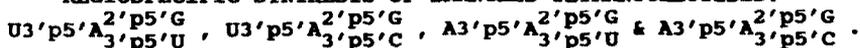


REGIOSPECIFIC SYNTHESIS OF BRANCHED TETRANUCLEOTIDES:



X-X. ZHOU¹, A. NYILAS^{1,2}, G REMAUD¹ and J. CHATTOPADHYAYA^{1*}

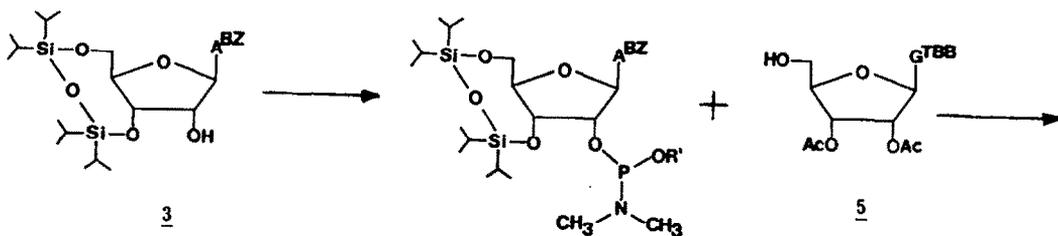
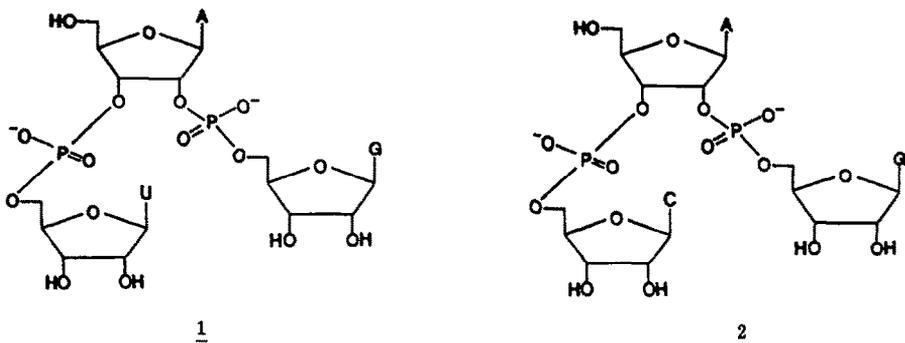
- (1) Department of Bioorganic Chemistry, Box 581, Biomedical Center, University of Uppsala, S-751 23 Uppsala, Sweden
- (2) Center for Agricultural Biotechnology, 2101 Gödöllő, Hungary

(Received in UK 21 July 1987)

Abstract: An efficient general strategy for the synthesis of branched tetranucleotides 14, 15, 24 and 25 is described using key intermediates 9 and 20 to give protected tetranucleotides 13 and 23 which could be specifically deprotected to give either 14 or 24 and 15 or 25 in good yields.

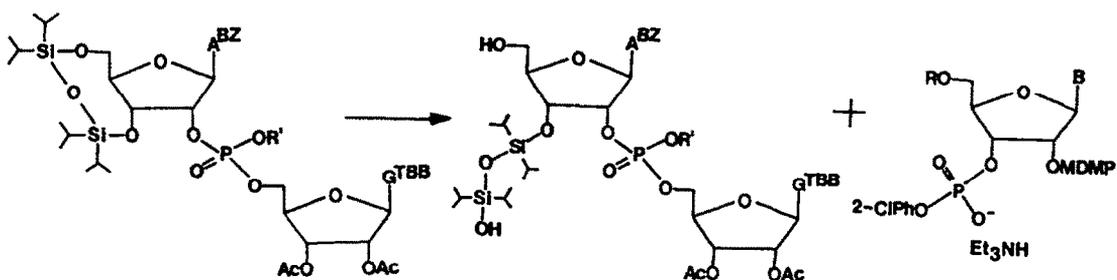
Precision in the chemical processing (splicing) of eukaryotic pre-mRNA, involving the excision of correct sequence of introns and ligation of exons is absolutely essential for the biological activity¹ of the resulting product. In group II and nuclear mRNA splicing, the scission of the intron and the subsequent ligation of exons is processed through the formation of a branched structure (lariat) with the 3'-exon attached giving a circular intron (lariat) and the ligated 5'- and 3'-exons². In such lariat structures, adenosine residue forms the branch point with an additional 2' → 5' nucleobase, which is invariably a guanine residue, while the 3' → 5' nucleobase is either a uracil or a cytosine residue^{2,3}. We have recently reported an unambiguous regiospecific synthesis of simple trimeric branched structures⁴ 1 and 2, which allowed us to carry out their 270 MHz and 500 MHz ¹H-NMR studies. Such conformational studies have revealed an unique structural feature of these branched trinucleotides 1 and 2 consisting of preferential 2' → 5' stacking⁵ (free-energy minimum) and a complete absence of 3' → 5' stacking.

We now report an expedient synthesis of four branched tetranucleotides 14, 15, 24 and 25 comprising of an additional either uridine or adenosine residue at the 5'-end of the branched molecules 1 and 2, in order to address the conformational influence of the fourth base residue at the 5'-terminus on the overall branched structures of 1 and 2. These branched tetranucleotides are naturally occurring and have been actually isolated from yeast cells^{2,3}. In this new procedure, we introduced 2-phenylsulfonyl ethyl (PSE)⁶ or 9-fluorenylmethyl (FM)⁷ phosphiteamidite function selectively at the 2'-OH of the building block 3 to give compound 4 in 86 % yield (³¹P-NMR: 149.9 and 149.7 ppm) and 16 in 55% yield (³¹P-NMR: 149.8 ppm) respectively which we coupled to an appropriate 5'-hydroxy block 5, in presence of tetrazole⁸, to give the fully protected dinucleotide 6 in 79 % yield (³¹P-NMR: -3.6 and -4.3 ppm) or 17 in 56 % yield (³¹P-NMR: -3.29 and -3.88 ppm). The 5'-hydroxyl functions of these dimers 6 and 17 were regiospecifically released⁹, using 0.2 M aqueous HCl in dioxane, to give compound 7 (³¹P-NMR: -3.25 and -3.9 ppm) in 86 % yield and compound 18 (³¹P-NMR: -3.27 ppm) in 70 % yield



4: R' = PSE

16: R' = FM



6: R' = PSE

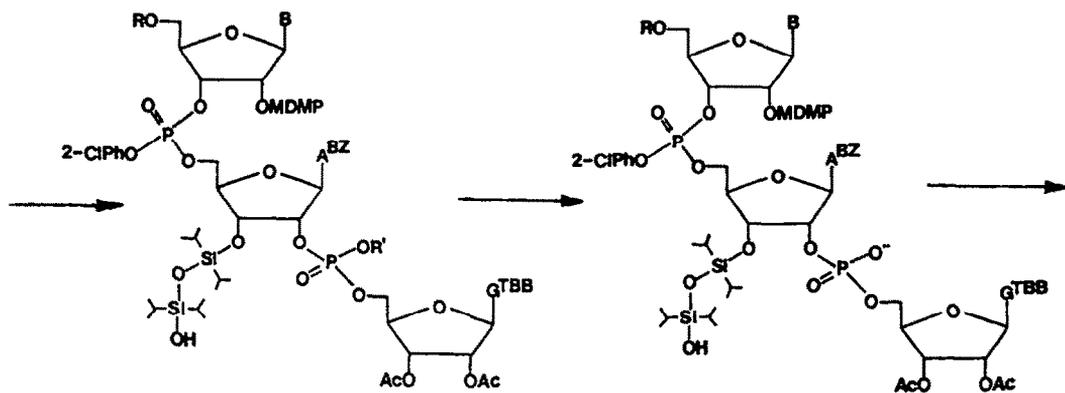
7: R' = PSE

8: B = U; R = DMT

17: R' = FM

18: R' = FM

19: B = A^{Bz}; R = Tol

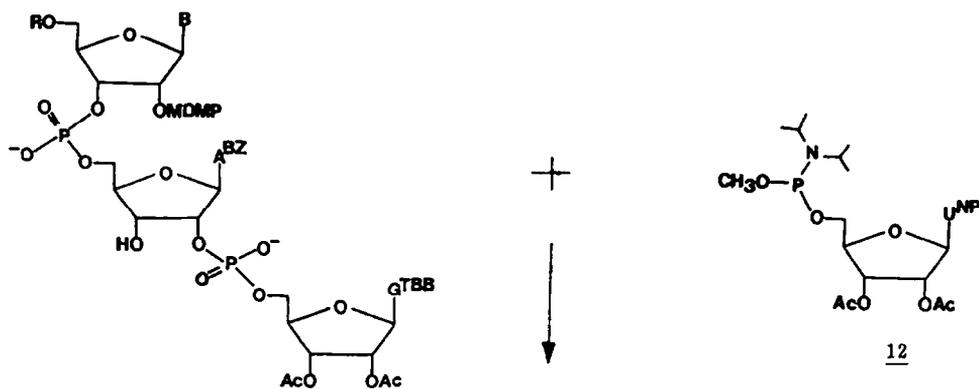


9: B = U; R = DMT; R' = PSE

10: B = U; R = DMT

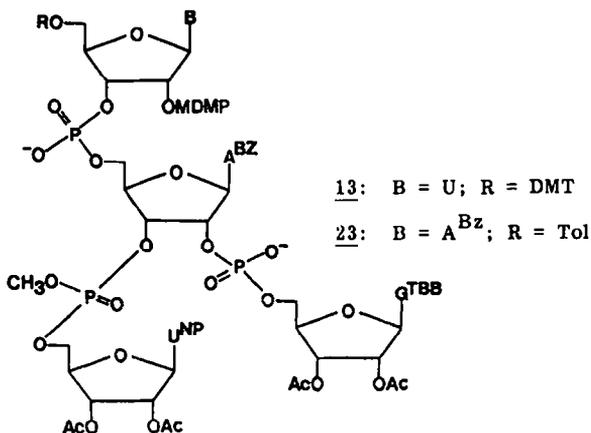
20: B = A^{Bz}; R = Tol; R' = FM

21: B = A^{Bz}; R = Tol



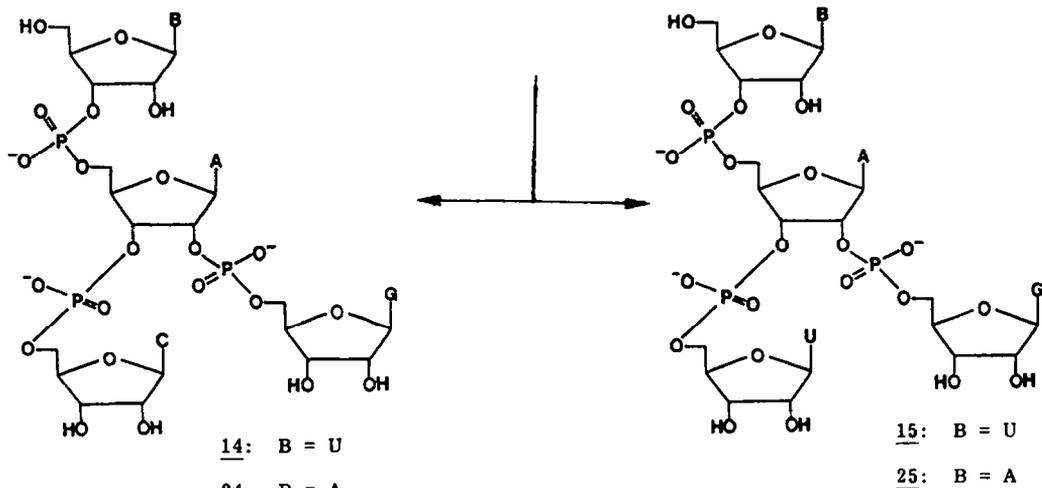
11: B = U; R = DMT

22: B = A^{Bz}; R = Tol



13: B = U; R = DMT

23: B = A^{Bz}; R = Tol

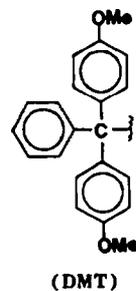
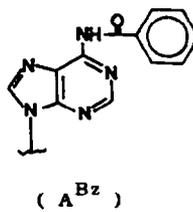
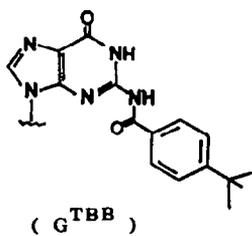
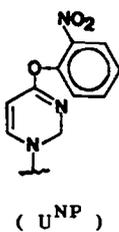
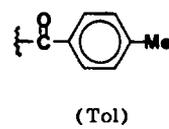
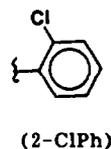
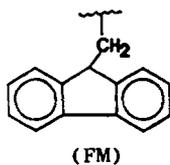
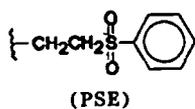
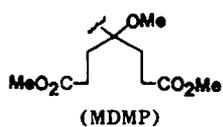


14: B = U

24: B = A

15: B = U

25: B = A



respectively. Chain elongation at the 5'-end was accomplished, using methodologies of the phosphotriester approach¹⁰, by coupling with an appropriate 5'-protected phosphodiester block **8** or **12** in presence of 1-mesitylenesulfonyl-(3-nitro-1,2,4-triazole) [MSNT]¹¹ to give the protected trimer **9** in 87 % yield (³¹P-NMR: -3.4 and -7.4 ppm) or **20** in 78 % yield (³¹P-NMR: -2.61, -2.66, -6.98 and -7.34 ppm). Then the 2-phenylsulphonyl (PSE) group⁶ from compound **9** and the 9-fluorenylmethyl (FM) group⁷ from compound **20** were selectively removed [Et₃N (10 equiv.) in dry pyridine at 20 °C for 2 h] in almost quantitative yields to give compound **10** (³¹P-NMR: -2.64 and -7.5 ppm) and compound **21** (³¹P-NMR: -2.34, -7.03 and -7.57 ppm), respectively. Both 3'-silyl and 2-chlorophenyl groups were then removed in one step using *n*-tetrabutylammonium fluoride¹² in moist tetrahydrofuran at 20 °C to give pure partially protected trimer **11** (³¹P-NMR: -0.7 and -0.8 ppm) in 87 % yield or **22** (³¹P-NMR: -0.59 and -0.68 ppm) in 90 % yield. Finally, compound **11** and **22** were coupled to a 5'-protected phosphiteamidite uridine derivative **12**, in presence

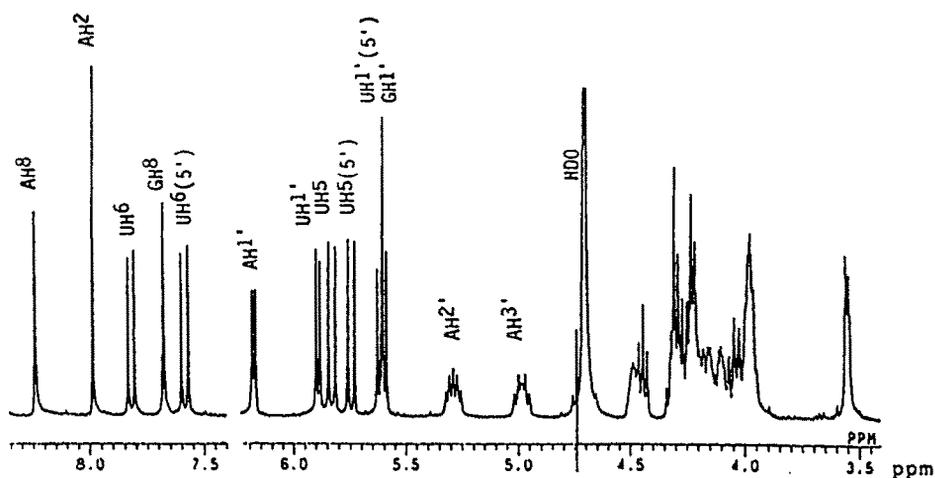


Fig 1: A 270 MHz ¹H-NMR spectrum of U3'p5'A²p5'G (14) in D₂O at 298 K.

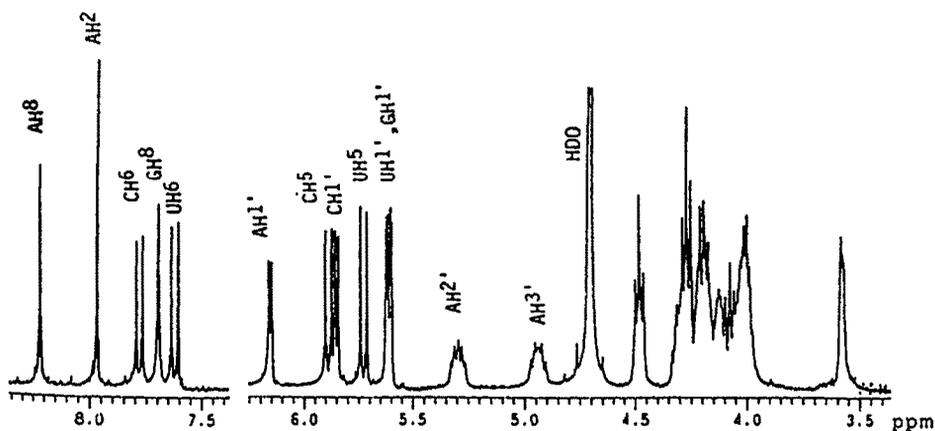


Fig 2: A 270 MHz ¹H-NMR spectrum of U3'p5'A²p5'C (15) in D₂O at 298 K.

of tetrazole, to give tetranucleotides **13** (96 %) and **23** (89 %) which were deprotected in two different ways¹³ to give **14** in 46 % yield (³¹P-NMR: -0.8, -1.0 and -1.2 ppm) and **24** in 41 % yield (³¹P-NMR: -0.83, -0.98 and -1.27 ppm) [liquid

NH_3 for 48 h, aqueous ammonia for 120 h, 80 % aqueous acetic acid for 5 h at 20°C] or to give **15** in 49 % yield (^{31}P -NMR: -0.9, -1.0 and -1.3 ppm) and **25** in 39 % yield (^{31}P -NMR: -0.83, -0.90 and -1.29 ppm) [4-nitrobenzaloximate¹¹ for 24 h, aq. NH_3 for 120 h, 80 % aqueous acetic acid for 5 h]. The 270 MHz 1D ^1H -NMR spectra, 2D ^1H correlations and $^{31}\text{P}/^1\text{H}$ correlation spectra of the tetranucleotides in D_2O are shown in Figures 1 - 12 which show that the desired regiospecificities of the phosphodiester linkages have indeed been achieved. Assignment of resonances with practical details will be described elsewhere.

EXPERIMENTAL

^1H -NMR spectra were recorded, in δ scale, with Jeol 90 Q and JNM GX 270 spectrometers at 90 and 270 MHz respectively, using TMS or acetonitrile (set at 2.0 ppm) as internal standards. ^{31}P -NMR spectra were recorded at 36 MHz in the same solvent using phosphoric acid as an external standard and chemical shifts quoted are in ppm. TLC was carried out using pre-coated silica gel F_{254} plate in following solvent systems: (A) methanol-dichloromethane [9.5:0.5 (v/v)], (B) methanol-dichloromethane [9:1 (v/v)], (C) methanol-dichloromethane [8:2 (v/v)], (D) ethylacetate-hexane-triethylamine [4:4:2 (v/v/v)]. The short column chromatographic separations were carried out using Merck G60 silica gel.

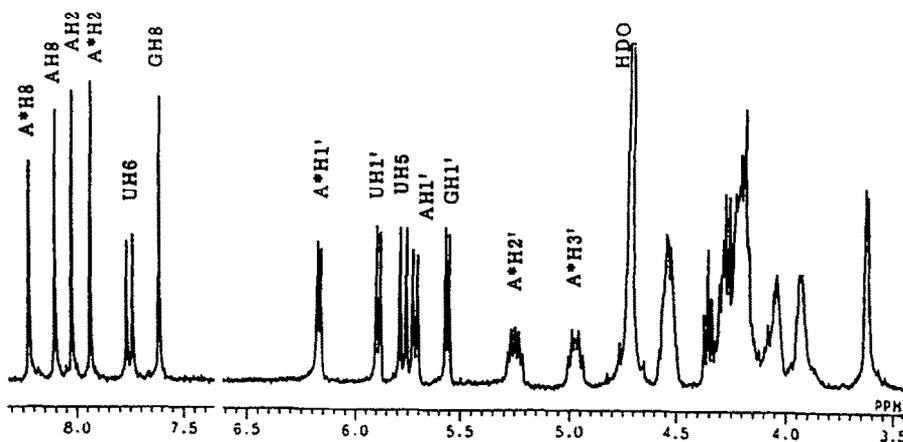


Fig. 3: 270 MHz ^1H -NMR spectrum of $\text{A}_3'\text{p}_5'\text{A}^{*2'}\text{p}_5'\text{G}_3'\text{p}_5'\text{U}$ in D_2O at 298 K.

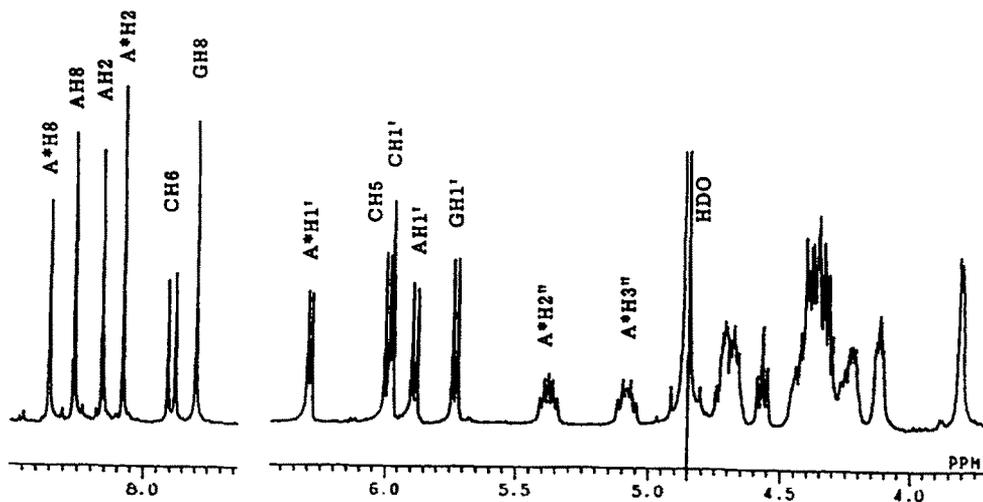
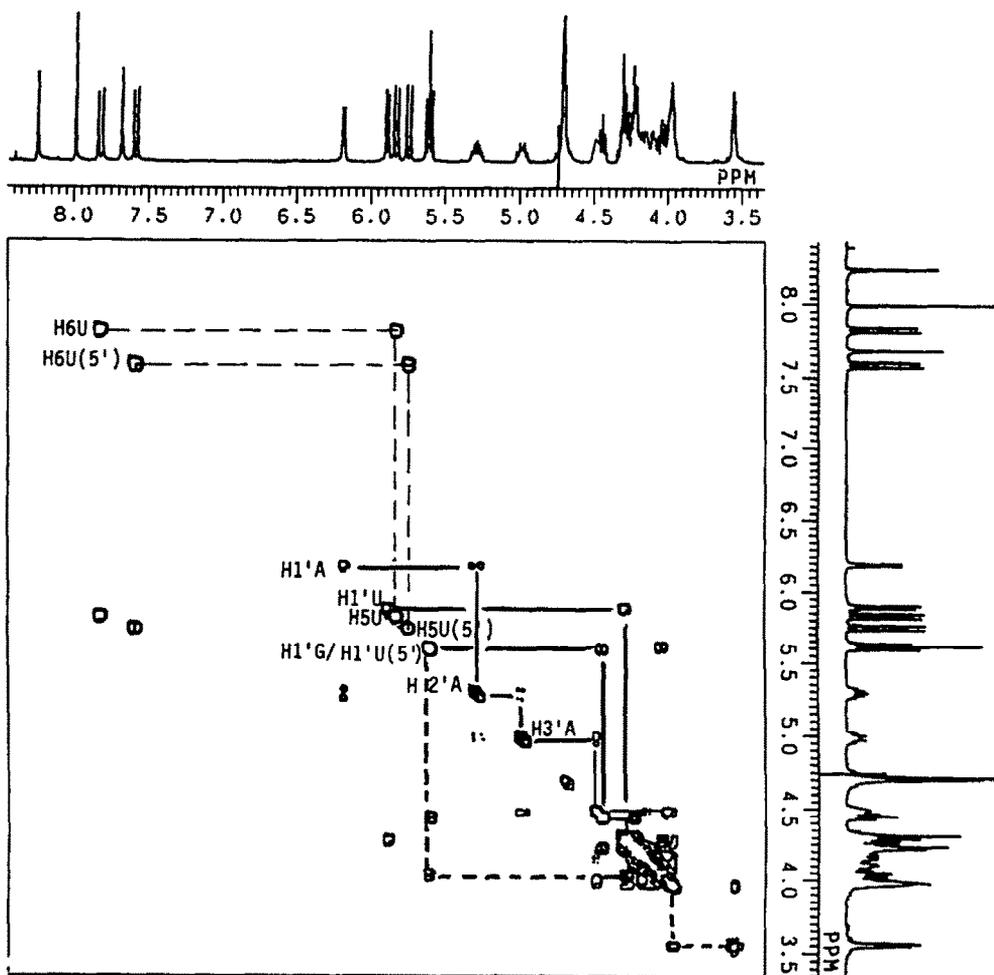


Fig. 4: 270 MHz ^1H -NMR spectrum of $\text{A}_3'\text{p}_5'\text{A}^{*2'}\text{p}_5'\text{G}_3'\text{p}_5'\text{C}$ in D_2O at 298 K.



DOUBLE QUANTUM FILTER COSY OF $U3'p5'A^{2'}p5'G$
 $3'p5'U$

Fig. 5

6-N-Benzoyl-3',5'-O-(tetraisopropyl-1,3-disiloxane-1,3-diyl)-3'-O-phosphiteamidite (4):

To a suspension of 2-phenylsulfonyl ethylphosphorodichloridite (2.29 g, 8 mmol) and 1,2,4-triazole (1.93 g, 28 mmol) in dry THF (20 ml) at -25°C , was added *N,N*-diisopropylethylamine (4.89 ml, 28 mmol). After stirring the reaction mixture for 12 min at -25°C , a solution of compound **3** (2.4 g, 4 mmol) in dry THF (20 ml) was added dropwise within 1 h and kept it stirring for 40 min, trimethylsilyl-*N,N*-dimethylamine (4.4 ml, 28 mmol) was then added. After 10 min, the reaction mixture was warmed up to room temperature, and kept stirring for a further period of 15 min. The reaction mixture was subsequently poured into saturated aqueous sodium chloride solution (150 ml), and extracted with ethylacetate (100 ml). The organic layer was washed with sodium chloride solution and water successively, and dried *in vacuo*. The residue was purified by short silica gel column chromatography, [hexane: dichloromethane:ethylacetate:triethylamine, 3:3:1:1 (v/v/v/v)], the product was precipitated from cooled hexane (-70°C). Yield: 2.95 g (86 %) $R_f = 0.6$

(solvent D). $^1\text{H-NMR}$ (mixture of two diastereomers) ($\text{CDCl}_3 + \text{pyridine-d}_5$): 8.74 and 8.73 (2 x \underline{s} , 1H) H-8; 8.33 and 8.32 (2 x \underline{s} , 1H) H-2; 8.08-7.19 (m, 10H) BZ and PSE; 6.05 (m, 1H) H-1'; 4.66 (m, 2H) H-2', H-3'; 4.15 (m, 5H) H-4', H-5', PSE; 3.42 (m, 2H) PSE; 2.61, 2.58, 2.51 and 2.48 (4 x \underline{s} , 6H) $(\text{CH}_3)_2\text{N-}$; 1.05 (m, 28H)

TIPDSi. $^{31}\text{P-NMR}$ (CDCl_3): 149.9 and 149.7.

Synthesis of fully protected dinucleoside monophosphate (6)

A mixture of compounds **4** (2.3 g, 3 mmol), **5** (527 mg, 1 mmol) and 1,2,3,4-tetrazole (1.4 g, 20 mmol) was dissolved in dry acetonitrile (20 ml). After 40 min, iodine solution (0.1 M) in pyridine: THF: water mixture (1:8:1, v/v/v) was added until the iodine color was not further discharged. After 20 min, the reaction mixture was worked up in the usual way. The product was purified by short silica gel column chromatography. Yield 1.07 g (79 %) $R_f = 0.4$ (solvent B). $^{31}\text{P-NMR}$ (CDCl_3): -3.56 and -4.27.

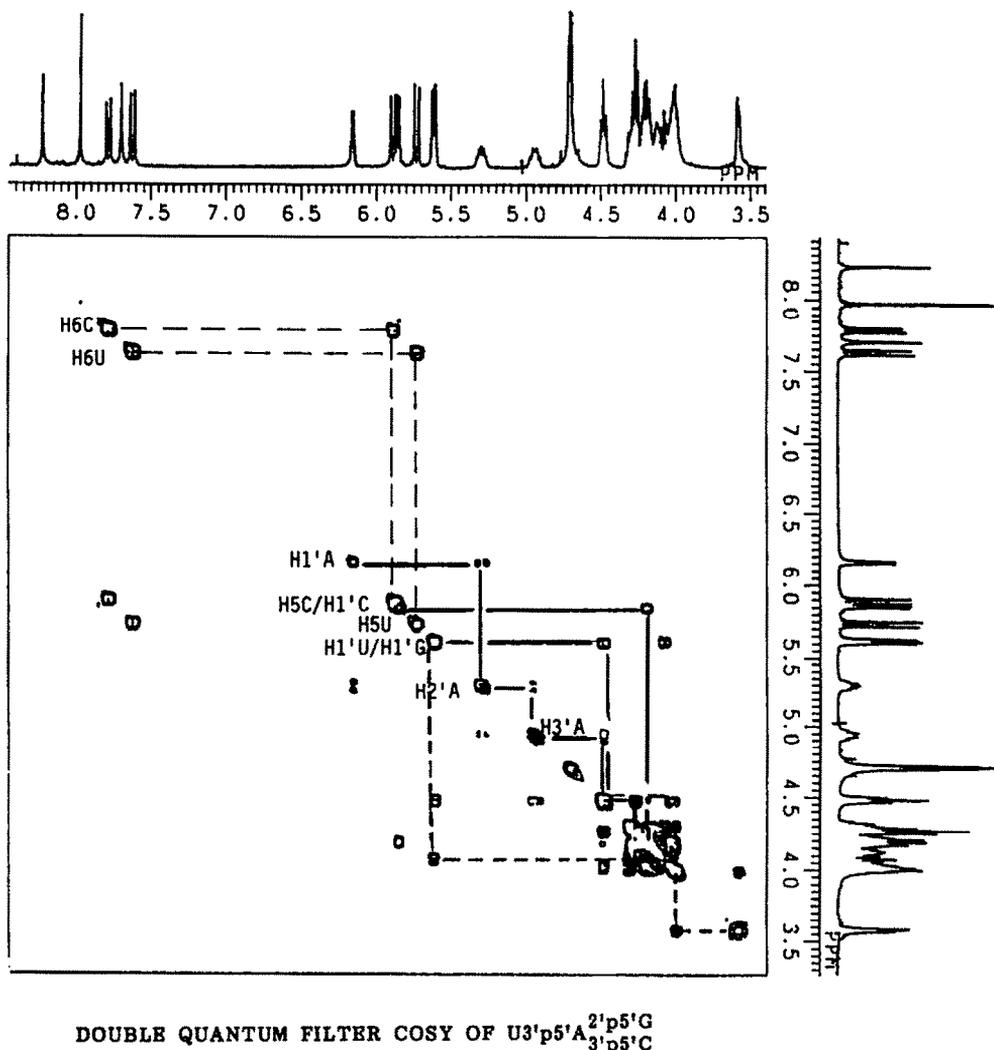


Fig. 6

Synthesis of partially protected dinucleoside monophosphate (7)

Compound **6** was dissolved in 0.2 M aqueous HCl in dioxane (14 ml) under stirring at 20 °C. A few drops of 0.2 M aqueous HCl were subsequently added until the reaction mixture became opalescent. After 1 h, the mixture was poured into saturated sodium hydrogen carbonate solution (100 ml) and extracted with dichloromethane (3 x 40 ml). The extract was concentrated and chromatographed by short silica gel column chromatography. Yield: 708 mg (86 %). $R_f = 0.35$ and 0.37 (solvent B).

$^{31}\text{P-NMR}$. (CDCl_3): -3.25 and -3.88.

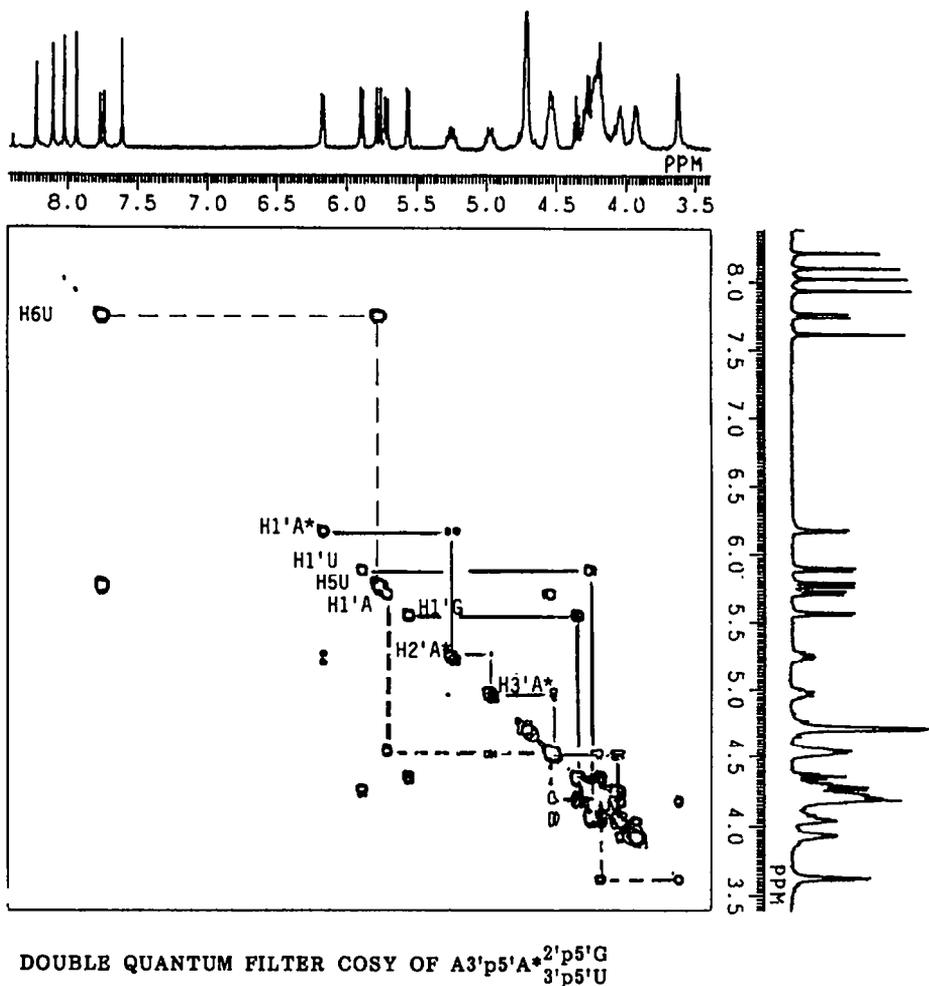


Fig. 7

Synthesis of fully protected trimer (9)

Compound **7** (487 mg, 0.35 mmol) and **8** (707 mg, 0.7 mmol) were dissolved in dry pyridine (10 ml). To this solution was added 1-mesitylsulfonyl-3-nitro-1,2,4-triazole (MSNT) (1.6 g, 4.9 mmol) at 20 °C. After 20 min, the reaction mixture was poured into a saturated sodium hydrogen carbonate solution (80 ml), and extracted with dichloromethane (3 x 30 ml). The product was subsequently isolated by short silica gel column chromatography. Yield: 692 mg (87 %). $R_f = 0.4$ (solvent B).

$^{31}\text{P-NMR}$ ($\text{CDCl}_3 + \text{pyridine-d}_5$): -3.39 and -7.37.

Synthesis of compound (10)

Compound **9** (680 mg, 0.3 mmol) was dissolved in pyridine (10.5 ml), followed by triethylamine (0.83 ml, 6 mmol). After 90 min at 20 °C, the reaction mixture was dried *in vacuo*, and purified by short silica gel column. Yield: 614 mg (99 %). $R_f = 0.3$ (solvent B). $^{31}\text{P-NMR}$ ($\text{CDCl}_3 + \text{pyridine-d}_5$): -2.64 and -7.49.

Synthesis of compound (11)

Compound **10** (600 mg, 0.29 mmol) was dissolved in THF (10 ml) and, subsequently, 1 M tetrabutylammonium fluoride in moist THF (1.74 ml) was added at 20 °C. After 1 h, the reaction mixture was dried *in vacuo* and purified by silica gel column chromatography. Yield: 480 mg (86 %). $R_f = 0.2$ (solvent C). $^{31}\text{P-NMR}$ ($\text{CDCl}_3 + \text{pyridine-d}_5$): -0.71 and -0.78.

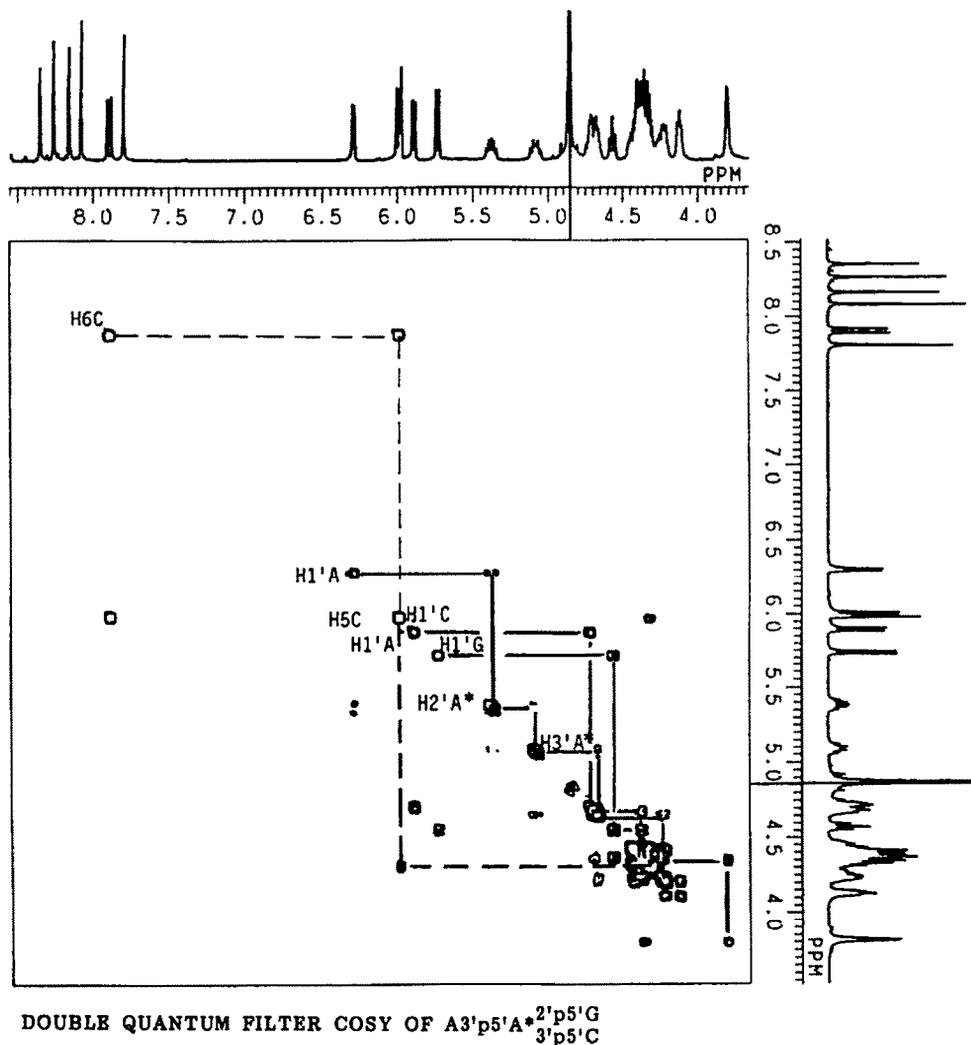


Fig. 8

Synthesis of compound (12)

O⁴-(2-nitrophenyl)-2',3'-O-diacetyluridine (1.35 g, 3 mmol) was dissolved in dichloromethane (25 ml) and subsequently *N,N*-diisopropylethylamine (2.1 ml, 12 mmol) and methoxy diisopropylaminochlorophosphine (1.14 ml, 6 mmol) were successively added. After 50 min, the reaction mixture was poured into a saturated sodium chloride solution (100 ml) and extracted with ethylacetate (4 x 50 ml). The product was subsequently purified by silica gel column chromatography. Yield:

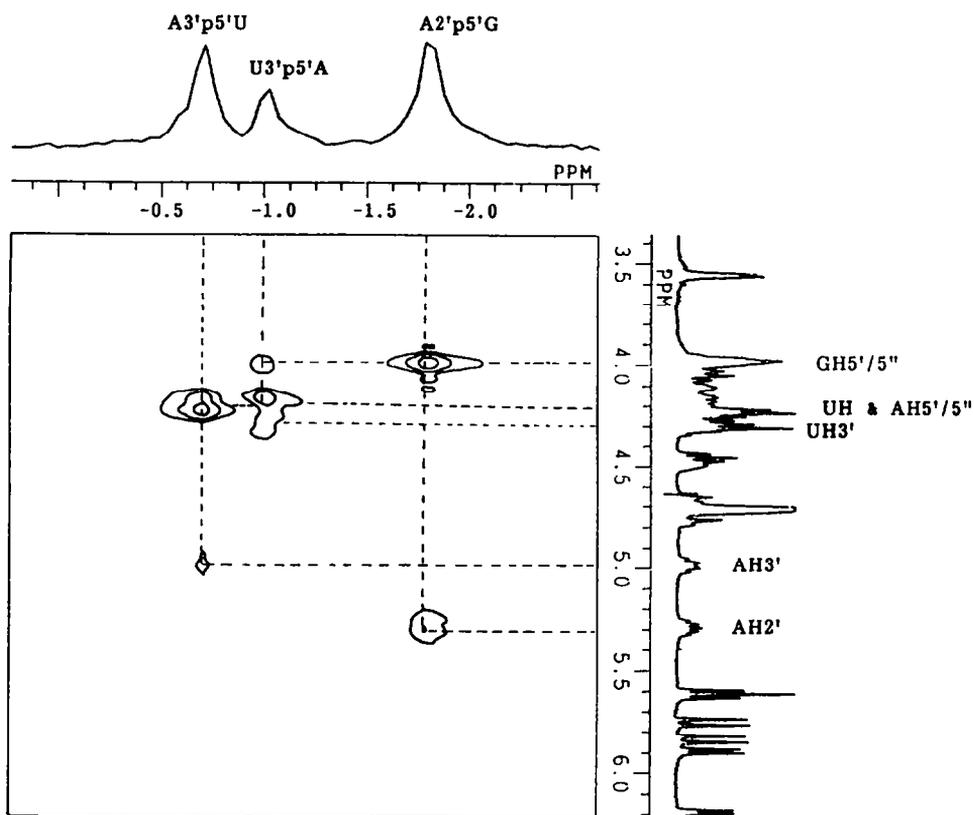
1.61 g (90 %). $R_f = 0.65$ (solvent D). $^{31}\text{P-NMR}$ (CDCl_3): 150.5 and 150.4.

Synthesis of compound (13)

Compound **11** (350 mg, 0.2 mmol) and **12** (596 mg, 1 mmol) and tetrazole (700 mg, 10 mmol) were dissolved in dry acetonitrile (7 ml). After 30 min, the reaction mixture was oxidized with 0.1 M iodine solution in THF-pyridine-water mixture (1:8:1, v/v/v) and then worked up in the usual way. The column chromatographic separation was carried out, first, by washing the column with 4 % methanol in dichloromethane and then the product was eluted out by 30 % methanol in dichloromethane. Yield: 428 mg (96 %). $R_f = 0.2$ (solvent C). $^{31}\text{P-NMR}$ ($\text{CDCl}_3 + \text{pyridine-d}_5$): -0.78, -0.85, -1.32, -1.42, -2.22, -2.78.

Deprotection of compound (13)

Procedure A: Compound **13** (200 mg, 0.09 mmol) was dissolved in dioxane-water mixture (30 ml, 8:2 v/v) and then *N,N,N,N*-tetramethylguanidine (0.36 ml, 2.79 mmol)



^{31}P - ^1H CORRELATION SPECTRUM OF $\text{U3}'\text{p5}'\text{A}^{2'}\text{p5}'\text{G}$

Fig. 9

and *syn*-*p*-nitrobenzaloxime (448 mg, 2.7 mmol) were added. After 24 h at 20 °C, concentrated ammonia (70 ml, $d = 0.9$) was added. The reaction mixture was kept stirring for 6 days at 20 °C, and then dried *in vacuo*. The residue was coevaporated with water once, and then treated with 80 % aqueous acetic acid (50 ml) for 5 h. The reaction mixture was dried *in vacuo*, residue dissolved in water and extracted with dichloromethane (6 x 20 ml). The aqueous layer was dried *in vacuo* and subjected to DEAE-Sephadex A25 column chromatographic separation using a linear gradient (0 to 0.5 M) of triethylammonium bicarbonate (TEAB) (pH 7.4) as an eluent. Appropriate fractions were pooled and concentrated. The residue was subsequently coevaporated with distilled water several times to remove traces of buffer to give pure compound **15**. Yield: 1940 A_{260} o.d. units (49%). ^{31}P -NMR (D_2O): -0.88, -0.98, -1.30.

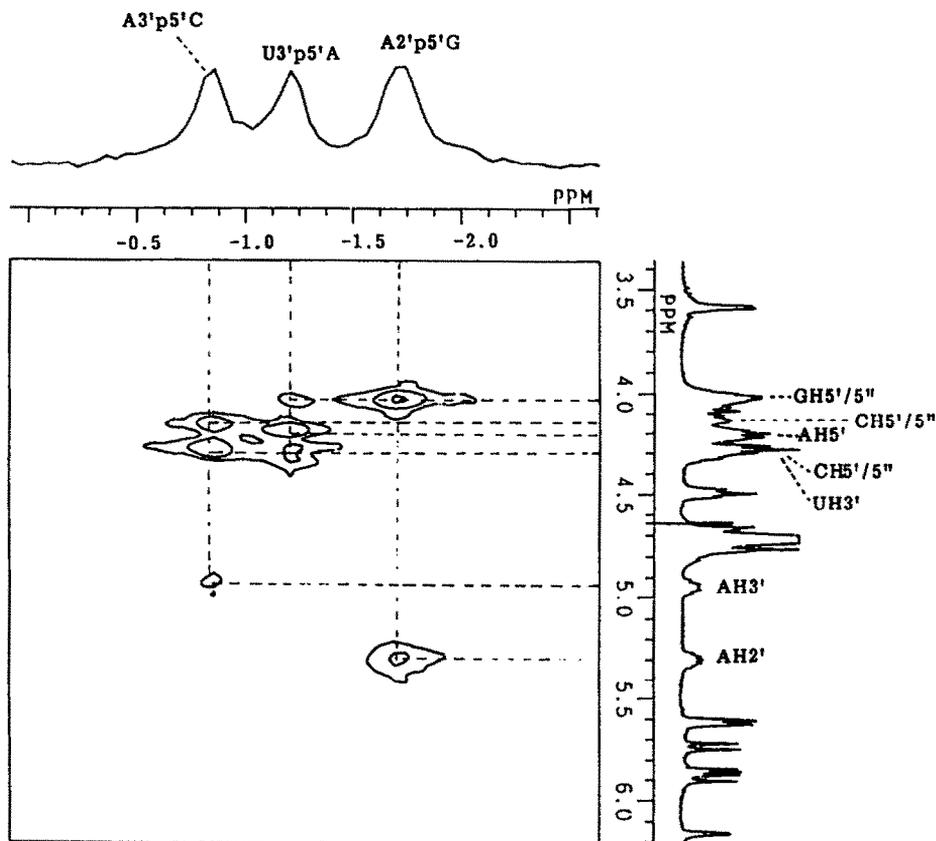
Procedure B: Compound **13** (200 mg, 0.09 mmol) was treated with liquid ammonia (30 ml) for 2 days at 20 °C, then the volatile matters were evaporated. The residue was then dissolved in concentrated ammonia (70 ml, $d = 0.9$). After 6 days, the solution was dried *in vacuo*. After a few coevaporations with water, it was treated with 80 % aqueous acetic acid (50 ml) for 5 h at 20 °C. The reaction mixture was dried *in vacuo*, the residue was partitioned between water (50 ml) and dichloromethane (50 ml). The aqueous layer was further extracted with dichloromethane (3 x 40 ml) and subsequently aqueous layer was concentrated and applied on a DEAE-sephadex A25 column using a linear gradient of 0 to 0.5 M TEAB solution (pH 7.4) as eluent. Appropriate fractions were pooled, concentrated, coevaporated with distilled water several times to give compound **14**. Yield: 1718 A_{260} o.d. units (46 %). ^{31}P -NMR (D_2O): -0.83, -0.98, -1.20.

Preparation of compound (16)

To the mixture of 9-fluorenylmethylphosphorodichloridite (1.78 g, 6 mmol) and 1,2,4-triazole (1.45 g, 21 mmol) in dry THF at -25 °C, was added *N,N*-diisopropylethylamine (3.66 ml, 21 mmol). After stirring for 10 min, a solution of compound **3**

(1.84 g, 3 mmol) in THF (10 ml) was added dropwise. The mixture was kept for 30 min and then trimethylsilyl-*N,N*-dimethylamine (3.3 ml, 21 mmol) was added. After 10 min, the reaction mixture was warmed up to room temperature, and kept stirring for a further period of 10 min. It was then worked up and separated by the procedure described for the preparation of compound **4**. Yield: 1.46 g (55 %).

^{31}P -NMR (CDCl_3): 149.8.



^{31}P - ^1H CORRELATION SPECTRUM OF $\text{U3'p5'A}^{2'p5'G}$
 $^{3'p5'C}$

Fig. 10

Preparation of compound (17)

Compound **16** (880 mg, 1 mmol) and **5** (220 mg, 0.5 mmol) were coupled in presence of 1,2,3,4-tetrazole (0.7 g, 2.4 mmol) in dry THF as described for the preparation of compound **6**. Yield: 500 mg (56 %). ^{31}P -NMR (CDCl_3): -3.29 and -3.88.

Preparation of compound (18)

Compound **17** (500 mg, 0.33 mmol) was treated with 0.2 M aqueous HCl in dioxane using a procedure as described for the preparation of compound **7**. Yield: 350 mg (70 %). ^{31}P -NMR (CDCl_3): -3.27.

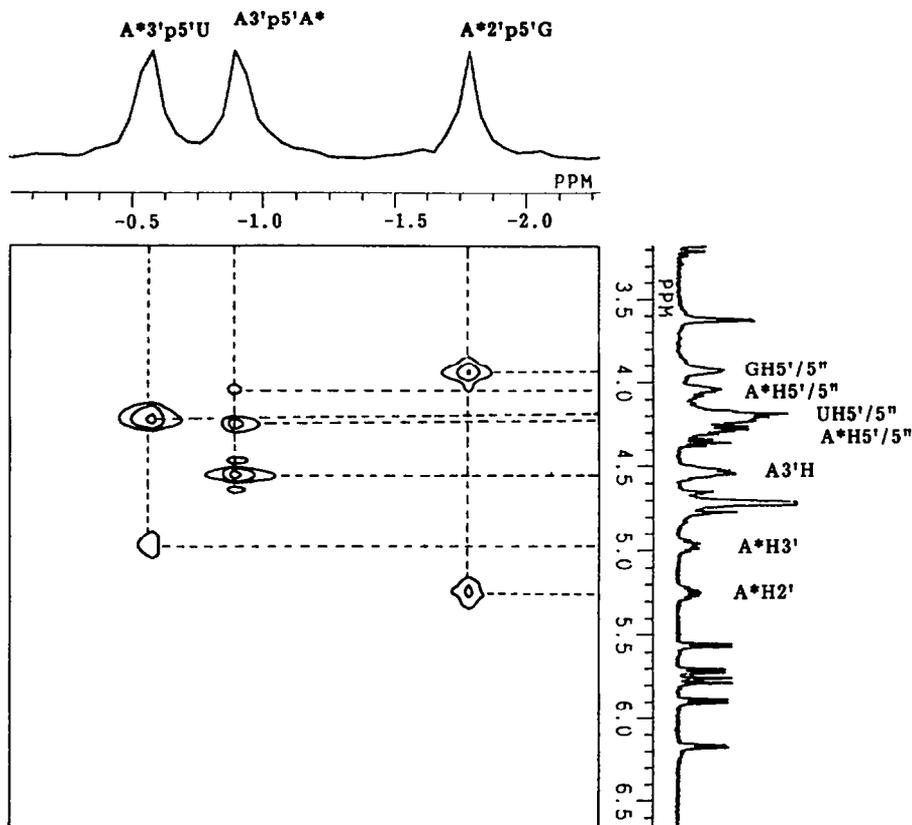
Preparation of compound (20)

Compound **18** (500 mg, 0.37 mmol) and **19** (725 mg, 0.74 mmol) and MSNT (746 mg, 2.2 mmol) were reacted in pyridine solution according to the method described for the preparation of compound **2**. Yield: 634 mg (77.5 %). $R_f = 0.38$ (solvent B).

^{31}P -NMR (CDCl_3): -2.61, -2.66, -6.98, -7.34.

Preparation of compound (21)

Compound **20** (630 mg, 0.28 mmol) was treated with triethylamine in dry pyridine in a similar way as described for the preparation of compound **10**. Yield: 601 mg (100 %). $R_f = 0.27$ (solvent B). $^{31}\text{P-NMR}$ (CDCl_3): -2.34, -7.03, -7.57.



$^{31}\text{P-}^1\text{H}$ CORRELATION SPECTRUM OF $\text{A}3'\text{p}5'\text{A}^*2'\text{p}5'\text{G}$
 $3'\text{p}5'\text{U}$

Fig. 11

Preparation of compound (22)

Compound **21** (598 mg, 0.28 mmol) was treated with tetrabutylammonium fluoride in dry THF in a similar way as described for the preparation of compound **11**. Yield: 449 mg (90 %). $R_f = 0.20$ (solvent C). $^{31}\text{P-NMR}$ (CDCl_3): -0.59 and -0.68.

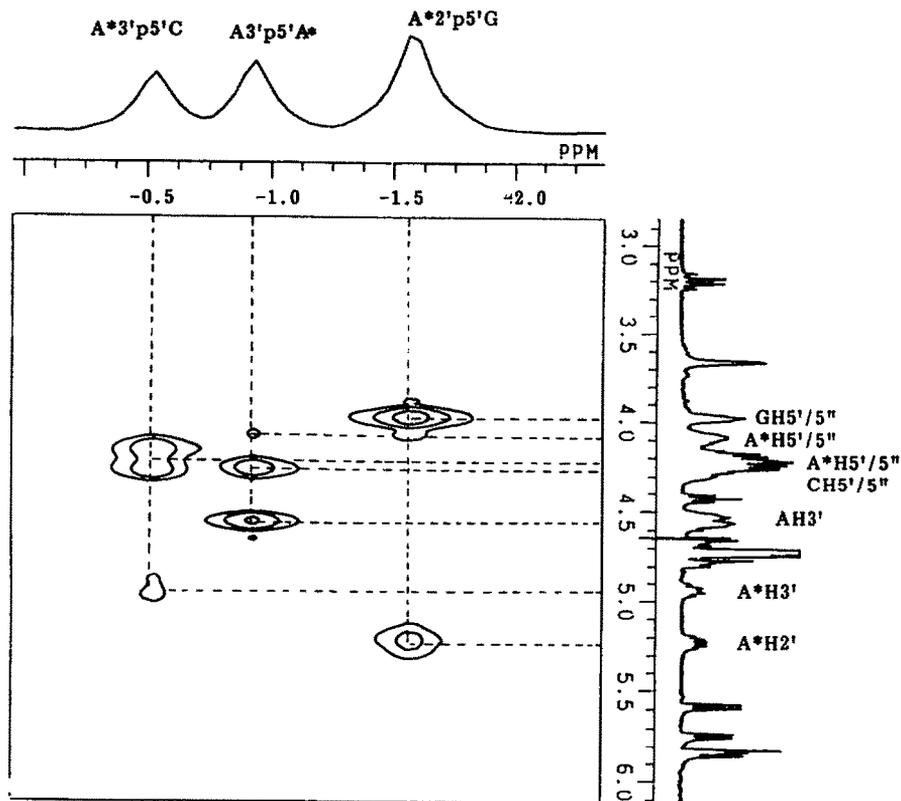
Preparation of compound (23)

Compound **22** (390 mg, 0.22 mmol) was condensed with compound **12** (655 mg, 1.1 mmol) in acetonitrile in presence of tetrazole (770 mg, 11 mmol) in the usual way. Yield: 472 mg (89 %). $R_f = 0.27$ (solvent C). $^{31}\text{P-NMR}$ (CDCl_3): -0.68, -0.78, -1.00, -1.17, -2.61, -2.71.

Deprotection of compound (23)

Procedure A: Compound **23** (204 mg, 0.085 mmol) was deprotected and purified in a similar way as described for the preparation of compound **15** to give compound **25**. Yield: 1641 A_{260} o.d. units (39 %). $^{31}\text{P-NMR}$ (D_2O): -0.83, -0.90, -1.29.

Procedure B: Compound **23** (204 mg, 0.085 mmol) was deprotected and purified in the similar way as preparation of compound **14** to get compound **24**. Yield: 1636 A_{260} units (41 %). $^{31}\text{P-NMR}$ (D_2O): -0.83, -0.98, -1.27.



^{31}P - ^1H CORRELATION SPECTRUM OF $\text{A}_3'\text{p}_5'\text{A}^*2'\text{p}_5'\text{G}$
 $3'\text{p}_5'\text{C}$

Fig. 12

ACKNOWLEDGEMENTS

The authors gratefully acknowledge financial supports from the Swedish Board for Technical Development and the Natural Science Research Council and thank Ms. Ingegård Schiller for her excellent secretarial assistance.

REFERENCES

- M.M. Konarska, P.J. Grabowski, R.A. Padgett, P.A. Sharp. *Nature (London)*, **313**, 552 (1985).
 - B. Ruskin, A.R. Krainer, T. Maniatis, M.R. Green. *Cell*, **39**, 317 (1984).
 - R.A. Padgett, P.J. Grabowski, M.M. Koarska, S. Seiler, P.A. Sharp. *Ann. Rev. Biochem.*, **55**, 1119 (1986).
 - B. Ruskin, M.R. Green. *Science*, **229**, 135 (1985).
 - T.R. Cech, B.L. Bass. *Ann. Rev. Biochem.*, **55**, 599 (1986).
- W. Keller. *Cell*, **39**, 423 (1984) and references cited therein.
 - T.R. Cech. *Cell*, **44**, 207 (1986) and references cited therein.
- M. Aebi, H. Hornig, R.A. Padgett, J. Reiser, C. Weissmann. *Cell*, **47**, 555 (1986).
 - H. Hornig, M. Aebi, C. Weissmann. *Nature (London)*, **324**, 589 (1986).
 - G.A. Freyer, J. Arenas, K.K. Perkins, H.M. Furneaux, L. Pick, B. Yound, R.J. Roberts, J. Hurnitz. *J. Mol. Biol.*, **262**, 4267 (1987).
- J.-M. Vial, N. Balgobin, G. Renaud, A. Nyilas and J. Chattopadhyaya. *Nucleosides & Nucleotides*, **6** (1,2), 209 (1987).

5. (a) G. Remaud, J-M. Vial, A. Nyilas, N. Balgobin and J. Chattopadhyaya. Tetrahedron, **43**, 947, (1987).
- (b) J-M. Vial, G. Remaud, N. Balgobin and J. Chattopadhyaya. Tetrahedron, in press.
- (c) L.H. Koole, N. Balgobin, H.M. Buck, W.H.A. Kuijpers, A. Nyilas, G. Remaud, J.M. Vial and J. Chattopadhyaya, J. Am. Chem. Soc. (submitted).
6. (a) N. Balgobin, S. Josephson and J. Chattopadhyaya. Tetrahedron Lett., **22**, 1915 (1981).
- (b) N. Balgobin and J. Chattopadhyaya. Acta Chem. Scand., **B39**, 883 (1985).
7. (a) C. Gioeli and J. Chattopadhyaya. Chemica Scripta, **19**, 235 (1982).
- (b) C.A.A. Claesen, R.P.A.M. Segers and G.I. Tesser. Rec. Trav. Chim. PaysBas, **104**, 209 (1985).
8. (a) M.H. Caruthers and S.L. Beaucage. Tetrahedron Lett., **22**, 1859 (1981).
- (b) L.J. McBride and M.H. Caruthers. Tetrahedron Lett., **24**, 245 (1983).
9. (a) C. Gioeli, M. Kwiatkowski, B. Öberg and J. Chattopadhyaya. Tetrahedron Lett., **22**, 1741 (1981).
- (b) C. Gioeli, J. Chattopadhyaya, B. Öberg and A.F. Drake. Chemica Scripta, **19**, 13 (1982).
10. C.B. Reese. Tetrahedron, **34**, 3143 (1978).
11. C.B. Reese, R.C. Titmus and L. Yau. Tetrahedron Lett., 2727 (1978).
12. G. Kumar, L. Celewicz and S. Chladek. J. Org. Chem., **47**, 634 (1982).
13. X-X. Zhou and J. Chattopadhyaya. Tetrahedron, **42**, 5149 (1986).