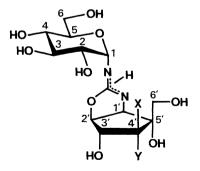
Total Synthesis and Trehalase-Inhibitory Activity of Trehalostatin and Its Diastereoisomer

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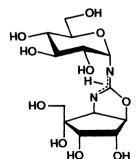
Complete synthesis of both diastereoisomeric structures, initially proposed for the trehalase inhibitor trehalostatin, and their biological assay have been carried out. The synthetic compounds were found to lack an inhibitory activity against trehalase, suggesting that the inhibitor would possess convincingly its 4'-epimeric structure later assigned for trehazolin.

In 1990, trehalostatin **1**, a potent and specific inhibitor against blowfly trehalase, was isolated by Murao *et al.*¹⁾ from the culture broth of *Amycolatopsis trehalostatica*, and the structure²⁾ was proposed as depicted mainly on the basis of the ¹H-NMR spectroscopic data. Later, Ando *et al.*³⁾ reported the isolation of trehalase inhibitor trehazolin **2** from the culture broth of *Micromonospora* strain SANK 62390, and showed it to be rather identical to **1** by comparison of physical and spectroscopic data. They, however, assigned a different structure to it. Very recently, conclusive synthesis⁴⁾ of the aminocyclitol moiety of **2**, followed by a total synthesis^{5,6)} of the whole molecule of the inhibitor, established the structure proposed for **2** and its absolute configuration as depicted. Therefore, the question still remains unanswered whether compounds **1** and **2** are identical or the former is in fact the 4'-epimer of **2**.



Trehalostatin 1: X = H, Y = OH

Trehazolin 2: X = OH, Y = H



Trehalostatin diastereoisomer 3

11a

This communication describes a complete synthesis of **1** and its diastereoisomer 3 *via* an unambiguous route similar to that⁵⁾ for **2** and evaluation of their inhibitory activity against silkworm trehalase.

The aminocyclitol moiety of **1** was first prepared from *O*-cyclohexylidene-*N*,*O*-isopropylidene derivative of (1,4,5/2,3)-5-acetamido-1,2,3,4-cyclopentanetetrol.⁴⁾ Thus, optical resolution⁵⁾ of the racemic compound was carried out by chromatographic separation of their (*S*)-acetylmandelates, giving the 1*R*-isomer **4a**, $[\alpha]_D^{29}$ +49° (*c* 1.2, CHCl₃) and 1*S*-isomer **4b**, $[\alpha]_D^{28}$ -45° (*c* 1.1, CHCl₃), in 41 and 45% yields, respectively. Compound **4a** was converted into (1R)-(1,2,3/4,5)- (**5a**) and (1*S*)-(1,4,5/2,3)-5-amino-1-*C*-hydroxymethyl-1,2,3,4-cyclopentanetetrol penta-*N*,*O*-acetates (**6a**) in 21 and 19% over-all yields, following essentially the five-step sequence⁴⁾ employed for the racemic compounds. Similarly, the antipodes **5b** and **6b** were obtained from **4b**. Hydrolysis of **5a** and **5b** with 2 mol dm-³ hydrochloric acid at 80 °C, followed by purification by a column of Dowex 50W-X2 (H+) resin with aq. 5% ammonia afforded the respective free amines **7a**, $[\alpha]_D^{23}$ -10° (*c* 0.8, H₂O), and **7b**, $[\alpha]_D^{22}$ +9° (*c* 1.0, H₂O), quantitatively.

Coupling of **7a** and 1.2 molar eq. of 2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosylisothiocyanate⁷⁾ (**8a**) in aq. 75% *N*,*N*-dimethylformamide for 4 h at room temperature afforded a thiourea⁸⁾ **9a**, $[\alpha]_D^{20}$ +138° (*c* 1.0, CHCl₃), IR (neat) 1540 cm⁻¹ (NH), in 91% yield. Treatment of **9a** with excess of mercury(II) oxide in acetone-ether (1:6) for 23 h resulted in a ready formation of oxazoline ring to give an isourea **10a**, $[\alpha]_D^{24}$ +39° (*c* 0.9, CHCl₃), IR (neat) 1665 cm⁻¹ (C=N), quantitatively. Deblocking of the benzyl ether groups of **10a** was effected by treatment with sodium in liquid ammonia to give the free base **1**, which, after conventional acetylation, was purely isolated as a sole octa-*N*,*O*-acetyl derivative **11a** (69% over-all yield). De-*N*,*O*-acetylation of **11a** with methanolic sodium methoxide afforded **1**, $[\alpha]_D^{22}$ +92° (*c* 0.6, H₂O), as a syrup in a quantitative yield. Interestingly, the ¹H NMR spectrum (270 MHz, D₂O) of **1** was rather similar to that⁵) of **2**. Therefore, it might have been very difficult to differentiate between two structures **1** and **2** only on the basis of ¹H NMR spectroscopic data.

Likewise, the diastereoisomer 3, $[\alpha]_D^{22}$ +117° (c 0.8, H_2O), was prepared in three steps ($9b \rightarrow 10b \rightarrow 11b \rightarrow 3$) in 69% over-all yield, starting from the thiourea 9b obtained, in 90% yield, by coupling of 7b and 8.

Biological assay⁹⁾ of the synthetic **1** and **3** showed no inhibitory activity against silkworm trehalase at the final concentration of 100 μ g/ml (*cf.* synthetic **2**⁵⁾: IC_{50} 0.016 μ g/ml). Consequently, the present results clearly suggest that the structure **1** or **3** initially proposed for trehalostatin is incorrect and should probably be revised to the structure **2**, being identical to that of trehazolin.¹⁰⁾

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- 8) All new compounds were characterized by 270 MHz ¹H NMR and IR spectrometric, and elemental analyses. Selected ¹H NMR data for 10a (in CDCl₃) δ 5.33 (1 H, br s, 1 H), 4.89 and 4.77 (each 1 H, ABq, J_{gem} 11 Hz, PhC \underline{H}_2), 4.83 (1 H, d, $J_{1',2'}$ 8.1 Hz, 2'-H), 4.76 and 4.54 (each 1 H, ABq, J_{gem} 11.3 Hz, PhC \underline{H}_2), 4.62 and 4.58 (each 1 H, ABq, J_{gem} 11 Hz, PhC \underline{H}_2), 4.44 and 4.40 (each 1 H, ABq, J 11.7 Hz, PhCH₂), 4.41 (1 H, d, 1'-H), 4.01 (1 H, d, J_{3',4'} 3.6 Hz, 3'-H). For 11a (in CDCl₃) δ 5.58 (1 H, d, J_{1.2} 4 Hz, 1-H), 5.51—5.46 (2 H, m, 3'-H, 4'-H), 5.40 (1 H, dd, J_{2.3} 10.3, $J_{3.4}$ 9.7 Hz, 3-H), 5.07 (1 H, dd, $J_{4,5}$ 9.9 Hz, 4-H), 5.06 (1 H, dd, $J_{1,2}$ 4 Hz, 2-H), 4.99—4.90 (2 H, m, 1'-H, 2'-H), 4.30 (1 H, ddd, J_{5,6a} 1.8, J_{5,6b} 4.8 Hz, 5-H), 4.22 (1 H, dd, J_{gem} 12.5 Hz, 6b-H), 4.09 (1 H, dd, 6a-H), 4.10 and 3.95 (each 1 H, ABq, J_{gem} 11.7 Hz, 6',6'-H), 3.58 (1 H, br s, OH), 2.66 (3 H, s, NAc), 2.14, 2.11, 2.10, 2.06, 2.03, 2.004, and 2.002 (each 3 H, 7 s, 7 Ac). For 1 (in D_2O) δ 5.17 (1 H, d, $J_{1,2}$ 5.1 Hz, 1-H), 4.85 (1 H, dd, $J_{1',2'}$ 8.4, $J_{2',3'}$ 1.2 Hz, 2'-H), 4.25 (1 H, d, 1'-H), 4.06 (1 H, dd, *J*_{3',4'} 5.1 Hz, 3'-H), 3.80 (1 H, d, 4'-H), 3.67 (1 H, dd, *J*_{5,6a} 2.7, *J*_{gem} 13 Hz, 6a-H), 3.58 (1 H, dd, *J*_{2,3} 8.8 Hz, 2-H), 3.66 and 3.50 (each 1 H, ABq, J_{gem} 12.1 Hz, 6',6'-H), 3.58 (1 H, dd, J_{5,6b} 5.9 Hz, 6b-H), 3.50 (1 H, dd, $J_{3,4}$ 9.9 Hz, 3-H), 3.42 (1 H, ddd, 5-H), 3.25 (1 H, dd, 4-H). For 3 (in D₂O) $\delta\,5.13\,(1\,\,\mathrm{H},\,\mathrm{d},\,J_{1,2}\,5.1\,\,\mathrm{Hz},\,1-\mathrm{H}),\,4.78\,(1\,\,\mathrm{H},\,\mathrm{dd},\,J_{1',2'}\,8.2,\,\,J_{2',3'}\,1.1\,\,\mathrm{Hz},\,2'-\mathrm{H}),\,4.19\,(1\,\,\mathrm{H},\,\mathrm{d},\,1'-\mathrm{H}),\,4.00$ (1 H, dd, $J_{3',4'}$ 5.1 Hz, 3'-H), 3.76 (1 H, d, 4'-H), 3.56 (1 H, dd, $J_{2,3}$ 9.4 Hz, 2-H), 3.58—3.44 (5 H, m, 3-H, 6,6-H, 6',6'-H), 3.39 (1 H, ddd, J_{4.5} 10.1, J_{5.6} 2.8 Hz, 5-H), 3.24 (1 H, dd, J_{3.4} 8.6 Hz, 4-H).
- 9) Dr. Shuji Takahashi, personal communication.
- 10) It is now very difficult to get each authentic sample for direct comparison, because of a minute quantity of natural trehalostatin.

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