SYNTHESIS OF D:A-FRIEDO-18 β ,19 α H-LUPAN-3-ONE AND D:B-FRIEDO-18 β ,19 α H-LUP-5-EN-3 β -OL AND A COMMENT ON THE STRUCTURE OF GUIMARENOL

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D:B-Friedo-18 β ,19 α H-lup-5-en-3 β -ol was synthesized from friedelin <u>via</u> D:A-friedo-18 β ,19 α H-lupan-3-one. The stereostructure of a 3α ,4 α -epoxide derived from the 3-one was determined by X-ray technique to show the 19 α H configuration for both the epoxide and the 5-en-3 β -ol. The disagreement of the spectral data of the synthetic 5-en-3 β -ol with those of guimarenol is described.

Although a number of migrated oleanane, ursane, and hopane derivatives were isolated and their structures were elucidated,¹⁾ only a few investigations on migrated lupane derivatives have been described, <u>viz</u>., the isolation of guimarenol (1) and lup-18-en-3β-ol from <u>Ceropegia dichotoma</u>²⁾ and the preparation of 3β,28-diacetoxy-18β,19αH-lupane derivatives from betulin.³⁾ The stereochemistry at C-13, C-14, C-17, C-18, and C-19 of guimarenol (1) remained undetermined.²⁾ A plausible stereostructure, D:B-friedo-18β,19αH-lup-5-en-3β-ol (2), can be suggested for guimarenol by means of biogenetic considerations⁴⁾ involving a sequence of migration of hydrides and methyl groups of an intermediate carbonium ion (A; path <u>a</u>) produced from squalene 2,3-oxide. We have recently reported the synthesis of two migrated lupane derivatives of a D:A-friedo type, D:A-friedo-18β-lup-19-en-3-one (3) and D:A-friedo-18β-lup-19-ene (4) from friedelin (5).⁵⁾ The present communication deals with a synthesis of D:A-friedo-18β,19αH-lupan-3-one (6) from 3 and a conversion of 6 into 2. Although this 3β-ol (2) could be proposed for the structure of guimarenol (1) on the basis of biogenetic considerations, the spectral data of the synthetic 38-ol (2) were not identical with those of guimarenol (1).

of the synthetic 3β-ol (2) were not identical with those of guimarenol (1). D:A-Friedo-18β-lup-19-en-3-one (3)⁵⁾ was treated with sodium borohydride in methanol and then with acetic anhydride in pyridine to yield D:A-friedo-18β-lup-19-en-3β-yl acetate (7) quantitatively. The acetate (7) was dissolved in acetic acid and hydrogenated in the presence of platinum catalyst under atmospheric pressure at 40 °C to give a single hydrogenation product (8) in 84% yield. Treatment of the saturated acetate (8) with lithium aluminium hydride in refluxing tetrahydrofuran afforded an alcohol (9), which was then subjected to dehydration with phosphoryl chloride in pyridine under reflux condition. A dehydration product (10), thus obtained in 84% yield, was treated with <u>m</u>-chloroperbenzoic acid in chloroform to afford a mixture of two epoxides in a ratio of 9:5 in 93% yield. ¹H-NMR measurement of each epoxide revealed that a major epoxide was $3\alpha,4\alpha$ -epoxide (11; δ 2.85, 1H, t, J = 2.5 Hz) and a minor one $3\beta,4\beta$ -epoxide (12; δ 2.90, 1H, t, J = 1 Hz).⁶

In order to determine the configuration of the isopropyl group on C-19 of § generated by the hydrogenation of 7, a single crystal of the $3\alpha,4\alpha$ -epoxide (11) was subjected to X-ray diffraction analysis. Crystals of 11 belong to a monoclinic space group P2₁ with the cell parameters of a = 13.996(6), b = 11.461(5), c = 8.199 (4) Å, $\beta = 106.34(6)^{\circ}$ and the two molecules are contained in a cell. A total of 2494 independent, non-zero reflections are measured on a Philips PW 1100 automatic diffractometer using graphite-monochromated Cu Ka radiation. The structure was solved by the direct method using program MULTAN⁷ and was refined by the block-diagonal least-squares calculation assuming the anisotropic thermal vibrations. The final R-factor was 9.9% excluding hydrogen atoms. Figure 1 is a computer-generated perspective drawing of the molecular structure of the epoxide (11), and showed a 19 α H-configuration as well as 18 β H-configuration. Thus it was found that the acetate (8) could be formulated as D:A-friedo-18 β ,19 α H-lupan-3 β -yl acetate with the 19 β -isopropyl group and that the hydrogenation of 7 had occurred from α -side preferentially to afford the 19 α H-isomer (8).

The alcohol (9) was subjected to oxidation with the Jones reagent to give D:A-friedo-18 β ,19 α H-lupan-3-one (6) quantitatively. The ketone (6) is a biogenetically interesting compound, because it represents the ultimate rearrangement product from the original posturated ion (A) by path <u>b</u> in the migrated lupane series, corresponding to a friedelin-type compound in the migrated oleanane series.

The ketone (6) was treated with benzoyl chloride at reflux temperature to give a Δ^3 -enol benzoate (13), which, on bromination with bromine in pyridine, gave a 4-bromo ketone (14) in 62% yield. According to the known procedure,⁸) the bromide (14) was treated with silver acetate to give a 1:2 mixture of two unsaturated ketones, (15) and (16), in 74% yield, which could not be separated by TLC nor HPLC. Reduction of the mixture with sodium borohydride in methanol at 0 °C gave a mixture of the corresponding 3β-alcohols and 3α-alcohols, which was subjected to preparative TLC and preparative HPLC to separate into D:B-friedo-18β,19αH-lup-5-en-3β-ol (2), -5-en-3α-ol (17), -5(10)-en-3β-ol (18), and -5(10)-en-3α-ol (19) in a ratio of 1:7:14:2. The structures of these unsaturated alcohols were determined by comparison of their ¹H-NMR spectra and TLC chromatograms with those of the corresponding D:B-friedooleanane⁹ and D:B-friedobaccharane derivatives.¹⁰

The melting point and the ¹H-NMR spectrum of the synthetic 5-en-3 β -ol (2) were found to be not identical with those of guimarenol (1)²) (vide infra). Oxidation of 2 with the Sarett reagent gave the ketone (15). The melting point and the ¹H-NMR spectrum of 15 were not in agreement with those of guimarenone (20) described previously.²) The structure of guimarenol (1) could not be represented by D:Bfriedo-18 β ,19 α H-lup-5-en-3 β -ol (2); further structural studies on guimarenol would be required.

Characterization of products is as follows; <u>D:B-friedo-18 β ,19 α H-lup-5-en-3 β ol (2): mp 166-167 °C, IR (Nujol) 3400, 825 cm⁻¹; ¹H-NMR¹¹⁾ δ 0.88, 1.03, 1.13,</u> 1.24 (each 3H, s; <u>t</u>-CH₃), 0.90 (6H, d, J = 6 Hz; $(CH_3)_2$ CH-), 0.97 (6H, s; 2 x <u>t</u>-CH₃, 3.45 (1H, m, W_{1/2} = 7 Hz; 3α-H), 5.65 (1H, m, W_{1/2} = 7 Hz; 6-H), MS m/e (%) 426 (M⁺; 7), 274 (100), MW¹²) 426.3852. [<u>cf</u>. guimarenol (1)²); mp 276-278 °C, ¹H-NMR δ 0.77, 0.89, 1.00, 1.03, 1.06, 1.13 (Me signals), 3.55 (1H, m, W_{1/2} = 6 Hz; -<u>CH</u>-OH), 5.60 (1H, m, -<u>CH</u>=C)]; <u>D:A-friedo-18β,19αH-lupan-3-one</u> (6): ¹³⁾ mp 260-262 °C, IR 1715 cm⁻¹, ¹H-NMR \Box Eu(fod)₃-<u>d₂₇/6</u> = 0.2 (M/M) in CDCl₃] δ 1.72 (3H, d, J = 7 Hz), MS m/e 426 (M⁺; 100); <u>D:A-friedo-18β-lup-19-en-3β-vl</u> acetate (7): ¹³⁾ mp 240-242.5 °C, IR 1738 cm⁻¹, ¹H-NMR δ 1.63, 1.67 (each 3H, br. s), 2.04 (3H, s), 4.92



(1H, m, $W_{1/2} = 7$ Hz), MS m/e 468 (M⁺; 60), 317 (100); <u>D:A-friedo-186,19 α H-lupan-36-yl acetate</u> (8):¹³⁾ mp 251-253 °C, IR 1735 cm⁻¹, ¹H-NMR δ 2.03 (3H, s), 4.94 (1H, m, $W_{1/2} = 6$ Hz), MS m/e 470 (M⁺; 58), 410 (100); <u>D:A-friedo-186,19 α H-lupan-36-ol</u> (9); mp 225-227 °C, IR 3440 cm⁻¹, ¹H-NMR δ 3.75 (1H, m, $W_{1/2} = 7$ Hz), MS m/e 428 (M⁺; 100), MW¹²) 428.3967; <u>D:A-friedo-186,19 α H-lup-3-ene</u> (10):¹³⁾ mp 169-170 °C, IR 795 cm⁻¹, ¹H NMR δ 5.18 (1H, m, W = 0.440 (M⁺; 65), 740 (100); 75 tr cm^{-1} , ¹H-NMR δ 5.18 (1H, m, W_{1/2} = 9 Hz), MS m/e 410 (M⁺; 65), 318 (100); <u>3 α ,4 α -</u> epoxy-D:A-friedo-18 β ,19 α H-lupane (11):¹³ mp 200-202 °C, ¹H-NMR δ 0.80-0.93 (15H), 0.96, 1.07, 1.19 (each 3H, s), 2.85 (1H, t, J = 2.5 Hz), MS m/e 426 (M⁺; 77), 411 (100); <u>3β,4β-epoxy-D:A-friedo-18β,19αH-lupane</u> (<u>12</u>):¹³⁾ mp 215-216 °C, ¹H-NMR δ 0.78-0.93 (15H), 0.96, 1.05, 1.17 (each 3H, s), 2.90 (1H, t, J = 1 Hz), MS m/e 426 (M⁺; 100); <u>3-benzoyloxy-D:A-friedo-18β,19αH-lup-3-ene</u> (13):¹³⁾ mp 255-256 °C, IR 1732 cm⁻¹; ¹H-NMR δ 1.49 (3H, s), 7.4-7.7 (3H, m), 8.0-8.3 (2H, m), MS m/e 530 (M⁺; 5), 105 (100); <u>4α-bromo-D:A-friedo-18β,19αH-lupan-3-one</u> (14): mp 192.5-194.5 ^OC, IR 1715 cm⁻¹, ¹H-NMR δ 1.71 (3H, s), 3.38 (1H, ddd, J = 20, J = 13, J = 8 Hz; axial 2α -H), MS m/e 506 (M⁺; 12), 504 (M⁺; 12), 123 (100), MW¹²⁾ 506.2946 and 504.2992; D:B-friedo-18β,19αH-lup-5-en-3-one (15): mp 149-153 °C, IR (Nujol) 1718 cm⁻¹, ¹H-NMR δ 0.82, 0.90 (each 3H, s; t-CH₃), 0.83 (6H, d, J = 7 Hz; (CH₃)₂CH-), 1.21 (12H, s; 4 x <u>t</u>-CH₃), 5.63 (1H, m, $W_{1/2} = 11$ Hz; -CH=C), ms m/e 424 (M⁺; 11), 274 (100), 123 (93), MW¹²) 424.3731 [<u>cf</u>. guimarenone (20)²): mp 196-200 °C, ¹H-NMR δ 0.78, 0.86, 0.92, 1.01, 1.06, 1.22, 1.25 (Me signals), 5.75 (1H, m, -CH=C)]; <u>D:B-friedo</u>-<u>18β,19αH-lup-5-en-3α-ol</u> (17): mp 181-182 ^OC, IR 3320, 825 cm⁻¹, ¹H-NMR δ 3.20 (1H, dd, J = 4, J = 10 Hz), 5.61 (1H, m, $W_{1/2} = 9 \text{ Hz}$), MS m/e 426 (M⁺; 10), 274 (100), MW¹²⁾ 426.3808; <u>D:B-friedo-18β,19αH-lup-5(10)-en-3β-ol</u> (<u>18</u>): mp 191-193 ^oC, IR 3350 cm⁻¹, ¹H-NMR δ 3.48 (1H, dd, J = 4, J = 10 Hz), MS m/e 426 (M⁺; 13), 408 (100), 205 (72), MW¹²⁾ 426.3846; <u>D:B-friedo-18β,19αH-lup-5(10)-en-3α-ol</u> (19): mp 195.5-196 °C, IR 3620 cm⁻¹, ¹H-NMR δ 3.48 (1H, dd, J = 3, J = 4.5 Hz), MS m/e 426 (M⁺; 10), 205 (100), MW¹²⁾426.3880.

References and notes

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