SYNTHESIS OF 8-AZAPURINES GLYCOSIDES STARTING FROM 1-AZIDOGLYCOSIDES[†]

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Abstract—An improved synthesis of 8-aza hypoxanthyl- α - (and - β -) mannopyranosides and - β -ribofuranosides starting from 1-azidoglycosides is reported. In the manno series, the anomeric configuration of the products is controlable and the mechanism of this control is discussed.

The 8-azapurines and their glycosides are interesting cytotoxic¹⁻⁸ compounds. The syntheses of the latter are carried out either by fusion reaction between an ose and a modified heterocycle⁹⁻¹¹ or by the construction of the heterocycle, starting from a 1-azidoglycoside.¹¹

Since we have devised a good method for obtaining these azido sugars,¹² we wish to demonstrate their usefulness in purine glycoside synthesis. This paper is devoted to the synthesis of 8 - azahypoxantyl - α - (and - β -) manno-pyranosides and - β -ribofuranosides (8azapurine) using this technique. These syntheses are characterised by yields which are higher than those of previously published procedures and by the fact that we are able to control the α/β stereochemistry of the azidosugar reaction, at least in the manno series.

RESULTS AND DISCUSSION

The access to 1-azidoglycosides lies in the direct conversion of the free anomeric hydroxyl group into the azido one owing to activation in the phosphonium salt, followed by a substitution by the mesityloxytris(dimethylamino)-phosphonium azide (scheme 1).

Formation of the vic-triazolo ring

In order to do this, we began by using Tolman's procedure¹³ based on the action of powdered potassium hydroxide on a solution of the azido compound and cyanoacetamide in aqueous dimethylformamide at room temperature for 6 hours. With the starting materials 1, 2

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and 3, which are of 1-2 *cis* configuration, we systematically obtained good yields of the 5 - amino - 4 - carboxamido - vic - triazologlycosides 4, 5 and 6, which are of 1-2 trans configuration (Table 1).

We originally considered decreasing the reaction time by previously preparing the cyanacetamido carbanion, with sodium hydride in anhydrous dimethylformamide, and by allowing it to react, in a second step, still in an anhydrous medium, on the 1-azidoglycoside. While this did not decrease so much the reaction time, we nevertheless observed an interesting fact. With the manno derivative 3, we obtained 7 as the major product, which is anomeric of 6 and corresponds to a retention of configuration. In order to understand this fact and to be able to prepare the 6 or 7 pure isomers at will (for they are not easily separated), we undertook a short study of the influences of the nature of the solvent and of the base. Our results are summarized in Table 2.

The best conditions for obtaining retention of configuration at the anomeric carbon atom involve sodium hydride in dimethylformamide as a solvent at 0°.

To account for the inversion in the case of the 2,3,5tribenzylarabinose series, Tolman¹³ proposed a two step mechanism, with the opening of the osidic ring. Below, we have adapted this to our case of the 1 - azido - 2,3; 5,6 - di - O - isopropylidene β - D-mannopyranoside **3** (scheme 2).

To explain the anomeric inversion, Tolman invoked the opening of the intermediate A, arising from the initial attack of the acetamido anion, into the intermediate B. This latter undergoes ring closure into C, with an opposite anomeric configuration, which leads to 6.



Scheme 1.

Table 1. Synthesis of 5-amino-4-carboxamido vic-triazolo nucleosides



STARTING MATERIAL	PRODUCT	YIELD 🔏 (a)
1	4	70
2	5	77
3	<u>6</u>	80

(a) PURE PRODUCT, ISOLATED BY CHROMATOGRAPHY



This mechanism seems fairly suited to explaining our results by making the supplementary assumption that the intermediate A can directly close itself into 7 as does C into 6. The intermediate C is probably thermodynamically more stable than A but depending upon the natures of the solvent and of the cation, A and B are more or less stabilized relative to each other. Study of Table 2 does not lead to an easy explanation for the nature of the stabilization. The hydrogen bonding between water and the oxygen atom of B can be suggested in runs number 1, 2 and 3, but it is then rather difficult to account for run 12 in which the same results are obtained in hexamethyl-phosphoramide, an aprotic solvent.

Alternatively, it seems attractive to call upon the "hard and soft theory" for the ions involved. On this basis a plausible assumption is the following. In order to avoid the formation of 6, the equilibrium $A \rightleftharpoons B$ must be displaced to the left. The A form is a harder anion than the C one due to the α effect.¹⁴ The harder the cation M⁺ the more the A form will be stabilized and a soft cation will stabilize rather the C form. In run number 3 the ratio 7/6 is higher than in run number 2. This result is in agreement with the fact that the cation Na⁺ (H₂O) is harder than K⁺ (H₂O).

In anhydrous solvents such as dimethylformamide (runs number 12, 13), the ratio 7/6 changes as previously, depending upon the nature of the cation.

In run number 15, in which the sodium cation is

complexed by a crown ether, and thus softer than the uncomplexed sodium cation, the amount of retention decreases. We do not fully understand the results with the lithium cation (runs number 4, 7, 11) which is known to be highly complexing.

This analysis is all the more difficult as the thermodynamic/kinetic control, we called upon above, plays a role only for the intermediate stages A, B and C. In effect, the paths from A to 7 and from C to 6, via the hydrolysis of an amidure, must be irreversible; in fact, compound 7 replaced in the reaction medium, overnight and at room temperature, does not lead to 6.

Finally, it must be pointed out that, under the best conditions, described above, we were not able to secure the products with retention of configuration starting from the ribo derivatives 1 and 2; this is probably due to the greater strain of the five membered ring which makes opening easier giving the intermediate B.

Pyrimidone ring formation

This was performed, using Tolman's method,¹³ by the reaction of diethoxymethylacetate followed by treatment with methanolic ammonia. Our results are given in Table 3.

The removal of the protective group

This is performed with aqueous trifluoroacetic acid (50%) at room temperature for the two manno deriva-

Table 2. Cyanoacetamid condensation: Results concerning the influences of the nature of the solvent and of the base



RUN	BASE	SOLVENT	TEMPE- RATURE	TIME (HOURS)	6 % (a)	7 % (a)	TOTAL YIELD	
1	KOH	DMF/H ₂ 0	R.T.	5	100	-	80	
2	*	•	0*	6	100	-	83	
3	N.OH		0*	2	65-70	30-35	76	
4	LiOH		0*	1.5	75-70	25-30	70	
5	КН	DMF	0*	0.5	40	60	75	
6	NaH	•	0*	2	10-15	85-90	92	
7	LiH	•	0*	6.5	40	60	75	
8	KH	DME	R.T.	6.5	30	70	80	
9	NaH	-	0*	20	15-20	85-80	95	
10	•		R.T.	6.25	20-25	80-75	95	
11	LiH	.	"	10	40-45	60-55	70	
12	КН	HMPA	0•	0.75	100	-	·70	
IJ	NaH		0*	0.5	30-35	70-65	80	
14	•	DMF+LiC104	0*	1	10-15	85-80	80	
15	"	DMF	0*	0.5	40	60	75	
		CROWN ETHER 18~5						

(a) FOUND BY ^{1}H NMR at 250 mHz after addition of $D_{2}0$

(b) AS A MIXTURE OF 6 AND 7 OBTAINED BY CHROMATOGRAPHY

tives 10 and 11 which give rise to 12 and 13, respectively. In the case of the ribo derivatives, the 8-azainosine 15 is obtained either starting from 8, by the successive actions of aqueous trifluoro acetic acid (50%) and a Dowex 50 H⁺ resin in a tetrahydrofuran/water (1/1, v/v) mixture or starting from 9 by the successive actions of a solution of tetrabutyl-ammonium fluoride in tetrahydrofuran and of a Dowex 50 H⁺ resin. In these latter two cases, the 2',3' di - O - isopropylidene - 8 - azainosine 14 is isolated and characterized.

Anomeric structural assignments

We rely upon optical rotations, ¹H NMR spectra and circular dichroism. For the manno derivatives we have both of the anomers, so that we are able to assign the α/β configuration without ambiguity. On the other hand, in the ribo series, we have only one anomer which is already known.¹¹ Knowledge of the specific optical rotation and of the $J_{1'-2'}$ coupling constants is not enough to ascertain unequivocally the configurations (Table 4).

Nevertheless, the 8-azainosine exhibits a specific optical rotation of -74.8° , which is little different from that reported by Tolman¹¹ (-80°). His ¹H NMR spectrum (60 MHz in dimethylsulfoxyde-d₆) includes an anomeric proton signal at $\delta = 6.18$ ppm with a coupling constant $J_{1'-2'} = 4.75$ Hz. These values are identical to those of the work described above. Thus, the compound 15 has the β configuration.

The compounds 4, 5, 8, 9 and 14 are precursors of 15 and the reactions from 4 to 15 do not allow epimerization. So we conclude that 4, 5, 8, 9 and 14 have the β configuration. Moreover, the difference between the two isopropylidene signals (C-2', C-3') is always more than 0.15 ppm; this is characteristic of a β configuration according to the rule proposed by Imbach.^{15,16}

Optical measurements, particularly those of circular dichroism, can theoretically offer valuable information

Table 3. Synthesis of 8-aza-hypoxanthyl glycosides

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SUBSTRATES	PRODUCTS	YIELD%(a)			
4	<u>&</u>	85			
<u>5</u>	2	95			
Ę.	10	95			
2	<u><u>µ</u></u>	93			

(*) PURE PRODUCT, ISOLATED BY CHROMATOGRAPHY



Scheme 2.



PRODUCT	[a] _D (c.so)LV.)	°H-1'	SOLVENT	J _{1'-2'} ^{HZ}	Δδ	CONFI- GURA- TION
é	+55.8 (0.22	CHC13)	6,68	PYRIDINE	0	-	α
Z	-			6,78	"	2.5	-	в
<u>10</u>	+44.2 (0.57	CHC13)	6,53	DMSO	1	-	a
<u>↓</u>	-20.5 (0.81	CHC13)	6,64	*	3,35	-	ß
12	+37 ((0.81	DMF)	5.5	n	0	-	α
13	+ 6.5 ((0.69	DMF)		"	0	-	в
4	-39.6 (1.2	CHC13)	5.94	CDC13	0	0.19	β
5	-49.7 (1.24	CHC13)	6.01	"	1.3	0.20	β
8	-25.5 (1.83	CHC13)		"	0		β
٩	-27.9 (1.18	CHC13)	6.55	*	2	0.24	β
14	-52.8 (1.06	H ₂ 0/MeOH)	6.37	"	1.3	0.18	β
15	-74,8 (1.21	DMF)	6.18	DMSO	4.75		β
	-80,9 C	1	DMF)	6.18	*	4,80		ß

Table 4. Specific optical rotations and J_{1'-2'} coupling constants of products 4-15

about the structures of nucleosides. We used circular dichroism to furnish additional structural proof, even though, the empiric rules devised to assign the anomeric configuration require careful use because the sign and the magnitude of the Cotton effect depend on several factors such as anomeric configuration, sugar ring conformation, heterocycle orientation with respect to the glycosidic bond, solvent, pH¹⁷⁻²⁰

In the manno series, the α derivatives 10 and 12 have a positive Cotton effect at 260 nm and a negative one at 235 nm. On the other hand, the β derivatives 6, 11 and 13 have two negative effects at 235 and 260 nm (Fig. 1 and 2).

2). The circular dichroism spectra of the 8-azainosine 15 and of the inosine, which is known to be in the β configuration, show the same general features; only the magnitude of the effect differs with the two negative effects merged in one at 245 nm in the inosine spectrum (Fig. 3). The spectra of 13 and 15 (Fig. 3) are identical. The negative Cotton effect at 260 nm indicates the β configuration, whatever the nature of the sugar moiety.

The derivatives 4 and 5 have a negative effect at 260 nm (Fig. 4), and only 14 has a positive one at the same wavelength (Fig. 4). In the spectra of 8 and 9 (Fig. 5) there is no negative effect at 260 nm. One can assume that the very intense positive effect at 250 nm hides the negative one at 260 nm probably due to interactions between the heterocycle and the triphenylmethyl and t-butyldiphenylsilyl groups at the C-5' carbon atom.

CONCLUSION

We believe that the work presented here provides a particularly efficient means of synthesis of vic-triazolo-(4,5,d)pyrimidone glycosides starting from a sugar with a free hemiacetalic hydroxyl. In the case of the 8-azainosine, we increased the total yield by more than 100% compared with the best one previously reported,¹¹ i.e. 30% with six steps against 13% with seven steps. We have also been able to control the anomeric configuration of the products in the manno series.

EXPERIMENTAL

U.V. spectra were recorded on a Jobin-Yvon model DUOS-PAC 203 ultraviolet spectrophotometer. Optical rotations were determined on a Perkin-Elmer model 141 digital readout polarimeter. The NMR spectra were measured with Perkin-Elmer R 12 B and Cameca 250 MHz spectrometers with T.M.S. as internal reference standard. C.D. spectra were recorded on a Jobin-Yvon III dichograph connected to a Tektronix 4051 (32 K) computer. M.ps are uncorrected. Microanalyses were carried out by the microanalytical laboratory of the C.N.R.S. Chromatography solvent mixture were v/v.

General procedure for formation of the vic-triazolo ring

Method 1. Glycosyl azide (1 mM) was added to a cooled solution of KOH (84 mg; 1.5 mM) and cyanoacetamide (126 mg; 1.5 mM) in H₂O (0.5 ml) and DMF (5 ml). The yellow solution was allowed to slowly warm up to room temperature. The reaction was monitored by tlc. When all the starting material had disappeared the mixture was evaporated to dryness in vacuo. The residue was extracted with ethyl acetate. The organic layers were collected, dried over MgSO₄ and evaporated to dryness. The residual syrup was analyzed chromatographically on a silica gel column.

Method 2. Glycosyl azide (1 mM) was added to a cooled solution of sodium cyanoacetamide in dry DMF (5 ml) (obtained from NaH in paraffin (72 mg) and cyanoacetamide (126 mg)). The solution was kept at 0° and worked up as described in method 1. 5 - Amino 4 - carboxamido 1 - (2',3' - O - isopropylidene - 5 - O

5 - Amino 4 - carboxamido 1 - (2',3' - O - isopropylidene - 5 - O - trityl β - D - ribofuranosyl) - vic triazole 4 Prepared by method 1 or 2 from 1 (457 mg; 1 mM). 4 (446 mg, 70%) was obtained by column chromatography with AcOEt as eluent. $[\alpha]_{25}^{25} = -39.6^{\circ}$ (c = 1.2, CHCl₃); m.p. (hexane-AcOEt) = 224°; ¹H NMR (δ CDCl₃ 250 MHz): 1.38 and 1.57 (6 H, 2 s, isopropylidene); 3.02 (2 H, m); 4.48 (1 H, m); 4.81 (1 H, q); 5.47 (2 H, m, ChH₂); 5.67 (1

H, d, H-2', $J_{2,3'} = 6$ Hz); 5.94 (1 H, s, H-1'); 7.30 (17 H, m, OCPh₃ + NH₂). UV (ethanol) $\epsilon_{259} = 7270$ (Found: C, 71.42; H, 4.78; N, 10.83; Calc. For $C_{38}H_{31}N_5O_5$ (637.7) C, 71.57; H, 4.90; H, 10.98%).

5 - Amino 4 - carboxamido - $1(2',3' - O - isopropylidene - 5 - O - t - butyldiphenyl silyl <math>\beta - D$ - ribofuranosyl) - vic triazole 5. Prepared from 2 (453 mg; 1 mM) by method 1 or 2. 5 (395 mg, 77%) was obtained by column chromatography with hexane/AcOEt 1/1 as eluent. $[\alpha]_{D}^{25} = -49.7^{\circ}$ (c = 1.24, CHCl₃); m.p. (hexane-AcOEt) = 190-192°; ¹H NMR (δ CDCl₃ 250 MHz): 1.0 (9 H, s, tBu); 1.37 and 1.57 (6 H, 2 s, isopropylidene); 3.53 (2 H, d, H-5' and H-5''); 4.4 (1 H, d.t, H-4', $J_{4'-5'} = J_{4'-5'} = 5.3$ Hz, $J_{4'-3'} = 2$ Hz); 4.91 (1 H, d.d, H-3', $J_{2'-3'} = 6.6$ Hz); 5.61 (2 H, s, CONH₂); 5.72 (1 H, d.d, H-2', $J_{1'-2'} = 1.33$ Hz); 6.01 (1 H, d. H-1', $J_{1'-2'} = 1.33$ Hz); 6-7 (2 H, NH₂); 7.48 (10 H, m, aromatic). UV (ethanol) $\epsilon_{204} = 12430, \epsilon_{233} = 14928, \epsilon_{221} = 29821.$ (Found: C, 58.35; H, 6.81; N, 13.67; Calc. for $C_{25}H_{35}O_5N_5SI$ (513.7) C, 58.46; H, 6.87; N, 13.63%).

5 - Amino 4 - carboxamido - 1 (2',3': 5',6' di - O - isopropylidène α - D - mannopyranosyl) vic - triazole 6. Prepared by method 1 from 3 (285 mg, 1 mM). 6 (295 mg, 80%) was obtained by column chromatography with hexane/AcOEt 1/4 as eluent. $[\alpha]_{5}^{25} = +55.8^{\circ}$ (c = 0.22, CHCl₃); m.p. (hexane-AcOEt) = 252°; ¹H NMR (δ pyridin D₅ 250 MHz) 1.36, 1.40, 1.46 1.56 (12 H, 4 s, isopropylidene); 3.5-5 (5 H, m); 5-12 (1 H, d, H-2', J_{2'-3'} = 5.5 Hz); 6.66 (1 H, s, H-1'); 7.52 (2 H, s, CONH₂); 8.24 and 8.51 (2 H, 2 s, NH₂). UV (ethanol) $\epsilon_{261} = 6200$, $\epsilon_{236} = 6700$. (Found; C, 48.65; H, 6.31; N, 18.87; Calc. for C₁₅H₂₃N₅O₆ (369.4) C, 48.77; H, 6.27; N, 18.96%).

5 - Amino 4 - carboxamido - 1 (2',3':5',6' di - () - isopropylidene



Fig. 1. CD spectra of 10, 11, 12 and 13 in EtOH.

β- D - mannopyranosyl) vic triazole 7. Prepared by method 2 from 3. 7 could not be isolated as a pure product. ¹H NMR (δ pyridin D₅ 250 MHz) 1.29, 1.45, 1.67 (12 H, 3 s, isopropylidene); 3.60 to 4.10 (3 H, m); 4.24 (1 H, d.d, H-4', J_{3'-4'} = 8 Hz); 4.60 (1 H, d.d, H-3', J_{2'-3'} = 5 Hz); 4.84 (1 H, d.d, H-2', J_{1'-2'} = 2.5 Hz); 6.78 (1 H, d, H-1'); 6.82 (2 H, s, CONH₂); 8.18 and 8.36 (2 H, 2 s, NH₂).

3 - 2,3' - O - isopropylidene - 5' - O - trityl β - D - ribofuranosyl) vic triazolo (4,5-d) pyrimid - 7 - one 8. 4 (637 mg, 1 mM) was heated to 100° with diethoxymethyl acetate (2 ml) for 3 hr. The orange solution was concentrated in vacuo, and the resulting amber syrup was dissolved in methanolic NH₃ (40 ml saturated at 0°) and allowed to stand in a sealed vessel at room temperature for 15 hr. The solution was evaporated to dryness and the residue was purified by column chromatography with AcOEt as eluent to give white crystals of 8 (440 mg, 85%). m.p. (hexane-AcOEt) = 128-130°; [α]_D²⁵ = -25.5° (c = 1.83, CHCl₃); ¹H NMR (δ CDCl₃ 60 MHz) 1.38 and 1.62 (6 H, 2 s, isopropylidene); 3.22 (2 H, m, H - 5' and H - 5''); 4.66 (1 H, m, H - 4'); 5.0 (1 H, d, H - 3''), 3_{y-4'} = 2.3 Hz, J_{2'-3'} = 6 Hz); 5.6 (1 H, d, H - 2'); 6.66 (1 H, s, H - 1'); 7.35 (15 H, m, aromatic); 7.51 (1 H, s, H - 5). UV (ethanol) ϵ_{257} = 12000. (Found: C, 67.15; H, 5.10; N, 13.01; Calc. for C₃₁H₂₉O₃N₅ (551.6) C, 67.51; H, 5.26; N, 12.70%).

3 - (2',3' - O - isopropylidene - 5' - O - terbutyldiphenylsilyl β - D - ribofuranosyl) vic triazolo (4,5 - d) pyrimid - 7 - one 9. 5 (513 mg, 1 mM) was treated as described above to give 9 (497 mg, 95%) after column chromatography with AcOEt as eluent. m.p. (hexane-AcOEt) = 80°; $[\alpha]_D^{2} = -27.9^{\circ}$ (c = 1.18, CHCl₃); ¹H NMR (δ CDCl₃ 250 MHz) 1.0 (9 H, s, tBu); 1.4 and 1.64 (6 H, 2 s, isopropylidene); 3.72 (2 H, m, H - 5' and H - 5''); 4.51 (1 H, td, H - 4', J_{4'-5'} = J_{4'-5'} = 7.3 Hz, J_{4'-3'} = 2.7 Hz); 5.10 (1 H, dd, H - 3', J_{2'-3'} = 7.3 Hz); 5.58 (1 H, dd, H - 2', J_{1'-2'} = 2 Hz); 6.55 (1 H, d, H - 1'); 7.45 (10 H, m, aromatic); 8.38 (1 H, s, H - 5); 10.10 (1 H, large s, N - H). UV (ethanol) $\epsilon_{256} = 81112, \epsilon_{206} = 11022.$ (Found: C, 59.91; H, 6.50; N, 13.37%).

3 - $(2',3';4',6' - di - O - isopropylidene \alpha - D - mannopyranosyl)$ vic triazolo (4,5-d) pyrimid - 7 - one 10. 6 (369 mg, 1 mM) was treated as described for 8 to afford 10 (360 mg, 95%) after column chromatography with AcOEt as eluent. m.p. (hexane-AcOEt) = 252° (with decomposition); $[\alpha]_{25}^{25} = +44.2°$ (c = 0.57, CHCl₃); ¹H NMR (δ DMSO D₆ 60 MHz) 1.42, 1.54 and 1.62 (12 H, 3 s, isopropylidene); 3.44 to 5.07 (7 H, m); 6.62 (1 H, s, H – 1'); 8.51 (1 H, s, H – 5). UV (ethanol) $\epsilon_{255} = 10940$. (Found: C, 50.25; H, 5.87; N, 18.15; Calc. for C₁₆H₂₁N₅O₆ (379.4) C, 50.65; H, 5.58; N, 18.46%).

3 - $(2^{'},3^{'},4^{'},6^{'}$ - di - O - *isopropylidene* β - D - *mannopyrannosyl*) vic triazolo (4,5-d) pyrimid - 7 - one 11. 7 was treated as described for 8 to afford 11 (95%) after column chromatography with AcOEt as eluent. m.p. (hexane-AcOEt) > 250° (with decomposition); $[\alpha]_{15}^{25} = -20.5^{\circ}$ (c = 0.81, CHCl₃); ¹H NMR (δ DMSO D₆ 60 MHz) 1.22, 1.35 and 1.54 (12 H, 3 s, isopropylidene); 3.29-4.78 (7 H, m); 6.64 (1 H, d, H - I', J_{1'-2'} = 3.35 Hz); 8.26 (1 H, d, H - 5, become s with D₂O); 12.77 (1 H, large s, NH). UV (ethanol) $\epsilon_{255} = 10276$. (Found: C, 49.39; H, 5.70; N, 18.35; Calc. for C₁₆H₂₁N₅O₆ (379.4); C, 50.65; H, 5.58; N, 18.46%).

3 - (α - D - mannopyranosyl) vic triazolo (4,5-d) pyrimid-7-one 12. 10 (379 mg, 1 mM) was dissolved in an aqueous trifluoroacetic acid solution (50%) (3 ml). The mixture was kept for 1 hr at room temperature and evaporated to dryness in vacuo. The crude crystals were stored over KOH overnight in vacuo in order to remove the trifluoroacetic acid. 12 was recrystallized in a mixture of H₂O/MeOH (293 mg, 98%). m.p. = 189-190°; [α]_D² = +37° (c = 0.81, DMF); ¹H NMR (δ DMSO D₆ 60 MH2) 6.09 (1 H, s, H - 1'); 8.33 (1 H, s, H - 5); 12.87 (1 H, s, NH). UV (ethanol) $\epsilon_{255} = 6353$. (Found C, 40.22; H, 4.40; N, 23.25; Calc. for C₁₀H₁₃O₆N₅ (299.2) C, 40.13; H, 4.34; N, 23.41%).

3-(β -D-mannopyranosyl) vic triazolo (4,5-d) pyrimid-7-one 13. 13 (290 mg, 97%) was obtained as described above from 11 (379 mg, 1 mM). m.p. = 172-173° (H₂O/MeOH); [a]₂₅²⁵ = +6.5° (c = 0.69, DMF); ¹H NMR (δ DMSO D₆ 250 MHz) 6.0 (1 H, s, H-1'); 8.23 (1 H, s, H-5). UV (ethanol) ϵ_{255} = 8400. (Found C, 39.80; H, 4.52; N, 23.31; Calc. for C₁₀H₁₃O₆N₅ (299.2) C, 40.13; H, 4.34; N, 23.41%).

3-(2',3'-O-isopropylidene β -D-ribofuranosyl) vic triazolo (4,5d) pyrimid-7-one 14. (a) From 8 (552 mg, 1 mM) was dissolved in an aqueous trifluoroacetic acid solution 50% (3 ml). The mixture was kept at room temperature and the reaction was monitored by tlc (AcOEt as eluent). The mixture was evaporated to dryness and purified by column chromatography to afford 14 (267 mg, 90%).



Fig. 2. CD spectra of 6 and 10 in EtOH.



Fig. 3. CD spectra of inosine, 8-azainosine 15 and 13 in EtOH

(b) From 9. 9 (524 mg, 1 mM) was added to a solution of tetrabutylammonium fluoride (300 mg) in tetrahydrofurane (4 ml) and kept at room temperature. Work up and purification as above give 14 (282 mg, 95%). M.p. 184–185° (H₂O/MeOH); $[\alpha]_{1}^{2}=-52.8^{\circ}$ (c = 1.06, H₂O–MeOH); ¹H NMR (8 DMSO D₆ 250 MHz) 1.35 and 1.53 (6 H, 2s, isopropylidene); 3.38 (2 H, m, H-5' and H-5''); 4.2 (2 H, m, H-4' and OH); 5.0 (1 H, d.d, H-3', J_{3'-4'} = 2 Hz); 5.53 (1 H, d.d, H-2', J_{2'-3'} = 6 Hz); 6.37 (1 H, d. H-1', J_{1'-2'} = 1.3 Hz); 7.69 (1 H, s, NH); 8.31 (1 H, s, H-5). UV (ethanol) $\epsilon_{255} = 9700$. (Found: C, 44.60; H, 4.89; N, 23.12; Calc. for C₁₁H₁₅N₅O₅ (297.3) C, 44.44; H, 5.09; N, 23.56%).

3-(β -O-ribofuranosyl) vic triazolo (4,5-d) pyrimid-7-one: 8azainosine 15. 14 (297 mg, 1 mM) was added to a suspension of resin (Dowex 50 H⁺) (400 mg) in a mixture of THF (2 ml) and H₂O (2 ml). The solution was kept at room temperature for 96 h. The resin was filtered and the filtrate was evaporated to dryness in vacuo. The crude product was recrystallized from MeOH-H₂O to give 15 (242 mg, 90%) as white crystals. M.p. = 220°; [a] β° = -74.8° (c = 1.21, DMF); [itt.¹¹ [a] β° = -80.9° (c = 1, DMF); ¹H NMR (δ DMSO D₆ 60 MHz) 6.18 (1 H, d, H-1', J₁-z' = 4.75 Hz); 8.31 (1 H, s, H-5). UV (ethanol) ϵ_{254} = 10000; (H₂O pH 7.5) ϵ_{256} = 10313.









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