C}_2\text{)}, \ 59.7 \ (\text{C}_3\text{)}, \ 54.2 \ (\text{C}_4\text{)}, \ 27.5 \ (\text{CH}_3\text{)}, \ 25.5 \ (\text{CH}_3\text{)}. \text{ FT-IR} \\ (\tilde{v}_{max}/\text{cm}): \ 3507 \ (\text{OH}), \ 2947 \ (\text{CH}_3\text{)}, \ 2820 \ (\text{CH}_3\text{)}, \ 1744 \ (\text{C}=\text{O}), \ 1383 \ (\text{gem dimethyl}), \ 1244 \ (\text{epox}), \ 1219 \ (\text{C-O}), \ 1188 \ (\text{C-O-C-O-C}), \ 1159 \ (\text{C}-\text{O-C-O-C}), \ 1101 \ (\text{C-O-C-O-C}), \ 1069 \ (\text{C-O-C-O-C}). \ \text{DI-MS m/z} \ (\text{Rel. Int}): \ 185 \ (18, \ [\text{M} - \text{Me}]^+), \ 142 \ (2), \ 100 \ (61), \ 85 \ (100), \ 71 \ (81). \ \text{HRMS} \ (\text{ESI}/ \ Q-\text{TOF}) \ \text{m/z}: \ [\text{M} + \text{Na}]^+ \ \text{Calcd for} \ [\text{C}_9\text{H}_12\text{O}_5\text{Na}]^+ \ 223.0577, \ found \ 223.0569. \ \text{C}-9. \$$

(55,65)-2,3-dihydroxy-5,6-isopropylidenedioxycyclohexa-2-ene-1-one (14).^[34]



Structure I could be in equilibrium with structure II. As HMBC experiments shows a long distance coupling of C1-H6 and C1-H6', as well as C₃-H₄, we propose that the main structure in a methanolic solution is II. White solid. The spectroscopic data of the compound were found to be in agreement with previous reports.^[34] ¹H NMR (400 MHz, CD₃OD) δ (ppm): 4.90-4.85 (m-signal partially overlapped with water signal, 1H, H₄), 3.97 (ddd, $J_{5-6'} = 11.6$ Hz, $J_{4-5} = 8.5$ Hz, $J_{5-6} = 5.3$ Hz, 1H, H₅), 2.66 (dd, $J_{\text{gem}} = 16.5$ Hz, $J_{5-6} = 5.3$ Hz, 1H, H₆), 2.44 (dd, J_{gem} = 16.5 Hz, J_{5-6'} = 11.6 Hz, 1H, H_{6'}), 1.64 (s, 3H, CH₃), 1.59 (s, 3H, CH₃). ¹³C NMR (101 MHz, CD₃OD) δ (ppm): 192.5 (C₁), 152.0 (C₃), 128.1 (C₂), 118.5 (C_{isopropylidene}), 82.1 (C₄), 71.1 (C₅), 43.4 (C₆), 26.9 (CH₃), 24.3 (CH₃). FT-IR (v_{max}/cm): 3441 (OH), 2992 (CH₃), 2936 (CH₂), 1771 (enodiol-enone), 1385 (gem dimethyl), 1221 (C-C(O)-C), 1152 (C-O-C-O-C), 1130 (C-O-C-O-C), 1192 (C-O-C-O-C), 1066 (C-O-C-O-C). DI-MS m/z (Rel. Int): 200 (63, [M].+), 142 (99), 114 (58), 71 (100), 68 (99). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₉H₁₂O₅Na]⁺ 223.0577, found 223.0575.

D-1,2-anhydro-5,6-O-isopropylidene-allo-inositol (15).[35]



3.000 g of epoxide 3 (12.15 mmol, 1 eq.) were dissolved in 97.5 mL of CH₃CN, and the system was cooled to 0 °C. 3.117 g of NalO₄ (14.57 mmol, 1.2 eq.) and 88 mg of RuCl₃·2H₂O (0.36 mmol, 0.03 eq.) were dissolved in 16.3 mL of water on a separated vial, and were also cooled to 0 °C. Once both solutions were at 0°C, the aqueous solution was added all at once over de organic solution, and the reaction was kept at 0°C under vigorous stirring for 5 minutes, and then 923 mg of NaBH₄ (24.29 mmol, 2 eq.) were added in small portions. The reaction was kept at 0°C for 10 minutes, and then it was left to stir at room temperature for another 10 minutes. The reaction was filtered through a SiO₂ pad, washed with abundant AcOEt and the eluted was evaporated rendering 2.830 mg of a brownish solid. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 10 % followed by 20 %, 30 %, 35 % and finally 40 % (v/v of AcOEt in Hexane). 1.470 g of 15 were obtained as white solid (60 % yield). $\alpha_D^{21.5} = +22$ (c = 1.41, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 4.58 (dt, $J_{5-6} = 5.8$ Hz, $J_{1-6} = J_{2-6} = 1.3$ Hz, 1H, H₆), 4.51 (ddd, $J_{5-6} = 5.8$ Hz, $J_{4-5} = 3.6$ Hz, $J_{1-5} = 1.3$ Hz, 1H, H₅), 4.25 - 4.19 (m, 1H, H₃), 4.04 (dtd, $J_{4-OH4} = 11.7$ Hz, $J_{4-5} = 3.6$ Hz, J = 3.6 Hz, J = 1.6 Hz, 1H, H₄), 3.53 (dq, $J_{1-2} = 4.0$ Hz, $J_{1-6} = 1.3$ Hz, J = J = 1.3 Hz, 1H, H₁), 3.38 (dt, $J_{1-2} = 4.0$ Hz, J = J = 1.4 Hz, 1H, H₂), 3.10 (d, $J_{3-OH3} = 9.4$ Hz, 1H, OH₃), 2.87 (d, $J_{4-OH4} = 11.8$ Hz, 1H, OH₄), 1.41 (s, 3H, CH₃), 1.34 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 110.1 (C_{isopropylidene}), 77.0 (C₅), 69.8 (C₆), 68.6 (C₄), 64.3 (C₃), 57.9 (C₁), 56.1 (C₂), 27.4 (CH₃), 25.1 (CH₃). FT-IR (\tilde{v}_{max} /cm): 3318 (OH), 3244 (OH), 2988 (C-H), 2945 (C-H), 2909 (C-H), 1385 (gem dimethyl), 1370 (gem dimethyl), 1225 (epox), 1169 (C-O-C-O-C), 1144 (C-O-C-O-C), 1088 (C-O-C-O-C), 1061 (C-O-C-O-C). DI-MS m/z (Rel. Int): 187 (100, [M - Me]⁺), 109 (72), 81 (52), 73 (53). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₉H₁₄O₅Na]⁺: 225.0733, found 225.0774.

(1S,2R,3S,4S)-5-bromo-3,4-isopropylidenedioxycyclohexa-5-en-1,2-diol (16).^[36]



White solid. The ¹H- and ¹³C-NMR spectra data of the compound were in agreement with previous reports.^[36] ¹H NMR (400 MHz, CDCl₃) δ (ppm): 6.26 (d, $J_{1-6} = 2.5$ Hz, 1H, H₆), 4.69 (d, $J_{3-4} = 6.1$ Hz, 1H, H₄), 4.21 (dd, $J_{2-3} = 7.8$ Hz, $J_{3-4} = 6.1$ Hz, 1H, H₃), 4.13 – 4.05 (m, 1H, H₁), 3.75 (t, $J_{1-2} = J_{2-3} = 7.8$ Hz, 1H, H₂), 1.55 (s, 3H, CH₃), 1.43 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 134.3 (C₆), 119.8 (C₅), 111.3 (C_{isopropylidene}), 77.7 (C₄), 77.2 (C₃), 73.3 (C₁), 71.0 (C₂), 28.2 (CH₃), 26.2 (CH₃).

D-1,2-anhydro-3,4-di-O-acetyl-5,6-O-isopropyliden-allo-inositol (17).



24 mg of compound 15 (0.12 mmol, 1 eq) were dissolved in 1.0 mL of CH₂Cl₂ under nitrogen atmosphere, and the system was cooled to 0 °C. Then 132 μ L of Et₃N (0.95 mmol, 8 eq), 45 μ L of Ac₂O (0.48 mmol, 4 eq) and a catalytic amount of 4-dimethylaminopyridine were added at 0°C. The reaction was kept at 0 °C for 10 minutes, then it was warmed up to room temperature and continued to stir during another 10 minutes. A saturated solution of NaHCO3 was added, and the aqueous solution was extracted with three portions of AcOEt. The combined organic layers were washed with three portions of saturated solution of CuSO₄ and one portion of brine, dried with Na₂SO₄, filtered and the solvent was removed under reduced pressure. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and eluting with a solution 20 % (v/v) of AcOEt in Hexane. 31 mg of $\boldsymbol{17}$ were obtained as a colorless oil (91 % yield). $\alpha_D^{21.5}$ = +57 (c = 0.93, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 5.51 (dd, J_{3-4} = 4.6 Hz, J_{2-3} = 2.1 Hz, 1H, H₃), 5.41 (td, $J_{3-4} = J_{4-5} = 4.6$ Hz, $J_{2-4} = 1.0$ Hz, 1H, H₄), 4.53 (d, $J_{5-6} = 5.7$ Hz, 1H, H₆), 4.25 (ddd, $J_{5-6} = 5.7$ Hz, $J_{4-5} = 4.6$ Hz, $J_{1-5} = 4.6$ Hz, $J_$ 1.4 Hz, 1H, H₅), 3.37 (ddt, $J_{1-2} = 4.0$ Hz, $J_{2-3} = 2.1$ Hz, $J_{2-4} = J_{j,2-6} = 3.1$ 3.28 (ddd, J₁₋₂ = 4.0 Hz, J₁₋₅ = 1.4 Hz, J₁₋₆ = 0.8 Hz, 1H), 2.09 (s, 3H, C(O)CH₃), 2.09 (s, 3H, C(O)CH₃), 1.45 (s, 3H, CH₃), 1.34 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 170.3 (OC(O)CH₃), 170.2 (OC(O)CH₃), 110.2 (C_{isopropylidene}), 74.3 (C₅), 70.4 (C₆), 66.6 (C₃), 65.5 (C₄), 53.6 (C₁), 52.1 (C₂), 27.7 (CH₃), 25.5 (CH₃), 21.0 (OC(O)CH₃), 20.9





L-4-azido-4-deoxy-1,2-O-isopropyliden-chiro-inositol (18).



700 mg of epoxide 15 (3.47 mmol, 1 eg) were dissolved in 7 mL of DMF under nitrogen atmosphere. 450 mg of NaN₃ (6.93 mmol, 2 eq) and 371 mg of NH₄Cl (6.93 mmol, 2 eq) were added, and the system was warmed up to 50 °C. The reaction was monitored by TLC, and once the starting material was consumed (approximately 12h) the solid (inorganic salts) was filtered through filter paper, and washed with methanol. The organic solution was evaporated at reduced pressure rendering 1.192 g of a white solid. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 30 % followed by 40 %, 50 %, 60 % and finally 70 % (v/v of AcOEt in Hexane). 739 mg of 18 were obtained as white solid (87 % yield). M.P = 153.2 – 154.1 °C. $\alpha_D^{21.5}$ = –122 (c = 1.27, MeOH). ¹H NMR (400 MHz, CD₃OD) δ (ppm): 4.22 (dd, $J_{1-2} = 5.4$ Hz, $J_{1-6} = 3.0$ Hz, 1H, H₁), 4.10 (t, $J_{1-6} = J_{5-6} = 3.0$ Hz, 1H, H6), 4.04 (dd, $J_{2-3} = 7.4$ Hz, $J_{1-2} = 5.4$ Hz, 1H, H₂), 3.58 (dd, J₄₋₅ = 10.0 Hz, J₅₋₆ = 3.0 Hz, 1H, H₅), 3.52 (t, J₃₋₄ = J₄₋₅ = 10.0 Hz, 1H, H₄), 3.42 (dd, J₃₋₄ = 10.0 Hz, J₂₋₃ = 7.4 Hz, 1H, H₃), 1.44 (s, 3H, CH_3), 1.33 (s, 3H, CH_3). ^{13}C NMR (101 MHz, CD_3OD) δ (ppm): 110.5 (C_{isopropylidene}), 80.5 (C₂), 78.7 (C₁), 76.2 (C₃), 71.7 (C₅), 70.4 (C₆), 66.5 (C₄), 28.4 (CH₃), 26.3 (CH₃). FT-IR (ṽ_{max}/cm): 3350 (OH), 2999 (C-H), 2986 (C-H), 2969 (C-H), 2914 (C-H), 2124 (N₃), 1387 (gem dimethyl), 1223 (C-O-C-O-C), 1063 (C-O-C-O-C). DI-MS m/z (Rel. Int): 230 (100, [M - Me]⁺), 187 (2), 100 (17), 85 (24), 73 (75), 60 (25). HRMS (ESI/Q-TOF) m/z: $[M + Na]^+$ Calcd for $[C_9H_{15}N_3O_5Na]^+$: 268.0904, found 268.0907.

D-5-azido-5-deoxy-1,2-O-isopropyliden-neo-inositol (19).



The same procedure described for compound **15** was applied to synthesize **19**. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase and performing the following gradient elution: 30 % followed by 40 %, 50 %, 60 %, 70 % and finally 80 % (v/v of AcOEt in Hexane). White solid (61 % yield). M.P = 139.7 - 140.5 °C. $\alpha_D^{21.5} = -14$ (c = 1.23, MeOH). ¹H NMR (400 MHz, CD₃OD) δ (ppm): 4.35 (t, $J_{1-2} = J_{2-3} = 4.6$ Hz, 1H, H₂), 4.04 (dd, $J_{1-6} = 7.9$ Hz, $J_{1-2} = 4.6$ Hz, 1H, H₁), 3.94 (ddd, $J_{3-4} = 10.6$ Hz, $J_{2-3} = 4.6$ Hz, $J_{2,3-57} = 0.9$ Hz, 1H, H₃), 3.89 – 3.84 (m, 2H, H₄ + H₅), 3.77 (dd, $J_{1-6} = 7.9$ Hz, $J_{5-6} = 3.0$ Hz, 1H, H₆), 1.48 (s, 3H, CH3), 1.34 (s, 3H, CH3). ¹³C NMR (101 MHz, CD₃OD) δ (ppm): 110.5 (C_{isopropylidene}), 80.3 (C₁), 77.5 (C₂), 72.8 (C₆), 71.1 (C₄), 70.0 (C₃), 69.7 (C₅), 28.7 (CH₃), 26.3 (CH₃). FT-IR (\tilde{v}_{max} /cm): 3399 (OH), 2988 (C-H), 2936 (C-H), 2911 (C-H), 2108 (N₃), 1375 (gem dimethyl), 1219 (C-O), 1163 (C-O-C-O-C), 1076 (C-O-C-O-C), 1053 (C-O-C-O-C). DI-MS m/z (Rel. Int): 230 (80, [M –

 $\begin{array}{l} Me]^+), \ 185 \ (2), \ 100 \ (18), \ 85 \ (26), \ 73 \ (100), \ 60 \ (31). \ HRMS \ (ESI/Q-TOF) \\ m/z: \ [M + Na]^+ \ Calcd \ for \ [C_9H_{15}N_3O_5Na]^+: \ 268.0904, \ found \ 268.0903. \end{array}$

D-2-azido-2-deoxy-5,6-O-isopropyliden-allo-inositol (20).



White solid (8 % yield). M.P = 140.2 - 141.1 °C. $\alpha_D^{21.5} = +23$ (c = 1.55, MeOH). ¹H NMR (400 MHz, CD₃OD) δ (ppm): 4.29 - 4.22 (m, 2H, H₅ + H₆), 4.08 - 4.00 (m, 2H, H₁ + H₃), 3.79 (dd, *J* = 5.0, 3.4 Hz, 1H, H₄), 3.64 (dd, *J* = 3.5, 2.5 Hz, 1H, H₂), 1.45 (s, 3H, CH₃), 1.35 (s, 3H, CH₃). ¹³C NMR (101 MHz, CD₃OD) δ (ppm): 110.2 (C_{isopropylidene}), 79.1 (C_{5or6}), 79.0 (C_{5or6}), 72.9 (C_{1or3or4}), 72.8 (C_{1or3or4}), 72.1 (C_{1or3}), 63.0 (C₂), 28.6 (CH₃), 26.4 (CH₃). FT-IR (\tilde{v}_{max} /cm): 3412 (OH), 2990 (C-H), 2938 (C-H), 2110 (N₃), 1383 (gem dimethyl), 1219 (C-O), 1163 (C-O-C-O-C), 1101 (C-O-C-O-C), 1067 (C-O-C-O-C), 1055 (C-O-C-O-C). DI-MS m/z (Rel. Int): 230 (24, [M - Me]⁺), 160 (15), 142 (9), 100 (61), 85 (33.9), 73 (100), 60 (44). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₉H₁₅N₃O₅Na]⁺: 268.0904, found 268.0904.

D-3,4,6-tri-O-acetyl-5-azido-5-deoxy-1,2-O-isopropyliden-neo-inositol (21).



70 mg of triol 19 (0.29 mmol, 1 eq) were dissolved in 0.5 mL of pyridine under nitrogen atmosphere, and the system was cooled to 0 °C. 162 µL of Ac₂O (1.71 mmol, 6 eq) and a catalytic amount of 4-dimethylaminopyridine were added. The system was left to react during 15 minutes at 0 °C, and was then warmed up to r.t. and left to stir for another 15 minutes. The reaction was guenched by addition of saturated aqueous solution of NH₄Cl, and extracted with three portions of AcOEt. The combined organic layers were washed with four portions of saturated aqueous solution of CuSO₄, one portion of brine, dried with Na2SO4, filtered and evaporated under reduced pressure. 107 mg of a colorless oil were obtained. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase and performing the following gradient elution: 10 % and then 20 % (v/v of AcOEt in Hexane). 103 mg of 21 were obtained as a colourless oil (97 % yield). $\alpha_D^{21.5} = -8$ (c = 0.91, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 5.44 (dd, J₃₋₄ = 10.4 Hz, J₂₋₃ = 4.1 Hz, 1H, H₃), 5.36 (dd, $J_{3-4} = 10.4$ Hz, $J_{4-5} = 3.0$ Hz, 1H, H₄), 5.07 (dd, J_{1-6} = 8.0 Hz, J_{5-6} = 3.0 Hz, 1H, H₆), 4.49 (t, J_{1-2} = J_{2-3} = 4.5 Hz, 1H, H₂), 4.29 (dd, J_{1-6} = 8.0 Hz, J_{1-2} = 4.5 Hz, 1H, H₁), 4.16 (t, J_{4-5} = $J_{5-6} = 3.0 \text{ Hz}, 1\text{H}, \text{H}_5), 2.14 \text{ (s, 3H, OC(O)CH}_3), 2.11 \text{ (s, 3H, OC(O)CH}_3),$ 2.09 (s, 3H, OC(O)CH₃), 1.53 (s, 3H, CH₃), 1.31 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 170.3 (OC(O)CH₃), 170.0 (OC(O)CH₃), 169.7 (OC(O)CH₃), 110.9 (C_{isopropylidene}), 75.6 (C₁), 73.0 (C₂), 71.9 (C₆), 69.0 (C₄), 68.0 (C₃), 61.8 (C₅), 28.0 (CH₃), 26.2 (CH₃), 20.9 (OC(O)CH₃), 20.9 (OC(O)CH_3), 20.7 (OC(O)CH_3). FT-IR ($\tilde{\nu}_{max}/cm)$: 2990 (C-H), 2938 (C-H), 2920 (C-H), 2110 (N₃), 1748 (C=O), 1433 (gem dimethyl), 1373 (gem dimethyl), 1225 (C-O-C-O-C), 1159 (C-O-C-O-C), 1007 (C-O-C-O-C), 1069 (=C-O-C), 1047 (=C-O-C). DI-MS m/z (Rel. Int): 356 (57, [M - Me]⁺), 328 (0.6), 184 (19), 166 (100), 142 (23), 124 (99), 109 (28), 73 (19). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₁₅H₂₁N₃O₈Na]⁺: 394.1221, found 394.1215.





meso-1,2,3,4,6-penta-O-acetyl-5-azido-5-deoxy-neo-inositol (22).



Compound 22 was synthesized from 21 following the same procedure described for the synthesis of compound 6 (but at room temperature). The crude was subjected to esterification following the same procedure described for the synthesis of compound 17. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase and performing the following gradient elution: 20 % and then 30 % (v/v of AcOEt in Hexane). White solid (72 % yield). $\alpha_D^{21.5} = 0$ (c = 0.67, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 5.57 (s, 1H, H₂), 5.30 (bs, 4H, H_{1 +} H_{3 +} H_{4 +} H₆), 4.35 (s, 1H, H₅), 2.14 (s, 3H, CH₃), 2.10 (s, 6H, 2xCH₃), 1.98 (s, 6H, 2xCH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 169.9 (2xAc), 169.9 (Ac), 169.5 (2xAc), 68.7 ($C_1 + C_3$ or $C_4 + C_6$), 68.1 (C_2), 67.3 ($C_4 + C_6$ or $C_1 + C_3$), 60.7 (C5), 20.8 (Ac), 20.7 (2xAc), 20.6 (2xAc). FT-IR (vmax/cm): 2990 (C-H), 2941 (C-H), 2106 (N3), 1748 (C=O), 1372 (C-N), 1225 (C-O), 1076 (= C-O-C), 1045 (=C-O-C). DI-MS m/z (Rel. Int): 356 (3, [M - OAc]⁺), 225 (35), 183 (54), 157 (46), 115 (100), 103 (35), 73 (26). HRMS (ESI/Q-TOF) m/z: $[M + Na]^+$ Calcd for $[C_{16}H_{21}N_3O_{10}Na]^+$: 438.1119, found 438.1138.

D-2-azido-1,3,4-tri-O-benzyl-2-deoxy-5,6-O-isopropyliden-neo-inositol (23).



The same procedure described for the synthesis of compound **5** was followed for synthesizing compound **23**, with the following modifications: 300 mg of compound **19** (1.22 mmol, 1 eq) were dissolved in 2 mL of DMF, 4.5 eq of NaH (55 % mineral oil dispersion) and 5 eq of BnBr were used. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 0 %, then 2 % and finally 5 % (v/v of AcOEt in Hexane). 619 mg of **23** were obtained as a colorless oil (98 % yield).



 $\begin{aligned} &\alpha_{D}^{25.5} = +18 \ (c = 1.16, \ MeOH: AcOEt \ 3:1). \ ^{1}H \ NMR \ (400 \ MHz, \ CDCl_3) \\ &\delta \ (ppm): \ 7.43 \ - \ 7.27 \ (m, \ 15H, \ 3xPh), \ 4.86 \ (d, \ J_{gem} = 12.2 \ Hz, \ 1H, \ OCH_2Ph), \ 4.85 \ (d, \ J_{gem} = 11.9 \ Hz, \ 1H, \ OCH_2Ph), \ 4.76 \ - \ 4.63 \ (m, \ 4H, \ 2xOCH_2Ph), \ 4.85 \ (d, \ J_{gem} = 11.9 \ Hz, \ 1H, \ OCH_2Ph), \ 4.76 \ - \ 4.63 \ (m, \ 4H, \ 2xOCH_2Ph), \ 4.22 \ (t, \ J_{4-5} = J_{5-6} = 4.7 \ Hz, \ 1H, \ H_5), \ 4.17 \ (dd, \ J_{1-6} = 7.6 \ Hz, \ J_{5-6} = 4.7 \ Hz, \ 1H, \ H_5), \ 4.17 \ (dd, \ J_{1-6} = 7.6 \ Hz, \ J_{5-6} = 4.7 \ Hz, \ 1H, \ H_2), \ 3.82 \ (dd, \ J_{3-4} = 9.6 \ Hz, \ J_{2-3} = 3.0 \ Hz, \ 1H, \ H_3), \ 3.45 \ (dd, \ J_{1-6} = 7.6 \ Hz, \ J_{1-2} = 3.0 \ Hz, \ 1H, \ H_1), \ 1.37 \ (s, \ 3H, \ CH_3), \ 1.32 \ (Ph_{quaternary}), \ 137.8 \ (Ph_{quaternary}), \ 128.6 \ (2xC_{Ph}), \ 128.0 \ (2xC_{Ph}), \ 128.0 \ (2xC_{Ph}), \ 127.8 \ (2xC_{Ph}), \ 128.0 \ (2xC_{Ph}), \ 128.0 \ (2xC_{Ph}), \ 127.8 \ (2xC_{Ph}), \ 109.6 \ (C_{isopropylidene}), \ 78.0 \ (C_6), \ 77.6 \ (C_1), \ 77.5 \ (C_3), \ 75.9 \ (C_4), \ 74.4 \ (C_5), \ 73.8 \ (C_{10}, \ 74.4 \ (C_{5}), \ 73.8 \ (C_{10}, \ 74.6 \ (C_{10}$

(OCH₂Ph), 73.7 (OCH₂Ph), 72.0 (OCH₂Ph), 62.5 (C₂), 28.3 (CH₃), 26.1 (CH₃). FT-IR (\tilde{v}_{max} /cm): 3032 (=C-H arom), 2987 (C-H), 2916 (C-H), 2872 (C-H), 2106 (N₃), 1954 (Ph), 1877 (Ph), 1811 (Ph), 1748 (Ph), 1497 (Ph), 1454 (Ph), 1381 (gem dimethyl), 1371 (gem dimethyl), 1219 (C-O-C-O-C), 1155 (C-O-C-O-C), 1094 (C-O-C-O-C), 1076 (C-O-C-O-C), 1051 (C-O-C-O-C), 737 (Ph), 698 (Ph). DI-MS m/z (Rel. Int): 500 (0.1, [M – Me]⁺), 424 (0.6), 396 (2), 106 (6), 91 (100), 65 (4). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₃₀H₃₃N₃O₅Na]⁺: 538.2312, found 538.2305.

D-2-azido-1,3,4-tri-O-benzyl-2-deoxy-5,6-O-methylen-neoinositol (24).



The general procedure for the synthesis of dioxolane under acidic conditions using pTsOH as catalyst was used, but changing DCM for CHCl₃ as solvent, in order to increase de boiling point temperature. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 5 %, then 10 % and finally 20 % (v/v of AcOEt in Hexane). Colorless oil (87 % yield). $\alpha_D^{25.5} = -18$ (c = 1.21, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.34 – 7.27 (m, 15H, Phx3), 5.07 (s, 1H, OCH₂O), 4.97 (s, 1H, OCH₂O), 4.83 (d, J_{gem} = 11.8 Hz, 1H, OCH₂Ph), 4.81 (d, J_{gem} = 11.8 Hz, 1H, OCH₂Ph), 4.77 (d, J_{gem} = 11.8 Hz, 1H, OCH₂Ph), 4.73 (d, J_{gem} = 12.2 Hz, 1H, OCH₂Ph), 4.68 (d, J_{gem} = 11.8 Hz, 1H, OCH₂Ph), 4.65 (d, J_{gem} = 12.2 Hz, 1H, OCH₂Ph), 4.30 (dd, J_{1-6} = 7.8 Hz, J_{6-5} = 4.6 Hz, 1H, H₆), 4.12 - 4.04 (m, 2H, H₄ + H₅), 3.94 (t, $J_{1-2} = J_{2-3} =$ 2.9 Hz, 1H, H₂), 3.80 (dd, J_{3-4} = 9.0 Hz, J_{2-3} = 2.9 Hz, 1H, H₃), 3.39 (dd, J_{1-6} = 7.8 Hz, J_{1-2} = 2.9 Hz, 1H, H₁). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 138.1 (Ph_{quaternary}), 138.0 (Ph_{quaternary}), 137.7 (Ph_{quaternary}), 128.7 (2xC_{Ph}), 128.6 (2xC_{Ph}), 128.5 (2xC_{Ph}), 128.1 (2xC_{Ph}), 128.0 $(1xC_{Ph})$, 128.0 $(1xC_{Ph})$, 127.9 $(3xC_{Ph})$, 127.8 $(2xC_{Ph})$, 95.0 (OCH_2O) , 77.5 (C₃), 77.4 (C₆), 75.9 (C_{1 or 4 or 5}), 75.8 (C_{1 or 4 or 5}), 75.6 (C_{1 or 4 or 5}), 73.6 (OCH₂Ph), 73.6 (OCH₂Ph), 72.2 (OCH₂Ph), 62.5 (C₂). FT-IR (vmax/ cm): 3030 (=C-H arom), 2974 (C-H), 2102 (N₃), 1956 (Ph), 1879 (Ph), 1813 (Ph), 1797 (Ph), 1454 (Ph), 1206 (C-O-C-O-C), 1098 (C-O-C-O-C), 945 (-OCH₂O), 737 (Ph), 696 (Ph). DI-MS m/z (Rel. Int): 458 (0.2), 396 (1, [M - Bn]+), 368 (2), 106 (6), 91 (100), 65 (5). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₂₈H₂₉N₃O₅Na]⁺: 510.1999, found 510.1968.

L-2-amino-2-deoxy-4,5-O-methylen-neo-inositol (25).[12]



213 mg of **24** (0.44 mmol, 1 eq) were dissolved in 8 mL of a solution THF:MeOH (1:1). 21 mg of Pd-C 10 % and 40 μ L of HCl_{cc} (0.48 mmol, 1.1 eq) were added, and hydrogen was bubbled into the system (P=1 atm). Once the reaction was completed (approximately 3 h), nitrogen was bubbled during 10 minutes to remove dissolved H₂, then the catalyst was removed by filtration and washed with a solution MeOH:H₂O (8:2, v/v). The solvent was evaporated under reduced pressure (absolute EtOH was used to remove H₂O by azeotropic distillation) rendering 100 mg of a very hygroscopic white





solid (gum). Pure product was obtained as the hydrochloride, then no purification was needed. Hvdrochloride: ¹H NMR (400 MHz, D₂O, with MeOH as internal reference at $\delta = 3.34 \text{ ppm}^{[37]}$ δ (ppm): 5.23 (s, 1H, OCH₂O), 4.81 (s, 1H, OCH₂O), 4.36-4.30 (m, 2H, H₄ + H₅), 4.21 (dd, $J_{1-6} = 7.9$ Hz, $J_{5-6} = 2.3$ Hz, 1H, H₆), 4.11 (dd, $J_{1-6} = 7.9$ Hz, $J_{1-2} = 6.5$ Hz, 1H, H₁), 4.01 (t, $J_{2-3} = J_{3-4} = 3.3$ Hz, 1H, H₃), 3.78 (dd, $J_{1-2} = 6.5$ Hz, $J_{2-3} = 3.3$ Hz, 1H, H₂). ¹³C NMR (101 MHz, D₂O, with MeOH as internal reference at $\delta = 49.50 \text{ ppm}^{[37]}$) δ (ppm): 95.5 (OCH₂O), 76.8 (C_{4 o 5}), 76.2 (C_{5 o 4}), 70.5 (C₆), 67.7 (C_{1 o 3}), 67.7 (C_{3 o} ₁), 50.9 (C₂). FT-IR (\tilde{v}_{max} /cm): 3402 (OH), 2930 (NH), 1630 (NH₃⁺), 1508 (NH₃⁺), 1080 (C-O), 957 (methylenedioxy). ID-MS m/z (Int. Rel.): 192 (1, M⁺), 191 (3), 173 (6), 126 (9), 101 (26), 88 (100), 73 (65). Free base: an ion exchange chromatography was employed for obtaining the free base. DOWEX 50WX8-100 resin was used as stationary phase, and a gradient elution was applied: first a solution 9:1 (MeOH:NH₄OH_{cc}), followed by a 8:2 solution (MeOH:NH₄OH_{cc}), then a 7:3 solution (MeOH:NH₄OH_{cc}), followed by a 6:4 solution (MeOH:NH₄OH_{cc}), and finally a 5:5 solution (MeOH:NH₄OH_{cc}). Methanol was removed under reduced pressure, and the remaining water was lyophilized rendering the free base aminocyclitol as a hygroscopic white solid. Spectroscopic data was in concordance with previously described values.^{[12] 1}H NMR (400 MHz, D₂O, HDO signal was used as internal reference at δ = 4.79 ppm) δ (ppm): 5.19 (s, 1H, OCH₂O), 4.93 (s, 1H, OCH₂O), 4.28 (dd, J₃₋₄ = 7.5 Hz, J₄₋₅ = 5.1 Hz, 1H, H₄), 4.18 (t, $J_{4-5} = J_{5-6} = 4.7$ Hz, 1H, H₅), 4.13 (dd, $J_{1-6} = 9.6$ Hz, $J_{5-6} = 4.3$ Hz, 1H, H₆), 3.87 (dd, $J_{1-6} = 9.6$ Hz, $J_{1-2} = 3.9$ Hz, 1H, H₁), 3.74 (dd, $J_{3-4} = 7.5$ Hz, $J_{2-3} = 3.5$ Hz, 1H, H₃), 3.40 (t, $J_{1-2} = J_{2-3} =$ 3.7 Hz, 1H, H₂). ¹³C NMR (101 MHz, D₂O, CH₂O signal of EtOH was used as internal reference at $\delta = 58.05 \text{ ppm}^{[37]}$) δ (ppm): 94.9 (OCH2O), 77.7 (C4), 77.5 (C5), 70.3 (C103), 70.2 (C301), 68.8 (C6), 55.1 (C2). FT-IR (vmax/cm): 3370 (OH), 2982 (C-H), 2929 (C-H), 1634 (N-H), 1389 (O-H), 1138 (C-O-C-O-C), 1088 (C-O-C-O-C), 1045 (C-O-C-O-C), 972 (N-H). HRMS (ESI/Q-TOF) m/z: [M + H]⁺ Calcd for [C₇H₁₄NO₅]⁺: 192.0866, found 192.0866.

(1R,2S,5S,6S)-2-azido-4-bromo-5,6-isopropylidenedioxycyclohexa-3-ene-1-ol (26).^[38]



6.008 g of epoxide 3 (24.32 mmol, 1 eq) were dissolved in 175 mL of a solution THF:EtOH:H₂O (3:2:2). 4.743 g of NaN₃ (72.97 mmol, 3 eq) and 2.863 g of NH₄Cl (53.51 mmol, 2.2 eq) were added. The system was heated to reflux temperature and kept stirring at that temperature until consumption of the starting material (approximately 2 h). Water was added, and the system was extracted with three portions of AcOEt. The combined organic layers were washed with brine, dried with Na₂SO₄, filtered and concentrated under reduced pressure, rendering 7 g of a white solid. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 10 %, then 15 % and finally 20 % (v/v of AcOEt in Hexane). 6.597 g of a white solid were obtained (94 % yield). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 6.12 (d, $J_{2-3} = 1.6$ Hz, 1H, H₃), 4.69 (dd, $J_{5-6} = 6.2$ Hz, $J_{2-5} = 1.6$ Hz, 1H, H₅), 4.17 (dd, J_{1-6} = 8.6 Hz, J_{5-6} = 6.2 Hz, 1H, H₆), 3.92 (dt, $J_{1-2} = 8.6$ Hz, $J_{2-3} = J_{2-5} = 1.6$ Hz, 1H, H₂), 3.71 (t, $J_{1-2} = J_{1-6} = 8.6$ Hz, 1H, H₁), 2.84 (bs, 1H, OH₁), 1.56 (s, 3H, CH₃), 1.44 (s, 3H, CH₃). ^{13}C NMR (101 MHz, CDCl_3) δ (ppm): 131.0 (C_3), 120.6 (C_4), 111.4 (Cisopropylidene), 77.9 (C5), 77.2 (C5, signal overlapped with the central peak of CDCl₃), 73.0 (C₄), 62.3 (C₂), 28.3 (CH₃), 26.0 (CH₃). FT-IR $\begin{array}{l} (\tilde{v}_{max}/cm): 3447 \ (OH), \ 2986 \ (C-H), \ 2943 \ (C-H), \ 2880 \ (C-H), \ 2112 \ (N_3), \\ 1377 \ (gem dimethyl), \ 1331 \ (gem dimethyl), \ 1252 \ (O-H), \ 1157 \ (C-O-C-O-C), \ 1084 \ (C-O-C-O-C), \ 1072 \ (C-O-C-O-C). \ DI-MS \ m/z \ (Rel. \ Int): \\ 276 \ (52, \ [M-Me]^+), \ 274 \ (53, \ [M-Me]^+), \ 188 \ (2), \ 101 \ (48), \ 59 \ (35), \ 43 \ (100). \ HRMS \ (ESI/Q-TOF) \ m/z: \ [M+Na]^+ \ Calcd \ for \ [C_9H_{12}BrN_3O_3Na]^+: \\ 311.9954, \ found \ 311.9954. \end{array}$

D-5-azido-5-deoxy-1,2-O-isopropyliden-myo-inositol (27).

The same procedure described for compound 15 was applied for synthesizing 27. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase and performing the following gradient elution: 30 % followed by 40 %, 50 %, 60 %, 70 %, 80 % and finally 90 % (v/v of AcOEt in Hexane). Colourless syrup (75 % yield). $\alpha_D^{25.5} = -52$ (c = 1.54, MeOH). ¹H NMR (400 MHz, (CD₃)₂CO) δ (ppm): 4.74 (d, J_{6-OH6} = 5.0 Hz, 1H, OH₆), 4.58 (d, J_{4-OH4} = 4.2 Hz, 1H, OH₄), 4.38 (dd, $J_{1-2} = 5.2$ Hz, $J_{2-3} = 3.9$ Hz, 1H, H₂), 4.07 - 3.99 (m, 2H, H₁ + OH₃), 3.79 (ddd, J₃₋₄ = 9.3 Hz, J_{3-OH3} = 5.8 Hz, J₂₋₃ = 3.9 Hz, 1H, H₃), 3.63 (td, $J_{3-4} = J_{4-5} =$ 9.3 Hz, $J_{4-OH4} =$ 4.2 Hz, 1H, H₄), 3.58 (ddd, $J_{5-6} = 10.3$ Hz, $J_{1-6} = 7.3$ Hz, $J_{6-OH6} = 5.0$ Hz, 1H, H₆), 3.18 (dd, $J_{5-6} = 10.3$ Hz, $J_{4-5} = 9.3$ Hz, 1H, H₅), 1.41 (s, 3H, CH₃), 1.29 (s, 3H, CH₃). ¹³C NMR (101 MHz, (CD₃)₂CO) δ (ppm): 109.8 (C_{isopropylidene}), 80.6 (C1), 77.1 (C2), 74.8 (C6), 72.6 (C4), 71.8 (C3), 68.9 (C5), 28.4 (CH3), 26.1 (CH₃). FT-IR ($\tilde{\nu}_{max}$ /cm): 3383 (OH), 2990 (C-H), 2938 (C-H), 2922 (C-H), 2110 (N₃), 1377 (O-H), 1221 (C-O), 1146 (C-O-C-O-C), 1099 (C-O-C-O-C), 1042 (C-O-C-O-C). DI-MS m/z (Rel. Int): 230 (100, [M -Me]+), 109 (13), 96 (12), 85 (22), 73 (88), 60 (35). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₉H₁₅N₃O₅Na]⁺: 268.0904, found 268.0912.

D-5-azido-3,4,6-tri-O-benzyl-5-deoxy-1,2-O-isopropyliden-myo-inositol (28).



The same procedure described for the synthesis of compound 23 was followed for synthesizing compound 28. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 0 %, then 2 %, 5 % and finally 10 % (v/v of AcOEt in Hexane). Colorless oil (74 % yield). $\alpha_D^{25.5} = +19$ (c = 1.37, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.44 – 7.27 (m, 15H, 3xPh), 4.91 – 4.70 (m, 6H, 3xOCH₂Ph), 4.26 (dd, J₁₋₂ = 5.5 Hz, J₂₋₃ = 3.6 Hz, 1H, H₂), 4.10 (dd, J₁₋₆ = 6.9 Hz, J_{1-2} = 5.5 Hz, 1H, H₁), 3.74 (t, J_{3-4} = J_{4-5} = 8.9 Hz, 1H, H₄), 3.68 (dd, $J_{3-4} = 8.9$ Hz, $J_{2-3} = 3.6$ Hz, 1H, H₃), 3.56 (dd, $J_{5-6} = 10.3$ Hz, $J_{1-6} =$ 6.9 Hz, 1H, H₆), 3.34 (dd, $J_{5-6} = 10.3$ Hz, $J_{4-5} = 8.9$ Hz, 1H, H₅), 1.51 (s, 3H, CH₃), 1.36 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 138.0 (Ph_{guaternary}), 138.0 (2xPh_{guaternary}), 128.6 (2xPh), 128.6 (2xPh), 128.5 (2xPh), 128.4 (2xPh), 128.2 (2xPh), 128.2 (2xPh), 128.1 (Ph), 128.0 (Ph), 127.9 (Ph), 110.2 (C_{isopropylidene}), 80.9 (C₆), 79.4 (C_{1 or 4}), 79.3 (C_{4 or 1}), 77.4 (C₃), 75.6 (OCH₂Ph), 74.6 (C₁), 73.7 (OCH₂Ph), 73.4 (OCH₂Ph), 65.9 (C₅), 28.0 (CH₃), 25.9 (CH₃). FT-IR (\tilde{v}_{max} /cm): 3063 (= C-H arom), 3032 (=C-H arom), 2988 (C-H), 2911 (C-H), 2874 (C-H), 2106 (N₃), 1952 (Ph), 1877 (Ph), 1811 (Ph), 1497 (Ph), 1454 (Ph), 1371 (gem dimethyl), 1265 (C-O), 1242 (C-O-C-O-C), 1219 (C-O-C-O-C), 1153 (C-O-C-O-C), 1072 (C-O-C-O-C), 1047 (C-O-C-O-C), 737 (Ph), 696 (Ph). DI-MS m/z (Rel. Int): 500 (0.3, [M - Me]⁺), 424 (0.4), 290 (1), 181 (2), 106 (6), 91 (100), 65 (4). HRMS (ESI/Q-TOF) m/z: [M + Na]⁺ Calcd for [C₃₀H₃₃N₃O₅Na]⁺: 538.2312, found 538.2278.





D-5-azido-3,4,6-tri-O-benzyl-5-deoxy-1,2-O-methylen-myoinositol (29).



The general procedure for the synthesis of dioxolane under acidic conditions using pTsOH as catalyst was used, but changing DCM for CHCl₃ as solvent, in order to increase de boiling point temperature. Purification was accomplished by flash chromatography, using SiO₂ as stationary phase, and performing the following gradient elution: 5 %, then 10 % (v/v of AcOEt in Hexane). Colorless oil (89 % yield). $\alpha_D^{25.5} = +10$ (c = 0.73, MeOH). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.42 - 7.26 (m, 15H, 3xPh), 5.18 (s, 1H, OCH₂O), 4.99 (s, 1H, OCH₂O), 4.86 (d, J_{aem} = 11.3 Hz, 1H, OCH₂Ph), 4.82 - 4.69 (m, 5H, OCH₂Ph), 4.20 - 4.12 (m, 2H, H₁ + H₂), 3.75 (dd, J_{3-4} = 8.3 Hz, J_{2-3} = 3.2 Hz, 1H, H₃), 3.69 (t, $J_{3-4} = J_{4-5} = 8.3$ Hz, 1H, H₄), 3.61 – 3.50 (m, 1H, H₆), 3.39 (dd, $J_{5-6} = 10.3$ Hz, $J_{4-5} = 8.3$ Hz, 1H, H₅). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 137.9 (2xPh_{quaternary}), 137.8 (Ph_{quaternary}), 128.6 (2xPh), 128.6 (2xPh), 128.5 (2xPh), 128.4 (2xPh), 128.3 (2xPh), 128.1 (Ph), 128.1 (Ph), 128.0 (2xPh), 128.0 (Ph), 95.5 (OCH2O), 79.3 (C4), 79.0 (C₆), 78.9 (C_{1 or 2}), 77.3 (C₃), 75.3 (C_{2 or 1}), 75.2 (OCH₂Ph), 73.9 (OCH₂Ph), 73.2 (OCH₂Ph), 65.8 (C₅). FT-IR (ṽ_{max}/cm): 3030 (=C-H arom), 2876 (C-H), 2106 (N₃), 1954 (Ph), 1877 (Ph), 1811 (Ph), 1497 (Ph), 1454 (Ph), 1207 (C-O-C-O-C), 1094 (C-O-C-O-C), 920 (-OCH2O-), 737 (Ph), 698 (Ph). DI-MS m/z (Rel. Int): 458 (0.2), 396 (0.6, [M - Bn]+), 262 (2), 106 (7), 91 (100), 65 (5). HRMS (ESI/Q-TOF) m/z: $[M + Na]^+$ Calcd for $[C_{28}H_{29}N_3O_5Na]^+$: 510.1999, found 510,1966.

D-5-amino-5-deoxy-1,2-O-methylen-myo-inositol (30).



The same procedure described for compound **25** was applied for synthesizing **30**. Colorless syrup (85 % yield). Hydrochloride: $\alpha_D^{25.5} = -42$ (c = 1.41, MeOH). ¹H NMR (400 MHz, D₂O, HDO signal was used as internal reference at $\delta = 4.79$ ppm) δ (ppm): 5.22 (s, 1H, OCH₂O), 4.98 (s, 1H, OCH₂O), 4.26 - 4.13 (m, 2H, H₁ and H₂), 3.95 - 3.86 (m, 1H, H₃), 3.71 (t, J₃₋₄ = J₄₋₅ = 9.9 Hz, 1H, H₄), 3.65 - 3.55 (m, 1H, H₆), 2.96 (t, J₄₋₅ = J₅₋₆ = 9.9 Hz, 1H, H₅). ¹³C NMR (101 MHz, D₂O, CH₃ signal of EtOH was used as internal reference at $\delta = 17.47$ ppm) δ (ppm): 95.4 (OCH₂O), 78.7 (C₁), 77.9 (C₂), 70.9 (C₃), 70.4 (C₆), 70.1 (C₄), 55.3 (C₅). FT-IR (\tilde{v}_{max} /cm): 3404 (OH), 2922 (CH₂), 2851 (C-H), 1636 (N-H), 1240 (C-N), 1165 (C-O-C-O-C), 1069 (C-O-C-O-C), 924 (-OCH₂O-). DI-MS m/z (Rel. Int): 191 (2, [M]⁻⁺), 173 (4), 126 (3), 113 (6), 101 (20), 88 (100) 72 (38), 60 (31). HRMS (ESI/Q-TOF) m/z: [M + H]⁺ Calcd for [C₇H₁₄NO₅]⁺: 192.0866, found 192.0866.

Acknowledgments

The authors are grateful to PEDECIBA and CSIC-UdelaR for support of this project. GC acknowledges Agencia Nacional de Investigación e Innovación (ANII) and Comisión Académica de Posgrado (CAP) for a PhD fellowship. The authors also acknowledge Prof. Alejandra Rodriguez for HRMS experiments. **Keywords:** Cyclitols · Biocatalysis · Amino alcohols · Dihydroxylation · Asymmetric synthesis

- [1] R. L. Mann, R. M. Gale, R. F. van Abeele, Antibiot. Chemother. 1953, 3, 1279–1282.
- [2] M. del Carmen Guerrero, J. Modolell, Eur. J. Biochem. 1980, 107, 409-414.
- [3] R. C. Pittenger, R. N. Wolfe, M. M. Hoehn, P. N. Marks, W. A. Daily, J. M. McGuire, Antibiot. Chemother. 1953, 3, 1268–1278.
- [4] S. Omura, A. Nakagawa, T. Fujimoto, K. Saito, K. Otoguro, J. C. Walsh, J. Antibiot. 1987, 40, 1619–1626.
- [5] A. Nakagwa, T. Fujimoto, S. Omura, J. C. Walsh, R. L. Stotish, B. George, J. Antibiot. 1987, 40, 1627–1635.
- [6] S. F. Hayashi, L. J. L. Norcia, S. B. Seibel, A. M. Silvia, J. Antibiot. 1997, 50, 514–521.
- [7] N. Chida, M. Ohtsuka, K. Nakazawa, S. Ogawa, J. Org. Chem. 1991, 56, 2976–2983.
- [8] a) T. J. Donohoe, A. Flores, C. J. R. Bataille, F. Churruca, Angew. Chem. Int. Ed. 2009, 48, 6507–6510; Angew. Chem. 2009, 121, 6629; b) T. J. Donohoe, A. Flores, C. J. R. Bataille, F. Churruca, Angew. Chem. Int. Ed. 2009, 48, 6507; Angew. Chem. 2009, 121, 6629–6632.
- [9] H.-J. Lo, Y.-K. Chang, T.-H. Yan, Org. Lett. 2012, 14, 5896–5899.
- [10] B. M. Trost, J. Dudash Jr., O. Dirat, Chem. Eur. J. 2002, 8, 259-268.
- [11] N. Palaniappan, S. Ayers, S. Gupta, E.-S. Habib, K. A. Reynolds, *Chem. Biol.* 2006, 13, 753–764.
- [12] B. P. Gurale, M. S. Shashidhar, R. G. Gonnade, J. Org. Chem. 2012, 77, 5801–5807.
- [13] N. Chida, M. Ohtsuka, K. Nakazawa, S. Ogawa, J. Chem. Soc., Chem. Commun. 1989, 436–438.
- [14] a) B. M. Trost, O. Dirat, J. Dudash Jr., E. J. Hembre, Angew. Chem. Int. Ed. 2001, 40, 3658–3660; Angew. Chem. 2001, 113, 3770; b) B. M. Trost, O. Dirat, J. Dudash, E. J. Hembre, Angew. Chem. Int. Ed. 2001, 40, 3658; Angew. Chem. 2001, 113, 3770–3772.
- [15] T. J. Donohoe, P. D. Johnson, R. J. Pye, M. Keenan, Org. Lett. 2005, 7, 1275–1277.
- [16] a) A. Bellomo, D. Gonzalez, *Tetrahedron Lett.* **2007**, *48*, 3047–3051; b) N. Thevenet, V. de la Sovera, M. A. Vila, N. Veiga, D. Gonzalez, G. Seoane, I. Carrera, *Org. Lett.* **2015**, *17*, 684–687.
- [17] C. Coronel, G. Arce, C. Iglesias, C. M. Noguera, P. R. Bonnecarrère, S. R. Giordano, D. Gonzalez, J. Mol. Catal. B 2014, 102, 94–98.
- [18] a) V. de la Sovera, P. Garay, N. Thevenet, M. A. Macias, D. Gonzalez, G. Seoane, I. Carrera, *Tetrahedron Lett.* **2017**, *57*, 2484–2487; b) J. A. Griffen, J. C. White, G. Kociok-Köhn, M. D. Lloyd, A. Wells, T. C. Arnot, S. E. Lewis, *Tetrahedron* **2013**, *69*, 5989–5997; c) S. Pilgrim, G. Kociok-Köhn, M. D. Lloyd, S. E. Lewis, *Chem. Commun.* **2011**, *47*, 4799–4801; d) B. J. Paul, J. Willis, T. A. Martinot, I. Ghiviriga, K. A. Abboud, T. Hudlický, J. Am. Chem. Soc. **2002**, *124*, 10416–10426.
- [19] a) M. Pazos, B. Gonzalez, L. Suescun, G. Seoane, I. Carrera, *Tetrahedron Lett.* **2017**, *58*, 2182–2185; b) C. Vitelio, A. Bellomo, M. Brovetto, G. Seoane, D. Gonzalez, *Carbohydr. Res.* **2004**, *339*, 1773–1778; c) A. Bellomo, C. Giacomini, B. Brena, G. Seoane, D. Gonzalez, *Synth. Commun.* **2007**, *37*, 3509.
- [20] a) A. Bellomo, J. B. Bonilla, J. López-Prados, M. Martín-Lomas, D. Gonzalez, *Tetrahedron: Asymmetry* **2009**, *20*, 2061–2064; b) G. Carrau, N. Veiga, L. Suescun, G. F. Giri, A. G. Suárez, R. Spanevello, D. González, *Tetrahedron Lett.* **2016**, *57*, 4791–4794.
- [21] a) K. A. Oppong, T. Hudlicky, F. Yan, C. York, B. V. Nguyen, *Tetrahedron* 1999, 55, 2875–2880; b) T. Hudlicky, J. Rouden, H. Luna, S. Allen, *J. Am. Chem. Soc.* 1994, *116*, 5099–5107; c) T. Hudlicky, H. F. Olivo, *Tetrahedron Lett.* 1991, *32*, 6077–6080.
- [22] a) V. de la Sovera, A. Bellomo, D. Gonzalez, *Tetrahedron Lett.* 2011, *51*, 430–433; b) V. de la Sovera, L. Suescun, A. Bellomo, D. Gonzalez, *Eur. J. Org. Chem.* 2017, 2017, 3912–3916.
- [23] G. Carrau, C. C. Drewes, A. L. B. Shimada, A. Bertucci, S. H. P. Farsky, H. A. Stefani, D. Gonzalez, *Bioorg. Med. Chem.* **2013**, *21*, 4225–4232.
- [24] M. A. Vila, M. Brovetto, D. Gamenara, P. Bracco, G. Zinola, G. Seoane, S. Rodríguez, I. Carrera, J. Mol. Catal. B 2013, 96, 14–20.
- [25] T. C. Nugent, T. Hudlicky, J. Org. Chem. 1998, 63, 510-520.
- [26] J. S. Bajwa, R. C. Anderson, Tetrahedron Lett. 1991, 32, 3021–3024.





- [27] K. Stott, J. Keeler, Q. N. Van, A. J. Shaka, J. Magn. Reson. 1997, 125, 302– 324.
- [28] L. Krause, R. Herbst-Irmer, G. M. Sheldrick, D. Stalke, J. Appl. Crystallogr. 2015, 48, 3–10.
- [29] G. Sheldrick, Acta Crystallogr., Sect. A 2015, 71, 3-8.
- [30] G. Sheldrick, Acta Crystallogr., Sect. C 2015, 71, 3-8.
- [31] C. B. Hubschle, G. M. Sheldrick, B. Dittrich, J. Appl. Crystallogr. 2011, 44, 1281–1284.
- [32] T. Hudlicky, F. Rulin, T. Tsunoda, H. Luna, C. Andersen, J. D. Price, Isr. J. Chem. 1991, 31, 229–238.
- [33] B. A. Johns, Y. T. Pan, A. D. Elbein, C. R. Johnson, J. Am. Chem. Soc. 1997, 119, 4856–4865.
- [34] T. Hudlicky, M. Mandel, J. Rouden, R. S. Lee, B. Bachmann, T. Dudding, K. J. Yost, J. S. Merola, J. Chem. Soc., Perkin Trans. 1 1994, 1553–1567.

- [35] B. Ananthan, W.-C. Chang, J.-S. Lin, P.-H. Li, T.-H. Yan, J. Org. Chem. 2014, 79, 2898–2905.
- [36] T. Hudlicky, H. Luna, H. F. Olivo, C. Andersen, T. Nugent, J. D. Price, J. Chem. Soc., Perkin Trans. 1 1991, 2907–2917.
- [37] G. R. Fulmer, A. J. M. Miller, N. H. Sherden, H. E. Gottlieb, A. Nudelman, B. M. Stoltz, J. E. Bercaw, K. I. Goldberg, *Organometallics* **2010**, *29*, 2176– 2179.
- [38] A. Bellomo, D. Gonzalez, H. A. Stefani, J. Organomet. Chem. 2008, 693, 1136–1142.

Received: September 24, 2018





Aminocyclitol Synthesis

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 Chemoenzymatic Synthesis of Hygromycin Aminocyclitol Moiety and its C2 Epimer



This article describes the synthesis of hygomycin A aminocyclitol moiety and its C2 epimer from bromobezene by a chemoenzymatic strategy. The sequence involved epoxidation, stereocontrolled oxirane ring opening, dihydroxylation of an electron-poor olefin, and introduction of a methylenedioxy group. The aminocyclitols were obtained in high chemical and optical purity.

DOI: 10.1002/ejoc.201801424