¹³C NMR SPECTRAL AND CONFORMATIONAL ANALYSIS OF 8-0-4' NEOLIGNANS

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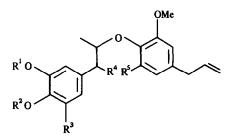
Abstract—The ¹³C NMR spectra of the *erythro* and *threo* forms of representative members of the 8-0-4' type of neolignans were recorded and the signals assigned. Based on these assignments and on the comparison with previously reported ¹H NMR data, the most probable conformations for the above mentioned stereoisomers are suggested.

INTRODUCTION

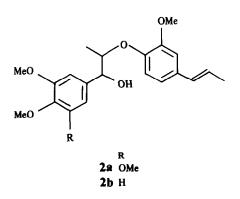
Of the great structural variety of neolignans, the 8-0-4' type represents a small group whose members were isolated exclusively from plants of the Myristicaceae [1]. Amongst them, those isolated from Myristica fragrans Houtt. [2] with two methoxyl groups on ring B, correspond to the erythro series (1a-1h) whereas those from Virola surinamensis (Rol.) Warb (2a and 2b) [3], carrying one methoxyl on ring B, to the threo series. By phenol oxidation coupling of arylpropenoids, mixtures of erythro and threo neolignans related to the natural products were also prepared and interestingly enough the β -O coupling of dimethoxyphenols, like 2.6-dimethoxy-4-propenylphenol, produces mainly the erythro isomer but from methoxyphenols, like isoeugenol, the major coupling product corresponds to the threo series, supporting the proposed biogenetic pathway [4, 5]. For the determination of the relative configurations of the synthetic as well as the natural products extensive use of ¹HNMR spectroscopy was made. In this connection, we decided to carry out an analysis of some synthetic members of this group, as part of a project on ¹³C NMR spectral analysis of lignans and related products [6, 7], that combined with previously reported ¹HNMR data will allow us to elucidate features of their stereochemistry and conformation.

RESULTS AND DISCUSSION

For carrying out the spectral analysis we selected two pairs of diastereoisomers, with one and two methoxyl groups on ring B (1i and 1j) as representative members of each group of 8-O-4' neolignans. These compounds were synthesized following the general method described previously [2, 8]. Treatment of the bromoketone 3 with the sodium salt of eugenol in DMF or with 2,6-dimethoxy-4allylphenol and potassium carbonate in 2-butanone, afforded ketones 4a and 4b, respectively. Sodium borohydride reduction of 4a gave, as described before [5, 8] a mixture of alcohols in which the *erythro* isomer is the major product (1i, *erythro*), based on the intensity of the

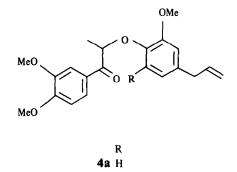


1a $R^{1} = R^{2} = -CH_{2} - ;$ $R^{3} = OMe$; $R^{4} = OH$; $R^{5} = OMe$ 1b $R^{1} = Me$; $R^{2} = Me$ $R^{3} = OMe$; $R^{4} = OH$; $R^{5} = OMe$ 1c $R^{1} = Me$; $R^{2} = H$ $R^{3} = H$; $R^{4} = OH$; $R^{5} = OMe$ 1d $R^{1} = Me$; $R^{2} = Ac$; $R^{3} = H$; $R^{4} = OAc$; $R^{5} = OMe$ 1e $R^{1} = R^{2} = -CH_{2} - ;$ $R^{3} = H$; $R^{4} = OAc$; $R^{5} = OMe$ 1f $R^{1} = Me$; $R^{2} = Me$; $R^{3} = OAc$ $R^{4} = OAc$; $R^{5} = OMe$ 1g $R^{1} = R^{2} = -CH_{2} - ;$ $R^{3} = H$; $R^{4} = OBc$; $R^{5} = OMe$ 1g $R^{1} = R^{2} = -CH_{2} - ;$ $R^{3} = H$; $R^{4} = OBc$; $R^{5} = OMe$ 1g $R^{1} = Me$; $R^{2} = Me$; $R^{3} = OMe$; $R^{4} = H$; $R^{5} = OMe$ 1h $R^{1} = Me$; $R^{2} = Me$; $R^{3} = H$; $R^{4} = OH$; $R^{5} = H$ 1j $R^{1} = Me$; $R^{2} = Me$; $R^{3} = H$; $R^{4} = OH$; $R^{5} = OMe$



MeO MeO

3



4b OMe

H-7 signal in the ¹H NMR spectrum of the mixture.* An attractive explanation for this stereochemical result, by analogy with the reduction of t-butyl α -alkoxy- β -keto carboxylates [9], is that the reaction proceeds via a chelate (Fig. 1) in which the sodium ion is coordinated by the oxygen atoms of the carbonyl and the aryloxy groups. The approach of hydride from the less hindered side of the CO generates predominantly the erythro isomer. Confirmatory evidence for the above explanation was obtained by reduction of ketone 4a with sodium borohydride in the presence of 15-crown-5. Under these conditions a 9:1 mixture of threo and erythro forms was obtained. Sodium borohydride reduction of 4b afforded a ca 1:1 mixture of both isomers (1j, erythro, threo) whereas in the presence of 15-crown-5, again the threo isomer predominates. As was previously reported [2], lithium aluminium hydride reduction of 4a and 4b led exclusively to the erythro form of 1i and 1j, respectively. For the spectral analysis described below the diastereoisomers were isolated in pure form by column chromatography from the mixtures where they are the major products

The stereochemistry and most probable conformation of the aryl ethers may be deduced from their IR and NMR spectra following the arguments utilized by Wallis [4]. The independence to concentration changes of the hydroxyl bands in the IR spectra of this type of compounds in chloroform solutions clearly indicated intramolecular hydrogen bonding of the benzylic hydroxyl and the aryloxy group. Of the possible staggered forms of the *erythro* and *threo* isomers, rotamers A, B and C (Fig. 2) are the only ones that would contribute significantly to the

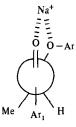


Fig. 1. Proposed reduction of 8-O-4' neolignans via a chelate.

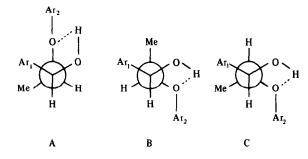


Fig. 2. Possible staggered forms of 8-O-4' neolignan diastereoisomers.

conformation of the aryl ethers, based on the ¹H NMR spectra of the diastereoisomers of **1i** and **1j**. The coupling constants $(J_{7,8})$ of 3.2 and 8 Hz are consistent with dihedral angles between H-7 and H-8 of 60° and 180°, respectively, allowing the assignment of conformer C, with the larger J values, to the *threo* form and conformers A or B (J = 3.2 Hz) to the *erythro* one. Although the signals for H-7 and H-8 in the ¹H NMR spectra of the *threo* isomers are, in all cases, at higher fields compared to the corresponding protons of the *erythro* forms, these data are not conclusive to decide upon which of the two *erythro* rotamers will provide the main contribution to the conformations of these isomers.

A comparison of C-7, C-8 and C-9¹³C NMR signals for the diastereoisomers of 1i and 1j shows downfield shifts for all of them in going from an erythro to a threo form and since there is a restricted rotamer population due to the intramolecular hydrogen bonding, the chemical shifts of C-9 could be calculated for rotamers A, B and C, as an extension of the method of Beirbeck et al. [10]. Such calculations gave values of 17.68, 13.1 and 17.7 ppm, respectively. Shifts of 13.1 and 17.7 are in excellent agreement with those observed for the erythro and threo forms, and indicate that conformer B should be the main contributor to the conformation of the erythro isomer. These appreciable differences in ¹³C chemical shifts between the erythro and threo forms will be a powerful additional probe toward the elucidation of stereochemical features of neolignans.

The carbon shifts of 1i (erythro and threo) and 1j (erythro and threo) assigned by comparison with those of 4a and 4b and related compounds previously reported [3, 11], and by the analysis of the generated CH/CH₃ and

^{*}The numbering used in this paper for the 8-O-4' neolignans is the one suggested in ref. [1].

Table 1. Carbon shifts of 8-0-4' neolignans

Carbon	li	1i	1 j	1j		a
No.	(erythro)	(threo)	(erythro)	(threo)	4a	4 b
1	132.4	132.3	132.5	135.5	127.2	127.9
2	1104	110.3	110.6	110.6	111.1	111.0
3	147.8	148.2	147.7	148.4	149.6	148.2
4	1509	150.0	148.6	148.6	153.3	152 5
5	109.2	109.6	109.1	109.9	109.9	109.5
6	1207	120.3	117.9	119.6	123.4	123.5
7	73 3	77.5	72.6	78.7	190.2	190.5
8	818	82.8	821	86.1	78.0	80.1
9	13.2	16.2	12.6	17.3	19.0	17.7
ſ	134.9	134.3	132.8	133.1	134.0	133.4
2'	112.2	111.9	105.3	105 2	112.5	104 9
3'	148.5	148.4	153.3	152.4	148 7	152 5
4′	144.6	145.3	1359	135.5	145.0	135.4
5'	118 2*	118 2*	153 3	152.4	115.8	152.5
6'	119 2*	119 3*	105 3	105.2	120.3	104.9
7′	39.6	39.3	40.3	40.2	39.6	39.9
8′	137.0	136.8	136.8	136.8	137.2	136.7
9′	115 5	115.2	115.9	115.8	115.5	115.5
OMe	55.5	55.2	55 7	55.5	55.7	55.4
			55.9	55.6	55 8	

*Signals within a column may be interchanged

 CH_2/q subspectra by spin-echo sequences utilizing the proton-flip method (APT) [12] are listed in Table 1.

EXPERIMENTAL

The ¹H NMR were recorded at 80.13 MHz and the ¹³C NMR spectra at 20.15 MHz in the Fourier transform mode and in CDCl₃ solns. Chemical shifts are expressed on the TMS scale according to TMS = δ CDCl₃ + 76.9 ppm. The asterisks on the Table indicate possible signal reversal. TLC was done on silica gel GF 254 and column chromatography on silica gel H.

1-(3',4'-Dimethoxyphenyl)-2-(2''-methoxy-4''-allyl-phenoxy) propan-1-one (4a). The Na salt of eugenol (2.88 g, 15.5 mmol) was added to a stirred soln of 1-(3,4-dimethoxyphenyl)-2bromopropan-1-one (3), prepared according to ref. [13] (3.77 g, 14 mmol), in dry DMF (55 ml). After being stirred for 48 hr the mixture was diluted with H₂O (50 ml) and extracted with Et₂O (2 \times 50 ml). The combined Et₂O extracts were washed with 0.2 N aq. NaOH, H₂O and dried (NaSO₄). The residue, 1.87 g of crude product, was recrystallized from 95% EtOH yielding 1.67 g (67%) of pure product, mp 86-88°. IR v_{max}^{KBr} cm⁻¹: 3120, 3090, 2095, 1680, 1650, 1600, 1525, 1450, 1275, 1245, 1030, 925, 915. ¹H NMR: δ 1.69 (3H, d, J = 7 2 Hz, H-3), 3.28 (2H, br d, J = 6.4 Hz, H-7"), 3.82, 3.91 and 3.93 (9H, s, 3 × MeO), 4.95 (1H, m, H-9"), 5.12 (1H, m, H-9"), 5.40 (1H, q, J = 7.2, H-8), 5.72-6.20 (1H, m, H-8"), 6.50–7.90 (6H, m, ArH). MS m/z (rel. int): 356 [M]⁺ (10), 191 (21), 165 (100), 115 (12), 103 (19), 91 (30), 77 (43), 55 (25), 51 (21).

 $\Delta^{8'}$,3,3',4-trymethoxy neolignan (1i, crythro). A soln of ketone 4a (0.30 g, 0.84 mmol) in dry Et₂O (25 ml) was added, dropwise, to a stirred suspension of LiAlH₄ (0.480 g, 12.6 mmol) in Et₂O (36 ml). After the addition was complete, the mixture was refluxed for 24 hr. Excess LiAlH₄ was carefully destroyed by addition of EtOAc-ice, and finally diluted with H₂O (60 ml), acidified (10% HCl) and extracted with Et₂O (3 × 60 ml). The combined Et₂O extracts were washed with 1 N aq. NaOH and H₂O, dried (Na₂SO₄) followed by concn affording a crude oil (0.277 g). CC (gradient of hexane-EtOAc) produced pure 1i, *erythro* (0.250 g, 82 %). IR v $\frac{film}{max}$ cm⁻¹: 3260, 3045, 3005, 2975-2940, 1650, 1620, 1520, 1475, 1275, 1170, 1040, 925, 820. ¹H NMR: δ 1.17 (3H, *d*, *J* = 6.4 Hz, H-9), 3.35 (2H, *d*, *J* = 6.4 Hz, H-7'), 3.85 (9H, br s, 3 × MeO), 4.33 (1H, *dq*, *J* = 6.4, 3.2 Hz, H-8), 4.84 (1H, *d*, *J* = 3.2 Hz, H-7), 5.01 (1H, br s, H-9'), 5.18 (1H, *m*, H-9'), 5.70-6.20 (1H, *m*, H-8'), 6.60-7.10 (6H, *m*, Ar<u>H</u>). MS *m/z* (rel. int.): 358 [M]⁺ (10), 194 (38), 192 (29), 167 (88), 164 (100), 149 (21), 139 (50), 121 (20), 107 (22), 103 (25), 91 (41), 77 (47), 57 (80), 43 (60).

NaBH₄ reduction of ketone 4a: $\Delta^{8'}$, 3, 3', 4-trimethoxy neolignan (1i, erythro + threo). Solid NaBH₄ (0.170 g, 4.5 mmol) was added in small portions to a stirred, cooled (0°) soln of ketone 4a (0.505 g, 1.5 mmol) in dry MeOH (22 ml). The mixture was stirred for 30 min at 0° and 4 hr at room temp, H₂O and a few drops of HOAc were then added and the mixture extracted with Et₂O (4 × 50 ml). The combined Et₂O extracts were washed with satd aq. NaHCO₃ soln and H₂O, dried (Na₂SO₄), decanted and evapd, yielding a 72:28 mixture of 1i, erythro threo (0.5 g, 92%).

 $NaBH_4 + 15$ -crown-5 reduction of ketone 4a. A soln of $NaBH_4$ (0.114, 3 mmol) in iso PrOH (10 ml) was added to a stirred soln of 15-crown-5 ether (0.72 g, 3.6 mmol) in dry isoPrOH (5 ml). After 6 hr, a soln of ketone 4a (0.356 g, 1 mmol) in dry MeOH (5 ml) was added and the mixture stirred for 4 hr at room temp. The same work up as before, gave a 1:9 mixture of crude 1i, erythro threo (0.33 g, 92 %). Pure 1i, threo (0.186 g) was obtained by CC of the mixture. IR ν_{max}^{film} cm⁻¹: 3250, 3045, 3005, 2995–2925, 1650, 1620, 1600, 1525, 1475, 1430, 1280, 1240, 1155, 1045, 925, 820. ¹H NMR $\cdot \delta$ 1.15 (3H, d, J = 6.4 Hz, H-9), 3.35 (2H, br d, J = 6.4 Hz, H-7'), 3.90, 3.91 and 3.93 (9H, s, 3 × MeO), 4.04 (1H, m, H-8), 4.64 (1H, d, J = 8 Hz, H-7), 4.99 (1H, m, H-9'), 5.16 (1H, m, mH-9'), 5.73-6.25 (1H, m, H-8'), 6.67-7 (6H, m, ArH). MS m/z (rel. int.): 358 [M]⁺ (11), 194 (26), 192 (23), 167 (80), 164 (100), 149 (22), 139 (69), 121 (33), 107 (39), 103 (45), 91 (71), 77 (76), 65 (43), 57 (70), 43 (59).

1-(3',4'-Dimethoxyphenyl)-2-(2",6"-dimethoxy-4"-allyl-phenoxy) propan-1-one (4b). 1-(3,4-Dimethoxyphenyl)-2-bromopropan-1-one (3) (2.62 g, 9.75 mmol), 6-methoxyeugenol (2 ml, 18 mmol) and dry K₂CO₃ (2.4 g) were heated under reflux, with stirring, in dry MeCOEt (37.5 ml) for 30 hr. The soln was cooled, diluted with H₂O (50 ml), acidified (HOAc) and extracted with Et_2O (2 × 100 ml). The combined Et_2O extracts were washed with 1 % aq. NaOH (1 \times 100 ml), H₂O (2 \times 50 ml), dried (Na₂SO₄) and concd to dryness. Crystallization of the crude product from MeOH, yielded pure ketone 4b (2.1 g, 60.5%) mp 71–73°. IR v_{max}^{KBr} cm⁻¹: 3095–2840, 1690, 1600, 1540, 1510, 1280, 1140, 1120, 1030. ¹H NMR: δ 1 56 (3H, d, J = 6.4 Hz, H-9), 3.31 (2H, d, J = 6.4 Hz, H-7'), 3.72, 3.92 (12H, s, $4 \times$ MeO), 4.98 (1H, m, H-9'), 5.17 (1H, m, H-9'), 5.25 (1H, q, J = 6.4 Hz, H-8),5.96 (1H, m, H-8'), 6.37 (2H, s, ArH), 6.87 (1H, d, J = 8 Hz, ArH), 7.75–7.95 (2H, m, Ar<u>H</u>). MS m/z (rel. int.): 386 [M]⁺ (30), 221 (30), 193 (54), 165 (100), 105 (22), 91 (38), 77 (52), 51 (19).

 $\Delta^{8'}$,3,3',4,5'-Tetramethoxy neolignan (1j, erythro). The same procedure, as for reduction of **4a** was used, ketone **4b** (0 324 g, 1 mmol), LiAlH₄ (0.48 g, 12.6 mmol) in dry Et₂O (36 ml). After CC pure 1j erythro was obtained (0.294 g, 90%) as a colorless oil. IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1.} 3520, 3090–2925, 1600, 1520, 1470, 1275, 1240, 1040, 925, 740 ¹H NMR: δ 1.13 (3H, d, J = 6.4 Hz, H-9), 3.36 (2H, br d, J = 6.4 Hz, H-7'), 3.86, 3.87 (12H, 4 × MeO), 4.36 (1H, d q, J = 6.4, 3 2 Hz, H-8), 4 81 (1H, d, J = 3.2 Hz, H-7), 5 3 (1H, m, H-9'), 6.0 (1H, m, H-9'), 5.75–6.23 (1H, m, H-8'), 6.47 (2H, s, Ar<u>H</u>), 6.78 (1H, s, Ar<u>H</u>), 6.96 (1H, s, Ar<u>H</u>). MS m/z (rel. int.)⁻ 388 [M]⁺ (20), 194 (100), 167 (50), 165 (23), 57 (19).

NaBH₄ reduction of ketone **4b**: $\Delta^{8'}$,3,3',4,5'-tetramethoxy neolignan (**1j**, erythro + threo). Following the same procedure as described for reduction of **4a** starting from ketone **4b** (0.194 g,

0.5 mmol) and NaBH₄ (0.057 g, 1.5 mmol) in MeOH (6.4 ml). The crude product obtained (0.184 g, 94%) was a 1:1 mixture of 1j, erythro: threo.

NaBH₄ + 15-crown-5 reduction of ketone 4b. The same procedure as described for ketone 4a was followed starting from ketone 4b (0.386 g, 1 mmol), NaBH₄ (0.114 g, 3 mmol) and 15-crown-5 (0.69 ml, 3.6 mmol). A 1:4 mixture of crude 1j, erythro: threo was obtained. CC of the mixture afforded pure 1j, threo (0.298 g, 76%) as a colorless oil. IR $v_{\text{CMC}}^{\text{CMC}}$ cm⁻¹: 3240, 2990–2930, 1600, 1470, 1350, 1140, 1040. ¹H NMR: δ 1.17 (3H, d, J = 6.4 Hz, H-9), 3.34 (2H, br d, J = 6.4 Hz, H-7'), 3.85 (12H, s, 4 × MeO), 4.03 (1H, m, H-8), 4.62 (1H, d, J = 8 Hz, H-7), 5.01 (1H, m, H-9'), 5.74–6.15 (1H, m, H-8'), 6.45 (2H, s, Ar<u>H</u>), 6.83–7.96 (3H, m, Ar<u>H</u>). MS m/z (rel. int.): 388 [M]⁺ (12), 194 (100), 167 (8), 165 (17), 57 (20).

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