November 1994 SYNTHESIS 1133

A Convenient Synthesis of Hispidin from Piperonal

Waldemar Adam,*a Chantu R. Saha-Möller, Markus Veit, Birgit Welkea

^a Institute of Organic Chemistry, University of Würzburg, Am Hubland, D-97074 Würzburg, Germany

^b Julius-von-Sachs-Institut für Biowissenschaften, University of Würzburg, Mittlerer Dallenbergweg 64, D-97082 Würzburg, Germany Received 13 June 1994

An improved synthesis of hispidin (overall yield 24%) starting from the commercially available piperonal is reported. This three-step method is more effective and advantageous compared to the known six-step preparation.

Styrylpyrones are natural products accumulated in fungi¹ and higher plants.² The first styrylpyrone glycoside, namely equisetumpyrone A, was isolated from gametophytes and fertile sprouts of Equisetum arvense L., whose structure was recently elucidated.3 Its biosynthesis is still unknown and we propose that this styrylpyrone glycoside is formed through caffeoyl-CoA, which combines with malonate, and by hydroxylation of the resulting hispidin and subsequent glucosilation of the hydroxyhispidin B. Hispidin, a naturally occurring styrylpyrone, whose structure was determined as 4-hydroxy-6-(3',4'-dihydroxystyryl)-2-pyrone (4)⁴ was first isolated from *Polyporus* hispidus in 1889.5 This biologically active trihydroxy-substituted styrylpyrone exhibits in vitro antimicrobial activity against Gram-positive organisms, and also the acidfast Mycobacterium smegmatis.6

To verify the proposed biosynthetic route of equisetum-pyrone, we required sufficient amounts of hispidin. A cumbersome, multistep synthesis of hispidin was reported earlier; unfortunately, we encountered difficulties with this synthetic sequence and were, therefore, obliged to work out a more convenient method. We report here a three-step synthesis of hispidin (4) (24% overall yield), which starts from the commercially available piperonal (1), as displayed in the Scheme.

The condensation of piperonal (1) with 4-methoxy-6-methyl-2-pyrone⁸ in the presence of magnesium methoxide, analogous to the reported preparation of 4-methoxy-6-(4'-methoxystyryl)-2-pyrone,⁸ afforded the methylenedioxy styrylpyrone 2 in 52 % yield. The selective demethylenation of 2 with boron trichloride in dichloromethane at 40 °C gave the dihydroxy styrylpyrone 3 in 74 % yield. The latter was converted to hispidin (4)

in 64% yield by demethylation of the pyrone ether group with sodium thioethoxide in dimethylformamide at reflux

The advantages of the present three-step route (Scheme) over the previous six-step one are:

a) instead of 3,4-di(methoxymethoxy)benzaldehyde, which is prepared by alkylation of 3,4-dihydroxybenzaldehyde with expensive and toxic chloromethyl methyl ether, commercially available piperonal (1) is employed as the carbonyl component for condensation with methylpyrone, which proceeds in much better yield (52% vs. 23%), and b) the methylenedioxy group of 2 is readily cleaved by boron trichloride to release the dihydroxysubstituted styrylpyrone methyl ether 3, which is effectively demethylated to hispidin (4) by sodium thioethoxide in dimethylformamide, 9 an ether cleaving reagent employed for the first time for such an application. In contrast, several steps are required for the deprotection of 4-methoxy-6-[3',4'-di(methoxymethoxy]-styryl-2-pyrone to hispidin, which was employed in the literature procedure.

Solvents were purified according to standard procedures. $\mathrm{CH_2Cl_2}$ was distilled from $\mathrm{P_2O_5}$ and DMF from $\mathrm{CaH_2}$. Melting points were determined on a Reichert Thermovar hot stage apparatus. IR spectra were recorded on a Perkin-Elmer 1420 spectrometer and the UV spectra on a Hitachi U-3200 spectrophotometer. $^1\mathrm{H}$ and $^{13}\mathrm{C}$ NMR spectra were acquired on a Bruker AC 200 (200 MHz) or AC 250 (250 MHz) spectrometer. Chemical shifts refer to CDCl₃, DMSO- d_6 , or $\mathrm{CD_3OD}$. The FAB mass spectrum was obtained on a Finnigan MAT 8430 instrument by using glycerol as matrix. 4-Methoxy-6-methyl-2-pyrone was prepared according to the literature procedure by methylation of the commercially available 4-hydroxy-6-methyl-2-pyrone with dimethyl sulfate. 8

4-Methoxy-6-(3',4'-methylenedioxystyryl)-2-pyrone (2):

To a suspension of Mg(OMe)₂ (60.0 mmol, prepared from 1.46 g

Short Papers SYNTHESIS

of Mg turnings) in anhydr. MeOH (40 mL) was added dropwise a solution of piperonal (1; 3.00 g, 20.0 mmol) and 4-methoxy-6-methyl-2-pyrone (3.36 g, 24.0 mmol) in MeOH (40 mL) under an Ar atmosphere. The mixture was stirred under gentle reflux for 7 h, the solvent was removed by rotoevaporation (40 °C/20 Torr), the residue was dissolved in CH₂Cl₂ (25 mL) and treated with 3.3 M AcOH (30 mL) in H₂O. The aqueous layer was separated and extracted with CH₂Cl₂ (5 × 25 mL). The combined organic layers were washed with H₂O (6 mL), sat. NaHCO₃ solution (12 mL) and brine (6 mL), and dried (Na₂SO₄). After removal of the solvent (20 °C/20 Torr) the residue was recrystallized from MeOH (1 L) to afford 2.82 g (10.4 mmol, 52 %) of styrylpyrone 2 as pale yellow needles; mp 234–235 °C (Lit. 10 mp 236 °C).

IR (KBr): v = 3070, 1715, 1625, 1605, 1550, 1505, 1440, 1405, 1250, 1155, 1030 cm⁻¹.

UV (MeOH): $\lambda_{\text{max}} (\log \varepsilon) = 223 (4.284), 251 (3.993), 363 \text{ nm} (4.325).$ ¹H NMR (CDCl₃, 250 MHz): $\delta = 3.82$ (s, 3 H, OCH₃), 5.47 (d, J = 2.2 Hz, 1 H, 3 -H), 5.89 (d, J = 2.2 Hz, 1 H, 5 -H), 5.99 (s, 2 H, OCH₂O), 6.40 (d, J = 15.9 Hz, 1 H, 8' -H), 6.80 (d, J = 8.0 Hz, 1 H, 5' -H), 6.97 (dd, $J_1 = 8.0 \text{ Hz}, J_2 = 1.5 \text{ Hz}, 1 \text{ H}, 6' \text{-H}), 7.00$ (d, J = 1.5 Hz, 1 H, 2' -H), 7.41 (d, J = 15.9 Hz, 1 H, 7' -H).

4-Methoxy-6-(3',4'-dihydroxystyryl)-2-pyrone (3):

A 1 M solution of BCl₃ in hexane (6.91 mL, 6.91 mmol) was added dropwise to a solution of styrylpyrone **2** (0.607 g, 2.23 mmol) in anhydr. CH_2Cl_2 (100 mL) under an N_2 atmosphere. Stirring was continued for 22 h at reflux and MeOH (10 mL) was added at 20 °C to the mixture. The solvent was removed (20 °C/20 Torr) and the residue was recrystallized from MeOH (25 mL) to give 0.431 g (1.65 mmol, 74%) of styrylpyrone **3** as yellow needles, mp 256–257 °C (Lit. 7 mp 257 °C).

IR (KBr): v = 3600 - 2900, 1645, 1615, 1585, 1530, 1440, 1395, 1335, 1285, 1240, 1135, 1095, 1025, 950 cm⁻¹.

UV (MeOH): $\lambda_{\rm max}$ (log ε) = 220 (4.372), 253 (4.133), 371 nm (4.405). ¹H NMR (DMSO- d_6 , 200 MHz): δ = 3.81 (s, 3 H, OCH₃), 5.58 (d, J = 2.1 Hz, 1 H, 3-H), 6.25 (d, J = 2.1 Hz, 1 H, 5-H), 6.66 (d, J = 16.0 Hz, 1 H, 8'-H), 6.76 (d, J = 8.2 Hz, 1 H, 5'-H), 6.94 (dd, J_1 = 8.2 Hz, J_2 = 1.7 Hz, 1 H, 6'-H), 7.03 (d, J = 1.7 Hz, 1 H, 2'-H), 7.15 (d, J = 16.0 Hz, 1 H, 7'-H).

¹³C NMR (DMSO- d_6 , 50 MHz): δ = 56.2 (q, CH₃), 87.9 (d, C-3), 100.0 (d, C-5), 114.0 (d, C-2'), 115.7 (d, C-5'), 116.0 (d, C-8'), 120.3 (d, C-6'), 126.6 (s, C-1'), 134.7 (d, C-7'), 145.5 (s, C-3'), 147.4 (s, C-4'), 158.9 (s, C-6), 162.7 (s, C-2), 170.9 (s, C-4).

Hispidin (4):

To a suspension of NaH (72.0 mg, 60% oil dispersion) in anhydr. DMF (1 mL) was added a solution of EtSH (107 mg, 1.73 mmol) in DMF (2 mL) under an N_2 atmosphere. The mixture was stirred for 5 min and a solution of styrylpyrone 3 (100 mg, 0.384 mmol) in DMF (3 mL) was added. Stirring was continued for 1 h at reflux, the mixture was cooled to 0° C, acidified (pH 6) with 10% aq HCl and extracted with EtOAc (5 × 5 mL). The combined extracts were dried (Na₂SO₄) and the solvent removed (40°C, 20 Torr). The crude product was recrystallized from aq MeOH to yield 60.0 mg (0.244 mmol, 64%) of hispidin (4) as yellow needles, mp 258–259°C (dec.) (Lit.6 mp 259°C).

MS (FAB, Glycerol): $m/z = 247 (M + H)^+$.

IR (KBr): v = 3600-2900, 1635, 1585, 1530, 1425, 1355, 1275, 1250, 1185, 1140, 1100, 950 cm⁻¹.

UV (MeOH): $\lambda_{\rm max}$ (log ε) = 220 (4.363), 252 (4.116), 369 nm (4.380).

¹H NMR (CD₃OD, 200 MHz): δ = 5.37 (d, J = 1.9 Hz, 1 H, 3-H), 6.07 (d, J = 1.9 Hz, 1 H, 5-H), 6.53 (d, J = 15.9 Hz, 1 H, 8'-H), 6.75 (d, J = 8.2 Hz, 1 H, 5'-H), 6.89 (dd, J₁ = 8.2 Hz, J₂ = 1.9 Hz, 1 H, 6'-H), 7.01 (d, J = 1.9 Hz, 1 H, 2'-H), 7.25 (d, J = 15.9 Hz, 1 H, 7'-H).

 $^{13}\text{C NMR}$ (CD₃OD, 50 MHz): $\delta = 90.3$ (d, C-3), 101.7 (d, C-5), 114.8 (d, C-2'), 116.5 (d, C-5'), 116.8 (d, C-8'), 122.0 (d, C-6'), 128.7 (s, C-1'), 137.2 (d, C-7'), 146.6 (s, C-3'), 148.6 (s, C-4'), 161.9 (s, C-6), 167.7 (s, C-2), 173.3 (s, C-4).

Financial support by Fonds der Chemischen Industrie is gratefully acknowledged. We thank Dr. L. Witte, Institut für Pharmazeutische Biologie, Technische Universität Braunschweig, Germany, for recording the FAB mass spectrum.

- (1) Gill, M.; Steglich, W. Progress in the Chemistry of Organic Natural Products 1987, 51, 1.
- Hegnauer, R. Chemotoxonomie der Pflanzen; Birkhäuser: Basel, 1986.
- (3) Veit, M.; Geiger, H.; Wray, V.; Abou-Mandour, A.; Rozdzinski, W.; Witte, L.; Strack, D.; Czygan, F.-C. Phytochemistry 1993, 32, 1029.
- (4) Edwards, R. L.; Lewis, D. G.; Wilson, D. V. J. Chem. Soc. 1961,
- (5) Zopf, W. Bot. Ztg. 1889, 47, 53.
- (6) Benedict, R.G.; Brady, L.R. J. Pharm. Sci. 1972, 61, 1820.
- (7) Edwards, R.L.; Wilson, D. V. J. Chem. Soc. 1961, 5003.
- (8) Bu'Lock, J.D.; Smith, H.G. J. Chem. Soc. 1960, 502.
- (9) Feutrill, G.I.; Mirrington R. N. Aust. J. Chem. 1972, 25, 1719.
- (10) Hansel, R.; Rimpler, H.Z. Analyt. Chem. 1965, 207, 270.