Chemical synthesis of the pyruvic acetal-containing trisaccharide unit of the species-specific glycopeptidolipid from *Mycobacterium avium* serovariant 8

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ABSTRACT

The functionalized, pyruvic acetal-containing haptenic trisaccharide, p-trifluoroacetamidophenyl 6-deoxy-2-O-{3-O-[4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]- α -Lrhamnopyranosyl]- α -L-talopyranoside (19), a component of the glycolipid from Mycobacterium avium serovar 8 was synthesized. For the preparation of the terminal pyruvic acetal-containing unit, benzyl 2-O-benzyl-3-O-methyl- β -D-glucopyranoside (6) was condensed with methyl 2,2-di(ethylthio)propionate (1) in the presence of SO₂Cl₂-CF₃SO₃H catalyst to yield benzyl 2-O-benzyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranoside (7S), which was then converted into the suitably substituted glycosyl donor 2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- α -D-glucopyranosyl trichloroacetimidate (11). The disaccharide glycosyl acceptor p-nitrophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-(2,4-di-O-benzyl- α -L-rhamnopyranosyl)- α -L-talopyranoside (15) was glycosylated with 11 in the presence of trimethyl trifluoromethanesulfonate to furnish the protected trisaccharide p-nitrophenyl 2-O-{3-O-[2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]-2,4-di-O-benzyl- α -L-rhamnopyranosyl]-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (16). After deprotection, this gave the spacer-armed unprotected haptenic trisaccharide 19.

INTRODUCTION

Besides Mycobacterium tuberculosis¹ and M. leprae^{2,3}, the agents of human tuberculosis and leprosy, respectively, another "atypical" mycobacterium⁴ may also cause infections in humans. The fact that up to 50% of patients with AIDS are infected with the M. avium complex⁵⁻⁷ has evoked a growing interest in the epidemiology, immunochemistry, and structure elucidation of Mycobacterium antigens, and their partial or complete syntheses.

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Member serovariants of the *M. avium* complex are distinguished by a set of highly characteristic tetrapeptide-containing surface glycolipids $(GPLs)^{8-10}$, and the general structure of a GPL is shown in A. The antigenic specificity is due to the structural variability at the nonreducing terminus of the haptenic oligosaccharide.

The structure of the haptenic oligosaccharide of the *M. avium* serovar. 8^{11} is: 2-O-{3-O-[4,6-O-(S)-1-carboxyethylidene-3-O-methyl- β -D-glucopyranosyl]- α -L-rhamnopyranosyl}-6-deoxy- α -L-talopyranose. We now report the synthesis of this trisaccharide carrying a *p*-trifluoroacetamidophenyl spacer.

RESULTS AND DISCUSSION

There are numerous methods¹²⁻²¹ for the synthesis of pyruvic acetals of carbohydrates, but the yield or the stereochemical outcome of these reactions is not satisfactory. We used a new reagent, methyl 2,2-di(ethylthio)propionate (1) for the synthesis of 4,6-O-(1-methoxycarbonylethylidene) acetals. This compound can be easily prepared from methyl pyruvate and ethanethiol in the presence of HCl, and then purified by distillation under reduced pressure. For the activation of 1, a SO_2Cl_2 -trifluoromethanesulfonic acid²² mixture (1:1, 1 M solution in 7:3 toluene-diethyl ether) was applied.

Benzyl 2-O-acetyl-3-O-methyl- β -D-glucopyranoside (2) was obtained from benzyl 2,4,6-tri-O-acetyl-3-O-methyl- β -D-glucopyranoside²⁴ by Zemplén transesterification (catalytic amount of NaOMe)²³. Compound 2 was selected since the anomeric center could be selectively deprotected, and the participating group at position 2 ensured the β -selectivity of the glycosylation. However, the coupling reaction of 2 with 1 in the presence of SO_2Cl_2 -trifluoromethanesulfonic acid gave a complex reaction mixture. This observation suggested the change of the O-acetyl function at position 2 to a temporary, easily removable, ether-type protecting group. Thus, benzyl 3-O-methyl- β -D-glucopyranoside²⁴ (3) was converted into its 4,6-O-isopropylidene derivative 4, using 2,2-dimethoxypropane and p-toluenesulfonic acid catalyst²⁵. Compound 4 was then benzylated, and subsequent acid hydrolysis of the isopropylidene group led to benzyl 2-O-benzyl-3-O-methyl- β -Dglucopyranoside (6). TLC analysis of the reaction of 6 with 1 in the presence of SO_2Cl_2 -trifluoromethanesulfonic acid revealed the presence of only two products, which were separated and identified as benzyl 2-O-benzyl-4.6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranoside (7S) and the corresponding diastereoisomer 7R. The yields were acceptable (45 and 24%, respectively), and an advantage of the catalyst employed is that, because of its strong acid character, the formation of the thermodynamically stable product (7S) is favoured.



Catalytic hydrogenation of 7S over 10% Pd–C and subsequent acetylation gave 1,2-di-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl-D-glucopyranose (9), and removal of the anomeric acetyl group with hydrazine acetate²⁶ afforded 2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl-D-glucopyranose (10). This was then transformed into the trichloroacetimidate donor 11 using the procedure described by Schmidt²⁷.

The preparation of the suitably protected crystalline disaccharide glycosyl acceptor 15 and its use in the synthesis of complex oligosaccharides have been reported previously^{28,29}. The synthesis of 15 was slightly modified in that a new rhamnosyl donor, ethyl 3-O-acetyl-2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside (14) was applied. For the preparation of this new donor, we utilized our earlier observation that phase-transfer alkylation of thioglycosides at position 2 proceeds more easily and with better selectivity than the alkylation of O-glycosides. In the present case, ethyl 4-O-benzyl-1-thio- α -L-rhamnopyranoside (12) was benzylated under phase-transfer catalysis conditions to produce ethyl 2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside (13) in a yield of 64%. Conventional acetylation of 13 then gave the rhamnosyl donor 14. Glycosylation of *p*-nitrophenyl *endo*-3,4-O-benzyl-idene- α -L-talopyranoside²⁹ with the donor 14 and subsequent deacetylation of O-3 of the prepared disaccharide, using NaOMe in MeOH, then yielded the crystalline 15.

Glycosylation of the "core" disaccharide acceptor 15 with 11 in the presence of trimethylsilyl trifluoromethanesulfonate promoter at -50° C furnished the trisac-



charide 16 (42%). The β configuration of the newly formed glycosidic linkage was proved by careful analysis of the ¹H and ¹³C NMR spectra of 16, which also revealed that, fortunately, no isomerization at the acetalic carbons occurred during the glycosylation.

The *p*-nitrophenyl aglycon of the trisaccharide 16 was converted into *p*-trifluoroacetamidophenyl (\rightarrow 17) by means of hydrogenation and subsequent treatment with trifluoroacetic anhydride. In accordance with earlier observations²³, removal of the 2-OAc group from 17 required vigorous conditions. The benzyl and the benzylidene groups were removed conventionally (H₂/10% Pd-C) to yield the unprotected, spacer-armed trisaccharide 19. The ¹³C NMR spectrum of 19 was well resolved, but the complete assignment of the ¹H NMR spectrum required various 1D and 2D techniques (DQ-COSY, single and double ¹H, ¹H RELAY, H, C COSY, and 1D NOE). The ¹H and ¹³C NMR data are shown in Table I.

The preparation of neoglycoprotein-type artificial antigens from p-trifluoroacetamidophenyl glycosides is well documented³⁰.

EXPERIMENTAL

General methods.—Melting points were determined on a Kofler apparatus. Optical rotations were measured in CHCl₃ at 20°C with a Perkin-Elmer 241

Atom	Chemical shift		
	¹³ C	¹ H	
1	99.34	5.60	
2	78.56	4.00	
3	73.60	3.63	
4	67.35	4.08	
5	69.24	3.76	
6(CH ₃)	16.74	1.22	
1'	104.28	5.02	
2'	71.58	4.15	
2'	82.14	3.81	
4'	72.48	3.58	
5'	70.64	3.77	
6'(CH ₃)	18.00	1.25	
1″	106.21	4.68	
2"	75.12	3.30-	
3″	83.61		
4″	78.38		
5″	66.93	-3.45	
6″a,b	66.19	4.03-4.11	

TABLE I

¹³C and ¹H NMR data (chemical shifts in ppm) for trisaccharide 19

polarimeter. Reactions were monitored by TLC on Kieselgel 60 F_{254} (Merck) with A, 8:2 hexane-EtOAc; and B, 7:3 hexane-EtOAc; and with detection by UV light and/or by charring with aq 50% H_2SO_4 . Column chromatography was performed on Kieselgel 60 (Merck, 70-230 mesh). All solvents were distilled from an appropriate drying agent. Evaporations were performed under reduced pressure at 40°C. The ¹H (200 MHz) and ¹³C NMR (50.3 MHz) spectra were recorded with a Bruker WP-200 SY spectrometer for solutions in CDCl₃ (internal Me₄Si).

Methyl 2,2-di(ethylthio)propionate (1).—Ethanethiol (16 mL) was added dropwise to chilled methyl pyruvate (10 mL), then HCl was bubbled through the mixture for 1 h. The reaction was monitored by GLC. The mixture was poured onto ice, neutralized with satd aq NaHCO₃, and extracted with CH₂Cl₂ (3 × 150 mL). The organic layers were combined and washed with water (3 × 250 mL), dried, then concentrated in vacuo. The yellow crude product was purified by distillation under diminished pressure, under Ar, to give 1 (16 g, 89%) as a colourless liquid, bp 110°C/8 mmHg; ¹H NMR: δ 3.78 (s, 3 H, COOCH₃), 2.70 (dd, 4 H, SCH₂CH₃), 1.81 (s, 3 H, CH₃), 1.13 (t, 6 H, SCH₂CH₃). Anal. Calcd for C₈H₁₆O₂S₂: C, 46.13; H, 7.74; S, 30.76. Found: C, 46.09; H, 7.71; S, 30.80.

Benzyl 2-O-acetyl-3-O-methyl- β -D-glucopyranoside (2).—Benzyl 2,4,6-tri-O-acetyl-3-O-methyl- β -D-glucopyranoside²⁴ (2.0 g, 4.5 mmol) was dissolved in MeOH (50 mL), and NaOMe (20 mg) was added. The solution was stirred for 5 h at room temperature, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off, the filtrate was concentrated in vacuo, and the crude syrup was

purified on a column of silica gel (9:1 CH₂Cl₂-acetone), to give 2 (670 mg, 42%); mp 90-92°C; $[\alpha]_D$ +158.7° (c 0.46, MeOH); ¹H NMR: δ 7.40-7.31 (m, 5 H, Ph), 5.02 (t, 1 H, $J_{1,2}$ 8.5, $J_{2,3}$ 9 Hz, H-2), 4.56 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 3.62 (s, 3 H, OCH₃), 2.70 and 2.10 (2 bs, each 1 H, HO-4,6). Anal. Calcd for C₁₆H₂₂O₇: C, 58.88; H, 6.79. Found: C, 58.87; H, 6.79.

Benzyl 4,6-O-*isopropylidene-3*-O-*methyl-*β-D-*glucopyranoside* (4).—To a stirred suspension of benzyl 3-O-methyl-β-D-glucopyranoside (3; 4.5 g, 16 mmol) in 2,2-dimethoxypropane (20 mL) was added *p*-toluenesulfonic acid (100 mg). The reaction was monitored by TLC (*A*), and after 1 h the mixture was diluted with CH₂Cl₂, washed with aq NaHCO₃, dried, and concentrated in vacuo to yield 4 (5 g, 97%); R_f 0.4 (*A*); $[\alpha]_D$ -64.0° (*c* 0.4); ¹H NMR: δ 7.35 (s, 5 H, Ph), 4.92 and 4.63 (d, 2 H, PhCH₂), 4.44 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 3.95 (dd, 1 H, $J_{6a,6b}$ 10.5, $J_{6a,5}$ 5.5 Hz, H-6a), 3.80 (t, 1 H, $J_{3,4} = J_{4,5} = 10$ Hz, H-4), 3.67 (t, 1 H, $J_{6a,6b} = J_{6b,5} = 10.5$ Hz, H-6b), 3.60 (s, 3 H, OCH₃), 3.50 (dt, 1 H, $J_{1,2} = J_{2,3} = 8$, $J_{2,OH}$ 2 Hz, H-2), 3.25 (m, 2 H, H-5,3), 2.55 (bs, 1 H, OH), 1.51 and 1.42 [2 s, 6 H, C(CH₃)₂]. Anal. Calcd for C₁₇H₂₄O₆: C, 62.95; H, 7.46. Found: C, 63.08; H, 7.47.

Benzyl 2-O-benzyl-4,6-O-isopropylidene-3-O-methyl-β-D-glucopyranoside (5).—To a stirred suspension of 4 (966 mg, 3 mmol) and KOH (3 g) in N,N-dimethylformamide (10 mL) was added benzyl chloride (411 µL, 1.2 equiv). The mixture was stirred for 10 min at room temperature, then diluted with EtOAc, washed with water until neutral, and concentrated in vacuo to give 5 (1.17 g, 94%); R_f 0.62 (A); $[\alpha]_D - 45.5^\circ$ (c 0.59); ¹H NMR: δ 7.30 (m, 10 H, 2 Ph), 4.92, 4.87, 4.72 and 4.64 (2 d, each 1 H, PhC H_2), 4.55 (d, 1 H, $J_{1,2}$ 7 Hz, H-1), 3.95 (dd, 1 H, $J_{6a,6b}$ 11, $J_{6a,5}$ 5.5 Hz, H-6a), 3.79 (t, 1 H, $J_{6a,6b}$ 11, $J_{6b,5}$ 10.5 Hz, H-6b), 3.62 (t, 1 H, $J_{3,4} = J_{4,5} = 10$ Hz, H-4), 3.58 (s, 3 H, OCH₃), 3.35 (m, 2 H, H-2,3), 3.21 (dt, 1 H, $J_{4,5}$ 10, $J_{5,6b}$ 10.5, $J_{5,6a}$ 5.5 Hz, H-5), 1.5 and 1.41 [2 s, 6 H, C(CH₃)₂]. Anal. Calcd for C₂₄H₃₀O₆: C, 69.55; H, 7.30. Found: C, 69.02; H, 7.22.

Benzyl 2-O-benzyl-3-O-methyl-β-D-glucopyranoside (6).—A solution of 5 (1.0 g, 2.4 mmol) in aq AcOH (70%, 15 mL) was kept at 50°C for 2 h, then concentrated in vacuo. The crystalline crude product (0.9 g) was recrystallized from cyclohexane to yield 6 (780 mg, 87%); R_f 0.07 (A); mp 84–85°C; $[\alpha]_D$ –42.0° (c 0.71); ¹H NMR: δ 7.32 (m, 10 H, 2 Ph), 4.97, 4.90, 4.71 and 4.67 (2 d, each 1 H, PhC H_2), 4.57 (d, 1 H, $J_{1,2}$ 7.5 Hz, H-1), 3.92 (dd, 1 H, $J_{6a,6b}$ 11, $J_{6a,5}$ 3 Hz, H-6a), 3.78 (dd, 1 H, $J_{6a,6b}$ 11, $J_{6b,5}$ 5 Hz, H-6b), 3.62 (s, 3 H, OCH₃), 3.53 (t, 1 H, $J_{4,5} = J_{3,4} = 10$ Hz, H-4), 3.37 (m, 2 H, H-2,5), 3.20 (t, 1 H, $J_{2,3} = J_{3,4} = 10$ Hz, H-3), 2.7 (bs, 1 H, OH). Anal. Calcd for C₂₁H₂₆O₆: C, 67.36; H, 7.0. Found: C, 67.30; H, 6.92.

Benzyl 2-O-benzyl-4,6-O-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranoside (7).—To a solution of 6 (500 mg, 1.4 mmol) in dry CH₂Cl₂ (7 mL) were added 1 (375 mg, 1.8 mmol) and activated 4A molecular sieves (500 mg), and the suspension was stirred under Ar for 40 min at room temperature. After cooling to -78° C, 1:1 CF₃SO₃H-SO₂Cl₂ (4 mL; 1 M in 7:3 toluene-diethyl ether) was added in one portion, and the mixture was allowed to warm up to room temperature. After 3 h, the mixture was neutralized by addition of satd aq NaHCO₃, then diluted with EtOAc (50 mL), and the organic layer was washed with water (3×20 mL), dried, and concentrated in vacuo. The resulting crude syrup was purified by column chromatography using 85:15 hexane-EtOAc as eluant. Yield: **7S**, 283 mg (45%); **7R**, 152 mg (24%).

7S: $[α]_D - 23.9^\circ$ (c 0.88); ¹H NMR: δ 7.3 (bs, 10 H, 2 Ph), 4.9, 4.83, 4.72, and 4.62 (2 d, each 1 H, PhCH₂), 4.56 (d, 1 H, $J_{1,2}$ 7 Hz, H-1), 4.08 (dd, 1 H, $J_{6a,6b}$ 11, $J_{6a,5}$ 5 Hz, H-6a), 3.81 (s, 3 H, COOCH₃), 3.71 (t, 1 H, $J_{6a,6b} = J_{6b,5} = 10$ Hz, H-6b), 3.62 (s, 3 H, OCH₃), 3.4 (m, 4 H, H-2,3,4,5), 1.55 (s, 3 H, CH₃); ¹³C NMR: δ 169.79 (COOCH₃), 102.47 (C-1), 98.70 (acetalic C), 25.15 (C-CH₃). Anal. Calcd for C₂₅H₃₀O₈: C, 65.49; H, 6.59. Found: C, 64.99; H, 6.46.

4,6-O-(S)-(1-Methoxycarbonylethylidene)-3-O-methyl-D-glucopyranose (8).—To a solution of 7 (1.0 g, 2.2 mmol) in EtOH (100 mL) was added Pd–C (10%, 100 mg), and the mixture was vigorously stirred under H₂ overnight at room temperature. The catalyst was filtered off, the cake was washed with EtOH (2 × 20 mL), and the washings were combined and concentrated in vacuo, to yield 8 (600 mg, 96%); R_f 0.3 (75:25 CH₂Cl₂-acetone); $[\alpha]_D$ +12.5° (c 0.65, MeOH); ¹H NMR: δ 5.21 (d, 0.5 H, $J_{1,2}$ 3 Hz, H-1 α), 4.68 (d, 0.5 H, $J_{1,2}$ 8 Hz, H-1 β), 4.99, 4.20, 3.18 and 2.20 (4 bs, each 0.5 H, 2 HO α + β), 3.84 (s, 3 H, COOCH₃), 3.65 (s, 3 H, OCH₃), 1.54 (s, 3 H, CH₃). Anal. Calcd for C₁₁H₁₈O₈: C, 47.48; H, 6.52. Found: C, 47.11; H, 6.33.

1,2-Di-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl-D-glucopyranose (9).—Compound 8 (600 mg, 2.1 mmol) was acetylated by using the standard pyridine (20 mL)-Ac₂O (2 mL) procedure, and the crude product was purified on a short column of silica gel (7:3 hexane-EtOAc) to yield 9 (750 mg, 96%); R_f 0.38 (B); mp 62-63°C; $[\alpha]_D$ + 61.2° (c 0.74); ¹H NMR: δ 6.20 (d, 0.5 H, $J_{1,2}$ 4 Hz, H-1 α), 5.66 (d, 0.5 H, $J_{1,2}$ 8 Hz, H-1 β), 3.84 and 3.86 (s, 3 H, COOCH₃), 3.55 and 3.59 (s, 3 H, OCH₃), 2.18, 2.09, 2.07 and 2.06 (4 s, 6 H, OAc), 1.54 and 1.56 (2 s, 3 H, CH₃). Anal. Calcd for C₁₅H₂₂O₁₀: C, 49.72; H, 6.12. Found: C, 49.75; H, 6.10.

2-O-Acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl-D-glucopyranose (10).—A solution of 9 (600 mg, 1.7 mmol) in dry N,N-dimethylformamide (10 mL) was cooled to 0°C, then hydrazine acetate (200 mg) was added. The mixture was stirred for 6 h at 0°C, diluted with EtOAc (100 mL), washed with satd aq NaCl $(2 \times 20 \text{ mL})$, dried, and concentrated in vacuo, and the resulting syrup was chromatographed on silica gel, using eluant B, to yield 10 (360 mg, 67%) and 9 (100 mg).

10: $R_f 0.17 (B); [\alpha]_D + 68.2^{\circ} (c \ 1.2); {}^{1}H NMR: \delta 5.37 (t, 0.5 H, <math>J_{1,2} = J_{1,OH} = 3.5$ Hz, H-1 α), 3.85 (s, 3 H, COOCH₃), 3.57 and 3.59 (s, 3 H, OCH₃), 3.05 (d, 0.5 H, $J_{1,OH}$ 3.5 Hz, OH α), 2.15 and 2.17 (s, 3 H, OAc), 1.55 and 1.56 (s, 3 H, CH₃). Anal. Calcd for C₁₃H₂₀O₉: C, 48.75; H, 6.29. Found: C, 48.70; H, 6.12.

2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- α -D-glucopyranosyl trichloroacetimidate (11).—Compound 10 (200 mg, 0.62 mmol) was dissolved in dry N,N-dimethylformamide (7 mL), then trichloroacetonitrile (1.3 mL) and DBU (310 μ L) were added to this solution. The mixture was stirred at room temperature for 10 min, then diluted with EtOAc (30 mL), washed with satd aq NaCl (3 × 20 mL), dried, and concentrated. The brown syrup was purified by column chromatography (A), to give 11 (190 mg, 66%) as a yellowish syrup, R_f 0.45 (B). The compound was immediately used for coupling.

Ethyl 2,4-di-O-*benzyl-1-thio-α-L-rhamnopyranoside* (13).—To a solution of ethyl 4-*O*-benzyl-1-thio-*α*-L-rhamnopyranoside³¹ (12; 2.45 g, 5.2 mmol) in CH₂Cl₂ (15 mL) were added tetrabutylammonium bromide (662 mg), benzyl bromide (1.5 mL, 1.5 equiv), and aq 20% NaOH (15 mL), and the mixture was vigorously stirred at room temperature for 12 h. It was then diluted with CH₂Cl₂, and the organic layer was washed with water, dried, and concentrated. Column chromatography (eluant *A*) of the crude product yielded amorphous 13 (2.05 g, 64%); $[\alpha]_D - 91.3^\circ$ (*c* 0.64); ¹H NMR: δ 7.30 (m, 10 H, 2 Ph), 5.33 (d, 1 H, $J_{1,2}$ 1.5 Hz, H-1), 4.90–4.50 (m, 4 H, PhC*H*₂), 4.04 (dd, 1 H, $J_{4,5}$ 10, $J_{5,6a}$ 6 Hz, H-5), 3.91 (dt, 1 H, $J_{3,4} = J_{3,OH} = 10, J_{2,3}$ 4 Hz, H-3), 3.82 (dd, 1 H, $J_{1,2}$ 1.5, $J_{2,3}$ 4 Hz, H-2), 3.36 (t, 1 H, $J_{3,4} = J_{4,5} = 10$ Hz, H-4), 2.55 (m, 2 H, CH₂), 2.40 (d, 1 H, $J_{3,OH}$ 10 Hz, OH), 1.3 (m, 6 H, 2 CH₃). Anal. Calcd for C₂₂H₂₈O₄S: C, 68.01; H, 7.26. Found: C, 67.56; H, 7.20.

Ethyl 3-O-acetyl-2,4-di-O-benzyl-1-thio-α-L-rhamnopyranoside (14).—Conventional acetylation of 13 (1.58 g, 4.07 mmol) with Ac₂O (3 mL) in pyridine (30 mL) and subsequent chromatographic purification (eluant A) gave 14 (1.70 g, 97%); $[\alpha]_D - 56.7^\circ$ (c 1.70); ¹H NMR: δ 7.30 (m, 10 H, 2 Ph), 5.25 (d, 1 H, $J_{1,2}$ 2 Hz, H-1), 5.12 (dd, 1 H, $J_{2,3}$ 3, $J_{3,4}$ 10 Hz, H-3), 4.70–4.50 (m, 4 H, PhCH₂), 4.11 (dd, 1 H, $J_{4,5}$ 10, $J_{5,6}$ 6 Hz, H-5), 3.96 (dd, 1 H, $J_{1,2}$ 2, $J_{2,3}$ 3 Hz, H-2), 3.65 (t, 1 H, $J_{3,4} = J_{4,5} = 10$ Hz, H-4), 2.55 (m, 2 H, CH₂), 1.91 (s, 3 H, acetyl CH₃), 1.3–1.10 (m, 6 H, 2 CH₃). Anal. Calcd for C₂₄H₃₀O₅S: C, 66.95; H, 7.02. Found: C, 66.72; H, 6.93.

p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-(2,4-di-O-benzyl- α -L-rhamnopyranosyl)- α -L-talopyranoside (15).—A solution of 14 (900 mg, 2.1 mmol) and p-nitrophenyl endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside²⁹ (1.1 g, 2.6 mmol) in dry CH₂Cl₂ (20 mL) containing 4A molecular sieves (3 g) was stirred for 30 min under Ar, then methyl triflate (1.6 mL, 14.7 mmol) was added. After 1.5 h, pyridine was injected, and the mixture was filtered through Celite, concentrated, and co-concentrated with toluene (2 × 20 mL). Column chromatography (eluant B) of the crude product gave p-nitrophenyl 2-O-(3-O-acetyl-2,4-di-O-benzyl- α -Lrhamnopyranosyl)-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (1.16 g, 72%) which was O-deacetylated with NaOMe (100 mg) in MeOH (20 mL) to yield 15 (900 mg, 92%). For physical data, see ref 28.

p-Nitrophenyl 2-O-{3-O-[2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]-2,4-di-O-benzyl- α -L-rhamnopyranosyl}-endo-3,4-Obenzylidene-6-deoxy- α -L-talopyranoside (16).—A mixture of 11 (150 mg, 0.32 mmol), the disaccharide 15 (250 mg, 0.34 mmol), and 4A molecular sieves (500 mg) in dry CH₂Cl₂ (10 mL) was stirred under Ar at room temperature for 30 min. It was then cooled to -40°C, Me₃SiOSO₂CF₃ (30 μ L in 2 mL of CH₂Cl₂) was added dropwise, and, after 10 min, the reaction was quenched by the addition of 1 mL of dry pyridine. The mixture was diluted with CH_2Cl_2 (50 mL), filtered, washed with water (2 × 20 mL), dried, and evaporated. The crude product was purified on a silica gel column (75:25 hexane–EtOAc) to afford 16 as a colourless syrup (131 mg, 42%); R_f 0.54 (6:4 hexane–EtOAc); $[\alpha]_D$ –82.9° (c 0.38); ¹H NMR: δ 8.20 and 7.10 (2 bd, 4 H, *p*-nitrophenyl) 7.70–7.20 (m, 15 H, 3 Ph), 5.82 (s, 1 H, PhC*H* endo), 5.60 (d, 1 H, $J_{1,2}$ 6.5 Hz, H-1), 4.97 (dd, 1 H, $J_{1,2}$ 8, $J_{2,3}$ 9 Hz, H-2″), 4.88 (d, 1 H, $J_{1,2}$ 1.5 Hz, H-1′), 4.80–4.50 (PhC H_2), 4.69 (d, 1 H, $J_{1,2}$ 8 Hz, H-1″), 3.81 (s, 3 H, COOCH₃), 3.48 (s, 3 H, OCH₃), 1.83 (s, 3 H, OAc), 1.50 (s, 3 H, pyruvyl CH₃), 1.25 (bd, 6 H, CH₃). Anal. Calcd for $C_{52}H_{59}NO_{19}$: C, 62.33; H, 5.93; N, 1.40. Found: C, 61.85; H, 5.86.

p-Trifluoroacetamidophenyl 2-O-{3-O-[2-O-acetyl-4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]-2,4-di-O-benzyl- α -L-rhamnopyranosyl}endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (17).—A solution of 16 (80 mg, 0.08 mmol) in EtOAc (10 mL) was vigorously stirred under H_2 and in the presence of Adam's catalyst (10 mg), at room temperature for 2 h. Pyridine (200 μ L) and trifluoroacetic anhydride (150 μ L) were added consecutively to the mixture, and stirring was continued for an additional 1 h. The catalyst was filtered off, the cake was washed with EtOAc (2×10 mL), and the filtrate was concentrated and then purified by column chromatography (99:1 CH₂Cl₂-EtOAc) to give 17 as a colourless syrup (80 mg, 94%); R_f 0.68 (92:8 CH₂Cl₂-EtOAc); $[\alpha]_D$ -76.31° (c 0.11); ¹H NMR: δ 7.30 and 7.10 (2 d, 4 H, *p*-trifluoroacetamidophenyl), 7.70-7.3 (m, 15 H, 3 Ph), 5.78 (s, 1 H, PhCH endo), 5.46 (d, 1 H, J_{1.2} 6.5 Hz, H-1), 4.96 (dd, 1 H, J_{1,2} 8, J_{2,3} 9 Hz, H-2"), 4.91 (d, 1 H, J_{1,2} 2 Hz, H-1'), 4.73 (d, 1 H, J_{1,2} 8 Hz, H-1"), 3.81 (s, 3 H, COOCH₃), 3.48 (s, 3 H, OCH₃), 1.86 (s, 3 H, OAc), 1.49 (s, 3 H, pyruvyl CH₃), 1.30 (bd, 6 H, CH₃). Anal. Calcd for C₅₄H₆₀NF₃O₁₈: C, 60.72; H, 5.66. Found: C, 60.11; H, 5.38.

p-Trifluoroacetamidophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-{2,4-di-O-benzyl-3-O-[4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]- α -L-talopyranoside (18).—Sodium methoxide (30 mg) was added to a solution of 17 (80 mg, 0.07 mmol) in MeOH (5 mL) and the mixture was stirred for 5 h. After neutralization (Amberlite IR-120; H⁺) and concentration of the mixture, the crude syrup was purified on a short column of silica gel (8:2 CH₂Cl₂-EtOAc) to give 18 as a colourless syrup (42 mg, 58%). The product was applied for the next step without characterisation.

p-Trifluoroacetamidophenyl 6-deoxy-2-O-{3-O-{4,6-O-(S)-(1-methoxycarbonylethylidene)-3-O-methyl- β -D-glucopyranosyl]- α -L-rhamnopyranosyl}- α -L-talopyranoside (19).—To a solution of 18 (40 mg, 0.04 mmol) in MeOH (10 mL) was added Pd-C catalyst (10%, 20 mg), and the suspension was vigorously stirred under H₂ for 4 h. The catalyst was filtered off, the filtrate was concentrated in vacuo, and the crude material was purified on a short column of silica gel (99:1 CH₂Cl₂-MeOH) to furnish 19 as a colourless glass (25 mg, 83%); R_f 0.45 (92:8 CH₂Cl₂-MeOH); $[\alpha]_{\rm D}$ – 77.6° (*c* 0.22, MeOH). Anal. Calcd for C₃₁H₄₂NF₃O₁₇: C, 49.14; H, 5.59; N, 1.86; F, 7.56. Found: C, 49.32; H, 5.23.

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