



BIOORGANIC & MEDICINAL CHEMISTRY LETTERS

Bioorganic & Medicinal Chemistry Letters 13 (2003) 2973-2976

Discovery and Evaluation of 3-(5-Thien-3-ylpyridin-3-yl)-1*H*indoles as a Novel Class of KDR Kinase Inhibitors

Mark E. Fraley,* Kenneth L. Arrington, Scott R. Hambaugh, William F. Hoffman, April M. Cunningham, Mary Beth Young, Randall W. Hungate, Andrew J. Tebben, Ruth Z. Rutledge, Richard L. Kendall, William R. Huckle, Rosemary C. McFall, Kathleen E. Coll and Kenneth A. Thomas

Departments of Medicinal Chemistry and Cancer Research, Merck Research Laboratories, West Point, PA 19486, USA

Received 29 January 2003; accepted 14 February 2003

Abstract—We have discovered 3-(5-thien-3-ylpyridin-3-yl)-1*H*-indoles as potent inhibitors of KDR kinase activity. This communication details the evolution of this novel class from a potent screening lead of vastly different structure with an emphasis on structural modifications that retained activity and provided improvements in key physical properties. The synthesis and in-depth evaluation of these inhibitors are described.

© 2003 Elsevier Ltd. All rights reserved.

Angiogenesis, the formation of new capillaries from established blood vessels, is an essential process for normal growth and development that has also been implicated in the pathogenesis of several diseases including diabetic retinopathy,1 rheumatoid arthritis,2 psoriasis,³ and cancer.⁴ Specifically, the growth and metastasis of solid tumors has been shown to be dependent on angiogenesis at an early stage,⁵ while tumors that lack adequate vascularization become necrotic or apoptotic and do not grow beyond a limited size.⁶ Interest in inhibition of angiogenesis as a new approach for the treatment of cancer has led to the elucidation of key underlying molecular mechanisms that control the angiogenic process. Angiogenesis is regulated by the expression of a variety of growth factors including vascular endothelial growth factor (VEGF), a selective mitogen for endothelial cells whose mitogenic signaling is mediated through the receptor tyrosine kinase KDR (VEGFR-2).⁷ Several lines of evidence indicate that expression and signaling of VEGF are critical for tumor angiogenesis. Among these, antibodies against VEGF⁸ and its receptor KDR⁹ as well as small molecule inhibitors of KDR kinase activity¹⁰ have been shown to inhibit angiogenesis in tumor xenograft models. Clinical trials have been initiated for KDR kinase inhibitors

*Corresponding author. Fax: +1-215-652-6345; e-mail: mark_fraley@merck.com

derived from a number of different structural classes, including indolin-2-ones, phthalazines, and quinazo-lines.¹¹

In our efforts to discover novel small molecule inhibitors of KDR kinase activity, a screening of a large nondirected library of 5-acetamido-2,4-diaryloxazoles produced lead compound 1 (Fig. 1). Compound 1 exhibited good intrinsic potency (KDR $IC_{50} = 16 \text{ nM})^{12}$ and reasonable cellular activity (cell $IC_{50} = 320 \text{ nM}$).¹³

However, we concluded from additional studies that the poor physical properties of **1**, including low aqueous solubility (<0.005 mg/mL) and high lipophilicity (logP > 4.1),¹⁴ would hinder its development. Toward improving these physical properties, we initially investigated more polar replacements for the 2-hydroxy-3-naphthyl moiety at C-2 of the oxazole ring system. Of



Figure 1. Screening lead 1.

0960-894X/03/\$ - see front matter \odot 2003 Elsevier Ltd. All rights reserved. doi:10.1016/S0960-894X(03)00627-9







the wide range of aryl and heterocyclic substituents examined at C-2, all were relatively inactive with the exception of a small subset of ortho-phenols and, more interestingly, the 3-pyridyl ring system found in compound 2a (Table 1).^{15,16} The 3-fold greater potency provided by the thienyl substituent of 2a compared to phenyl group within **2b** and dramatic loss in potency upon substitution of this phenyl group (cf 2c-f) were reminiscent of the SAR established for the pyrazolo[1,5*a*]pyrimidine class of KDR kinase inhibitors.¹⁷ We proposed a similar binding mode that placed the pyridyl ring in the adenine binding region of the ATP active site and allowed the pyridyl nitrogen to engage in a hydrogen bond with Cys 919 of the hinge region.¹⁸ We then envisioned the thienyl group fitting into the sterically confined hydrophobic region I and the oxazolyl moiety filling hydrophobic region II.

Because compound 2a showed little improvement in physical properties (aq soln $< 0.005 \text{ mg/mL}, \log P > 4.0$) over 1, we continued to focus on major structural modifications. Based on our limited understanding of the binding mode of 2a, we speculated that potency would be maintained by replacement of the oxazolyl appendage at C-3 of the pyridyl ring system with a planar substituent of lower molecular weight. A survey of a set of aryl and heterocyclic groups at C-3 culminated in the identification of the 3-indolyl substituent (3a, Table 2) as the optimal group in terms of retaining activity and providing an appropriate point of attachment for solubilizing functionality, via the indolyl nitrogen. We took advantage of this key feature in the design and synthesis of compounds **3b-h** (Scheme 1). Briefly, palladium-catalyzed cross-coupling of 3-bromo-5-thien-3
 Table 2.
 KDR kinase activity of compounds 3a-h





ylpyridine and 1-[(4-methylphenyl)sulfonyl]-1H-indol-3ylboronic acid¹⁹ under Suzuki conditions proceeded smoothly to give 1-[(4-methylphenyl)sulfonyl]-3-(5thien-3-ylpyridin-3-yl)-1H-indole. Hydrolysis of the tosyl group with potassium hydroxide afforded **3a**. Deprotonation of **3a** with sodium hydride and alkylation of the resultant anion with the corresponding mustard-like amines proceeded in moderate to excellent yield to furnish compounds **3b–h**.²⁰



Scheme 1. Synthesis of compounds 3a-h.



Figure 2. Proposed binding mode of 3f.

The addition of the alkyl amines to 3a was well-tolerated for compounds 3b and 3c, and modestly potencyenhancing for 3d-h, with 3f exhibiting the greatest potency (KDR $IC_{50} = 16 \text{ nM}$). The three-carbon methylene spacer in 3e and 3g conferred 2-fold greater potency relative to the two-carbon linker in analogues 3c and 3b, respectively. That inhibitory activity was relatively insensitive to the nature of the basic side chain is consistent with the proposed binding mode depicted in Figure 2 wherein the side chain extends away from the binding cleft and is solvent exposed. Further SAR studies supported this model. As was the case in the oxazolyl series (compounds 2a-f), replacement of the thienyl group with a substituted phenyl ring led to nearly complete loss of activity in this series, presumably the result of steric clashes within hydrophobic region I. The indolyl ring system proved more tolerant of substitution in terms of retaining activity. For example, placing a chloro, cyano, or methoxyl group at the indolyl C-5 position of 3f resulted in only a 3-fold decrease in potency.

Cell data indicated that the basic side chain was required for achieving submicromolar levels of inhibitory activity in this series (Table 3). A modest shift (5-to 20-fold) was observed in cellular IC_{50} compared to the corresponding biochemical IC_{50} value. This effect

Table 3. Cell activity for selected compounds

KDR IC50 (nM)	Cell IC ₅₀ (nM)	
126	37% inhib. @ 2500 nM	
60	1410	
38	495	
16	328	
46	256	
	KDR IC ₅₀ (nM) 126 60 38 16 46	

Table 4. Physical properties of compounds 3d-f

Compd LogP		Solubility @ pH 5.2 (mg/mL)	Solubility @ pH 7.4 (mg/mL)	
3d	3.3	0.024	0.005	
3e	N.D.	0.177	< 0.005	
3f	3.7	18.17	> 0.205	

N.D., not determined.

Table 5. Rat pharmacokinetic data for compounds 3d-f^a

Compd	Cl (mL/min/kg)	$t_{1/2}$ (h)	Vdss (L/kg)	%F	
3d	65	1.9	7.2	35	
3e	15	1.6	1.8	28	
3f	13	1.8	1.7	33	

^aCompound dosed 5 mg/kg iv as a solution in DMSO and 20 mg/kg po as a suspension in aqueous 0.5% methyl cellulose (pH 5).

Table 6. KDR kinase selectivity (fold)^a of 3f

PDGFRβ	Flt-1	Flt-4	FGFR-1	FGFR-2	c-Src
7	20	3	246	136	147

^aExpressed as the biochemical IC₅₀ (nM) ratios to KDR.

may be due, in part, to high protein binding, as measured for 3f (99.8% bound to human plasma protein).

Physical properties were determined for selected compounds (Table 4). Gratifyingly, compounds **3d**-f showed greater aqueous solubility and lower lipophilicity relative to compounds **1** and **2a**, leading us to evaluate their pharmacokinetic properties.

Compounds **3d–f** exhibited moderate pharmacokinetic behavior in rats with $t_{1/2}$ ranging from 1.6 to 1.9 h and oral bioavailability ca. 30% (Table 5).

The selectivity of **3f** for inhibition of KDR kinase versus several closely related receptor tyrosine kinases and a non-receptor tyrosine kinase (c-Src) is presented in Table 6 and representative of the class. In general, these compounds showed low levels of selectivity for KDR against the highly KDR-homologous kinases PDGFR β , Flt-1, and Flt-4 and higher levels against FGF-1, FGF-2, and c-Src.

In conclusion, we have described the evolution of a novel 3-(5-thien-3-ylpyridin-3-yl)-1H-indolyl class of KDR kinase inhibitors from a potent oxazole-based lead. Compounds within this new series retained high activity and exhibited improved physical properties and reasonable pharmacokinetics in rats.

Acknowledgements

We thank Dr. Art Coddington, Dr. Chuck Ross, and Dr. Harri Ramjit for mass spectral analyses, Matt Zrada and Ken Anderson for solubility and logP determinations, and Dr. George Hartman and Elaine Walker for editing and assistance in the preparation of this manuscript.

References and Notes

1. Adamis, A. P.; Shima, D. T.; Yeo, K. T.; Yeo, T. K.; Brown, L. F.; Berse, B.; D'Amore, P. A.; Folkman, J. *Biochem. Biophys. Res. Commun.* **1993**, *193*, 631.

2. Giatromanolaki, A.; Sivridis, E.; Athanassou, N.; Zois, E.; Thorpe, P. E.; Brekken, R. A.; Gatter, K. C.; Harris, A. L.; Koukourakis, I. M.; Koukourakis, M. I. J. Pathol. 2001, 194, 101.

- 3. Detmar, M. Dermatol. Sci. 2000, 24, S78.
- 4. For reviews, see: (a) Carmeliet, P.; Jain, R. K. Nature 2000, 407, 249. (b) Folkman, J. Nat. Med. 1995, 1, 27.
- 5. Zetter, B. R. Ann. Rev. Med. 1998, 49, 407.

6. (a) Hanahan, D.; Folkman, J. Cell **1996**, *86*, 353. (b) Holmgen, L.; O'Reilly, M. S.; Folkman, J. Nat. Med. **1995**, *1*, 149.

7. (a) Veikkola, T.; Karkkainen, M.; Claesson-Welsh, L.; Alitalo, K. *Cancer Res.* **2000**, *60*, 203. (b) Thomas, K. A. *J. Biol. Chem.* **1996**, *271*, 603.

8. Kim, K. J.; Li, B.; Winer, J.; Armanini, M.; Gillett, N.; Phillips, H. S.; Ferrara, N. *Nature* **1993**, *362*, 841.

9. Witte, L.; Hicklin, D.; Zhu, Z.; Pytowski, B.; Kotanides, H.; Rockwell, P.; Bohlen, P. *Cancer Metastasis Rev.* **1998**, *17*, 155.

 (a) Fong, T. A. T.; Shawver, L. K.; Sun, L.; Tang, C.; App, H.; Powell, T. J.; Kim, Y. H.; Schreck, R.; Wang, X.; Risau, W.; Ullrich, A.; Hirth, K. P.; McMahon, G. *Cancer Res.* **1999**, *59*, 99. (b) Drevs, J.; Hofmann, I.; Hugenschmidt, H.; Wittig, C.; Madjar, H.; Müller, M.; Wood, J.; Martiny-Baron, G.; Unger, C.; Marmé, D. *Cancer Res.* **2000**, *60*, 4819.
 (c) Wedge, S. R.; Ogilvie, D. J.; Dukes, M.; Kendrew, J.; Chester, R.; Jackson, J. A.; Boffey, S. J.; Valentine, P. J.; Curwen, J. O.; Musgrove, H. L.; Graham, G. A.; Hughes, G. D.; Thomas, A. P.; Stokes, E. S. E.; Curry, B.; Richmond, G. H. P.; Wadsworth, P. F.; Bigley, A. L.; Hennequin, L. F. *Cancer Res.* **2002**, *62*, 4645.

11. For recent reviews, see: (a) Connell, R. D. Expert Opin. Ther. Pat. 2002, 12, 1763. (b) Boyer, S. J. Curr. Top. Med. Chem. 2002, 2, 973. (c) Bilodeau, M. T.; Fraley, M. E.; Hartman, G. D. Expert Opin. Investig. Drugs 2002, 11, 737.

12. The KDR IC₅₀ value represents biochemical inhibition of phosphorylation of poly-Glu/Tyr (4:1) peptide substrate by isolated KDR kinase (cloned and expressed as a GST-fusion protein); see: Kendall, R. L.; Rutledge, R. Z.; Mao, X.; Tebben, A. J.; Hungate, R. W.; Thomas, K. A. J. Biol. Chem. **1999**, 274, 6453. Values are reported as the average of at least two determinations, except for compound **3c** (n=1). Standard deviations are typically within 25% of the IC₅₀ value.

13. The cell IC_{50} value represents the inhibition of VEGF-stimulated mitogenesis as determined in human umbilical vein endothelial cells. Refer to US Patent 6,265,403 for assay details. Values are reported as the average of three determinations for compounds 1 and 3d, and single determinations for 3a, 3e, 3f, and 3h.

14. Partition coefficients were determined by HPLC analysis in octanol/water.

15. All target compounds were fully characterized by 1 H NMR and mass spectroscopy.

16. Compounds **2a–f** were prepared by the following sequence; for related chemistry; see: Lipshutz, B. H.; Hungate, R. W.; McCarthy, K. E. J. Am. Chem. Soc. **1983**, 105, 7703.



17. (a) Fraley, M. E.; Hoffman, W. F.; Rubino, R. S.; Hungate, R. W.; Tebben, A. J.; Rutledge, R. Z.; McFall, R. C.; Huckle, W. R.; Kendall, R. L.; Coll, K. E.; Thomas, K. A. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 2767. (b) Fraley, M. E.; Rubino, R. S.; Hoffman, W. F.; Hambaugh, S. R.; Arrington, K. A.; Hungate, R. W.; Bilodeau, M. T.; Tebben, A. J.; Rutledge, R. Z.; Kendall, R. L.; McFall, R. C.; Huckle, W. R.; Coll, K. E.; Thomas, K. A. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 3537.

18. For a description of the ATP binding region of protein tyrosine kinases, see: Traxler, P.; Furet, P. *Pharmacol. Ther.* **1999**, *82*, 195.

(a) Conway, S. C.; Gribble, G. W. *Heterocycles* 1990, *30*, 627. (b) Yang, Y.; Martin, A. R. *Heterocycles* 1992, *34*, 1169. (c) Zheng, Q.; Yang, Y.; Martin, A. R. *Tetrahedron Lett.* 1993, *34*, 2235.

20. Additional examples as well as experimental details of the synthetic methods employed in this work can be found in US Patent 6,265,403.