

# N-ACETOXY-4-METHOXYANILINE, A MODEL COMPOUND FOR THE ULTIMATE CARCINOGEN OF THE PHENACETIN RELATED 4-ETHOXYANILINE

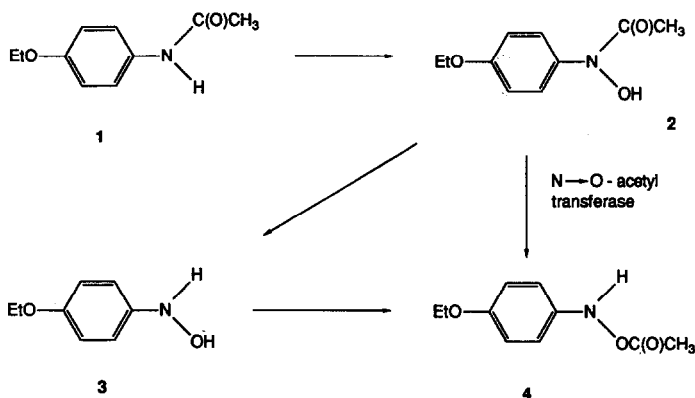
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**Summary:** We prepared and trapped in situ with N-methylaniline and deoxyguanosine (dG) the three N-acyloxy-4-methoxyanilines 7a-c. Furthermore, we synthesized N-acetoxy-4-methoxyaniline 7a and reacted it with dG. Especially 7a is a model for the ultimate carcinogen of phenacetin 1, namely N-acetoxy-4-ethoxyaniline 4.

Phenacetin 1<sup>[1]</sup> was introduced as a drug in 1898 because of its analgetic and antipyretic effects, and its low acute toxicity<sup>[2]</sup>. Abuse, however, led to damages<sup>[3]</sup>: phenacetin 1 reduces the lifetimes of erythrocytes<sup>[4]</sup>, and causes anaemia<sup>[5]</sup> and methaemoglobinemia<sup>[6]</sup>. An important side-effect is the formation of nephritis<sup>[7]</sup>. Furthermore, in some cases 1 led to kidney necrosis<sup>[8]</sup>, kidney carcinomas<sup>[9]</sup>, bladder cancer<sup>[10]</sup> and tumors in the urinary system<sup>[11]</sup>. The carcinogenicity of 1 was also demonstrated with animals. When male and female Sprague-Dawley rats were fed with diet containing 2.5 (1.5)% phenacetin 1, 96.3 (90.9)% of the male and 77.8 (76.9)% of the female rats showed neoplasmas<sup>[12]</sup>. Because of these non-negligible side-effects phenacetin 1 was forbidden as part of mixed analgetics in the Federal Republic of Germany in 1986<sup>[13,14]</sup>.

Scheme I

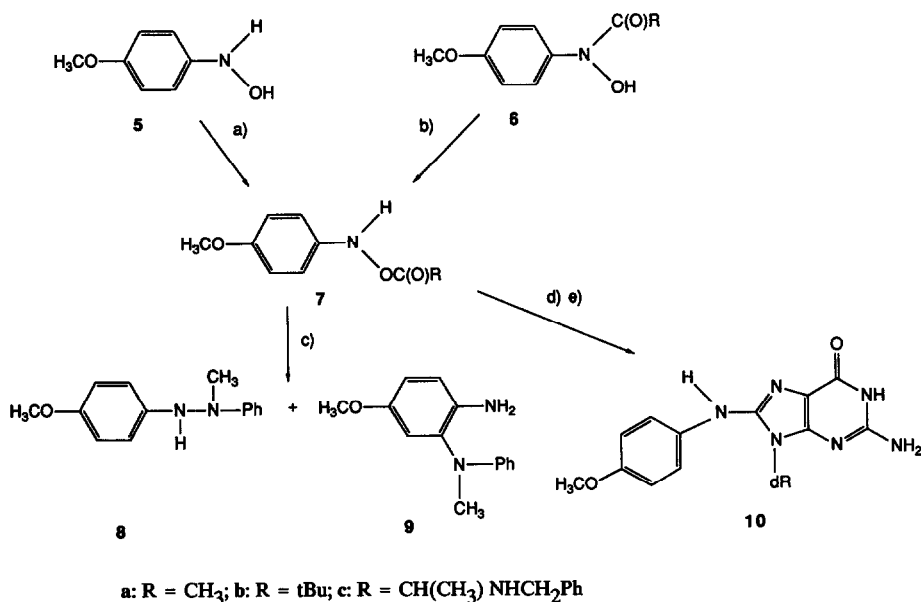


Among the investigations of the pathways leading to toxic, mutagenic and carcinogenic metabolites of 1 the following are relevant to the work reported in this communication. As in the case of other mutagenic and carcinogenic aromatic amines<sup>[15]</sup>, N-oxidation of 1 by means of microsomal mixed function oxidases leading to N-hydroxyphenacetin 2 is a first important step<sup>[16]</sup>. The transformation of 2 in mutagen (Ames test) and nucleic acid binding metabolites by N -> O acetyltransferase suggests the intermediate formation of N-acetoxy-4-ethoxy-aniline 4 as a reactive metabolite<sup>[17]</sup>. 4 might also result from O-acetylation of N-hydroxy-4-ethoxyaniline 3<sup>[15]</sup> (Scheme I). So far, however, there is no report in the literature on the synthesis of 4 or the characterization of a (bio)nucleophile adduct with this metabolite of 1.

In the following we report on model compounds of **4**: the intermediate formation of the N-acyloxy-4-methoxyanilines **7a-c**, their *in situ* trapping reactions with the nucleophile N-methylaniline to give the adducts **8** and **9** and with the bionucleophile deoxyguanosine (dG) to give the adduct **10**, as well as the synthesis of the extremely labile N-acetoxy-4-methoxyaniline **7a** and its reaction with dG to give **10**.

We have chosen these model compounds and model reactions for the following reasons: (1) p-methoxyphenylhydroxylamine **5** as a starting compound is more easily accessible and more stable than p-ethoxyphenylhydroxylamine **3**; at the same time the donor quality of the p-methoxy substituent is essentially the same as that of the p-ethoxy substituent which is responsible for the pharmacological and toxic properties of phenacetin **1**; (2) it is generally assumed that adduct formation of the ultimate carcinogen of an aromatic amine with the DNA base deoxyguanosine finally leads to cancer<sup>[15,18]</sup>. It is therefore important to know whether a putative ultimate carcinogen like **7** reacts with deoxyguanosine; (3) N-methylaniline has shown to be a successful nucleophile in testing the electrophilic character of nitrogen electrophiles like **7**<sup>[15,18,20]</sup>.

Scheme II



Reaction conditions: a) CH<sub>3</sub>C(O)CN, THF, -78°C, 5 min; b) NEt<sub>3</sub>, EtOH:CHCl<sub>3</sub>:H<sub>2</sub>O = 7:4:3, 85°C, 3 d; c) from **6**: b) plus N-methylaniline; d) from **6**: b) plus dG; e) **7a** in THF (cooled to -78°) added to dG in H<sub>2</sub>O, 20°C

*In situ trapping of 7a-c.* As shown earlier<sup>[15,18]</sup> we can model *in vitro* the *in vivo* N → O transacylation leading from N-acyl-N-arylhydroxamic acids to N-acyloxy-N-arylamines by means of an amine catalyst. Applying this methodology to the hydroxamic acids **6a-c** and triethylamine (route b) in Scheme II), trapping of **7a-c** with N-methylaniline (route c) in Scheme II) gave the hydrazine **8** and the "ortho adduct" **9**<sup>[19]</sup>, see Table 1. It is noteworthy that ortho adducts of the type **9** are only formed in the case of reactive N-acyloxy-N-arylamines derived from amines with carcinogenic potential like 4-aminobiphenyl<sup>[20a]</sup>, 2-naphthylamine<sup>[20b]</sup>, 2-amino-fluorene<sup>[20c]</sup> or p-toluidine<sup>[15]</sup>. In similar trapping reactions of **7a-c** with deoxyguanosine (dG) (route d) in Scheme II) the dG-adduct **10** was formed, see Table 1. The yields of **10** are higher by a factor of two if compared, e.g. with the yields in the trapping reactions of N-acyloxy-N-4-methylanilines with dG<sup>[15]</sup>. This indicates that the N-acyloxy-4-methoxyaniline intermediates **7a-c** are very reactive due to the p-methoxy donor substituent.

Table 1. Yields [%] of adducts 8 and 9 (reactions with N-methylaniline) and adduct 10 (reactions with deoxyguanosine dG) starting from the p-methoxyphenyl-hydroxyl amine derived hydroxamic acids 6a-c and N-acetoxy-p-methoxyaniline 7a, respectively.

	8	9	10
6a	28	30	5.2
6b	21	28	6.3
6c	28	30	6.3
7a	0	0	4.2

**7a: Synthesis and reactions.** N-acetoxy-4-methoxyaniline 7a was prepared according to a procedure first introduced by Lobo et al.<sup>[21]</sup> and successfully applied to other labile N-acetoxy-N-arylamines by us<sup>[15,18,20]</sup>: reaction of the hydroxylamine 5 with acetylcyanide in THF (route a) in Scheme II). Because of the extreme thermal lability of 7a this reaction had to be performed at -78°C within 5 min. All attempts to isolate 7a failed because of decomposition, however, when 7a was prepared in THF-D<sub>8</sub> it could be characterized by means of its <sup>1</sup>H-nmr spectrum<sup>[22]</sup>. From the reactions of 7a with N-methylaniline we were unable to isolate either 8 or 9. The reaction with dG, however, led to the C8 adduct 10 (route e) in Scheme II). When both nucleophiles, N-methylaniline and dG, reacted simultaneously with 7a, again only the dG-adduct 10 was isolated. Although we have no explanation for these interesting observations at the moment, the formation of 10 from 7a and dG proves that 7a is also formed from 6a via transacylation. Judging from the yields of the adducts 8, 9 and 10, the N-acyloxy-4-methoxyanilines 7b and 7c should similarly result from the hydroxamic acids 6b and 6c, respectively, by N -> O transacylation, and react with N-methylaniline and dG, respectively.

The model N-acyloxy-4-methoxyanilines 7a-c, prepared from the hydroxamic acids 6a-c and trapped in situ by N-methylaniline to give 8 and 9, and by dG to give 10, in high yields thus indicate the strong electrophilic character of the phenacetin 1 related ultimate carcinogen N-acetoxy-4-methoxyaniline 4. The extreme lability of N-acetoxy-4-methoxyaniline 7a, prepared at -78°C, is in accord with these observations.

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## References

- [1] J. von Mering, *Therap. Monatsh.* **7** (1893) 577.
- [2] E.Mutschler in "Arzneimittelwirkungen", 5. Aufl., Wissenschaftliche Verlagsgesellschaft, Stuttgart, 1986, S. 189.
- [3] P. K. Smith in "Acetophenetidine, a Critical Bibliographical Review", Interscience Publ., New York, London, 1958.
- [4] P. Miescher, A. Pletscher, *Schweiz. Med. Wochenschr.* **88** (1958) 1056.
- [5] a) F. Wuhrmann, B. Jasinski, *Dtsch. Med. Wochenschr.* **80** (1955) 1632; b) H.R. Marti, *Schweiz. Med. Wochenschr.* **88** (1958) 1054; c) Editorial, Hemolytic Reactions to Drugs, *Brit. Med. J.* 841 (1959/I).
- [6] M.E.M. Thomas, *Brit. Med. J.* 1020 (1959/II).
- [7] a) O. Spühler, H.U. Zollinger, *Z. Klin. Med.* **151** (1953) 1; b. O. Gsell, H.K. von Rechenberg, P. Mieschler, *Dtsch. Med. Wochenschr.* **82** (1957) 1673 and 1718; c) R. Sarre, A. Moench, R. Kluthe in "Phenacetinabusus und Nierenschädigungen",

- G. Thieme Verlag, Stuttgart, 1958; d) H. Hengstmann, *Med. Welt*, 529 (1962).
- [8] K.G. Thiele, H. Berning, *Münch. Med. Wochenschr.* **111** (1969) 1673.
- [9] W. Leistenschneider, R. Ehrmann, *Schweiz. Med. Wochenschr.* **103** (1973) 433.
- [10] R.A. Mannion, D. Susmano, *J. of Urol.* **106** (1971) 692.
- [11] M.J. Mihatsch, T. Manz, C. Knüsli, H.O. Hofer, M. Rist, R. Guetg, R. Ruitsbauer, H.U. Zollinger, *Schweiz. Med. Wochenschr.* **110** (1980) 255.
- [12] H. Isaka, H. Yoshii, A. Otsuyi, M. Koike, Y. Nagai, M. Koura, K. Sugiyasu, T. Kanabayashi, *J. Jpn. Cancer Soc. (Gann)* **70** (1979) 29.
- [13] M.J. Mihatsch, M. Molzahn, E. Ritz, *Dtsch. Med. Wochenschr.* **110** (1986) 1416.
- [14] In other countries it has been taken from the market much earlier.
- [15] C. Meier, G. Boche, *Tetrahedron Lett.* **31** (1990), accompanying communication and the ref. cited therein.
- [16] a) A. Klutch, M. Harfeinst, A.H. Conney, *J. Med. Chem.* **9** (1966) 63; b) H. Uehleke, *Naunyn-Schmiedeberg's Arch. Pharmacol.* **264** (1969) 434; c) J.A. Hinson, J. R. Mitchell, *Drug Metab. and Disposition* **4** (1976) 430; d) T. Nohmi, K. Yoshikawa, M. Nakadate, M. Ishidate, *Biochem. Biophys. Res. Commun.* **110** (1983) 746; e) R. Nery, *Xenobiotica* **1** (1971) 339; b) J. A. Hinson, *Environ. Health Perspect.* **49** (1983) 71.
- [17] a) J.B. Vaught, P.B. McGarvey, M.-S. Lee, C.D. Garner, C.Y. Wang, E.M. Linsmaier-Bednar, C.M. King, *Cancer Res.* **41** (1981) 3424; b) J. W. Oldham, R.F. Preston, J.D. Paulson, *J. Appl. Tox.* **6** (1986) 237.
- [18] G. Boche, F. Bosold, S. Schröder, *Angew. Chem.* **100** (1988) 965; *Angew. Chem. Int. Ed. Engl.* **27** (1988) 973.
- [19] The hydroxamic acids **6a-c** as well as the adducts **8**, **9**, and **10** are stable compounds which have been fully characterized by nmr and mass spectroscopy as well as elemental analysis.
- [20] a) *4-Aminobiphenyl*: M. Famulok, F. Bosold, G. Boche, *Angew. Chem.* **101** (1989) 349; *Angew. Chem. Int. Ed. Engl.* **28** (1989) 337.  
 b) *2-Naphthylamine*: M. Famulok, F. Bosold, G. Boche, *Tetrahedron Lett.* **30** (1989) 321.  
 c) *2-Aminofluorene*: F. Bosold, G. Boche, *Angew. Chem.* **102** (1990), in print.  
 d) *Aniline*: M. Famulok, G. Boche, *Angew. Chem.* **101** (1989) 470; *Angew. Chem. Int. Ed. Engl.* **28** (1989) 468.
- [21] A.M. Lobo, S. Prabhakar, M.M. Marques, *Tetrahedron Lett.* **23** (1982) 1391; A. M. Lobo, M.M. Marques, S. Prabhakar, H.S. Rzepa, *J. Chem. Soc., Chem. Commun.* **1113** (1985); A. M. Lobo, M.M. Marques, S. Prabhakar, H.S. Rzepa, *J. Org. Chem.* **52** (1987) 2925.
- [22]  $^1\text{H}$  nmr (400 MHz; THF- $\text{D}_8$ ;  $^3\text{J}[\text{Hz}]$ :  $\delta$  = 2.10 (s, 3H,  $\text{CH}_3$ ), 3.71 (s, 3H,  $\text{OCH}_3$ ), 6.82 (d, 2H,  $\text{H}^{2,6}$ , 8.94), 6.98 (d, 2H,  $\text{H}^{3,5}$ , 8.80), 9.57 (s, broad, 1 H, NH).

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