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Photocytotoxicity of Oligothiophenyl-functionalized Chelates that Sensitize Ln<sup>III</sup> Luminescence and Generate <sup>1</sup>O<sub>2</sub>Katherine R. Johnson,<sup>[a]</sup> Vincent C. Lombardi,<sup>[b]</sup> and Ana de Bettencourt-Dias\*<sup>[a]</sup>

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**Abstract:** Three new compounds containing a heptadentate lanthanide (Ln<sup>III</sup>) ion chelator functionalized with oligothiophenes, **n**Thept(COOH)<sub>4</sub> (*n* = 1, 2, or 3), were isolated. Their Ln<sup>III</sup> complexes not only display the characteristic metal-centred emission in the visible or near-infrared (NIR) but also generate singlet oxygen (<sup>1</sup>O<sub>2</sub>). Luminescence efficiencies ( $\phi^{Ln}$ ) for [Eu**1**Thept(COO)<sub>4</sub>]<sup>-</sup> and [Eu**2**Thept(COO)<sub>4</sub>]<sup>-</sup> are  $\phi^{Eu}$  = 3% and 0.5% in TRIS buffer and 33% and 3% in 95% ethanol, respectively. **3**Thept(COO)<sub>4</sub><sup>4-</sup> does not sensitize Eu<sup>III</sup> emission due to its low-lying triplet state. NIR luminescence is observed for all NIR-emitting Ln<sup>III</sup> and ligands with efficiencies of  $\phi^{Yb}$  = 0.002%, 0.005% and 0.04% for [Yb**n**Thept(COO)<sub>4</sub>]<sup>-</sup> (*n* = 1, 2, or 3), and  $\phi^{Nd}$  = 0.0007%, 0.002% and 0.02% for [Nd**n**Thept(COO)<sub>4</sub>]<sup>-</sup> (*n* = 1, 2, or 3) in TRIS buffer. In 95% ethanol, quantum yields of NIR luminescence increase and are  $\phi^{Yb}$  = 0.5%, 0.31% and 0.05% for [Yb**n**Thept(COO)<sub>4</sub>]<sup>-</sup> (*n* = 1, 2, or 3), and  $\phi^{Nd}$  = 0.40%, 0.45% and 0.12% for [Nd**n**Thept(COO)<sub>4</sub>]<sup>-</sup> (*n* = 1, 2, or 3). All complexes are capable of generating <sup>1</sup>O<sub>2</sub> in 95% ethanol with  $\phi_{1O2}$  efficiencies which range from 2% to 29%. These complexes are toxic to HeLa cells when irradiated with UV light ( $\lambda_{exc}$  = 365 nm) for 2 minutes. IC<sub>50</sub> values for the Ln<sup>III</sup> complexes are in the range 15.2 – 16.2  $\mu$ M; the most potent compound is [Nd**2**Thept(COO)<sub>4</sub>]<sup>-</sup>. The cell death mechanisms are further explored using an Annexin V – propidium iodide assay which suggests that cell death occurs through both apoptosis and necrosis.

performance.<sup>[7]</sup> To circumvent these issues, there is a fundamental interest in the development of new types of <sup>1</sup>O<sub>2</sub> generators that are just as effective.<sup>[8]</sup> Ideal photosensitizers for PDT need to be chemically pure, photostable, and effective at low doses for advanced cancer therapy. Compounds with additional properties, such as luminescence for *in situ* or *in vivo* imaging, have an additional advantage.<sup>[9]</sup>

Lanthanide ion (Ln<sup>III</sup>) luminescence is highly desirable for imaging purposes due to characteristic emission wavelengths, large pseudo-Stokes shifts of sensitized emission, and long decay lifetimes. The latter enable time-gated detection of the emission resulting in improved signal-to-noise ratios.<sup>[10]</sup> The metal-centred emission is based on transitions within the 4f orbitals. These intraconfigurational transitions are parity-forbidden, which poses a challenge for direct excitation. To overcome this, coordinated ligands are used to transfer energy to the Ln<sup>III</sup>. This sensitization process is referred to as the antenna effect (Figure 1).<sup>[10]</sup>

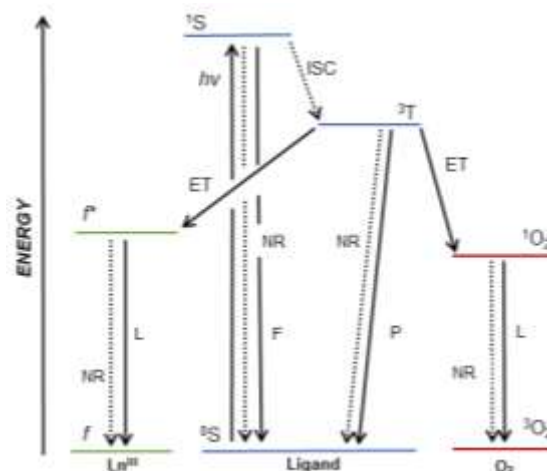


Figure 1. Energy level diagram showing the energy transfer (ET) pathways for both Ln<sup>III</sup> sensitization and <sup>1</sup>O<sub>2</sub> generation. Energy *hν* is absorbed by the ligand to populate a singlet excited state (<sup>1</sup>S). Intersystem crossing (ISC) leads to population of a triplet excited state (<sup>3</sup>T). This state can then transfer energy to populate the emissive <sup>1</sup>\* excited state which decays by luminescence (L) to the ground state, f. Alternatively, the energy transfer leads to <sup>1</sup>O<sub>2</sub> generation, which

## Introduction

Singlet oxygen (<sup>1</sup>O<sub>2</sub>) is a reactive oxygen species directly linked to cellular death and tissue damage.<sup>[1]</sup> Photosensitizers that efficiently generate <sup>1</sup>O<sub>2</sub> are useful in photodynamic therapy (PDT).<sup>[2]</sup> [3] Porfimer sodium, or Photofrin, was approved by the U.S. Food and Drug Administration in 1993 for the treatment of bladder cancer and has since been approved for several other cancers, including endobronchial and esophageal cancers.<sup>[4]</sup> Most compounds studied for PDT applications are porphyrin-based. However, they suffer from many drawbacks, including aggregation, which shortens the lifetime of the excited state resulting in lower <sup>1</sup>O<sub>2</sub> generation quantum yields,<sup>[5]</sup> and poor solubility.<sup>[6]</sup> These drawbacks limit dosage concentration and

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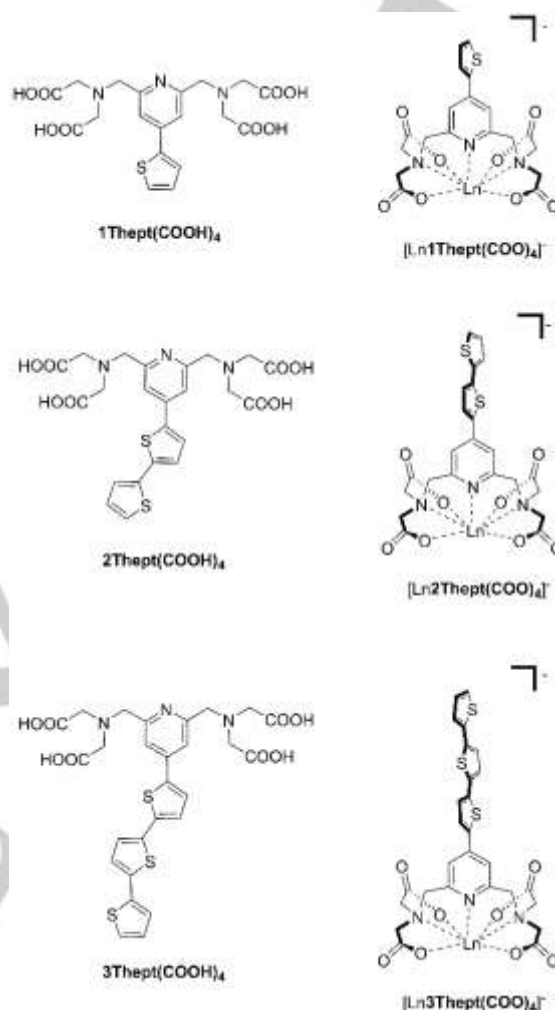
decays to triplet oxygen  $^3\text{O}_2$  by emitting at 1270 nm. Nonradiative (NR, dash-dot lines) pathways lead to quenching of excited states. Competing radiative processes are fluorescence (F) and phosphorescence (P). Energy levels are not drawn to scale.

$\text{Ln}^{\text{III}}$ -based systems work well as multifunctional platforms with imaging capabilities.<sup>[11],[12],[13]</sup> Patra and coworkers described  $\text{Eu}^{\text{III}}$  and  $\text{Tb}^{\text{III}}$  complexes with dipyrro[3,2-d:2',3'-f]quinoxaline (dbq) and 4,4,4-trifluoro-1-(2-naphthyl)-1,3-butanedione (tfnb) sensitizers. These complexes penetrate the cell membrane of H460 and MCF-7 cells. The  $\text{IC}_{50}$  value of the most phototoxic complex,  $[\text{Tb}(\text{dpq})(\text{tfnb})_3]$ , was reported as  $7.94 \pm 0.65 \mu\text{M}$  (H460 cells).<sup>[14]</sup> Although the authors mention that the cells remained viable in the presence of the complexes under dark conditions,  $\text{IC}_{50}$  values for dark toxicity were not reported. A  $\text{Tb}^{\text{III}}$  DOTA-based complex that displays a green  $\text{Tb}^{\text{III}}$ -centered emission with an emission efficiency ( $\phi^{\text{Tb}}$ ) of 24% and  $^1\text{O}_2$  generation efficiency ( $\phi_{1\text{O}_2}$ ) of 12% was used to image NIH-3T3 cells.<sup>[15]</sup> While  $\phi^{\text{Tb}}$  is high,  $\phi_{1\text{O}_2}$  falls well below the efficiencies of naturally occurring organic photosensitizers.<sup>[8]</sup> Ung, Gasser, and coworkers described  $\text{Tb}^{\text{III}}$  complexes and their photocytotoxicity against HeLa and MRC-5 cells. After treatment, the cell morphology of HeLa cells changed, which, coupled with cell viability studies, indicates compound toxicity.  $\text{IC}_{50}$  values were reported between 14.3 and 32.3  $\mu\text{M}$  (HeLa cells).<sup>[16]</sup> An  $\text{Er}^{\text{III}}$  porphyrin-based complex that displays  $\text{Er}^{\text{III}}$ -centred emission in the near-infrared and  $^1\text{O}_2$  generation was used to image HeLa cells and shown to localize in the cell mitochondria.<sup>[17]</sup> Although this complex displayed photocytotoxicity, the reported  $\phi_{1\text{O}_2}$  of 10% is much lower than reported efficiencies of other photosensitizers.<sup>[8]</sup>

Our group has recently described naphthalimide-based complexes that sensitize visible ( $\text{Eu}^{\text{III}}$ ) and near-infrared ( $\text{Yb}^{\text{III}}$ ,  $\text{Nd}^{\text{III}}$ ) emitting ions in addition to generating  $^1\text{O}_2$  with  $\phi_{1\text{O}_2}$  in the range 41 – 64%.<sup>[18]</sup> However, these compounds are not water-soluble, and thus, not useful for imaging and therapy in biological systems. Bithiophene and terthiophene are biologically active natural products present in a variety of plants. Their toxicity and ability to generate  $^1\text{O}_2$  with high  $\phi_{1\text{O}_2}$  have been well documented.<sup>[19]</sup> Oligothiophene-based systems have shown promise in photodynamic therapy for the treatment of bladder carcinomas<sup>[20]</sup> and fibrosarcoma cells.<sup>[21]</sup> Our group recently described terthiophene-based, luminescent  $\text{Ln}^{\text{III}}$  complexes with wavelength-dependent  $^1\text{O}_2$  generation.<sup>[22]</sup> Thus, we aimed to isolate  $\text{Ln}^{\text{III}}$  complexes based on three different oligothiophenes that display luminescence and generate  $^1\text{O}_2$  in aqueous systems.

As  $\text{Ln}^{\text{III}}$  can be toxic to living systems,<sup>[23]</sup> chelators with high complex stability are used and we selected 2,2',2'',2'''-[(2,6-pyridinediylbis(methylenenitrilo)]tetraacetic acid. This compound is known for its stability in aqueous solution with a  $\text{pGd}$  value, which is  $-\log[\text{concentration}]$  of free  $\text{Gd}^{\text{III}}$  in solution in the presence of the chelator, of 17.5, which is comparable to other metal chelators such as diethylenetriamine pentaacetate under the same conditions ( $\text{pGd} = 19.1$ ).<sup>[24]</sup> We report the isolation of oligothiophenyl-derivatized of 2,2',2'',2'''-[(2,6-pyridinediylbis(methylenenitrilo)]tetraacetic acid-based photosensitizers,  $n\text{Thept}(\text{COOH})_4$  (Scheme 1), and the luminescence and cytotoxic properties of their  $\text{Ln}^{\text{III}}$  complexes ( $\text{Ln}^{\text{III}} = \text{Eu}^{\text{III}}$ ,  $\text{Yb}^{\text{III}}$ , or

$\text{Nd}^{\text{III}}$ ), along with the mechanism of cell death. These complexes are water-soluble and show  $\text{Ln}^{\text{III}}$ -centred emission in either the visible ( $\text{Eu}^{\text{III}}$ ) or near infrared (NIR) range ( $\text{Yb}^{\text{III}}$  or  $\text{Nd}^{\text{III}}$ ) while generating  $^1\text{O}_2$ .



Scheme 1. The compounds  $n\text{Thept}(\text{COOH})_4$  and  $[\text{Ln}n\text{Thept}(\text{COO})_4]^-$  ( $n = 1, 2$ , or  $3$ ) studied here. The coordination sphere of the  $\text{Ln}^{\text{III}}$  ions is completed by molecules that are not shown (see text).

## Results and Discussion

$n\text{Thept}(\text{COOH})_4$  ( $n = 1, 2$ , or  $3$ ; Scheme 1) were synthesized through a Pd-catalyzed Suzuki-Miyaura cross-coupling between 4-bromo-2,6-bis[N,N-bis(ethoxycarbonylmethyl)aminomethyl]pyridine and the respective borolanes to yield the diethyl ester derivatives in 33 – 56% yield (Figure S1 Supporting Information). The esters were saponified to yield the water-soluble compounds,  $n\text{Thept}(\text{COOH})_4$  ( $n = 1, 2$ , or  $3$ ), in 90 – 94% yield. Their isolation was confirmed by NMR spectroscopy and mass spectrometry (Figures S2 – S10).

$\text{Ln}^{\text{III}}$  complexes were prepared by deprotonating the ligands with  $\text{K}_2\text{CO}_3$  and mixing in a 1:1 stoichiometry with the  $\text{LnCl}_3$  salt ( $\text{Ln}^{\text{III}} = \text{Gd}^{\text{III}}$ ,  $\text{Eu}^{\text{III}}$ ,  $\text{Yb}^{\text{III}}$ , or  $\text{Nd}^{\text{III}}$ ) in 1:1 water: methanol and heated

## FULL PAPER

to 60 °C. After solvent evaporation, the complexes were recovered as white, yellow, and brown salts for  $K[Ln1Thept(COO)_4]$ ,  $K[Ln2Thept(COO)_4]$ , and  $K[Ln3Thept(COO)_4]$ , respectively, in 89 – 96% yield. The complexes were characterized using mass spectrometry (Figures S14 – S24). For spectroscopy, the  $Ln^{III}$  complexes were prepared in solution and not isolated.

Speciation studies of  $1Thept(COO)_4^{4-}$ ,  $2Thept(COO)_4^{4-}$ , and  $3Thept(COO)_4^{4-}$  with  $LnCl_3$  ( $Ln^{III} = Yb^{III}$  or  $Eu^{III}$ ) (Figures S11 – S13) confirm the formation of 1:1 complexes, as expected.<sup>[24-25]</sup>

**Table 1.** Quantum yields of fluorescence ( $\phi^F$ ) and of  $Ln^{III}$  luminescence ( $\phi^{Ln}$ ) for  $nThept(COO)_4^{4-}$  ( $n = 1, 2$ , or  $3$ ) and their  $Ln^{III}$  complexes, and luminescence lifetime ( $\tau^{Eu}$ ), intrinsic quantum yield ( $\phi^{Eu}$ ), and sensitization efficiency ( $\eta_{sens}$ ), for  $[Eu1Thept(COO)_4]^-$  measured at  $25.0 \pm 0.1$  °C.

	solvent	$\phi^F$ no $Ln^{III}$ (%)	$\phi^F$ $Ln^{III} = Gd^{III}$ (%)	$\phi^{Yb}$ (%)	$\tau^{Yb}$ (ms)	$\phi^{Nd}$ (%)	$\phi^{Eu}$ (%)	$\tau^{Eu}$ (ms)	$\phi^{Eu}$ (%)	$\eta_{sens}$ (%)	$q$ ( $Eu^{III}$ )
$[Ln1Thept(COO)_4]^-$	TRIS buffer	5.3 ± 0.2	32 ± 2	0.002 ± 0.000	[a]	0.0007 ± 0.0000	3.1 ± 0.0	0.39 ± 0.01	10	31	1.9 ± 0.1
	95% EtOH	19 ± 0	19 ± 0	0.50 ± 0.01	4.03 ± 0.04	0.40 ± 0.04	32.6 ± 0.2	1.3 ± 0.00	10	82	
$[Ln2Thept(COO)_4]^-$	TRIS buffer	6.3 ± 0.8	20 ± 0	0.005 ± 0.000	[a]	0.002 ± 0.000	0.53 ± 0.01	[a]	[a]	[a]	
	95% EtOH	18 ± 1	13 ± 0	0.31 ± 0.01	4.19 ± 0.00	0.45 ± 0.05	3.0 ± 0.0	0.39 ± 0.01	10.3	29	
$[Ln3Thept(COO)_4]^-$	TRIS buffer <sup>[b]</sup>	4.7 ± 0.2	27 ± 1	0.04 ± 0.00	[a]	0.02 ± 0.00	[c]	[c]	[c]	[c]	
	95% EtOH	13 ± 0	19 ± 0	0.05 ± 0.00	3.99 ± 0.00	0.12 ± 0.00	[c]	[c]	[c]	[c]	

[a] Luminescence was too weak; [b] In TRIS buffer with 15% DMSO; [c]  $Eu^{III}$  luminescence was not observed.

All deprotonated ligands ( $nThept(COO)_4^{4-}$ ) display absorptions with maxima at 300 nm, 360, 400 nm for  $n = 1, 2$  or  $3$  in TRIS buffer. Exciting  $nThept(COO)_4^{4-}$  ( $n = 1, 2$ , or  $3$ ) at these absorbance maxima results in structureless fluorescence bands in the UV-visible region with maxima at 365, 460, and 525 nm, respectively (Figures S25 – S27). The bathochromic shifts observed in the absorbance and emission spectra from  $n = 1$  to  $3$  are consistent with increased  $\pi$ -conjugation and decreased HOMO-LUMO gap.<sup>[26]</sup> The profiles of the absorption, excitation, and emission spectra of the  $Gd^{III}$  complexes,  $[GdnThept(COO)_4]^-$ , exhibit analogous behavior to the free ligands in TRIS buffer (Figures S28 – S30), but with a significant increase in emission intensity (Figures S31 – S33). This is consistent with planarization of the ligand upon coordination<sup>[27]</sup> and some phosphorescence contribution due to improved ISC. Fluorescence quantum yields ( $\phi^F$ ) were determined from the emission spectra (Figures S31–S33) for the free ligands and  $Gd^{III}$  complexes.  $nThept(COO)_4^{4-}$  efficiencies are 5.3% for  $n = 1$ , 6.3% for  $n = 2$ , and 4.7 % for  $n = 3$ .  $\phi^F$  of  $[GdnThept(COO)_4]^-$  are 32% for  $n = 1$ , 20% for  $n = 2$ , and 27% for  $n = 3$  (Table 1).

The  $[Ln nThept(COO)_4]^-$  complexes emit in the visible ( $Ln^{III} = Eu^{III}$ ) and NIR ( $Ln^{III} = Nd^{III}$  or  $Yb^{III}$ ) in TRIS buffer. The emission spectra of  $[Eu1Thept(COO)_4]^-$ ,  $[Yb1Thept(COO)_4]^-$ , and  $[Nd1Thept(COO)_4]^-$ , shown in Figure 2, display the characteristic metal-centred narrow transitions. For  $[Eu1Thept(COO)_4]^-$ , the presence of the  $^5D_0 \rightarrow ^7F_0$  band is consistent with a low symmetry environment around the metal ion.<sup>[28]</sup>  $2Thept(COO)_4^{4-}$  also sensitizes  $Eu^{III}$ , but with poor efficiency, as evidenced by residual ligand emission (Figure S43). A similar behavior is observed in ethanol, and, in this solvent, we were able to determine the sensitization efficiency  $\eta_{sens}$  (Table 1), which is low at 29% (*vide infra*). This is consistent with a  $^3T$  state that is very close in energy to the emissive state of  $Eu^{III}$ .<sup>[18]</sup> Quantum yields of  $Eu^{III}$  emission ( $\phi^{Eu}$ ) were 3.1% and 0.53% for  $[Eu1Thept(COO)_4]^-$  and  $[Eu2Thept(COO)_4]^-$ , respectively (Table 1). The luminescence lifetime ( $\tau^{Eu}$ ) of  $[Eu1Thept(COO)_4]^-$  is 0.39 ms and was fit as a single-exponential decay (Figure S46), consistent with a unique coordination environment around the  $Eu^{III}$  ions.<sup>[10, 29]</sup> We were unable to measure the emission lifetime of the  $[Eu2Thept(COO)_4]^-$  complex due to the weak emission.



## FULL PAPER

Complexes containing NIR emitting  $\text{Ln}^{\text{III}}$  ions,  $[\text{Yb}n\text{Thept}(\text{COO})_4]^-$  and  $[\text{Nd}n\text{Thept}(\text{COO})_4]^-$ , display the characteristic  $\text{Ln}^{\text{III}}$  emission bands (Figures 2 and S43-S44). Quantum yields of  $\text{Yb}^{\text{III}}$  and  $\text{Nd}^{\text{III}}$  emission ( $\phi^{\text{Yb}}$  and  $\phi^{\text{Nd}}$ ) are summarized in Table 1.  $[\text{Yb}1\text{Thept}(\text{COO})_4]^-$  and  $[\text{Nd}1\text{Thept}(\text{COO})_4]^-$  display the lowest quantum efficiencies of the series at 0.002% and 0.0007%, respectively. The efficiencies for  $[\text{Yb}2\text{Thept}(\text{COO})_4]^-$  and  $[\text{Nd}2\text{Thept}(\text{COO})_4]^-$  are 0.005% and 0.002%, respectively. The largest efficiencies are observed for  $[\text{Yb}3\text{Thept}(\text{COO})_4]^-$  and  $[\text{Nd}3\text{Thept}(\text{COO})_4]^-$  and are 0.04% and 0.02%, respectively. These results are similar to what has been observed for other  $\text{Ln}^{\text{III}}$  species.<sup>[25b, c]</sup>

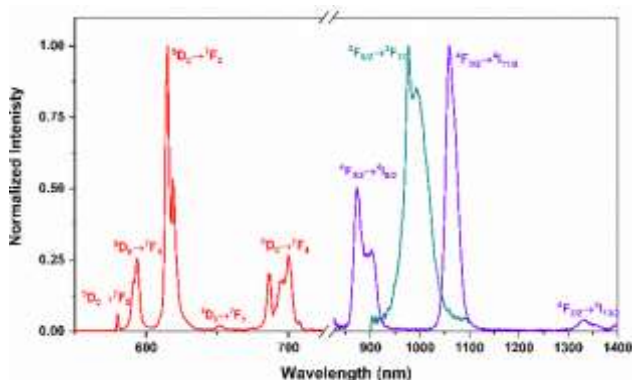


Figure 2. Normalized emission spectra of  $[\text{Ln}(1\text{Thept}(\text{COO})_4)]^-$  ( $\text{Ln}^{\text{III}} = \text{Eu}^{\text{III}}$  (red),  $\text{Yb}^{\text{III}}$  (teal), or  $\text{Nd}^{\text{III}}$  (purple)) in TRIS buffer. ( $\lambda_{\text{exc}} = 310 \text{ nm}$ ; [compound] =  $1 \times 10^{-4} \text{ M}$ ).

As the deprotonated ligand is heptadentate, we expect the coordination sphere of the  $\text{Ln}^{\text{III}}$  ions to be completed by coordinated water molecules. Comparison of the emission lifetimes for  $\text{Eu}^{\text{III}}$  in  $\text{H}_2\text{O}$  and  $\text{D}_2\text{O}$  (Figures S47 and S48),<sup>[30]</sup> indicated that the number of coordinated water molecules  $q$  for  $[\text{Eu}(1\text{Thept}(\text{COO})_4)]^-$  is  $\sim 2$  (Table S1). This corresponds to a coordination number of 9, and is consistent to what is reported for this chelator in other  $\text{Ln}^{\text{III}}$  ion complexes.<sup>[24, 25c, 31]</sup> Due to the similar sizes of the  $\text{Ln}^{\text{III}}$  ions, we expect a similar coordination number for all complexes.

In 95% ethanol, the absorption, emission, and excitation spectra of  $n\text{Thept}(\text{COO})_4^{4-}$  and  $[\text{Gd}n\text{Thept}(\text{COO})_4]^-$  display similar profiles to the spectra collected in TRIS buffer (Figures S34 – S39). A significant increase in fluorescence emission intensity is observed for each  $\text{Gd}^{\text{III}}$  metal complex compared to the respective free ligand, as was observed in TRIS buffer (Figures S40 – S42).

The emission spectra of  $[\text{Eu}1\text{Thept}(\text{COO})_4]^-$ ,  $[\text{Yb}1\text{Thept}(\text{COO})_4]^-$ , and  $[\text{Nd}1\text{Thept}(\text{COO})_4]^-$  in 95% ethanol (Figure S49a) are analogous to the ones in TRIS buffer.  $2\text{Thept}(\text{COO})_4^{4-}$  also sensitizes  $\text{Eu}^{\text{III}}$ , but with poor efficiency, as evidenced by residual ligand emission (Figure S49b). Quantum yields of  $\text{Eu}^{\text{III}}$  emission in 95% ethanol ( $\phi^{\text{Eu}}$ ) significantly increase from 3.1% and 0.53% (TRIS buffer) to 33% and 3% for  $[\text{Eu}1\text{Thept}(\text{COO})_4]^-$  and  $[\text{Eu}2\text{Thept}(\text{COO})_4]^-$ , respectively (Table 1).  $\tau^{\text{Eu}}$  of  $[\text{Eu}1\text{Thept}(\text{COO})_4]^-$  and  $[\text{Eu}2\text{Thept}(\text{COO})_4]^-$  are 1.3 and

0.39 ms, respectively (Figures S53 and S55, Table S2). These lifetimes are longer than those measured in TRIS buffer, consistent with the reduced quenching effect by O – H oscillators in 95% ethanol compared to water (Figure S58).<sup>[10]</sup> The lifetimes enabled us to calculate the  $\eta_{\text{sens}}$  for these two complexes as 82 and 29%, respectively, consistent with the observed residual ligand emission mentioned above in the latter case.

For the same reason,  $\phi^n$  for the NIR emitting complexes,  $[\text{Yb}n\text{Thept}(\text{COO})_4]^-$  and  $[\text{Nd}n\text{Thept}(\text{COO})_4]^-$ , were higher in 95% ethanol as well (Table 1). This enabled measuring the emission lifetimes ( $\tau^{\text{Yb}}$ ) of  $[\text{Yb}n\text{Thept}(\text{COO})_4]^-$ , which were 4.03  $\mu\text{s}$ , 4.19  $\mu\text{s}$ , and 3.99  $\mu\text{s}$  for  $n = 1, 2$ , and  $3$ , respectively (Figures S54, S56, and S57, Table S2). They are comparable to other  $\text{Yb}^{\text{III}}$  complexes.<sup>[32]</sup>

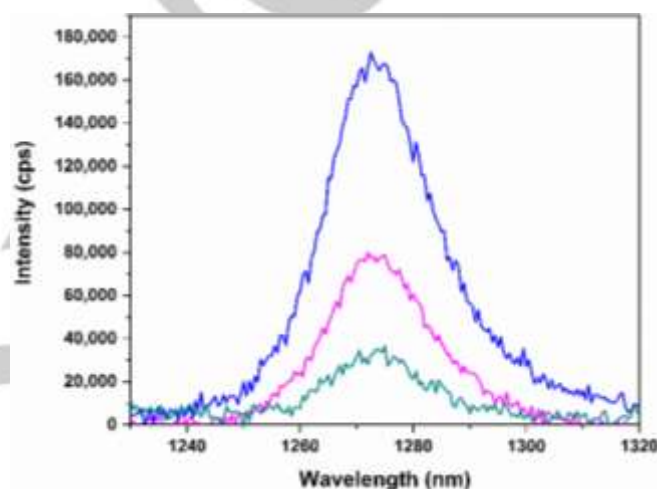


Figure 3. Phosphorescence spectra of  $^1\text{O}_2$  for solutions in 95% ethanol of  $3\text{Thept}(\text{COO})_4^{4-}$  (pink) and  $[\text{Ln}3\text{Thept}(\text{COO})_4]^-$  ( $\text{Ln}^{\text{III}} = \text{Gd}^{\text{III}}$  (blue),  $\text{Yb}^{\text{III}}$  (teal)). ( $\lambda_{\text{exc}} = 400 \text{ nm}$ ; [compound] =  $5 \times 10^{-5} \text{ M}$ ).

$^1\text{O}_2$  phosphorescence at 1270 nm, used to determine efficiency of  $^1\text{O}_2$  generation,<sup>[18, 22a]</sup> was low in TRIS buffer (Figure S45), most likely due the low dissolved  $\text{O}_2$  content (water;  $\chi_{\text{O}_2} = 0.2 \times 10^{-4}$ ).<sup>[33]</sup> In ethanol, which has a higher dissolved oxygen content ( $\chi_{\text{O}_2} = 5.71 \times 10^{-4}$ ),<sup>[34]</sup> the phosphorescence is more intense (Figures 3, S51 and S52) and allowed determination of  $\phi_{1\text{O}_2}$ . These values are summarized in Table 2 and are 15% for  $n = 1$ , 12% when  $n = 2$ , and 24% when  $n = 3$ , and are comparable to values reported for other thienyl-based compounds.<sup>[22a]</sup> The  $\text{Gd}^{\text{III}}$  complexes have the largest  $\phi_{1\text{O}_2}$  at 27% for  $[\text{Gd}1\text{Thept}(\text{COO})_4]^-$ , 17% for  $[\text{Gd}2\text{Thept}(\text{COO})_4]^-$ , and 29% for  $[\text{Gd}3\text{Thept}(\text{COO})_4]^-$ . This observed increase in  $\phi_{1\text{O}_2}$  from the organic photosensitizers to their  $\text{Gd}^{\text{III}}$  complexes was observed by Maury and coworkers<sup>[35]</sup> and us<sup>[18, 22a]</sup> as well, and is attributed to the heavy atom effect.<sup>[36]</sup>  $\phi_{1\text{O}_2}$  was determined to be 16% for  $[\text{Eu}2\text{Thept}(\text{COO})_4]^-$ , 6% for  $[\text{Yb}2\text{Thept}(\text{COO})_4]^-$ , and 2% for  $[\text{Yb}3\text{Thept}(\text{COO})_4]^-$ . Low  $^1\text{O}_2$  emission intensity is observed for  $[\text{Ln}1\text{Thept}(\text{COO})_4]^-$  ( $\text{Ln}^{\text{III}} = \text{Eu}^{\text{III}}$ ,  $\text{Yb}^{\text{III}}$ , or  $\text{Nd}^{\text{III}}$ ), which prevented us from determining  $\phi_{1\text{O}_2}$ .  $^1\text{O}_2$  phosphorescence is also observed for the  $\text{Nd}^{\text{III}}$  complexes with  $2\text{Thept}(\text{COO})_4^{4-}$  and  $3\text{Thept}(\text{COO})_4^{4-}$ ; however, due to the intensity of the nearby  $^4\text{F}_{3/2} \rightarrow ^4\text{I}_{13/2}$  transition,  $\phi_{1\text{O}_2}$  could not be quantified.

## FULL PAPER

**Table 2.** Quantum yields of  $^1\text{O}_2$  generation ( $\phi_{^1\text{O}_2}$ ) for  $n\text{Thept}(\text{COO})_4^{4-}$  and  $[\text{Ln}n\text{Thept}(\text{COO})_4]^-$  at  $25.0 \pm 0.1$  °C in 95% ethanol.

	no Ln <sup>III</sup>	Gd <sup>III</sup>	Eu <sup>III</sup>	Yb <sup>III</sup>	Nd <sup>III</sup>
$[\text{Ln}1\text{Thept}(\text{COO})_4]^-$	15 ± 0	27 ± 1	[a]	[a]	[a]
$[\text{Ln}2\text{Thept}(\text{COO})_4]^-$	12 ± 0	17 ± 1	16 ± 1	6 ± 0	[a]
$[\text{Ln}3\text{Thept}(\text{COO})_4]^-$	24 ± 0	29 ± 1	[b]	2 ± 0	[a]

[a] Luminescence emission was observed but was too low to quantify; [b] not studied

We studied the cytotoxicity of  $n\text{Thept}(\text{COO})_4^{4-}$  ( $n = 1$  and  $2$ ) and their Ln<sup>III</sup> complexes towards HeLa cells (Figures 4 and S61).  $3\text{Thept}(\text{COO})_4^{4-}$  and its complexes were poorly soluble in TRIS buffer and required 15% DMSO to completely dissolve, thus their cytotoxicity was not studied.<sup>[37]</sup>

HeLa cells are an ideal candidate for PDT studies due to their robust characteristics.<sup>[38]</sup> The phototoxicity and dark toxicity of our compounds were determined using an MTT metabolic activity assay.<sup>[39]</sup> As seen in Figure 4, treatment of HeLa cells with solutions of  $[\text{Ln}2\text{Thept}(\text{COO})_4]^-$  or  $2\text{Thept}(\text{COO})_4^{4-}$  in 3.13 μM, 6.25 μM, 12.5 μM, or 25 μM concentration in the dark does not appreciably change cell viability. However, at 50 μM and 100 μM, cell viabilities decrease below 90%, indicating some dark cytotoxicity at these higher concentrations.

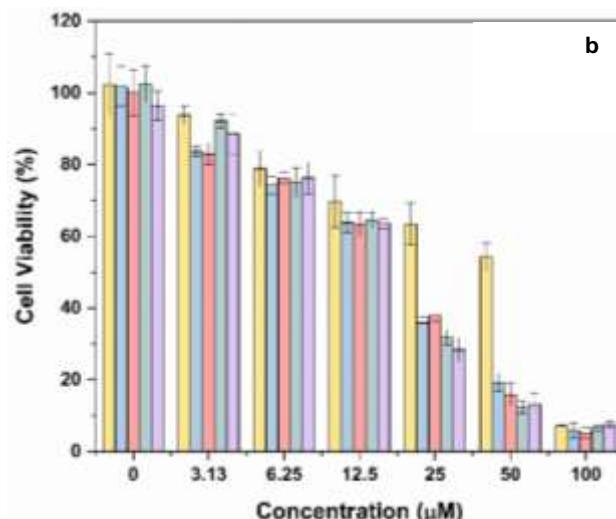
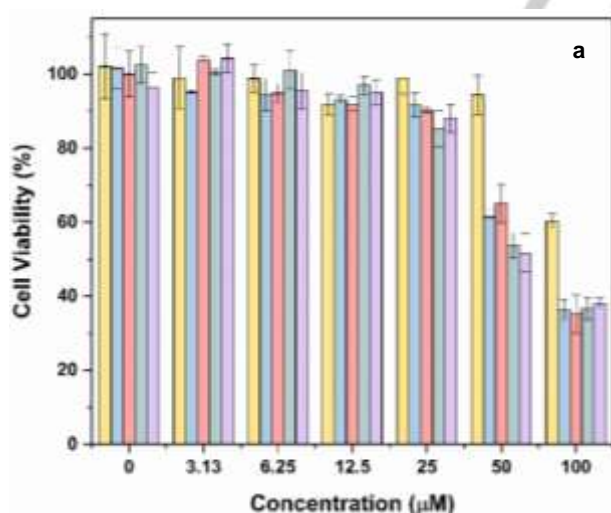


Figure 4. (a) Cell viability (%; MTT assay) as a function of concentration of  $2\text{Thept}(\text{COO})_4^{4-}$  (yellow) or  $[\text{Ln}2\text{Thept}(\text{COO})_4]^-$ , in the dark; (b) cell viability (%; MTT assay) as a function of concentration of  $2\text{Thept}(\text{COO})_4^{4-}$  or  $[\text{Ln}2\text{Thept}(\text{COO})_4]^-$ , after irradiating for 2 minutes with 365 nm light. The control experiments were performed under the same experimental conditions and correspond to the entries at 0 μM. [Ln<sup>III</sup> = Gd<sup>III</sup> (blue), Eu<sup>III</sup> (salmon), Yb<sup>III</sup> (green) or Nd<sup>III</sup> (purple)].

In contrast, the viability of the cells irradiated at 360 nm for 2 minutes decreases with increasing concentration of compound. Cell viability also decreases with increasing irradiation time (Figure S62). As irradiating the cells without added compounds does not lead to a decrease in cell viability, these results suggest that, at concentrations between 3.13 μM and 25 μM, our compounds are photocytotoxic. At the highest concentration tested, cell viabilities range from 8 to 15%.

A dose response fitting was applied to the above data for each compound (Figure S63 – S72, Tables S3 and S4), and the resulting IC<sub>50</sub> values are presented in Table 3. These values indicate that the metal complexes are more photocytotoxic than the free ligand,  $2\text{Thept}(\text{COO})_4^{4-}$ . Also, these compounds are less cytotoxic in the dark than the well-known photodynamic agent, Photofrin, in HeLa cells.<sup>[40]</sup> Cell viability of  $1\text{Thept}(\text{COO})_4^{4-}$  and all  $[\text{Ln}1\text{Thept}(\text{COO})_4]^-$  was also studied and shows that these compounds are photocytotoxic (Figure S61), although were not able to fit the data to meaningful IC<sub>50</sub> values. The photocytotoxicity was not unexpected, as these compounds are capable of generating  $^1\text{O}_2$  as well.

Table 3. IC<sub>50</sub> values of  $2\text{Thept}(\text{COO})_4^{4-}$  and  $[\text{Ln}2\text{Thept}(\text{COO})_4]^-$ , where Ln<sup>III</sup> = Gd<sup>III</sup>, Eu<sup>III</sup>, Yb<sup>III</sup>, or Nd<sup>III</sup>, in HeLa cells compared to Photofrin.

	IC <sub>50</sub> (μM) hv	IC <sub>50</sub> (μM) no hv
$2\text{Thept}(\text{COO})_4^{4-}$	33.9 ± 4.6	113.4 ± 7.1

## FULL PAPER

[Gd2Thept(COO) <sub>4</sub> ] <sup>-</sup>	16.1 ± 0.7	70.2 ± 3.7
[Eu2Thept(COO) <sub>4</sub> ] <sup>-</sup>	16.2 ± 0.8	71.7 ± 3.5
[Yb2Thept(COO) <sub>4</sub> ] <sup>-</sup>	15.8 ± 0.6	65.4 ± 4.2
[Nd2Thept(COO) <sub>4</sub> ] <sup>-</sup>	15.2 ± 0.7	64.7 ± 4.3
Photofrin <sup>[40]</sup>	7.1	> 41

We investigated the nature of the cell death mechanism for the **2Thept(COO)<sub>4</sub>** series using flow cytometry on Annexin V-FITC/PI-labelled HeLa cells in the dark and when exposed to 360 nm light (Figure 5). The cell population of viable cells (FITC<sup>-</sup>/PI<sup>-</sup>), early stage apoptotic cells (FITC<sup>+</sup>/PI<sup>-</sup>), late stage apoptotic cells (FITC<sup>+</sup>/PI<sup>+</sup>), and necrotic cells (FITC<sup>-</sup>/PI<sup>+</sup>) was measured to indicate phases of cell death. In the control groups (a- in the dark, b- light irradiation), less than 10.0% of combined cell death phases were observed at all conditions. Incubating the cells with **2Thept(COO)<sub>4</sub>** in the dark yields results similar to the control experiments. HeLa cells treated with [Ln**2Thept(COO)<sub>4</sub>**]<sup>-</sup> (Ln<sup>III</sup> = Gd<sup>III</sup> or Yb<sup>III</sup>) show a slight increase in cell death, which indicates

some dark toxicity. For the Gd<sup>III</sup> complex, 16.3% of cells were apoptotic (4.0% early stage and 3.6% late stage), and 5.8% of cells were necrotic, for a total of 13.4% of cell population death in the dark. Similarly, for the Yb<sup>III</sup> complex, 5.8% of cells were apoptotic (3.1% early stage and 2.7% late stage), and 4.6% of cells were necrotic, which shows a total of 10.4% of cell population death in the dark. These total cell death percentages are similar to those obtained through the MTT assay for dark toxicity (25 μM) (Figure 4).

In contrast, after irradiating the cells incubated with **2Thept(COO)<sub>4</sub>** or [Ln**2Thept(COO)<sub>4</sub>**]<sup>-</sup>, both necrotic and apoptotic cell death pathways are observed. For **2Thept(COO)<sub>4</sub>** 16.3% of cells were apoptotic (10.1% early stage and 6.2% late stage), and 7.6% of cells were necrotic. Combined, these values indicate that 24% of the cell population undergoes light-activated cell death.

For the Gd<sup>III</sup> complex, 55% of the cell population shows light-induced cell death (4.0% early stage apoptosis, 19.4% late stage apoptosis, and 31.6% necrosis). Similarly, for the Yb<sup>III</sup> complex, 49% of the cell population shows light-induced cell death (2.2% early stage apoptosis, 20.5% late stage apoptosis, and 26.2% necrosis). In analogy to the free ligand, these data suggest both apoptotic and necrotic pathways as well, yet twice the population of dead cells is observed. These findings are consistent to the MTT assay results for phototoxicity at 25 μM.

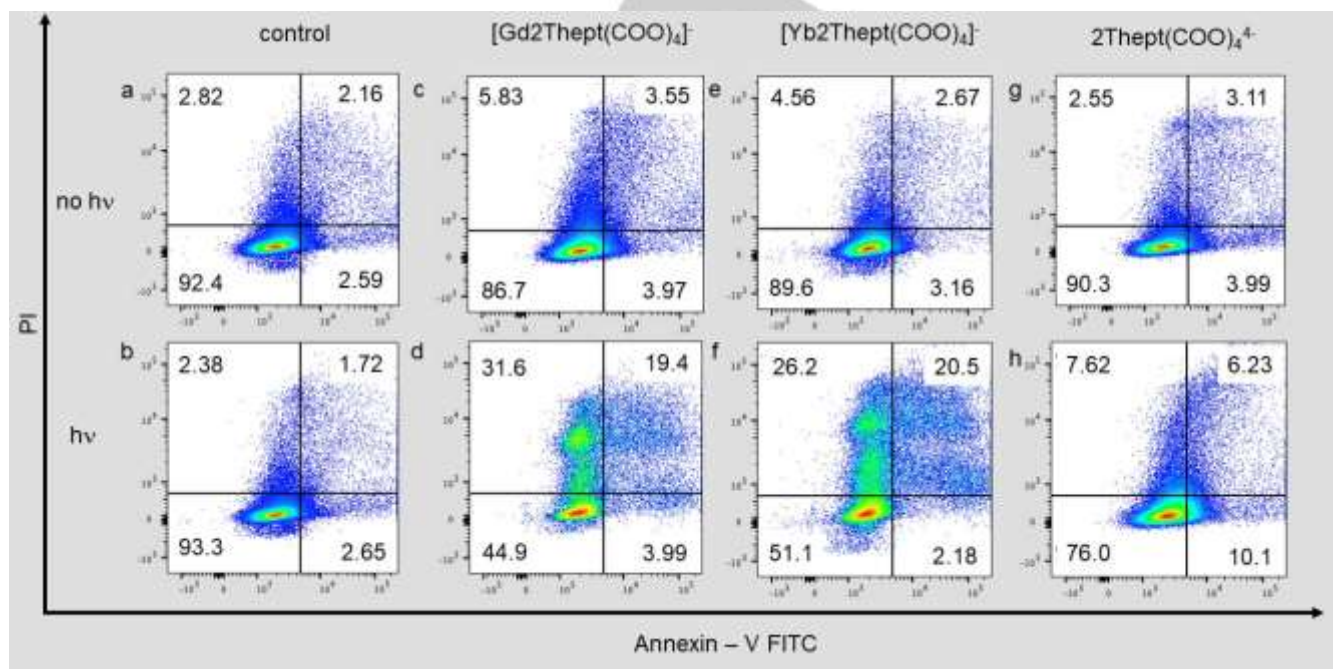


Figure 5. Flow cytometry quantification of Annexin V/PI labelled HeLa cells treated in the dark or with light irradiation (2 minutes of 365 nm): (a) control in the dark; (b) control under light irradiation; (c) [Gd**2Thept(COO)<sub>4</sub>**]<sup>-</sup> (25 μM) in the dark; (d) [Gd**2Thept(COO)<sub>4</sub>**]<sup>-</sup> (25 μM) irradiated with light; (e) [Yb**2Thept(COO)<sub>4</sub>**]<sup>-</sup> (25 μM) in the dark; (f) [Yb**2Thept(COO)<sub>4</sub>**]<sup>-</sup> (25 μM) irradiated with light; (g) **2Thept(COO)<sub>4</sub>** (25 μM) in the dark; (h) **2Thept(COO)<sub>4</sub>** (25 μM) irradiated with light. PBS buffer was used as a control to accommodate changes in volume during sample preparation.

## Conclusions

In this work, we have isolated a series of new complexes containing <sup>1</sup>O<sub>2</sub>-generating oligothiophene moieties. We have shown that these compounds are capable of Ln<sup>III</sup>-centred



## FULL PAPER

emission, and of generating  $^1\text{O}_2$  with efficiencies comparable to known compounds. We have also explored their phototoxicity and dark toxicity with HeLa cells, and determined  $\text{IC}_{50}$  values under irradiation at 360 nm of 15.2 to 16.2  $\mu\text{M}$ . Further investigation using flow cytometry indicates the presence of both necrotic and late apoptotic cells. In the dark, the compounds are much less cytotoxic towards HeLa cells, with decreases in cell viability to <10%. Although we attribute the cytotoxicity to the formation of  $^1\text{O}_2$ , we cannot rule out the formation of other ROS species that could contribute as well to the observed cell death.

The dual functionality of these compounds allows for a variety of applications where  $^1\text{O}_2$  plays a vital role and provides simultaneous tracking of the complexes by  $\text{Ln}^{\text{III}}$ -centred luminescence. While these complexes are water-soluble, attempts to image luminescence within HeLa cells were not fruitful, and is likely due to the low emission efficiencies in aqueous environment. Nonetheless, these compounds provide a design basis for the isolation of compounds with dual functionality, that generate both  $^1\text{O}_2$  and display the characteristic  $\text{Ln}^{\text{III}}$  emission.

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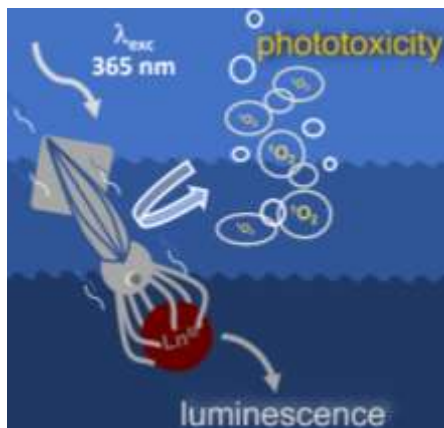
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## Entry for the Table of Contents



Oligothieryl-functionalized heptadentate chelators for  $\text{Ln}^{\text{III}}$  coordination were isolated and their phototoxicity towards HeLa cells studied. These complexes are luminescent in the visible ( $\text{Eu}^{\text{III}}$ ) or near-infrared region ( $\text{Nd}^{\text{III}}$  and  $\text{Yb}^{\text{III}}$ ). While dark cytotoxicity is low,  $\text{IC}_{50}$  values range from 15.2 to 16.2  $\mu\text{M}$  for all compounds upon irradiation of the incubated cells with 365 nm light.