Headline Articles

An Anion Receptor Based on Poly-Substituted Cobalticinium Complexes

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Various derivatives of poly-substituted cobalticinium complexes have been synthesized by using substituted cyclopentadienes with a convertible carboxylic group. By $^1\text{H}\,\text{NMR}$ titration experiments, these complexes were shown to behave as a receptor for anions such as Cl^- , Br^- , AcO^- , and $p\text{-TsO}^-$. Planar-chiral anilide derivative 3a showed chiral recognition toward camphor-10-sulfonate though the difference of the association constants was 10%. The molecular structure including the absolute configuration of 3a have been established by an X-ray crystallographic analysis. The results suggest the hydrophobic cavity constructed by the substituents on the cyclopentadienyl ligands may be an important component for the successful anion complexation.

Organic and inorganic anions are known to be important in chemical and biochemical processes, and their recognition by abiotic receptors is an area of current interest.¹⁾ In contrast to a large variety of host compounds which form host-guest complexes with cations, there are only a few abiotic receptors for organic anions.²⁾ For example, Lewisacid-containing ligands3) and quaternary ammonium salts4) have been reported so far. In particular, metallocene compounds such as ferrocene⁵⁾ and cobalticinium salts⁶⁾ have received much attention as ion-receptors based on their redox-active moieties. Beer et al. have prepared mono- or di-substituted cobalticinium amide complexes, and demonstrated electrochemical recognition for anions such as halides and nitrates.⁶⁾ Cobalticinium cations are, however, chemically inactive toward electrophilic substitutions on the cyclopentadienyl ring and there are few reports⁷⁾ about polysubstituted cobalticinium complexes in comparison with ferrocene derivatives.

During the last few years, we have reported the syntheses of several planar-chiral cobalticinium complexes bearing poly-substituted cyclopentadienyl ligands. Our method for preparing such poly-substituted cobalticinium complexes is based on use of a trisubstituted cyclopentadiene having a convertible ester group and we have obtained planar-chiral complexes in an enantiomerically pure form. Now, we have investigated their behavior toward anions and found that they act as an anion receptor for not only inorganic but also organic anions. The results of HNNR titration experiments and an X-ray structural analysis have indicated that the hydrophobic cavity constructed by the substituents on

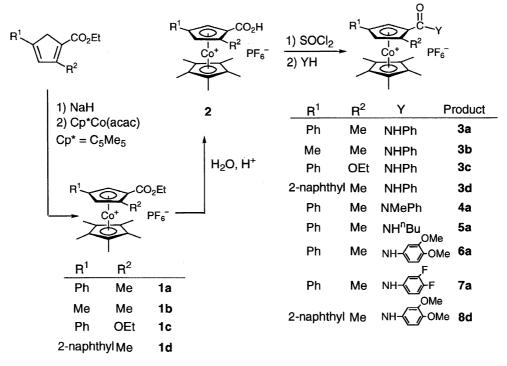
the cyclopentadienyl ligands is an important component for the anion complexation. We report here the properties as an anion receptor of poly-substitued cobalticinium complexes having an amide group on the cyclopentadienyl ligand.

Results and Discussion

Synthesis of Planar-Chiral Cobalticinium Complexes.

Cobalticinium complexes containing poly-substituted cyclopentadienyl ligands may be good candidates for anion receptors because the substituents as well as the cationic metal center may potentially construct a hydrophobic cavity which would be suitable to accommodate organic anions. Although the preparative method of poly-substitued cobalticinium complexes is limited because of the low reactivity of cobalticinium complexes toward electrophilic substitution reactions on the cyclopentadienyl ligand, they can be rather easily prepared by our new method.⁸⁾ Thus, we have now prepared ester (1), carboxylic acid (2), and amide derivatives (3-8) as shown in Scheme 1. Moreover planar chiral cobalticinium complexes are also prepared by our method and in some cases (3a, 3b) enantiomerically pure complexes have been isolated. Our method has an advantage for preparing various derivatives having a functional group via the carboxylic acid. Complexes (3—8) have also been prepared to evaluate the effects of substituents on the anion complexation (vide infra).

¹H NMR Titration Studies. Beer et al. reported that mono- or di-substituted cobalticinium amide derivatives act as an anion receptor. ^{6,11)} Their results showed that simple monoester derivatives of cobalticinium complexes had no



Scheme 1. Syntheses of poly-substituted cobalticinium complexes.

interaction with anions. ^{11a)} We examined the behavior of our ethyl ester derivative $\mathbf{1a}$ ($\mathbf{R}^1 = \mathrm{Ph}$, $\mathbf{R}^2 = \mathrm{Me}$) as a receptor by a ¹H NMR titration method. Thus, addition of tetraethylammonium p-toluenesulfonate (NEt_4^+p - TsO^-) to $\mathbf{1a}$ resulted in a down-field shift of the proton (H^a) at 5-position on the cyclopentadienyl ligand. The titration curve obtained from the experiment using CDCl₃ as a solvent is shown in Fig. 1, in which the change of chemical shift due to H^a is plotted against the molar equivalents of added p- TsO^- . The association constant (K_a) has been estimated at 7.7×10^2 M⁻¹ by use of a least-squares fit computer program ($1 \mathrm{M} = 1 \mathrm{mol} \, \mathrm{dm}^{-3}$).

Similar NMR titration experiments were done for complexes 1, 3, 4, and 5, and the association constants estimated for the complexation with anions such as halides and p-TsO are summarized in Table 1. It should be noted that in our case a substantial binding was observed between ester derivative 1a and p-TsO $^-$, though Beer et al. detected no interaction for a simple mono-substituted cobalticinium complex having an ester group, suggesting that a hydrophobic cavity constructed by R^1 and R^2 together with methyl groups on the pentamethylcyclopentadienyl ligand was an important component for successful complexation with an anion.

Anilide **3a** was found to form host–guest complexes with a variety of inorganic and organic anions. The titration experiment for **3a** with p-TsO $^-$ showed a large value of K_a (1.4×10⁴ M $^-$ 1) in CDCl₃. The titration curve of **3a** with p-TsO $^-$ traced by 1 H NMR is illustrated in Fig. 2, which shows the formation of a 1:1 host–guest complex. It is noteworthy that the complexation causes large downfield shifts of the amide proton (H b) and the proton (H a) on the trisubstituted cyclopentadienyl ligand. For example, shifts

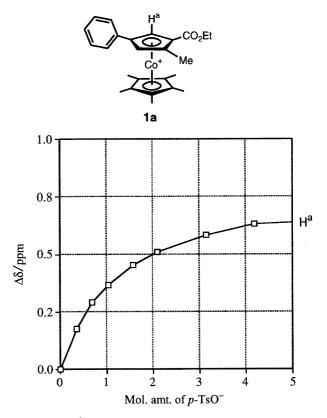


Fig. 1. 1 H NMR titration curve of **1a** with p-TsO ${}^{-}$.

of $\Delta \delta = 1.8$ ppm for the H^b and 1.0 ppm for H^a were observed on addition of equimolar amount of p-TsO⁻, implying that a CO–NH···X⁻ hydrogen bonding is a significant contributing factor to the overall anion complexation process. In the titra-

Table 1. Association Constants with Anions (25 °C, CDCl₃)

Host complex	Guest anion	K_a/M^{-1}
3a	Br ^{- a)}	ca. 10 ^{4 c)}
3a	Cl ^{- a)}	ca. $10^{4 \text{ c}}$
3a	AcO ^{- a)}	ca. 10 ^{5 c)}
3a	p -TsO $^{- \text{ b}}$	1.4×10^{4} c)
3b	p -TsO $^-$	$8.8 \times 10^{3} \text{ c}$
3c	p -TsO $^-$	8.6×10^{2} c)
4 a	p -TsO $^-$	3.8×10^{2} d)
5a	p -TsO $^-$	2.6×10^{3} c)
1a	p-TsO ⁻	7.7×10^{2} d)

a) Tetrabutylammonium salt was used. b) Tetraethylammonium salt was used. c) The value was calculated for H^b . c) The value was calculated for H^a .

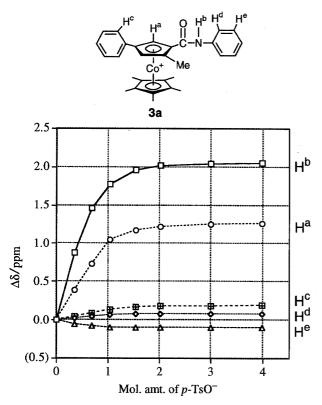


Fig. 2. 1 H NMR titration curves of **3a** with p-TsO ${}^{-}$.

tion experiments of 3a with Cl⁻, Br⁻, and AcO⁻, the same phenomenon was observed ($\Delta\delta$ for H^b: Cl⁻, 2.3; Br⁻, 1.8; AcO⁻, 3.4 ppm). The binding site is, therefore, undoubtedly the same in all the cases and is presumably located near the H^a and H^b groups in a hydrophobic cavity surrounded by a cobalt atom and substituents on two cyclopentadienyl ligands.

To find details about the anion complexation process, we did titration experiments for several derivatives **3**—**5**. Complexes **3a**, **3b**, and **5a** bearing an appending amide NH unit, which is capable to form a favorable hydrogen bond with a guest anion, afforded larger values of K_a than those of **1a**, **3c**, and **4a**. The poor association ($K_a = 8.6 \times 10^2 \text{ M}^{-1}$) between **3c** and $p\text{-TsO}^-$ may be due to the contribution of an

intramolecular hydrogen bond between NH and the neighboring OEt group which prevents the hydrogen bonding of NH with a guest anion (Fig. 3). This may be supported by the infrared spectrum (taken for a sample in solid state) in which 3c showed an amide NH stretching absorption at 3380 cm⁻¹ lower wavenumber than **3a** at 3410 cm⁻¹. Furthermore, in the ¹H NMR spectrum, the resonance due to the NH of 3c (8.84 ppm) was observed in a lower field relative to that of 3a (8.49 ppm). These results suggest an intramolecular hydrogen bonding between NH and OEt group. N-Butyl amide 5a gave a smaller value of K_a than that of anilide 3a. This may be due to the weaker hydrophobic effect of 5a in comparison with anilide derivative 3a. From these results, tentative model of the host-guest complex [3a·p-TsO-] is shown in Fig. 4. This may be supported by the X-ray crystallographic analysis of **3a** (vide infra).

X-Ray Structural Analysis of (+)-3a. To establish the molecular structure of 3a including the absolute configuration, an X-ray diffraction study of (+)-3a has been done. Recrystallization of (+)-3a from ethanol gave single crystals suitable for an X-ray analysis. The measurement was done with Cu $K\alpha$ radiation. The cobalt anomalous dispersion term for Cu $K\alpha$ ($\Delta f'' = 3.608$)¹²⁾ is much larger than that for Mo $K\alpha$ ($\Delta f'' = 0.973$) and this effect has an advantage even considering the problem due to the higher crystal absorption when using $Cu K\alpha$ radiation. The Bijvoet pairs were treated as independent reflections. In the course of the refinement with all observed reflections, two refinements were used. The refinement with the configuration shown in this article with anisotropic thermal parameters for non-H atoms gave the R factor 0.045 and $R_{\rm w}$ 0.054. On the other hand in the model with the opposite enantiomer they were 0.117(R) and $0.154 (R_{\rm w})$, even though the calculation was repeated from the isotropic stage. From these results, we have identified the absolute configuration of (+)-3a around the Cp-M moiety to be R. The molecular structure is illustrated in Fig. 5 together with atom labeling scheme. All the bond lengths are typical. The two Cp rings are planar, nearly parallel to

Fig. 3. Tentative model of 3c (top view).

Me O

$$Co^{+}$$
 $SO_{3}^{-} = p \cdot TsO^{-}$

Fig. 4. Tentative model of host–guest complex [3a·p-TsO⁻] (side view).

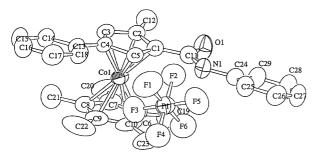


Fig. 5. ORTEP drawing of (1R)-(+)-3a with atom labelling scheme. Hydrogen atoms have omitted for clarity.

one another (dihedral angle, 2.67°). The conformation is midway between the fully eclipsed and staggered. The dihedral angles between trisubstituted cyclopentadiene and two phenyl rings (C18—23, C24—29) are 10.11° and 11.92°, respectively. It is noteworthy that there seems to be a cavity constructed by anilide and phenyl (R¹) together with methyl groups on the pentamethylcyclopentadienyl ligand, and the amide proton (H¹b) and the proton (H¹a) at the 5-position on the cyclopentadienyl ligand are located closely. As predicted by the large downfield shift of both H¹a and H¹b protons in the NMR spectra, the binding site may exist in the neighborhood of these protons.

Chiral Recognition. As our cobalticinium complexes have a planar chirality, a chiral recognition may be expected in a host-guest complexation. Thus, tetrabutylammonium (+)-camphor-10-sulfonate [(+)-camph] was used as a chiral guest toward the host cobalticinium complex (\pm) -3a. Addition of the chiral sulfonate to a solution of (\pm) -3a in CDCl₃ also caused a large shift (1—2 ppm downfield) of proton signals due to H^a and H^b. However, the resultant titration curve of 3a was significantly different from that with p-TsO⁻, and the NH signal of 3a was split into two peaks (the difference of chemical shift: 19 Hz). The splitting indicates the formation of diastereomeric host-guest complexes [(+)-camph·(+)-3a and (+)-camph·(-)-3a]. The two peaks were crossed with each other before reaching an equilibrium state, suggesting the different K_a between the diastereomeric complexes of (+)-3a and (-)-3a. In the case of (\pm)-3b with (+)-camph, the H^a signal as well as H^b was split into two peaks (the difference of chemical shift: 17 Hz), but they were not crossed with each other.

Although complexes 4a and 5a, which showed small association constants with camphor-10-sulfonate, did not induce splitting of the NMR signals, 6—8d with large association constants afforded similar titration curves to 3a. The association constants between cobalticinium complexes 3—8 and camphorsulfonate are summarized in Table 2. The association constants of 6a and 7a, which have electron-donating or -withdrawing substituents on the anilide group, indicate almost no electric effects of the substituents on complexation with the anion.

A quantitative study was done to estimate the degree of chiral recognition. Enantiomeric pure (+)-3a was prepared from carboxylic acid (+)-2a by the method reported

Table 2. Association Constants with Camphor-10-sulfonate (25 °C, CDCl₃)

Host-complex	K_a/M^{-1}
3a	1.8×10 ⁴ b)
3b	1.1×10^{4} b)
3c	1.9×10^{3} b)
4a	6.1×10^{2} a)
5a	5.8×10^{3} a)
6a	$1.8 \times 10^{4 \text{ b}}$
7a	$2.5 \times 10^{4 \text{ b}}$
8d	1.7×10^{4} b)

a) The value was calculated for H^a . b) The value was calculated for H^b .

previously.^{8b)} The association constants for the complexation of (+)-**3a** with (+)-and (-)-camphorsulfonate were calculated by the ¹H NMR titration method in a 1:1 mixture of CDCl₃/CD₃CN at 25 °C. The association constants K_a and K'_a , $(K_a = [(+)-3\mathbf{a}\cdot(+)-\text{camph}]/[(+)-3\mathbf{a}]\cdot[(+)-\text{camph}]$, have been estimated to be 1900 and 1700 M⁻¹, respectively. Consequently cobalticinium complex (+)-**3a** is able to recognize the chirality of camphorsulfonate, though the ability of recognition is not high.

In conclusion, we have studied the properties as an anion receptor based on poly-substituted cobalticinium complexes. These complexes, which have a hydrophobic cavity constructed by substituents on cyclopentadienyl ligands, are capable of receiving anions such as p-TsO $^-$ and have potentials to recognize the chirality of guest anions. One of features of our cobalticinium anion receptor is to have a convertible function which may be appropriately designed for a binding site.

Experimental

All the reactions except for hydrolysis were done under an atmosphere of nitrogen or argon. Melting points are uncorrected.

¹H NMR spectra were measured in CDCl₃ or CD₃OD with SiMe₄ as an internal standard and recorded on a Bruker AM360 or JEOL EX-270 spectrometer. Upfiled shifts are quoted as negative. IR spectra were recorded on a Hitachi 295 spectrometer. Mass spectrometry was performed with a Shimadzu QP-2000 GC-MS (EI, 70 eV) or JEOL JMX-DX300 (FAB) spectrometer. Elemental analysis was done by using a Perkin–Elmer 240C. Optical rotatory powers were measured on a JASCO DIP-370 digital polarimeter. Solvents were dried in usual manners and distilled. Unless stated to the contrary, commercial grade chemicals were used without further purification. Trisubstituted cyclopentadienes and complexes 1a, 1b, 2a, 2b, 3a, and 3b were prepared by the reported methods.

^{8b)}

Synthesis of Ethylester Derivatives (1c, 1d). The reaction of Co(acac)₂ (2.62 g, 10 mmol) with 1 equimolar amount of C₅Me₅Li in THF (50 mL) at -78 °C afforded C₅Me₅Co(acac). To this reaction mixture was added a solution of sodium 2-ethoxy-1-ethoxycarbonyl-4-phenylcyclopentadienide (Cp'Na) generated from Cp'H (2.58 g, 10 mmol) and NaH (60% in mineral oil; 0.44 g, 11 mmol) in THF (50 mL) at 0 °C. The mixture was allowed to warm to room temperature and stirred for 24 h, then was poured over 100 mL of 6 M hydrochloric acid. Ether (50 mL) was added

to the resulting solution, the aqueous layer was separated and then washed with ether to remove unreacted cyclopentadienes. To the aqueous solution dropwise addition of a saturated solution of ammonium hexafluorophosphate (4.89 g, 30 mmol) in water afforded compound 1c as yellow precipitates. Recrystallization from MeOH gave pure 1c as yellow powders in 30% yield (1.80 g).

1c: Mp 206.0—206.5 °C; IR (Nujol) 1740, 1240 cm⁻¹;
¹H NMR (360 MHz, CDCl₃) δ =7.70 (d, 2H, J=7.2 Hz), 7.56—7.46 (m, 3H), 6.30 (d, 1H, J = 2.2 Hz), 5.51 (d, 1H, J = 1.9 Hz), 4.47—4.37 (m, 2H), 4.35—4.26 (m, 2H), 1.65 (s, 15H), 1.50 (t, 3H, J=7.1 Hz), 1.45 (t, 3H, J=7.1 Hz); Mass (EI) m/z 451 (M⁺—PF₆). Found: C, 52.42; H, 5.41; P, 4.93; F, 19.03%. Calcd for C₂₆H₃₂O₃CoPF₆: C, 52.36; H, 5.41; P, 5.19; F, 19.11%.

Complex **1d** was prepared following the method for **1c** starting with 1-ethoxycarbonyl-2-methyl-4-naphthyl-1,3-cyclopentadiene.

1d: Orange powder (Yield 51%); mp 276.5—277.0 °C (decomp); IR (Nujol) 1725, 1255, and 1240 cm $^{-1}$; 1 H NMR (270 MHz, CDCl₃) δ = 8.23 (s, 1H), 8.10—7.54 (m, 6H), 6.03 (d, 1H, J=2.0 Hz), 5.89 (d, 1H, J=2.0 Hz), 4.44 (dq, 2H, J=7.2, 2.0 Hz), 2.25 (s, 3H), 1.61 (s, 15H), 1.49 (t, 3H, J=7.1 Hz); Mass (EI) m/z 471 (M $^{+}$ -PF₆). Found: C, 56.26; H, 5.03; P, 5.14; F, 18.74%. Calcd for C₂₉H₃₂O₂CoPF₆: C, 56.50; H, 5.23; P, 5.02; F, 18.49%.

Hydrolysis of 1c and 1d. Ethylester derivative **1c** (2.78 g, 4.7 mmol) was suspended in 300 mL of concentrated hydrochloric acid and heated for 24 h at 80 °C with constant stirring. The reaction mixture was evaporated to give yellow solids which were dissolved in 50 mL of hot water. Dropwise addition of ammonium hexafluorophosphate (3.83 g, 24 mmol) in water produced pale yellow precipitates, which were purified by recrystallization from EtOH. Orange needles (2.51 g) of carboxylic acid **2c** were obtained in 95% yield.

2c: Mp 196.0—197.0 °C (decomp); IR (Nujol) 1720 cm⁻¹; 1 H NMR (360 MHz, CD₃OD) δ = 7.77—7.75 (m, 2H), 7.58—7.53 (3H, m), 6.10 (d, 1H, J = 2.0 Hz), 6.97 (d, 1H, J = 2.0 Hz), 4.21 (q, 1H, J = 6.8 Hz), 4.19 (q, 1H, J = 7.0 Hz), 1.66 (s, 15H), 1.50 (t, 3H, J = 7.1 Hz); Mass (FAB) m/z 423 (M⁺—PF₆). Found: C, 50.73; H, 4.91; P, 5.30; F, 20.09%. Calcd for C₂₄H₂₈O₃CoPF₆: C, 50.72; H, 4.97; P, 5.45; F, 20.06%.

The same treatment of 1d gave 2d, which was used for the next reaction without further purification.

2d: Orange powder (Yield 83%); mp 240.0—243.0 °C (decomp); IR (Nujol) 3700—3200, 1720 cm⁻¹; 1 H NMR (360 MHz, CD₃OD) δ = 8.32 (s, 1H), 8.05—7.59 (m, 6H), 6.21 (s, 1H), 5.96 (s, 1H), 2.29 (s, 3H), 1.66 (s, 15H).

Synthesis of Amide Derivatives. Carboxylic acid 2c (0.57 g, 1.0 mmol) was refluxed with thionyl chloride (20 mL) for 3 h to give the corresponding acid chloride. After removal of thionyl chloride in vacuo, the resultant solid was dissolved in acetonitrile (20 mL). The solution was added to a solution of aniline (0.47 g, 5.0 mmol) in acetonitrile (10 mL). The mixture was stirred at room temperature overnight and the solvent was removed in vacuo. The crude product was dissolved in MeOH and ammonium hexafluorophosphate (0.49 g, 3.0 mmol) was added. Evaporation of MeOH and washing with water gave orange solids, which were purified by recrystallization from MeOH. Orange plates (0.52 g) were obtained in 82% yield.

3c: Orange plates (Yield 82%); mp 248.0—249.5 °C (decomp); IR (Nujol) 3380, 1680, and 1605 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ = 8.84 (s, 1H), 7.69—7.41 (m, 10H), 6.24 (d, 1H, J = 2.0 Hz), 5.69 (d, 1H, J = 1.8 Hz), 4.61—4.45 (m, 2H), 1.66 (s, 15H), 1.65 (t, 3H, J = 7.2 Hz); Mass (FAB) m/z 498 (M⁺ – PF₆) Found: C, 56.22; H, 4.89; N, 2.31; P, 4.60; F, 17.54%. Calcd for C₃₀H₃₃NO₂CoPF₆:

C, 56.00; H, 5.17; N, 2.18; P, 4.81; F, 17.71%.

Anilide derivative 3d was prepared following the method for 3c starting with 2d.

3d: Orange-red needles (Yield 56%); mp 260 °C (decomp); IR (Nujol) 1675, 1600, and 1535 cm⁻¹; 1 H NMR (360 MHz, CDCl₃) δ = 8.85 (s, 1H), 8.29 (s, 1H), 8.13—7.13 (m, 11H), 6.72 (s, 1H), 5.24 (s, 1H), 2.11 (s, 3H), 1.54 (s, 15H); Mass (EI) m/z 471 (M⁺-PF₆). Found: C, 60.04; H, 5.05; N, 2.33; P, 4.39; F, 16.98%. Calcd for C₃₃H₃₃NOCoPF₆: C, 59.74; H, 5.01; N, 2.11; P, 4.67; F, 17.18%.

N-Methylanilide derivative **4a** was prepared following the method for **3c** starting with **2a** and *N*-methylaniline.

4a: Yellow needles (Yield 72%); mp 274.0—275.0 °C (decomp); IR (Nujol) 1650, 1600 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ = 7.45—7.43 (m, 3H), 7.34—7.30 (m, 5H), 7.07 (d, 2H, J = 7.2 Hz), 5.50 (d, 1H, J = 1.6 Hz), 4.83 (d, 1H, J = 1.8 Hz), 3.48 (s, 3H), 2.05 (s, 3H), 1.66 (s, 15H); Mass (EI) m/z 482 (M⁺ –PF₆). Found: C, 57.25; H, 5.19; N, 2.31; P, 4.73; F, 18.29%. Calcd for C₃₀H₃₃NOCoPF₆: C, 57.42; H, 5.30; N, 2.23; P, 4.94; F, 18.17%.

N-Butyl amide derivative **5a** was prepared following the method for **3c** starting with **2a** and *N*-butyl amine.

5a: Orange plates (Yield 89%); mp 202.0—203.0 °C (decomp); IR (Nujol) 3445, 1670 cm $^{-1}$; 1 H NMR (360 MHz, CDCl $_{3}$) δ = 7.64 (d, 2H, J = 6.5 Hz), 7.55—7.45 (m, 3H), 7.11 (s, 1H), 6.20 (d, 1H, J=1.7 Hz), 5.19 (d, 1H, J=1.7 Hz), 3.56—3.46 (m, 1H), 3.39—3.30 (s, 3H), 2.26 (s, 3H), 1.72—1.66 (m, 2H), 1.63 (s, 15H), 1.46—1.36 (m, 2H), 0.96 (t, 3H, J = 7.3 Hz); Mass (EI) m/z 448 (M $^{+}$ -PF $_{6}$). Found: C, 54.70; H, 5.77; N, 2.14; P, 5.47; F, 19.06%. Calcd for C $_{27}$ H $_{35}$ NOCoPF $_{6}$: C, 54.64; H, 5.94; N, 2.36; P, 5.22; F, 19.21%.

3,4-Dimethoxyanilide derivative 6a was prepared following the

Table 3. Experimental Parameters for the X-Ray Diffraction Study of (1R)-(+)-3a

Formula	C ₂₉ H ₃₁ NOPF ₆ Co
Molecular weight	613.47
Crystal data	
Crystal color, habit	Yellow, prismatic
Crystal system	Orthorhombic
Space group	$P2_12_12_1$
$a/ m \AA$	17.346(2)
$b/{ m \AA}$	19.965(2)
$c/ m \AA$	8.131(2)
$V/\text{\AA}^3$	2816.0(7)
\boldsymbol{z}	4
$D_{ m calcd}/{ m gcm}^{-1}$	1.447
Measurement	
Crystal size/mm	$0.35 \times 0.25 \times 0.15$
Radiation	Cu $K\alpha$ (1.54178 Å, Ni filtered)
No. of reflections	
independent	4463
used $F_0 \geq 3\sigma F_0 $	3344
Absorption	
μ /cm ⁻¹	58.80
correction ψ scan	
correction factor	max 1.0
	min 0.70
Decay/I	No
Refimement	
R	0.045(0.117) ^{a)}
$R_{ m w}$	0.054(0.154) a)

a) Values in parentheses are for the enantiomer, (S)-3a.

method for 3c starting with 2a and 3,4-dimethoxyaniline.

6a: Orange needles (Yield 70%); mp 220.5—221.0 °C (decomp); IR (Nujol) 3425, 1680, and 1610 cm⁻¹; 1 H NMR (360 MHz, CDCl₃) δ = 8.27 (s, 1H), 7.65—7.60 (m, 3H), 7.44—7.40 (m, 3H), 7.32 (d, 1H, J = 2.2 Hz), 6.86 (d, 1H, J = 8.8 Hz), 6.11 (s, 1H), 5.35 (s, 1H), 3.98 (s, 3H), 3.89 (s, 3H), 2.18 (s, 3H), 1.57 (s, 15H); Mass (EI) m/z 528 (M⁺-PF₆). Found: C, 55.41; H, 5.06; N, 2.24; P, 4.84; F, 17.04%. Calcd for C₃₁H₃₅NO₃CoPF₆: C, 55.28; H, 5.24; N, 2.08; P, 4.60; F, 16.92%.

3,4-Diffuoroanilide derivative **7a** was prepared following the method for **3c** starting with **2a** and 3,4-diffuoroaniline.

7a: Orange needles (Yield 86%); mp 262.0—263.0 °C (decomp); IR (Nujol) 3420, 1695, and 1620 cm $^{-1}$; 1 H NMR (360 MHz, CDCl $_{3}$) δ = 8.57 (s, 1H), 8.03—7.97 (m, 1H), 7.67 (d, 2H, J = 7.6 Hz), 7.50—7.47 (m, 4H), 7.18 (q, 1H), 6.28 (s, 1H), 5.31 (s, 1H), 2.24 (s, 3H), 1.61 (s, 15H); Mass (EI) m/z 504 (M $^{+}$ -PF $_{6}$). Found: C, 53.38; H, 4.21; N, 2.07; P, 4.45; F, 23.42%. Calcd for $C_{29}H_{29}NOCoPF_{6}$: C, 53.63; H, 4.50; N, 2.16; P, 4.77; F, 23.40%.

3,4-Dimethoxyanilide derivative **8d** was prepared following the method for **3c** starting with **2d** and 4-aminoveratrole.

8d: Orange needles (Yield 54%); mp 269.0—272.0 °C (decomp); IR (Nujol) 3420, 1680, and 1600 cm⁻¹; ¹H NMR (360

MHz, CDCl₃) δ = 8.28—6.87 (m, 11H), 6.25 (s, 1H), 5.34 (s, 1H), 4.02 (s, 3H), 3.92 (s, 3H), 2.12 (s, 3H), 1.55 (s, 15H); Mass (EI) m/z 578 (M⁺ – PF₆). Found: C, 57.93; H, 5.02; N, 1.93; P, 4.23; F, 15.51%. Calcd for C₃₅H₃₇NO₃CoPF₆: C, 58.10; H, 5.15; N, 1.94; P, 4.28; F, 15.75%.

General Procedure for NMR Titration and Determination of Association Constants. For an example, the titration experiment of anilide 3a with p-toluenesulfonate is as follows. A 0.0022 M solution of 3a in CDCl₃ (2.68 mg in 2 mL) was prepared in a 2 mL volumetric flask. 0.50 mL of the solution was taken into a 5 mm NMR tube. A 0.054 M solution of Et₄N⁺p-TsO⁻ in CDCl₃ (32.3 mg in 2.00 mL) was prepared in a 2 mL volumetric flask. An initial NMR spectrum of the solution of 3a was taken, and the initial chemical shifts both of the Ha and Hb were measured to be 6.29 and 8.49 ppm. $^{13)}$ The solution of $\mathrm{Et_4N^+}p\text{-TsO}^-$ was then added, initially in 7 µL portions and the chemical shift of the protons was recorded after each addition. After 1 equimolar amount of guest was added, the sample size was increased to 10 µL. After a total of 41 µL had been added, the sample size was increased to 20 µL. The portions were added until no further change in the chemical shift of the protons was observed (usually 4-10 mol. amt.). The temperature of the NMR probe was 25 °C. The association constant

Table 4. Atomic Coordinates with Equivalent Isotropic Temperature Factors for (1R)-(+)-3a

P(1) 0.4	x 28769(4)	у	z	$B_{\rm eq}^{\rm a)}$	Atom				- a)
P(1) 0.4	28769(4)		~	$\boldsymbol{D}_{\mathrm{eq}}$	Atom	x	У	Z	$B_{ m eq}{}^{ m a)}$
		0.29629(3)	0.43319(8)	4.18(1)	C(26)	0.5699(3)	0.3821(2)	1.0786(6)	6.1(1)
F(1) 04	49291(7)	0.15187(7)	0.8470(2)	5.77(3)	C(27)	0.5772(3)	0.4509(3)	1.0720(7)	6.9(1)
1 (1)	4720(2)	0.2034(1)	0.9027(4)	8.11(8)	C(28)	0.5252(3)	0.4874(2)	0.9838(7)	6.6(1)
F(2) 0.4	4476(2)	0.1419(2)	0.6771(5)	10.4(1)	C(29)	0.4664(2)	0.4572(2)	0.9003(6)	5.6(1)
F(3) 0.4	4472(2)	0.0941(1)	0.9230(6)	11.8(1)	H(1)	0.4031	0.3055	0.8230	5.9095
F(4) 0.5	5571(2)	0.1045(2)	0.7867(5)	11.9(1)	H(2)	0.1329	0.2809	0.4701	5.2962
F(5) 0.5	5359(2)	0.1661(2)	1.0112(4)	9.07(9)	H(3)	0.3342	0.2321	0.7060	4.8301
F(6) 0.5	5358(2)	0.2136(2)	0.7667(4)	10.9(1)	H(4)	0.2072	0.4455	0.6187	8.0446
O(1) 0.3	3204(2)	0.4381(1)	0.7439(5)	6.76(9)	H(5)	0.1690	0.4220	0.4561	8.0446
N(1) 0.3	3982(2)	0.3526(2)	0.8249(5)	4.89(9)	H(6)	0.1262	0.4127	0.6220	8.0446
C(1) 0.2	2831(2)	0.3284(2)	0.6715(5)	4.49(10)	H(7)	0.1198	0.1713	0.3930	6.4789
C(2) 0.2	2115(2)	0.3449(2)	0.5897(5)	4.43(9)	H(8)	0.0934	0.0573	0.3544	8.2456
C(3) 0.	1795(2)	0.2848(2)	0.5310(5)	4.47(10)	H(9)	0.1686	-0.0235	0.4798	8.0976
C(4) 0.2	2270(2)	0.2299(2)	0.5775(5)	3.92(9)	H(10)	0.2704	0.0084	0.6467	7.8415
C(5) 0.3	2920(2)	0.2569(2)	0.6620(5)	4.06(9)	H(11)	0.2966	0.1215	0.6946	6.1414
C(6) 0.3	3952(2)	0.2910(3)	0.3342(6)	6.4(1)	H(12)	0.4811	0.2339	0.4283	13.0228
C(7) 0.3	3620(3)	0.3550(3)	0.3031(6)	5.8(1)	H(13)	0.5104	0.3022	0.3626	13.0228
C(8) 0.3	2943(3)	0.3448(2)	0.2123(6)	5.6(1)	H(14)	0.4671	0.2986	0.5294	13.0228
C(9) 0.3	2845(3)	0.2746(3)	0.1882(5)	5.8(1)	H(15)	0.4380	0.4188	0.4090	11.6065
C(10) 0.3	3474(3)	0.2418(2)	0.2646(6)	6.1(1)	H(16)	0.4016	0.4482	0.2504	11.6065
C(11) 0.3	3357(2)	0.3780(2)	0.7479(6)	4.8(1)	H(17)	0.3543	0.4461	0.4123	11.6065
C(12) 0.	1747(3)	0.4130(2)	0.5695(7)	6.5(1)	H(18)	0.2586	0.4409	0.1768	10.5748
C(13) 0.3	2116(2)	0.1577(2)	0.5485(5)	4.41(9)	H(19)	0.2400	0.3951	0.0279	10.5748
C(14) 0.	1505(2)	0.1382(2)	0.4463(6)	5.4(1)	H(20)	0.1902	0.3909	0.1861	10.5748
C(15) 0.	1352(3)	0.0704(2)	0.4233(7)	6.8(1)	H(21)	0.1881	0.2723	0.0531	12.1903
C(16) 0.	1792(3)	0.0228(2)	0.4972(7)	6.8(1)	H(22)	0.2467	0.2173	0.0028	12.1903
C(17) 0.3	2393(3)	0.0416(2)	0.5959(7)	6.4(1)	H(23)	0.1981	0.2091	0.1618	12.1903
C(18) 0.3	2554(2)	0.1090(2)	0.6233(6)	5.1(1)	H(24)	0.3642	0.1536	0.3856	12.4212
C(19) 0.4	4699(3)	0.2801(4)	0.4214(8)	11.0(2)	H(25)	0.3221	0.1441	0.2189	12.4212
C(20) 0.3	3920(4)	0.4232(3)	0.3482(8)	10.2(2)	H(26)	0.4102	0.1575	0.2222	12.4212
C(21) 0.3	2405(3)	0.3986(3)	0.1449(7)	9.2(2)	H(27)	0.5057	0.3042	0.9980	6.5305
C(22) 0.3		0.2404(3)	0.0934(7)	9.8(2)	H(28)	0.6054	0.3560	1.1398	7.3349
C(23) 0.	3633(4)	0.1669(3)	0.2732(8)	10.3(2)	H(29)	0.6178	0.4730	1.1285	8.2196
		0.3884(2)	0.9073(5)	4.5(1)	H(30)	0.5302	0.5348	0.9801	7.8156
C(25) 0.	5104(2)	0.3514(2)	0.9954(6)	5.4(1)	H(31)	0.4313	0.4835	0.8373	6.9359

a) $B_{eq} = \frac{8}{3}\pi^2(U_{11}(aa^*)^2 + U_{22}(bb^*)^2 + U_{33}(cc^*)^2 + 2U_{12}aa^*bb^*\cos\gamma + 2U_{13}aa^*cc^*\cos\beta + 2U_{23}bb^*cc^*\cos\alpha).$

was obtained using Eqs. 1 and 2^{14} Here, x is the

$$K_{\rm a} = x/([{\rm M}]_0 - x)([{\rm A}]_0 - x)$$
 (1)

$$x = [\mathbf{M}]_0 (\Delta \delta / \Delta \delta c) \tag{2}$$

concentration of the 1:1 complex in the equilibrium, $[M]_0$ and $[A]_0$ are initial concentrations of $\bf 3a$ and p-TsONEt₄, respectively, $\Delta \delta$ is the induced change in chemical shift of $\bf H^a$ or $\bf H^b$ proton signal, and $\Delta \delta c$ is the limiting shift, corresponding to the induced change after 100% complexation. From a series of measured $\Delta \delta$

$$\Delta \delta = [1/K_a + [M]_0 + [A]_0 - \{(1/K_a + [M]_0 + [A]_0)\}^2$$

$$-4[M]_0[A]_0\}^{1/2}]\Delta\delta c/(2[M]_0)$$

together with known $[A]_0$ and $[M]_0$ values, the K_a could be calculated by nonlinear least-squares method (modified Newton–Gauss type). Eight different concentrations of $[A]_0$ were measured.

In the case of the titration of (+)-3a with (+) or (-)-camphor sulfonate, we did the experiment for 4 times (in CDCl₃/CD₃CN=1/1) and their average was adapted.

Preparation of Tetrabutylammonium (+)-(S)-Camphor-10-sulfonate. To a solution of (+)-(S)-camphor-10-sulfonic acid monohydrate 1.93 g (7.7 mmol) in water (50 mL) was added 1.0

Table 5. Selected Bond Distances (Å) and Bond Angles (°) in (1R)-(+)-3a

		Te 5. Selected Bolla	Distances (21)	and Bond Angles ()	m (11t) (1)	Ju	
a) Bond distances				C(7)-C(6)-C(10)	107.7(6)	C(7)-C(6)-C(19)	123.6(7)
Co(1)–C(1)	2.041(6)	Co(1)-C(2)	2.071(5)	C(10)-C(6)-C(19)	128.7(7)	Co(1)-C(7)-C(6)	69.7(4)
Co(1)-C(3)	2.047(5)	Co(1)-C(4)	2.059(5)	Co(1)-C(7)-C(8)	69.5(4)	Co(1)-C(7)-C(20)	128.3(4)
Co(1)-C(5)	2.020(5)	Co(1)-C(6)	2.040(6)	C(6)-C(7)-C(8)	108.2(6)	C(6)-C(7)-C(20)	126.7(7)
Co(1)-C(7)	2.043(6)	Co(1)-C(8)	2.040(6)	C(1)- $Co(1)$ - $C(2)$	40.8(2)	C(1)- $Co(1)$ - $C(3)$	68.2(2)
Co(1)-C(9)	2.032(6)	Co(1)-C(10)	2.037(5)	C(1)- $Co(1)$ - $C(4)$	69.0(2)	C(1)- $Co(1)$ - $C(5)$	41.4(2)
P(1)-F(1)	1.536(4)	P(1)-F(2)	1.606(4)	C(1)- $Co(1)$ - $C(6)$	109.7(2)	C(1)- $Co(1)$ - $C(7)$	133.4(2)
P(1)-F(3)	1.605(5)	P(1)-F(4)	1.549(4)	C(1)- $Co(1)$ - $C(8)$	172.7(3)	C(1)- $Co(1)$ - $C(9)$	145.9(3)
P(1) - F(5)	1.559(4)	P(1)-F(6)	1.581(4)	C(1)- $Co(1)$ - $C(10)$	115.4(3)	C(2)-Co(1)-C(3)	40.1(2)
O(1)-C(11)	1.230(6)	N(1)– $C(11)$	1.356(7)	C(2)-Co(1)-C(4)	68.3(2)	C(2)- $Co(1)$ - $C(5)$	68.8(2)
N(1)-C(24)	1.412(6)	C(1)-C(2)	1.434(7)	C(2)- $Co(1)$ - $C(6)$	116.9(2)	C(2)- $Co(1)$ - $C(7)$	110.6(2)
C(1)-C(5)	1.436(7)	C(1)-C(11)	1.487(7)	C(2)- $Co(1)$ - $C(8)$	133.0(3)	C(2)- $Co(1)$ - $C(9)$	171.3(3)
C(2)-C(3)	1.410(6)	C(2)-C(12)	1.513(7)	C(2)- $Co(1)$ - $C(10)$	148.1(3)	C(3)- $Co(1)$ - $C(4)$	40.6(2)
C(3)-C(4)	1.423(7)	C(4)-C(5)	1.420(7)	C(3)-Co(1)-C(5)	68.3(2)	C(3)– $Co(1)$ – $C(6)$	147.9(3)
C(4)-C(13)	1.483(7)	C(6)-C(7)	1.412(8)	C(3)– $Co(1)$ – $C(7)$	116.3(2)	C(3)-Co(1)-C(8)	109.2(2)
C(6)-C(10)	1.421(8)	C(6)-C(19)	1.510(9)	C(3)-Co(1)-C(9)	131.9(3)	C(3)- $Co(1)$ - $C(10)$	170.5(3)
C(7)-C(8)	1.423(8)	C(7)-C(20)	1.518(8)	C(4)-Co(1)-C(5)	40.7(2)	C(4)- $Co(1)$ - $C(6)$	171.2(2)
C(8)–C(9)	1.421(9)	C(8)-C(21)	1.460(9)	C(4)-Co(1)-C(7)	146.5(2)	C(4)-Co(1)-C(8)	113.9(2)
C(9)–C(10)	1.402(9)	C(9)-C(22)	1.525(9)	C(4)-Co(1)-C(9)	107.4(2)	C(4)-Co(1)-C(10)	131.2(2)
C(10)-C(23)	1.484(8)	C(13)-C(14)	1.416(7)	C(5)- $Co(1)$ - $C(6)$	132.7(2)	C(5)-Co(1)-C(7)	172.6(3)
C(13)-C(18)	1.368(7)	C(14)-C(15)	1.395(8)	C(5)-Co(1)-C(8)	144.9(2)	C(5)-Co(1)-C(9)	113.3(3)
C(15)-C(16)	1.370(9)	C(16)-C(17)	1.368(9)	C(5)-Co(1)-C(10)	108.1(2)	C(6)-Co(1)-C(7)	40.5(2)
C(17)-C(18)	1.389(7)	C(24)-C(25)	1.384(7)	C(6)-Co(1)-C(8)	68.5(3)	C(6)-Co(1)-C(9)	68.4(2)
C(24)-C(29)	1.385(7)	C(25)-C(26)	1.379(7)	C(6)-Co(1)-C(10)	40.8(2)	C(7)-Co(1)-C(8)	40.8(2)
C(26)-C(27)	1.379(8)	C(27)-C(28)	1.359(8)	C(7)-Co(1)-C(9)	68.5(3)	C(7)-Co(1)-C(10)	68.2(2)
C(28)-C(29)	1.380(8)	` , ` ,	` ′	C(8)-Co(1)-C(9)	40.8(3)	C(8)-Co(1)-C(10)	68.3(3)
` , ` ,	. ,			C(9)-Co(1)-C(10)	40.3(3)	F(1)-P(1)-F(2)	89.6(2)
b) Bond angles				F(1)-P(1)-F(3)	89.6(3)	F(1)-P(1)-F(4)	93.0(3)
F(1)-P(1)-F(5)	92.6(3)	F(1)-P(1)-F(6)	176.8(3)	C(8)-C(7)-C(20)	125.1(7)	Co(1)-C(8)-C(7)	69.7(4)
F(2)-P(1)-F(3)	88.2(2)	F(2)-P(1)-F(4)	177.2(3)	Co(1)-C(8)-C(9)	69.3(4)	Co(1)-C(8)-C(21)	129.2(4)
F(2)-P(1)-F(5)	89.2(2)	F(2)-P(1)-F(6)	87.6(2)	C(7)-C(8)-C(9)	107.5(6)	C(7)-C(8)-C(21)	128.3(7)
F(3)-P(1)-F(4)	90.6(3)	F(3)-P(1)-F(5)	176.6(3)	C(9)-C(8)-C(21)	124.1(7)	Co(1)-C(9)-C(8)	69.9(3)
F(3)-P(1)-F(6)	88.8(3)	F(4)-P(1)-F(5)	91.8(3)	Co(1)-C(9)-C(10)	70.0(4)	Co(1)-C(9)-C(22)	126.1(5)
F(4)-P(1)-F(6)	89.8(3)	F(5)-P(1)-F(6)	88.8(3)	C(8)-C(9)-C(10)	108.2(6)	C(8)-C(9)-C(22)	128.3(8)
C(11)-N(1)-C(24)	127.5(5)	Co(1)-C(1)-C(2)	70.7(3)	C(10)-C(9)-C(22)	123.5(8)	Co(1)-C(10)-C(6)	69.7(3)
Co(1)-C(1)-C(5)	68.5(3)	Co(1)-C(1)-C(11)	125.6(4)	Co(1)-C(10)-C(9)	69.7(3)	Co(1)-C(10)-C(23)	127.9(5)
C(2)-C(1)-C(5)	107.3(5)	C(2)-C(1)-C(11)	124.7(5)	C(6)-C(10)-C(9)	108.4(6)	C(6)-C(10)-C(23)	124.6(8)
C(5)-C(1)-C(11)	128.0(6)	Co(1)-C(2)-C(1)	68.5(3)	C(9)-C(10)-C(23)	126.9(7)	O(1)-C(11)-N(1)	123.6(5)
Co(1)-C(2)-C(3)	69.1(3)	Co(1)-C(2)-C(12)	128.6(4)	O(1)-C(11)-C(1)	120.1(5)	N(1)-C(11)-C(1)	116.3(5)
C(1)-C(2)-C(3)	107.4(5)	C(1)-C(2)-C(12)	128.5(5)	C(4)-C(13)-C(14)	119.4(5)	C(4)-C(13)-C(18)	121.6(5)
C(3)-C(2)-C(12)	124.1(5)	Co(1)-C(3)-C(2)	70.9(3)	C(14)-C(13)-C(18)	119.0(5)	C(13)-C(14)-C(15)	119.5(6)
Co(1)-C(3)-C(4)	70.2(3)	C(2)-C(3)-C(4)	109.7(4)	C(14)-C(15)-C(16)		C(15)-C(16)-C(17)	120.4(6)
Co(1)-C(4)-C(3)	69.3(3)	Co(1)-C(4)-C(5)	68.1(3)	C(16)-C(17)-C(18)	120.5(6)	C(13)-C(18)-C(17)	120.5(5)
Co(1)-C(4)-C(13)	128.8(4)	C(3)-C(4)-C(5)	106.8(4)	N(1)-C(24)-C(25)	117.3(5)	N(1)-C(24)-C(29)	124.2(5)
C(3)-C(4)-C(13)	127.3(5)	C(5)-C(4)-C(13)	125.9(5)	C(25)-C(24)-C(29)	118.5(5)	C(24)-C(25)-C(26)	121.4(5)
Co(1)-C(5)-C(1)	70.1(3)	Co(1)– $C(5)$ – $C(4)$	71.1(3)	C(25)-C(26)-C(27)	119.4(6)	C(26)-C(27)-C(28)	119.5(6)
C(1)-C(5)-C(4)	108.7(5)	Co(1)–C(6)–C(7)	69.9(3)	C(27)-C(28)-C(29)	121.8(6)	C(24)-C(29)-C(28)	119.5(6)
Co(1)-C(6)-C(10)	69.5(3)	Co(1)-C(6)-C(19)	128.0(5)				

equimolar amount of a 40% water solution of tetrabutylammonium hydroxide (5.00 g). The resulting mixture was stirred for 30 min at room temperature. The solvent was evaporated in vacuo, and the resulting solid was recrystallized from hexane-ethyl acetate. Colorless needles (3.03 g) were obtained in 83% yield.

Mp 133.0—134.0 °C; IR (Nujol) 1740 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ =3.35—3.27 (m, 8H), 2.91—0.91 (m, 9H), 1.72—1.61 (m, 8H), 1.45 (q, 8H, J=7.3 Hz), 1.14 (s, 3H), 1.01 (t, 12H, J=7.3 Hz), 0.83 (s, 3H); $[\alpha]_D^{15}$ =+25.1° (CHCl₃, c 1.19). Found: C, 65.68; H, 10.73; N, 2.76; S, 6.65%. Calcd for C₂₆H₅₁NO₄S: C, 65.92; H, 10.85 N, 2.96; S, 6.77%.

Similarly tetrabutylammonium (-)-(R)-camphor-10-sulfonate was prepared in 86% yield.

Mp 134.5—135.0 °C; $[\alpha]_D^{25} = -25.1$ ° (CHCl₃, c 1.08). Found: C, 66.10; H, 10.81; N, 2.76; S, 6.58%. Calcd for $C_{26}H_{51}NO_4S$: C, 65.92; H, 10.85 N, 2.96; S, 6.77%.

X-Ray Crystallographic Analysis for (+)-3a. Enantiomeric complex (+)-3a was prepared following the method for 3c from starting with (+)-2a. (+)-3a gave the same IR, ¹H NMR, and mass spectral data as the racemic isomers. ^{8b)}

(+)-3a: Orange needles (Yield 86%), mp 280.5—282.0 °C (decomp); $[\alpha]_D^{25}$ =+47.5° (MeOH, c 0.390). Found: C, 56.73; H, 5.30; N, 2.18; P, 5.00; F, 18.52%. Calcd for C₂₉H₃₁ONCoPF₆: C, 56.78; H, 5.09; N, 2.28; P, 5.05; F, 18.58%.

The experimental conditions and the summary of structure analysis are shown in Table 3. For (+)-3a, X-ray diffraction date was collected on a Rigaku AFC5R four-circle diffractometer. The structure was solved by heavy-atom Patterson methods (SAPI 91). Hydrogen atoms were placed in appropriate trigonal or tetrahedral positions. All calculations were done using the teXsan crystallographic software package of Molecular Structure Corporation. Fractional coordinates are listed in Table 4 and bond distances and angles in Table 5. ¹⁵⁾ Additional data, including anisotropic temperature factors are available from the authors.

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